

[Chem. Pharm. Bull.]
35(3) 970-979 (1987)

Syntheses of 24,25-Dihydroxyvitamin D₂, 24,25-Dihydroxy-22-dehydrovitamin D₃, 25-Hydroxy-24-oxo-22-dehydrovitamin D₃ and 22,24,25-Trihydroxyvitamin D₃¹⁾

KOTOMI KATSUMI,^a TOSHIO OKANO,^a YURIE ONO,^a EMIKO MAEGAKI,^a
KUMIKO NISHIMURA,^a MIZUE BABA,^a TADASHI KOBAYASHI,*-^a
OKIKO MIYATA,^b TAKEAKI NAITO,^b
and ICHIYA NINOMIYA^b

*Departments of Hygienic Sciences^a and Medicinal Chemistry,^b
Kobe Women's College of Pharmacy, Motoyamakita-machi,
Higashinada-ku, Kobe 658, Japan*

(Received July 28, 1986)

Four vitamin D₂ and D₃ metabolites, 24,25-dihydroxyvitamin D₂ (**8**), 24,25-dihydroxy-22-dehydrovitamin D₃ (**10**), 25-hydroxy-24-oxo-22-dehydrovitamin D₃ (**12**) and 22,24,25-trihydroxyvitamin D₃ (**14**), were synthesized from ergosterol (**1**) via the hydroxyketone (**4**) as a common key intermediate.

Keywords—24,25-dihydroxyvitamin D₂; 24,25-dihydroxy-22-dehydrovitamin D₃; 25-hydroxy-24-oxo-22-dehydrovitamin D₃; 22,24,25-trihydroxyvitamin D₃; vitamin D; ergosterol; high-performance liquid chromatography

There are two of vitamin D,²⁾ namely D₂ and D₃, which differ in the side chain structure but have practically the same biological activity in mammals, including human beings. These two vitamins are known to be similarly metabolized to 25-hydroxyvitamin D (25-OH-D) in the liver and subsequently to 1 α ,25-dihydroxyvitamin D [1 α ,25-(OH)₂-D] or 24R,25-dihydroxyvitamin D [24R,25-(OH)₂-D] in the kidney when plasma calcium concentrations are lower or higher than normal.³⁾ Many studies have been done on the further metabolism of 25-OH-D₃ and 1 α ,25-(OH)₂-D₃. The 26,23-lactone,⁴⁾ 23-hydroxy,⁵⁾ 23,24-dihydroxy,⁶⁾ 24,26-dihydroxy,⁶⁾ 23-oxo,⁶⁾ 24-oxo^{6,7)} and 23-hydroxy-24-oxo⁸⁻¹⁰⁾ derivatives of 25-OH-D₃ and 26,23-lactone,¹¹⁾ 23-hydroxy¹²⁾ and 24-oxo^{13,14)} derivatives of 1 α ,25-(OH)₂-D have been isolated and identified as *in vivo* or *in vitro* metabolites. The mechanisms of metabolism have been well discussed^{15,16)} and some of the isolated metabolites were chemically synthesized.¹⁷⁻²¹⁾ However, few reports have appeared on the metabolism of vitamin D₂, and the metabolic fate of the double bond at the 22-position and the methyl group at the 24S-position in the side chain of vitamin D₂ remains to be clarified. We now report the syntheses of four potential metabolites, 24,25-(OH)₂-D₂ (**8**), 24,25-dihydroxy-22-dehydrovitamin D₃ [4²²-24,25-(OH)₂-D₃] (**10**), 25-hydroxy-24-oxo-22-dehydrovitamin D₃ [4²²-24-oxo-25-OH-D₃] (**12**) and 22,24,25-trihydroxyvitamin D₃ [22,24,25-(OH)₃-D₃] (**14**).

First, we investigated the synthesis of 24,25-(OH)₂-D₂ (**8**). Though the synthesis of **8** from stigmasterol was reported by Jones *et al.*,^{22,23)} their synthetic route is rather complicated. Therefore, we have developed an improved synthesis of **8** by modification of their procedure. As shown in Chart 1, ergosterol (**1**) was converted into the known 20-aldehyde (**3**) via a route involving protection of the 5,7-diene group and ozonolysis according to Barton *et al.*²⁴⁾ Aldol condensation of **3** with 3-methyl-3-(tetrahydropyran-2-yl)butan-2-one proceeded very smoothly to afford the hydroxyketone (**4a** and **4b**) as a diastereomeric mixture; **4a**

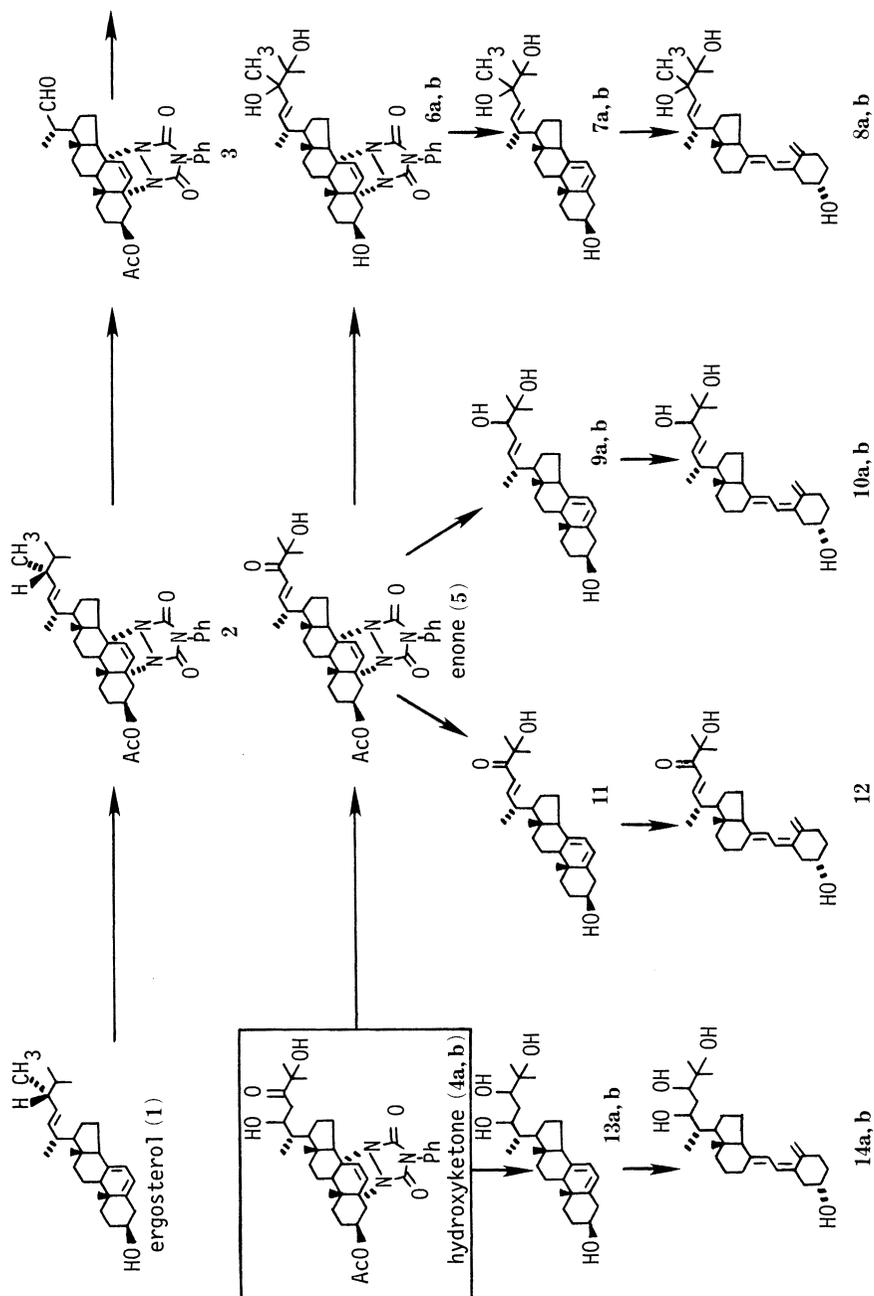


Chart 1. Synthetic Scheme

and **4b** were separated, both as colorless crystals, by preparative thin layer chromatography (TLC) using silica gel. The yields of the less polar (**4a**: $R_f=0.33$) and more polar (**4b**: $R_f=0.19$) 22-isomers were 18 and 21%, respectively, though their stereochemistry at the 22-position has not been clarified. Dehydration of a mixture of **4a** and **4b** with *p*-toluenesulfonic acid afforded the known enone (**5**) in 90% yield, which exhibited the proton nuclear magnetic resonance ($^1\text{H-NMR}$) signals of two newly formed olefinic protons at 6.36 (d, $J=15$ Hz, 23-H) and 7.03 (dd, $J=15, 8$ Hz, 22-H). The conversion of **3** to **5** was carried out according to Eyley and Williams,²⁵ who obtained **5** directly without isolation of **4**. Methylation of **5** with methyllithium afforded the methylated 24,25-glycols (**6**) in 60% yield, and these were refluxed with lithium aluminum hydride (LiAlH_4) in tetrahydrofuran (THF) to afford the desired 24,25-dihydroxyprovitamin D_2 [24,25-(OH) $_2$ -pro- D_2 , **7a** and **7b**] as a mixture of diastereomers in 70% yield.

The two diastereomers were completely separated by high-performance liquid chromatography (HPLC) on a Zorbax SIL column with 2.5% isopropanol in *n*-hexane as the mobile phase, as shown in Fig. 1, to afford the respective diastereomers in almost equal amounts; the less polar (peak 1) was confirmed to be (24*S*)-24,25-(OH) $_2$ -pro- D_2 (**7a**) and the more polar compound (peak 2) to be the (24*R*)-isomer (**7b**) by converting them into the corresponding 24,25-(OH) $_2$ - D_2 (**8a** and **8b**) by ultraviolet (UV) irradiation followed by thermal isomerization. Figure 2 shows the HPLC profiles of **7a** and **7b** before and after photochemical and thermal isomerization as representatives of the provitamin forms (**7**, **9**, **11** and **13**).

When **7a** and **7b** were irradiated by a monochromatic UV ray at 295 nm obtained from a spectroirradiator, the peaks decreased while those of the respective previtamin forms (**15a** and **15b**) with retention times of 46.3 and 49.4 min increased. Upon thermal isomerization by refluxing the irradiated ethanolic solutions for 2 h, the previtamin peaks were converted to those of the vitamin D forms (**8a** and **8b**) with retention times of 32.0 and 34.0 min, respectively. These products were purified by HPLC. The eluates corresponding to the respective peaks were carefully collected and the solvent was evaporated off under reduced pressure to give the respective vitamins (**8a** and **8b**) in pure form.

On co-chromatography (HPLC) with authentic **8a** and **8b**, kindly donated by Dr. Y. Mazur and Dr. G. Jones, the purified vitamins showed retention times of 32.0 and 34.0 min, and were confirmed to be (24*S*)-24,25-(OH) $_2$ - D_2 (**8a**) and (24*R*)-24,25-(OH) $_2$ - D_2 (**8b**), respectively.

Secondly, we synthesized Δ^{22} -24,25-(OH) $_2$ - D_3 (**10a** and **10b**). Δ^{22} -24,25-(OH) $_2$ -pro- D_3 (**9a** and **9b**) was produced from the enone (**5**) by a double reduction procedure. Reduction of **5**

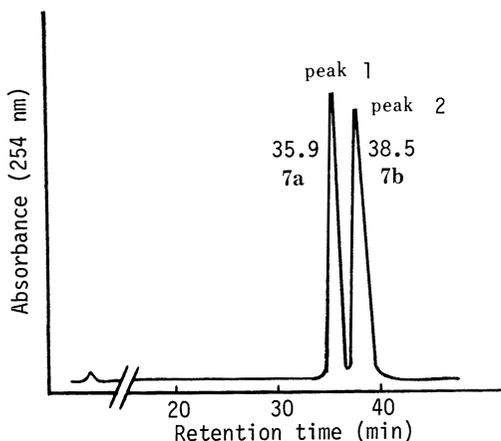


Fig. 1. HPLC Separation of the Diastereomeric Mixture of 24,25-(OH) $_2$ - D_2 (**7a** and **7b**)

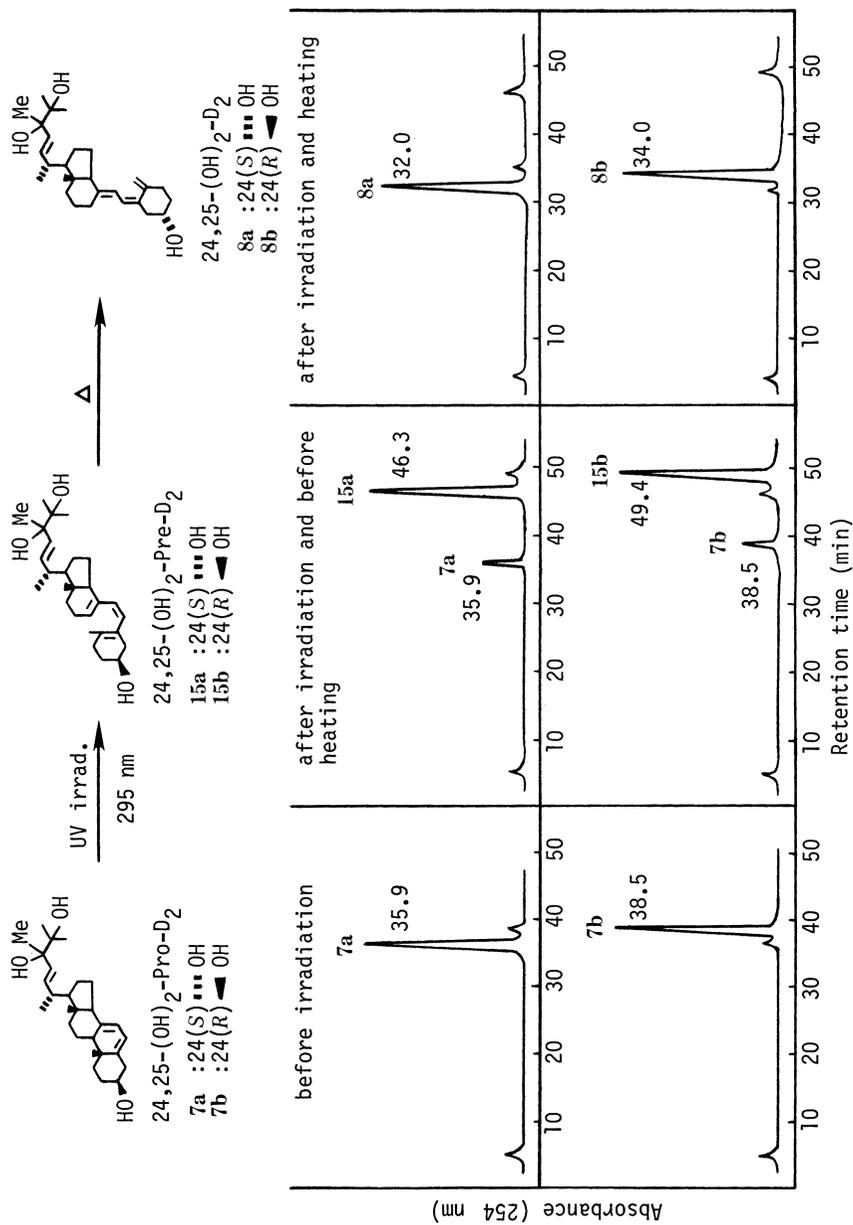


Fig. 2. HPLC Profiles of 7a and 7b before and after Photochemical and Thermal Isomerization

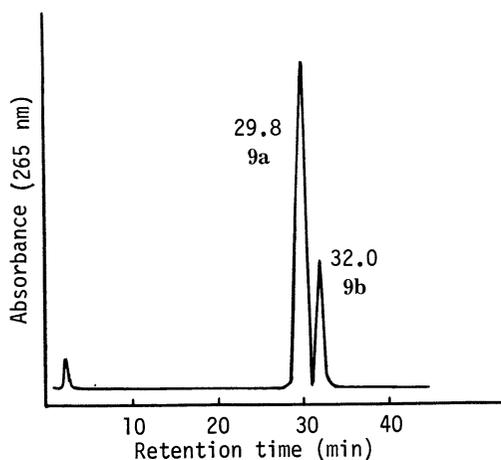


Fig. 3. HPLC Separation of the Diastereomeric Mixture of the 24-ol Provitamins (**9a** and **9b**)

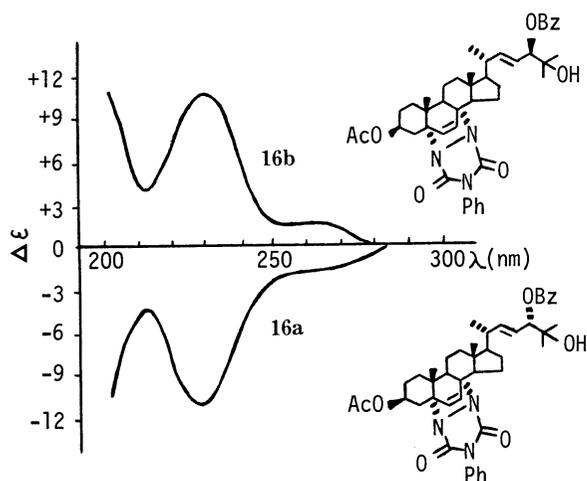


Fig. 4. CD Spectra of the 24-Benzoates (**16a** and **16b**)

with LiAlH_4 in THF afforded the two provitamins (**9a** and **9b**) in the ratio of 3 : 1. On the other hand, chemoselective reduction of the carbonyl group at the 24-position in **5** with lithium tri-*sec*-butylborohydride (L-selectride) in THF at -78°C followed by deprotection of the triazolone ring with LiAlH_4 gave **9a** and **9b** in the ratio of 6 : 1; these products were separated by HPLC on a Zorbax SIL column with 5.5% isopropanol in *n*-hexane as a mobile phase, as shown in Fig. 3. The absolute configurations at the 24-position of **9a** and **9b** were assigned as 24*S* and 24*R*, respectively, on the basis of the following considerations.

Since the 24*S*-isomer of 24,25-(OH)₂-pro-D₂ (**7a**) was eluted faster than the 24*R*-isomer (**7b**) on HPLC, as shown in Fig. 1, we considered that the faster and later peaks observed in Fig. 3 might also be due to the 24*S*- and 24*R*-isomers of the provitamin forms (**9a** and **9b**), respectively. The faster elution of the 24*S*-isomers (**7a** and **9a**) than the respective 24*R*-isomers (**7b** and **9b**) is in good agreement with the results on the analogous epimers of 24-hydroxyderivatives of the vitamin D₃ series reported by Sai *et al.*²⁶⁾ The provitamin D forms (**9a** and **9b**) were converted into the respective benzoates (**16a** and **16b**), and their circular dichroism (CD) spectra were measured. As shown in Fig. 4, a strongly negative Cotton curve in **16a** and a positive one in **16b** were observed. Therefore, the prediction of the "exciton chirality method" developed by Gonnella *et al.*²⁷⁾ is that **9a** and **9b** have 24*S* and 24*R*

configurations, respectively.

As mentioned above, the reduction of **5** with the two hydride reagents resulted in the preferential formation of the 24*S*-ol (**9a**) rather than the 24*R*-ol (**9b**). Sai *et al.*²⁶⁾ have also reported that the reduction of the Δ^{22} -24-one system in the vitamin D₃ series with sodium borohydride gave the (24*S*)-24-alcohol as the major product. The preferential formation of **9a** over **9b** can be explained as follows. The most stable conformation for the enone (**5**) may be that shown in Chart 2, in which the 20-hydrogen is *syn* to the *s-cis* enone system involving hydrogen bonding with the 25-hydroxyl group. Hydride ion would attack the carbonyl group preferentially from the *re*-face due to the presence of the more bulky steroidal skeleton than the methyl group in **5**. When we used L-selectride as a bulky hydride agent, stereoselectivity in the reduction of the enone system increased, as described above. Corey *et al.*²⁸⁾ reported the same effect on the chirality at the α , β -unsaturated enone system.

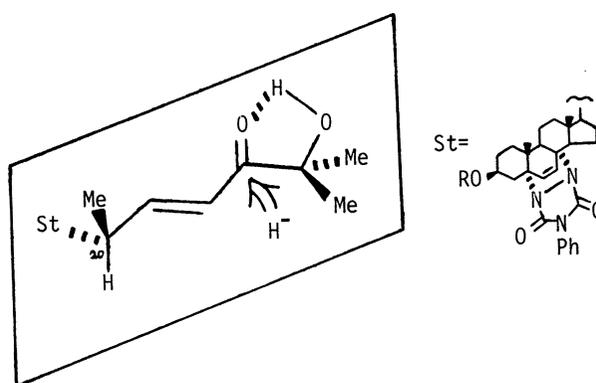


Chart 2. Presumed Mechanism of the Reduction of **5**

As in the syntheses of **8a** and **8b**, the provitamins (**9a** and **9b**) were converted into the respective vitamin forms (**10a** and **10b**) upon UV irradiation followed by thermal isomerization.

Finally, we also synthesized Δ^{22} -25-OH-24-oxo-D₃ (**12**) and 22,24,25-(OH)₃-D₃ (**14a** and **14b**). The enone (**5**) was heated at 120 °C in ethylene glycol in the presence of anhydrous K₂CO₃ to afford Δ^{22} -25-OH-24-oxo-pro-D₃ (**11**) as a result of retro-1,4-cycloaddition. On the other hand, reduction of each of the two separated hydroxyketones (**4a** and **4b**) with LiAlH₄ in THF afforded the respective 22,24,25-(OH)₃-pro-D₃ (**13a** and **13b**). The compounds were characterized spectrally except for their absolute configurations at the 22- and 24-positions. The provitamin D forms (**11**, **13a** and **13b**) thus obtained were similarly converted into the respective vitamin D forms (**12**, **14a** and **14b**) upon UV irradiation followed by thermal isomerization.

Experimental

All melting points were measured on a micro hot-stage apparatus (Yanagimoto) and are uncorrected. ¹H-NMR spectra were obtained on a Varian XL-200 spectrometer in CDCl₃ with tetramethylsilane as an internal standard. Mass spectra (MS) were recorded on a JEOL LMS-01SG or a Hitachi M-80 spectrometer. UV spectra were obtained on a Hitachi 323 spectrophotometer in ethanol and infrared (IR) spectra were recorded on a Hitachi IR-215 spectrometer in CHCl₃. CD spectra were obtained on a JASCO J500C spectropolarimeter in ethanol. Preparative TLC was carried out on precoated plates of silica gel (Kieselgel 60F₂₅₄, 2 or 0.5 mm thickness, Merck). HPLC was performed on a Shimadzu LC-3A or LC-4A high-performance liquid chromatograph equipped with a Shimadzu SPD 2AS detector (set at 265 nm, 0.005 absorbance unit full scale) or a Shimadzu UVD-2 (set at 254 nm, 0.001 absorbance

unit full scale) and a Zorbax SIL column (4.6 i.d. \times 250 mm or 6.2 i.d. \times 250 mm, DuPont) with 2.5 or 5.5% isopropanol in *n*-hexane as a mobile phase. The following abbreviations are used: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, br = broad, THF = tetrahydrofuran, DHP = dihydropyran, LiAlH₄ = lithium aluminum hydride.

3 β -Acetoxy-22,25-dihydroxy-5 α ,8 α -(3,5-dioxo-4-phenyl-1,2,4-triazolidino)cholesta-6-en-24-one (4a and 4b)—A solution of *n*-butyllithium (0.3 ml of 15% solution in *n*-hexane) was added to a cooled (-78°C) solution of diisopropylamine (75 mg, 0.7 mmol) in dry THF (3 ml) with stirring. A solution of 3-methyl-3-(tetrahydropyran-2-yloxy)butan-2-one²⁵ (68 mg, 0.67 mmol) in dry THF (2 ml) was then added dropwise at -78°C over 15 min. The mixture was stirred at the same temperature for 1 h and then a solution of the 20-aldehyde (3)²⁴ (200 mg, 0.36 mmol) in dry THF (4 ml) was added. After being stirred at -78°C for 2 h, the reaction mixture was brought back to room temperature. Diethyl ether (Et₂O) and water were added and the whole was vigorously shaken, then allowed to stand at room temperature. The separated organic layer was washed with brine, dried (Na₂SO₄) and evaporated. The resulting residue was dissolved in THF (10 ml) containing 1.5 N HCl (2 ml) and allowed to stand at room temperature for 3 h. Et₂O was added, and the solution was washed with 5% Na₂CO₃ solution and brine, dried (Na₂SO₄), and evaporated to give crude 4, which was subjected to preparative TLC (developing solvent: Et₂O-acetone, 97:3). The band of *R*_f 0.32 was scraped off and extracted with CHCl₃-MeOH. Removal of the solvent under reduced pressure gave the less polar hydroxyketone (4a) as colorless crystals (42 mg, 18%): mp 143–145.5 $^\circ\text{C}$ (Et₂O-*n*-hexane). High-resolution MS *m/z*: Calcd for C₂₇H₄₆O₃ (M⁺ - PhC₂N₃O₂ - CH₃COOH): 412.2974. Found: 412.2970. ¹H-NMR δ : 0.85 (3H, s, 13-Me), 1.00 (3H, s, 10-Me), 1.00 (3H, d, *J* = 6 Hz, 20-Me), 1.38 (6H, s, 25-Me₂), 2.03 (3H, s, 3-OAc), 4.22 (1H, m, 22-H), 5.48 (1H, m, 3 α -H), 6.30, 6.46 (each 1H, d, *J* = 8 Hz, 6- and 7-H), 7.44 (5H, m, Ar).

The band of *R*_f 0.19 on preparative TLC was scraped off and extracted with CHCl₃-MeOH to give the more polar hydroxyketone (4b) (49 mg, 21%): mp 153–154 $^\circ\text{C}$ (Et₂O-*n*-hexane) as colorless crystals. High-resolution MS *m/z*: Calcd for C₂₇H₄₆O₃ (M⁺ - PhC₂N₃O₂ - CH₃COOH): 412.2975. Found: 412.2974. ¹H-NMR δ : 0.82 (3H, s, 13-Me), 0.99 (3H, d, *J* = 6 Hz, 20-Me), 1.00 (3H, s, 10-Me), 1.38 (6H, s, 25-Me₂), 2.03 (3H, s, 3-OAc), 2.50 (1H, dd, *J* = 17, 2.5 Hz, 23-H), 2.92 (1H, dd, *J* = 17, 9 Hz, 23-H), 4.27 (1H, br d, *J* = 9.5 Hz, 22-H), 5.48 (1H, m, 3-H), 6.28, 6.46 (each 1H, d, *J* = 8 Hz, 6- and 7-H), 7.42 (5H, m, Ar).

(22E)-3 β -Acetoxy-25-hydroxy-5 α ,8 α -(3,5-dioxo-4-phenyl-1,2,4-triazolidino)cholesta-6,22-dien-24-one (5)—A mixture of the hydroxyketones (4a and 4b) (100 mg, 0.15 mmol) was dissolved in benzene and a small amount of *p*-toluenesulfonic acid was added. This mixture was stirred at room temperature for 2 h, and solid K₂CO₃ was added for neutralization. The mixture was extracted with benzene and the extract was dried (Na₂SO₄). Removal of the solvent gave the enone (5) as a colorless glass (90 mg, 90%). High-resolution MS *m/z*: Calcd for C₂₇H₃₈O₂ (M⁺ - PhC₂N₃O₂ - CH₃COOH): 394.2869. Found: 394.2860. IR ν_{max} cm⁻¹: 3400, 1750, 1730, 1700. ¹H-NMR δ : 0.83 (3H, s, 13-Me), 0.98 (3H, s, 10-Me), 1.12 (3H, d, *J* = 6 Hz, 20-Me), 1.38 (6H, s, 25-Me₂), 2.02 (3H, s, 3-OAc), 3.24 (1H, m, 9-H), 5.45 (1H, m, 3 α -H), 6.25, 6.42 (each 1H, d, *J* = 8 Hz, 6- and 7-H), 6.36 (1H, d, *J* = 15 Hz, 23-H), 7.03 (1H, dd, *J* = 15, 8 Hz, 22-H), 7.42 (5H, m, Ar).

(22E)-3 β ,24,25-Trihydroxy-5 α ,8 α -(3,5-dioxo-4-phenyl-1,2,4-triazolidino)ergosta-6,22-diene (6)—A solution of methylolithium (0.6 mmol/ml) in Et₂O (2 ml) was added with stirring to a solution of the enone (5) (100 mg, 0.16 mmol) in THF (5 ml) at 0 $^\circ\text{C}$ under argon. The solution was brought back to room temperature, stirred for 0.5 h and then treated with 5% HCl solution. The reaction mixture was extracted with ethyl acetate, and the extract was dried (Na₂SO₄) and evaporated. The crude product was purified by preparative TLC (developing solvent: CH₂Cl₂-MeOH = 9:1) to give the 24,25-glycol (6) (57 mg, 60%) as a colorless glass. High-resolution MS *m/z*: Calcd for C₂₈H₄₄O₃ (M⁺ - PhC₂N₃O₂): 428.3287. Found: 428.3229. IR ν_{max} cm⁻¹: 3500, 1730, 1700. ¹H-NMR δ : 0.84 (3H, s, 13-Me), 0.99 (3H, s, 10-Me), 1.07 (3H, d, *J* = 6 Hz, 20-Me), 1.20 (6H, s, 25-Me₂), 1.28 (3H, s, 24-Me), 4.48 (1H, m, 3 α -H), 5.60 (2H, m, 22- and 23-H), 6.26, 6.42 (each 1H, d, *J* = 8 Hz, 6- and 7-H), 7.46 (5H, m, Ar).

(22E)-3 β ,24,25-Trihydroxyergosta-5,7,22-triene (7a and 7b)—The 24,25-glycol (6) (20 mg, 0.03 mmol) in dry THF (5 ml) was treated with LiAlH₄ (21 mg, 0.56 mmol) under reflux for 3 h. After cooling, the excess reagent was destroyed by adding a few drops of water. The mixture was extracted with ethyl acetate, dried (Na₂SO₄) and evaporated. The crude product was purified by HPLC on a Zorbax SIL column (4.6 i.d. \times 250 mm) with 2.5% isopropanol in *n*-hexane as a mobile phase at a flow rate of 2.0 ml/min to give (24S)-24,25-(OH)₂-pro-D₂ (7a) (4.2 mg, 33%) as a colorless glass and (24R)-24,25-(OH)₂-pro-D₂ (7b) (4.3 mg, 34%) as a colorless glass.

7a: High-resolution MS *m/z*: Calcd for C₂₈H₄₄O₃: 428.3289. Found: 428.3271. UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm: 264, 272, 281, 292. IR ν_{max} cm⁻¹: 3500, 1600. ¹H-NMR δ : 0.64 (3H, s, 13-Me), 0.96 (3H, s, 10-Me), 1.07 (3H, d, *J* = 6 Hz, 20-Me), 3.66 (1H, m, 3 α -H), 5.42 (1H, m, 6- or 7-H), 5.60 (3H, m, 22,23-H and 6- or 7-H).

7b: High-resolution MS *m/z*: Calcd for C₂₈H₄₄O₃: 428.3289. Found: 428.3273. UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm: 264, 272, 281, 292. IR ν_{max} cm⁻¹: 3500, 1600. ¹H-NMR δ : 0.64 (3H, s, 13-Me), 0.96 (3H, s, 10-Me), 1.07 (3H, d, *J* = 6 Hz, 20-Me), 3.67 (1H, m, 3 α -H), 5.42 (1H, m, 6- or 7-H), 5.60 (3H, m, 22,23-H and 6- or 7-H).

(24S)-24,25-Dihydroxyvitamin D₂ (8a)—A solution (4 ml) of (24S)-24,25-(OH)₂-pro-D₂ (7a) in ethanol (0.5 mg/ml) was placed in a quartz cell (10 \times 10 \times 40 mm) in a spectroirradiator and irradiated with monochromatic light at 295 nm. The irradiated solution was refluxed for 2 h and the solvent was evaporated off. The crude product was purified by HPLC under the same conditions as above to give (24S)-24,25-(OH)₂-D₂ (8a) (0.2 mg, 10%) as a

colorless glass: High-resolution MS m/z : Calcd for $C_{28}H_{44}O_3$: 428.3288. Found: 428.3307. UV $\lambda_{\max}^{\text{EtOH}}$ nm: 265; $\lambda_{\min}^{\text{EtOH}}$ nm: 228. $^1\text{H-NMR}$ δ : 0.50 (3H, s, 13-Me), 0.98 (3H, d, $J = 6$ Hz, 20-Me), 1.14, 1.16, 1.20 (each 3H, s, 24-Me and 25-Me₂), 3.88 (1H, m, 3 α -H), 4.76, 4.99 (each 1H, br s, 19-H₂), 5.50 (2H, m, 22- and 23-H), 5.97, 6.20 (each 1H, d, $J = 11$ Hz, 6- and 7-H).

(24R)-24,25-Dihydroxyvitamin D₂ (8b)—As described for the conversion of **7a** to **8a**, (24R)-24,25-(OH)₂-pro-D₂ (**7b**) (2.0 mg, 0.005 mmol) was converted into (24R)-24,25-(OH)₂-D₂ (**8b**) (0.2 mg, 10%) as a colorless glass: High-resolution MS m/z : Calcd for $C_{28}H_{44}O_3$: 428.3287. Found: 428.3271. UV $\lambda_{\max}^{\text{EtOH}}$ nm: 265; $\lambda_{\min}^{\text{EtOH}}$ nm: 228. $^1\text{H-NMR}$ δ : 4.84, 5.08 (each 1H, br s, 19-H₂), 5.64 (2H, m, 22- and 23-H).

(22E)-3 β ,24,25-Trihydroxycholesta-5,7,22-trienes (9a and 9b) Reduction with LiAlH₄—The enone (**5**) (60 mg, 0.1 mmol) in dry THF was treated with LiAlH₄ (70 mg, 1.8 mmol) under reflux for 3 h. After cooling, the excess reagent was destroyed by adding a few drops of water. The mixture was extracted with ethyl acetate, and the extract was dried (Na₂SO₄) and evaporated. The crude product was purified by HPLC on a Zorbax SIL column (4.6 i.d. \times 250 mm) with 5.5% isopropanol in *n*-hexane as a mobile phase at a flow rate of 1.0 ml/min to give (24S)- Δ^{22} -24,25-(OH)₂-pro-D₃ (**9a**) (18.5 mg, 45%) and (24R)- Δ^{22} -24,25-(OH)₂-pro-D₃ (**9b**) (7 mg, 17%) as colorless crystals.

9a: mp 203–204 °C (MeOH-*n*-hexane). High-resolution MS m/z : Calcd for $C_{27}H_{42}O_3$: 414.3131. Found: 414.3112. UV $\lambda_{\max}^{\text{EtOH}}$ nm: 292, 280, 272, 262. $^1\text{H-NMR}$ δ : 0.64 (3H, s, 13-Me), 0.94 (3H, s, 10-Me), 1.06 (3H, d, $J = 6$ Hz, 20-Me), 1.13 and 1.18 (each 3H, s, 25-Me₂), 3.62 (1H, m, 3 α -H), 3.82 (1H, d, $J = 7$ Hz, 24-H), 5.42 (2H, m, 22- or 23- and 6- or 7-H), 5.62 (2H, m, 22- or 23- and 6- or 7-H).

9b: mp 206–207 °C (MeOH-*n*-hexane). High-resolution MS m/z : Calcd for $C_{27}H_{42}O_3$: 414.3132. Found: 414.3111. UV $\lambda_{\max}^{\text{EtOH}}$ nm: 292, 280, 272, 262. $^1\text{H-NMR}$ δ : 0.64 (3H, s, 13-Me), 0.95 (3H, s, 10-Me), 1.08 (3H, d, $J = 6$ Hz, 20-Me), 1.16 and 1.20 (each 3H, s, 25-Me₂), 3.64 (1H, m, 3 α -H), 3.86 (1H, d, $J = 7$ Hz, 24-H), 5.42 (2H, m, 22- or 23- and 6- or 7-H), 5.60 (2H, m, 22- or 23- and 6- or 7-H).

(22E)-3 β ,24,25-Trihydroxycholesta-5,7,22-trienes (9a and 9b). Reduction with *l*-Selectride and LiAlH₄—The enone (**5**) (15 mg, 0.024 mmol) was dissolved in dry THF (2 ml) and 0.03 ml (0.03 mmol) of 1.0 M *l*-selectride solution in THF was slowly added at -78 °C. The reaction mixture was kept at -78 °C for 2 h with stirring, then brought back to room temperature, and the hydrolyzed with 0.01 ml of 3 N NaOH solution. The organoborane was then decomposed with 0.01 ml of 30% H₂O₂. The reaction mixture was extracted with Et₂O, washed with water, dried (Na₂SO₄) and evaporated to give 3 β ,24,25-trihydroxy-5 α ,8 α -(3,5-dioxo-4-phenyl-1,2,4-triazolidino)cholesta-6,22-diene (**17**) (10 mg, 71%) as a colorless glass. $^1\text{H-NMR}$ δ : 0.83 (3H, s, 13-Me), 0.98 (3H, s, 10-Me), 1.06 (3H, d, $J = 6$ Hz, 20-Me), 1.13 and 1.20 (each 3H, s, 25-Me₂), 3.82 (6/7H, d, $J = 7$ Hz, 24-H), 3.86 (1/7H, d, $J = 7$ Hz, 24-H), 4.46 (1H, m, 3-H), 5.46 (1H, dd, $J = 15, 7$ Hz, 23-H), 5.65 (1H, dd, $J = 15, 8$ Hz, 22-H), 6.28 and 6.44 (each 1H, d, $J = 8$ Hz, 6- and 7-H), 7.40 (5H, m, Ar).

This triol (**17**) (5 mg, 0.008 mmol) in dry THF (5 ml) was treated with LiAlH₄ (9.3 mg, 0.24 mmol) under reflux for 3 h. The crude product was purified by HPLC using a Zorbax SIL column (4.6 i.d. \times 250 mm) with 5.5% isopropanol in *n*-hexane as a mobile phase at a flow rate of 1.0 ml/min to give **9a** (1.9 mg, 57%) and **9b** (0.3 mg, 9%).

3 β -Acetoxy-24-benzoyloxy-25-hydroxy-5 α ,8 α -(3,5-dioxo-4-phenyl-1,2,4-triazolidino)cholesta-6,22-dienes (16a and 16b)—Methanol (1 ml) and NaBH₄ (1.7 mg, 0.045 mmol) were added to a solution of the enone (**5**) (15 mg, 0.024 mmol) in dry THF (1 ml). The mixture was kept at room temperature for 2 h with stirring, then extracted with ethyl acetate. The extract was evaporated to give a residue, which was dissolved in CHCl₃ (2 ml). Benzoyl chloride (0.03 ml) was added, and the resulting reaction mixture was stirred in the presence of *N,N*-diisopropyl ethylamine at room temperature for 3 h. The usual work-up (CHCl₃ extraction) and purification by HPLC on a Zorbax SIL column (4.6 i.d. \times 250 mm) with 8% isopropanol in *n*-hexane as a mobile phase at a flow rate of 0.4 ml/min gave the 24S-benzoate (**16a**) (4 mg, 24%) as a colorless glass and the 24R-benzoate (**16b**) (3.8 mg, 22%) as a colorless glass.

16a: $^1\text{H-NMR}$ δ : 2.04 (3H, s, 3-OAc), 5.32 (1H, d, $J = 8$ Hz, 24-H), 5.50 (2H, m, 22- or 23-H and 3 α -H), 5.76 (1H, m, 22- or 23-H), 6.26 and 6.42 (2H, d, $J = 8$ Hz, 6- and 7-H), 7.48 (8H, m, Ar), 8.10 (2H, d, $J = 8$ Hz, Ar).

16b: $^1\text{H-NMR}$ δ : 0.82 (3H, s, 13-Me), 1.00 (3H, s, 10-Me), 1.06 (3H, d, $J = 6$ Hz, 20-Me), 1.28 (6H, s, 25-Me₂), 2.02 (3H, s, 3-OAc), 5.32 (1H, d, $J = 8$ Hz, 24-H), 5.50 (2H, m, 22- or 23-H and 3 α -H), 5.76 (1H, m, 22- or 23-H), 6.28 and 6.44 (each 1H, d, $J = 8$ Hz, 6- and 7-H), 7.48 (8H, m, Ar), 8.10 (2H, d, $J = 8$ Hz, Ar).

Upon reduction with LiAlH₄, the benzoates (**16a** and **16b**) thus isolated were unambiguously converted into the respective alcohols (**9a** and **9b**), which were identical with the respective alcohols prepared directly by reduction of the enone (**5**) with LiAlH₄.

(22E)-24,25-Dihydroxy-22-dehydrovitamin D₃ (10a and 10b)—In the same manner as described for the synthesis of **8**, (24S)- Δ^{22} -24,25-(OH)₂-pro-D₃ (**9a**) and (24R)- Δ^{22} -24,25-(OH)₂-pro-D₃ (**9b**) (each 4 mg, 0.01 mmol) were converted into (24S)- Δ^{22} -25-(OH)₂-D₃ (**10a**) as a colorless glass (0.6 mg, 15%) and (24R)- Δ^{22} -(24R)-24,25-(OH)₂-D₃ (**10b**) as a colorless glass (0.6 mg, 15%), respectively.

10a: High-resolution MS m/z : Calcd for $C_{27}H_{42}O_3$: 414.3131. Found: 414.3125. UV $\lambda_{\max}^{\text{EtOH}}$ nm: 265; $\lambda_{\min}^{\text{EtOH}}$ nm: 228. $^1\text{H-NMR}$ δ : 0.56 (3H, s, 13-Me), 1.04 (3H, d, $J = 6$ Hz, 20-Me), 1.16 and 1.20 (each 3H, s, 25-Me₂), 3.86 (1H, d, $J = 7$ Hz, 24-H), 3.96 (1H, m, 3 α -H), 4.84 and 5.07 (each 1H, br s, 19-H₂), 5.44 (1H, dd, $J = 15, 7$ Hz, 23-H), 5.64 (1H, dd, $J = 15, 8$ Hz, 22-H), 6.04 and 6.26 (each 1H, d, $J = 11$ Hz, 6- and 7-H).

10b: High-resolution MS m/z : Calcd for $C_{27}H_{42}O_3$: 414.3132. Found: 414.3132. UV $\lambda_{\max}^{\text{EtOH}}$ nm: 265; $\lambda_{\min}^{\text{EtOH}}$ nm:

228. ¹H-NMR δ : 0.56 (3H, s, 13-Me), 1.05 (3H, d, $J=6$ Hz, 20-Me), 1.16 and 1.20 (each 3H, s, 25-Me₂), 3.83 (1H, d, $J=7$ Hz, 24-H), 3.96 (1H, m, 3 α -H), 4.83 and 5.06 (each 1H, br s, 19-H₂), 5.42 (1H, dd, $J=15, 7$ Hz, 23-H), 5.60 (1H, dd, $J=15, 8$ Hz, 22-H), 6.05 and 6.25 (each 1H, d, $J=11$ Hz, 6- and 7-H).

(22E)-3 β ,25-Dihydroxycholesta-5,7,22-trien-24-one (11)—A mixture of the enone (5) (20 mg, 0.03 mmol), anhydrous K₂CO₃ and furan (2.8 g, 0.04 mmol) in ethylene glycol was heated at 120 °C for 5 h under N₂. The mixture was then extracted with ethyl acetate, and the extract was washed with 5% HCl, saturated aqueous Na₂CO₃ and brine, and dried (Na₂SO₄). After evaporation of the solvent, the crude product was purified by HPLC on a Zorbax SIL column (4.6 i.d. \times 250 mm) with 2.5% isopropanol in *n*-hexane as a mobile phase at a flow rate of 1.5 ml/min to give the Δ^{22} -25-OH-24-oxo-pro-D₃ (11) (7 mg, 54%) as a colorless glass. High-resolution MS m/z : Calcd for C₂₇H₄₀O₃: 412.2974. Found: 412.2969. UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm: 292, 282, 272, 262. ¹H-NMR δ : 0.51 (3H, s, 13-Me), 0.93 (3H, s, 10-Me), 1.02 (3H, d, $J=6$ Hz, 20-Me), 1.38 and 1.40 (each 3H, s, 25-Me₂), 3.64 (1H, m, 3 α -H), 5.40 and 5.58 (each 1H, m, 6- and 7-H), 6.40 (1H, d, $J=15$ Hz, 23-H), 7.10 (1H, dd, $J=15, 9$ Hz, 22-H).

(22E)-25-Hydroxy-24-oxo-22-dehydrovitamin D₃ (12)—In the same manner as described for the synthesis of 8, Δ^{22} -25-OH-24-oxo-pro-D₃ (11) (0.5 mg, 0.001 mmol) was converted into Δ^{22} -25-OH-24-oxo-D₃ (12) (0.015 mg, 3%) as a colorless glass. High-resolution MS m/z : Calcd for C₂₇H₄₀O₃: 412.2976. Found: 412.2989. UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm: 265; $\lambda_{\text{min}}^{\text{EtOH}}$ nm: 228.

3 β ,22,24,25-Tetrahydroxycholesta-5,7-diene (13a and 13b)—A solution of the hydroxyketone (4a) (40 mg, 0.06 mmol) in dry THF (10 ml) was treated with LiAlH₄ (70 mg, 1.8 mmol) under reflux for 3 h. After cooling, excess reagent was destroyed by adding a few drops of water. The mixture was extracted with ethyl acetate, and the extract was dried (Na₂SO₄) and evaporated. The crude product was purified by HPLC on a Zorbax SIL column (6.2 i.d. \times 250 mm) with 15% isopropanol in *n*-hexane as a mobile phase at a flow rate of 1.7 ml/min to give the 22,24,25-(OH)₃-pro-D₃ (13a) (18 mg, 70%) as a colorless glass. High-resolution MS m/z : Calcd for C₂₇H₄₄O₄: 432.3236. Found: 432.3234. UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm: 292, 280, 272, 262. ¹H-NMR δ : 0.64 (3H, s, 13-Me), 0.94 (3H, s, 10-Me), 0.98 (3H, d, $J=6$ Hz, 20-Me), 1.18 and 1.21 (each 3H, s, 25-Me₂), 3.60 (2H, m, 3 α - and 22-H), 3.94 (1H, m, 24-H), 5.40 and 5.58 (each 1H, m, 6- and 7-H).

In the same manner as described for 13a, the hydroxyketone (4b) (40 mg, 0.06 mmol) was converted into 22,24,25-(OH)₃-pro-D₃ (13b) (18 mg, 70%) as a colorless glass. High-resolution MS m/z : Calcd for C₂₇H₄₄O₄: 432.3237. Found: 432.3230. UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm: 292, 280, 272, 262. ¹H-NMR δ : 0.63 (3H, s, 13-Me), 0.94 (3H, s, 10-Me), 0.98 (3H, d, $J=6$ Hz, 20-Me), 1.16 and 1.20 (each 3H, s, 25-Me₂), 3.60 (2H, m, 3 α - and 22-H), 3.96 (1H, d-like, $J=7$ Hz, 24-H) 5.42 and 5.58 (each 1H, m, 6- and 7-H).

22,24,25-Trihydroxyvitamin D₃ (14a and 14b)—In the same manner as described for the synthesis of 8, 22,24,25-(OH)₃-pro-D₃ (13a) (4 mg, 0.01 mmol) was converted into 22,24,25-(OH)₃-D₃ (14a). The crude product was purified by HPLC on a Zorbax SIL column (4.6 i.d. \times 250 mm) with 5.5% isopropanol in *n*-hexane as a mobile phase at a flow rate of 1.7 ml/min to give 14a as a colorless glass. High-resolutions MS m/z : Calcd for C₂₇H₄₄O₄: 432.3237. Found: 432.3217. UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm: 265; $\lambda_{\text{min}}^{\text{EtOH}}$ nm: 228. ¹H-NMR δ : 0.56 (3H, s, 13-Me), 0.97 (3H, d, $J=6$ Hz, 20-Me), 1.20 and 1.24 (each 3H, s, 25-Me₂), 3.60 (1H, m, 22- or 24-H), 4.00 (2H, m, 22- or 24-H and 3 α -H), 4.85 and 5.08 (each 1H, br s, 19-H₂), 6.07 and 6.27 (each 1H, d, $J=11$ Hz, 6- and 7-H).

Similarly, 13b (4 mg, 0.01 mmol) was converted into 14b as a colorless glass. High-resolution MS m/z : Calcd for C₂₇H₄₄O₄: 432.3236. Found: 432.3235. UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm: 265; $\lambda_{\text{min}}^{\text{EtOH}}$ nm: 228. ¹H-NMR δ : 0.54 (3H, s, 13-Me), 0.98 (3H, d, $J=6$ Hz, 20-Me), 1.17 and 1.21 (each 3H, s, 25-Me₂), 3.73 (3H, m, 3 α -, 22- and 24-H), 4.84 and 5.06 (each 1H, br s, 19-H₂), 6.05 and 6.25 (each 1H, d, $J=11$ Hz, 6- and 7-H).

Acknowledgements The authors wish to thank Dr. Yehuda Mazur of the Weizmann Institute of Science, Israel, and Dr. Glenville Jones of Queen's University, Canada, for kind gifts of authentic samples of (24S)- and (24R)-24,25-(OH)₂-D₂ (8a and 8b) and valuable advice. The authors also wish to thank Dr. Makiko Sugiura and Dr. Kayoko Saiki of Kobe Women's College of Pharmacy for ¹H-NMR and MS measurements, and Dr. Yasuho Nishii and Dr. Isao Matsunaga of Chugai Pharmaceutical Co., Ltd., for their continuous encouragement and valuable advice.

References and Notes

- 1) A part of this work was presented in a preliminary report: K. Katsumi, T. Okano, T. Kobayashi, O. Miyata, T. Naito, and I. Ninomiya, *Chem. Pharm. Bull.*, **32**, 3744 (1984).
- 2) Vitamin D is used as a general name for vitamins D₂ and D₃.
- 3) H. F. DeLuca, *J. Lab. Clin. Med.*, **87**, 7 (1976).
- 4) J. K. Wichmann, H. F. DeLuca, H. K. Schnoes, R. L. Horst, R. M. Shepard, and N. A. Jorgensen, *Biochemistry*, **18**, 4775 (1979).
- 5) Y. Tanaka, H. F. DeLuca, H. K. Schnoes, N. Ikekawa, and T. Eguchi, *Proc. Natl. Acad. Sci. U.S.A.*, **78**, 4805 (1981).
- 6) J. K. Wichmann, H. K. Schnoes, and H. F. DeLuca, *Biochemistry*, **20**, 7385 (1981).

- 7) Y. Takasaki, N. Horiuchi, N. Takahashi, E. Abe, T. Shinki, and T. Suda, *Biochem. Biophys. Res. Commun.*, **95**, 177 (1980).
- 8) E. Meyer, G. S. Reddy, J. R. Kruse, G. Popjak, and A. W. Norman, *Biochem. Biophys. Res. Commun.*, **109**, 370 (1982).
- 9) E. Meyer, G. S. Reddy, R. A. S. Chandratna, W. H. Okamura, J. R. Kruse, G. Popjak, J. E. Bishop, and A. W. Norman, *Biochemistry*, **22**, 1798 (1983).
- 10) G. Jones, M. Kung, and K. Kano, *J. Biol. Chem.*, **258**, 12920 (1983).
- 11) N. Ohnuma, K. Bannai, H. Yamaguchi, Y. Hashimoto, and A. W. Norman, *Arch. Biochem. Biophys.*, **204**, 387 (1980).
- 12) R. L. Horst, P. M. Wovkulich, E. G. Baggiolini, M. R. Uskokovic, G. W. Engstrom, and J. L. Napoli, *Biochemistry*, **23**, 3937 (1984).
- 13) S. Yamada, M. Ohmori, H. Takayama, Y. Takasaki, and T. Suda, *J. Biol. Chem.*, **258**, 457 (1983).
- 14) E. Meyer, J. E. Bishop, R. A. S. Chandraratna, W. H. Okamura, J. R. Kruse, S. Popjak, N. Ohnuma, and A. W. Norman, *J. Biol. Chem.*, **258**, 13458 (1983).
- 15) Y. Tanaka, J. K. Wichmann, H. E. Paaren, H. K. Schnoes, and H. F. DeLuca, *Proc. Natl. Acad. Sci. U.S.A.*, **77**, 6411 (1980).
- 16) N. Ohnuma and A. W. Norman, *J. Biol. Chem.*, **257**, 8261 (1982).
- 17) Y. Takasaki, T. Suda, S. Yamada, M. Ohmori, and H. Takayama, *J. Biol. Chem.*, **257**, 3732 (1982).
- 18) D. S. Morris, D. H. Williams, and A. F. Morris, *J. Org. Chem.*, **46**, 3422 (1981).
- 19) N. Ikekawa, T. Eguchi, and Y. Hirano, *J. Chem. Soc., Chem. Commun.*, **1981**, 1157.
- 20) Y. Hirano, N. Ikekawa, Y. Tanaka, and H. F. DeLuca, *Chem. Pharm. Bull.*, **29**, 2254 (1981).
- 21) N. Ikekawa, Y. Hirano, M. Ishiguro, J. Oshida, T. Eguchi, and S. Miyasaka, *Chem. Pharm. Bull.*, **28**, 2852 (1980).
- 22) G. Jones, A. Rosenthal, D. Segev, Y. Mazur, F. Frolow, Y. Haffon, D. Rabinovich, and Z. Shakked, *Biochemistry*, **18**, 1094 (1979).
- 23) G. Jones, A. Rosenthal, D. Segev, Y. Mazur, F. Frolow, Y. Haffon, D. Rabinovich, and Z. Shakked, *Tetrahedron Lett.*, **1979**, 177.
- 24) D. H. R. Barton, T. Shioiri, and D. A. Widdowson, *J. Chem. Soc. C*, **1971**, 1968.
- 25) S. C. Eyley and D. H. Williams, *J. Chem. Soc., Perkin Trans. 1*, **1976**, 727.
- 26) H. Sai, S. Takatsuto, N. Ikekawa, Y. Tanaka, C. Smith, and H. F. DeLuca, *Chem. Pharm. Bull.*, **32**, 3866 (1984).
- 27) N. C. Gonnella, K. Nakanishi, V. S. Martin, and B. Sharpless, *J. Am. Chem. Soc.*, **104**, 3775 (1982).
- 28) E. J. Corey, S. M. Albonico, U. Koelliker, T. K. Schaaf, and R. K. Varma, *J. Am. Chem. Soc.*, **93**, 1491 (1971).