Dynamic Article Links

Organic & Biomolecular Chemistry

Cite this: Org. Biomol. Chem., 2012, 10, 4448

PAPER www.rsc.org/obc

Preparation of modified peptides: direct conversion of α-amino acids into **β-amino** aldehydes†‡

Carlos J. Saavedra, Alicia Boto* and Rosendo Hernández*

Received 28th February 2012, Accepted 18th April 2012 DOI: 10.1039/c2ob25433f

A direct method for the transformation of α -amino acids into β -amino aldehydes was developed, and applied to the modification of the C-terminal residue of peptides. The method takes place in good yields and under mild conditions. The application of this methodology to the preparation of small peptides with γ-amino alcohol units, which are precursors of analogues of peptaibol antibiotics, is also described.

Introduction

The replacement of amino acid residues in peptides by amino aldehyde units has provided several peptide analogues with remarkable biological activities. Some derivatives presenting an α-amino aldehyde unit are potent inhibitors of proteases, such as papain, thrombin, trypsin, calpain, caspases and viral proteases. Their applications range from antiviral drugs to antithrombotic, anticataract or antitumoral agents.2

On the other hand, there are few examples of peptide derivatives with β-amino aldehyde units.³ In most cases, an amino glyoxal residue⁴ is introduced, as occurs with the caspase-3 inhibitor 1^{5a} (Fig. 1) and the collagen-degradation inhibitor 2.^{5b}

COOH

Fig. 1 Bioactive peptides with β -amino aldehyde units.

Instituto de Productos Naturales y Agrobiología del CSIC, Avda. Astrofísico Francisco Sánchez 3, 38206-La Laguna, Tenerife, Spain. E-mail: alicia@ipna.csic.es, rhernandez@ipna.csic.es;

Fax: +34 922260135

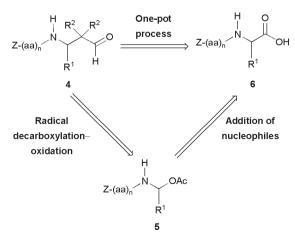
† Dedicated to Prof. Gerry Pattenden.

‡ Electronic supplementary information (ESI) available: ¹H and ¹³C NMR spectra of the amino acids 8, 14, 16, 18, 19, 21, 24, and 26-29, the peptide aldehydes 38-48 and the amino alcohol derivatives 52-57. See DOI: 10.1039/c2ob25433f.

Some bioactive peptides also present α -unsubstituted β -amino aldehyde units, ⁶ such as product 3, a potent inhibitor of interleukin 1-β converting enzyme, used as a drug lead for the treatment of inflammatory diseases.6a

The development of new derivatives, especially those with α-substituted β-amino aldehyde residues, could be useful to discover new drug leads. We reasoned that a peptide aldehyde 4 (Scheme 1) could be formed by addition of carbon nucleophiles (such as vinyl ethers or silvlenol ethers) to the N,O-acetals 5.7 These acetals, in turn, could be generated by radical decarboxylation-oxidation of α-peptides such as substrate 6.8 The direct formation of peptide aldehydes 4 from α-peptides 6 would be particularly interesting. With the one-pot process, a single α-peptide 6 could be transformed into a variety of peptide aldehydes with different substituents, allowing the generation of libraries of derivatives 4 to study structure–activity relationships.

In previous work from our group,8 the oxidative decarboxylation of α-amino acids followed by addition of different carbon nucleophiles had proven useful to obtain unnatural amino acids,



Scheme 1 Formation of peptide aldehydes from α -peptides.

One-pot oxidative radical decarboxylation-alkylation

Entry	DIB/I ₂ (equiv)	Nucleophile/Lewis acid (equiv) ^a	T (°C)	Products (%) ^b
1	2.0/1.0	A (5)/BF ₃ ·OEt ₂ (2)	0	8 (38)
2	1.5/0.5	$A (3)/BF_3 \cdot OEt_2 (2)$	0	8 (53)
3	1.5/0.3	$A (3)/BF_3 \cdot OEt_2 (2)$	0	8 (61)
4	1.5/0.3	A(3)/TMSOTf(2)	0	8 (58)
5	1.5/0.3	A (3)/SnCl ₄ (2)	-78	8 (51)
6	1.5/0.3	A (3)/TiCl ₄ (2)	-78	8 (25)
7	1.5/0.3	B (3)/BF ₃ ·OEt ₂ (2)	0	9 (41)
8	1.5/0.3	B (3)/TMSOTf (2)	0	9 (40)
9	1.5/0.3	$C(3)/BF_3 \cdot OEt_2(2)$	0	9 (60)

 a A = (TMSO)-CH=C(Me)₂; B = (TMSO)CH=CH₂; C = (EtO)-CH=CH₂. ^b Yield for products purified by chromatography.

alkaloid precursors and α,β-peptide hybrids. In the current article, a variation of this methodology is described, to allow the direct conversion of α -amino acids or α -peptides into β -amino aldehydes or peptide \(\beta\)-aldehydes, under mild conditions and good overall yields.

Results and discussion

The initial studies on the one-pot decarboxylation-alkylation process were carried out with the DL-phenyl alanine derivative 7 (Table 1), 8e which was treated with (diacetoxyiodo)benzene (DIB) and iodine under irradiation with visible light, to induce the oxidative decarboxylation.8 The scission generated an N,Oacetal intermediate, which was not isolated, but treated with a Lewis acid and a nucleophile (entries 1-9). When 1-(trimethylsilyloxy)-2-methyl-1-propene was used as the nucleophile, the β-aminoaldehyde 8 was formed, while the use of 1-(trimethylsilyloxy)-1-ethene or ethyl vinyl ether as nucleophiles provided the β-amino aldehyde 9.9

The amount of the scission reagents was important to obtain satisfactory yields (entries 1–3). The best result was obtained with a substrate/DIB/I₂ ratio of 1/1.5/0.3 (entry 3). Different Lewis acids were also tried (entries 3-6 and 7-8); boron trifluoride and TMSOTf provided the best yields. The nucleophiles B [1-(tert-butyldimethylsilyloxy)-1-ethene] and C [1-ethoxy-1ethene] were then compared (entries 7 and 9), and the vinyl ether C proved superior to the silyl enol ether B, probably due to increased reagent stability.

The best conditions for each nucleophile were then used with other amino acids, such as DL-substrates 10-138e,10 (Table 2), to afford the β-amino aldehyde derivatives **14–21**. 11

Interestingly, the oxidative decarboxylation-alkylation of the ornithine derivative 13 (Table 2, entry 10) afforded the proline analogue 21, probably due to intramolecular cyclization of the intermediate N,O-acetal to give the N,N-acetal 20, 12 which then reacted with the Lewis acid to give a five-membered acyliminium ion, followed by the addition of the nucleophile.

Table 2 One-pot oxidative radical decarboxylation-alkylation

Entry	Substrate	Nu (equiv) ^b	Products ^c (%)
	Bz N CO ₂ H		Bz N R H
1 2 3	Bz-Ala-OH 10	A (3) B (3) C (3)	14 R = Me (69) 15 R = H (45) 15 R = H (79)
	Bz N CO ₂ H		Bz N R H
4 5 6	Bz-Leu-OH 11	A (3) B (3) C (3)	16 R = Me (60) 17 R = H (55) 17 R = H (56)
	Bz-N CO ₂ H		Bz-N R H
7 8 9	Bz-Glu(OEt)-OH 12	A (3) B (3) C (3)	18 R = Me (66) 19 R = H (51) 19 R = H (45)
	MeO ₂ C-N CO ₂ H		MeO O
10	13	A (3)	$20 \rightarrow 21 (83)$ $20 X = NHCO_2Me$ $21 X = C(Me)_2$ -CHO

^a DIB, I₂, hv, then 0 °C, nucleophile, Lewis acid. ^b A = (TMSO)- $CH = C(Me)_2$; $B = (TMSO)CH = CH_2$; $C = (EtO)CH = CH_2$; $BF_3 \cdot OEt_2$ was used as the Lewis acid. ^cYield for products purified by chromatography.

Similar products were obtained when the DL-proline and hydroxy L-proline derivatives 22¹³ (Table 3, entries 1-3) and 23¹⁴ (entries 4–6) underwent the decarboxylation–alkylation reaction. In the case of the proline derivative 22, the process took place in 82-86% yields, affording products 24 (R = Me) or 25 (R = H). 15a-d When hydroxyproline substrate 23 was used, the process afforded compounds 26-27 (R = Me) and 28-29(R = H). ^{15e} Remarkably, the 2,4-cis products predominated over the 2,4-trans isomers, due to an stereoelectronic effect described by Woerpel and coworkers. 16

The one-pot process was then tried with peptides, using compounds 30–36¹⁷ (Scheme 2) as substrates. Interestingly, with peptides TMSOTf proved superior to boron trifluoride as the Lewis acid (conversions $30 \rightarrow 37$ and $31 \rightarrow 38$).

In the case of substrates 30 and 31, whose N-terminal residue was an α,α-disubstituted amino acid, the process afforded a racemic mixture of the peptide aldehydes 37 and 38, respectively. In the case of substrates 32–36, the reacting residue was attached to a chiral amino acid, so the process was stereoselective (dr from 3:2 to 3:1), affording compounds **39–46** in good overall yields.

Table 3 One-pot oxidative radical decarboxylation—alkylation

	•	•	•
Entry	Substrate	Nu (equiv) ^b	Products ^c (%)
	Cbz N CO ₂ H		Cbz N1 R R N1 R
1 2 3	22	A (3) B (3) C (3)	24 R = Me (86) 25 R = H (82) 25 R = H (85)
	Cbz N CO ₂ H TBSO		TBSO O
4	23	A (3)	26 (2 <i>R</i>) (63) 27 (2 <i>S</i>) (19)
			Cbz N * H
5	23	B (3)	28 (2 <i>R</i>) (34)
6	23	C (3)	29 (2 <i>S</i>) (20) 28 (2 <i>R</i>) (27) 29 (2 <i>S</i>) (13)

^a DIB, I₂, hv, then 0 °C, nucleophile, Lewis acid. ^b A = (TMSO)-CH=C(Me)₂; B = (TMSO)CH=CH₂; C = (EtO)CH=CH₂. c Yield for products purified by chromatography.

Their stereochemistry was unambiguously determined by correlation to known compounds, as will be commented later.

The aldehydes are not only potential drug candidates, but also useful precursors of other compounds, such as diamines, amino alcohols, etc. Especially interesting are the amino alcohol derivatives (Scheme 3) which are precursors of analogues of the peptaibol antibiotics. 18 The peptaibols present a short peptidic chain with a C-terminal β-amino alcohol unit. The presence of nonproteinogenic amino acids (in particular Aib, α-amino isobutyric acid) is key for their potent activity against bacteria and fungi. These antibiotics present interesting folding patterns which can allow the creation of holes in the bacterial membrane.

Our scission-alkylation methodology, followed by mild reduction of the aldehydes, would allow the preparation of precursors of peptaibol analogues, such as compounds 47-56 (Scheme 3). These precursors present unusual C-terminal β , β -dimethyl γ-amino alcohol units (instead of the peptaibol β-amino alcohol units), and with their β , β -substitution they also resemble an Aib-derived amino alcohol. This new terminal residue could modify the conformational and biological properties of the derivatives, and thus be useful to understand SAR relationships.

The reduction of peptide aldehydes 39-46 was tried under different conditions (DIBAL-H, NaBH4, or LiBH4 in different solvents), and the best results were obtained with LiBH4 in ¹PrOH, affording the amino alcohols in good yields (69–79%).

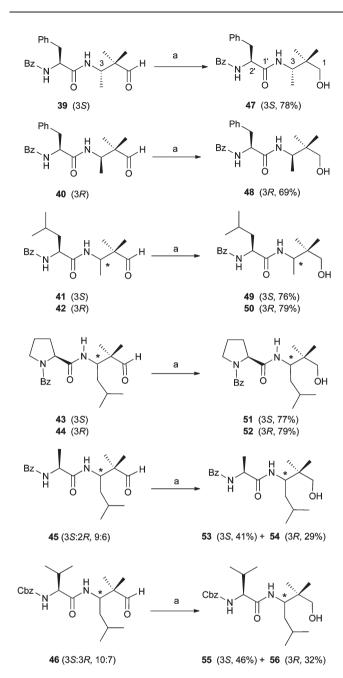
To our satisfaction, in the case of aldehyde mixtures, such as compounds 45 and 46, the reduction allowed the separation of the diastereomeric products.

Scheme 2 Conversion of α -dipeptides into derivatives with a C-terminal β-amino aldehyde. Reaction conditions: [a] DIB, I₂, hv, then 0 °C, BF₃·OEt₂, (TMSO)CH=C(Me)₂; [b] DIB, I₂, hv, then 0 °C, TMSOTf, $(TMSO)CH=C(Me)_2.$

36 Z = Cbz, R = CHMe₂

The stereochemistry of the amino aldehydes and the amino alcohols was determined by correlation to known compounds

46 R = CHMe₂, 76%, d.r. 10:7



Scheme 3 Conversion of dipeptides with a β -amino aldehyde unit into alcohol derivatives. Reaction conditions: [a] LiBH₄, ⁱPrOH.

(Scheme 4).8e Thus, the acids 32 and 33 were transformed into the known β-amino esters 52-53 and 54-55, respectively, using a sequential scission–alkylation process. 8e

The known esters 52-55 were then reduced with DIBAL-H to give the corresponding aldehydes 39-42 or the amino alcohols **47–50**, ¹⁹ confirming the assigned stereochemistries. In a similar way, the amino ester 57^{8e} (Scheme 5) was reduced to give the γ -amino alcohol derivative 51. In all the cases, the major isomer presented the "natural" (3S) configuration.

The formation of peptaibol analogues from these simpler precursors is under way, and together with their conformational properties, will be reported in due course.

Scheme 4 Determination of the stereochemistry of the aldehydes by correlation to known compounds. Reaction conditions: [a] DIBAL-H, CH₂Cl₂, -78 °C; [b] DIBAL-H, CH₂Cl₂, 0 °C.

Scheme 5 Determination of the stereochemistry of the aldehydes by correlation to known compounds. Reaction conditions: [i] DIBAL-H, CH₂Cl₂, 0 °C.

Conclusions

An efficient and mild one-pot process for the conversion of α-amino acid derivatives into β-amino aldehydes has been developed, and applied to the selective modification of the C-terminal residue in small peptides, giving peptide aldehydes in good overall yields. With this procedure, a single α -peptide could be transformed into a library of α,β-peptidyl aldehydes with different α-substituents. The aldehydes are useful precursors of other compounds, as illustrated by transformation of several peptide aldehydes into precursors of peptaibol analogues, which present γ-hydroxyamino units.

Experimental section

General remarks

Commercially available reagents and solvents were analytical grade or were purified by standard procedures prior to use.²⁰ All reactions involving air- or moisture-sensitive materials were carried out under nitrogen atmosphere. Three alternative spray reagents for TLC analysis were used: (a) 0.5% vanillin in H₂SO₄-EtOH (4:1); (b) 0.25% ninhydrin in ethanol; and (c) Fleet's reagent [Ce(SO₄)₂ (0.5 g) and ammonium phosphomolybdate hydrate (2.5 g) in H₂SO₄ (5 mL) and water (65 mL)]. Once sprayed, the TLC was heated until development of color. Merck silica gel 60 PF₂₅₄ and 60 (0.063-0.2 mm) were used for rotatory chromatography and column chromatography, respectively. Melting points were determined with a hot-stage apparatus and are uncorrected; the term "net" is used for crystals resulting from evaporation of the chromatography eluents. Optical rotations were measured at the sodium line at ambient temperature (26 °C). NMR spectra were determined at 500 or 400 MHz for ¹H and 125.7 or 100.6 MHz for ¹³C in CDCl₃ as solvent and at 25 °C ($\delta_{\rm H}$ 7.26; $\delta_{\rm C}$ 77.0), unless otherwise stated.

Amino aldehyde derivatives 9, 15, 17, and 25 are known but some are partially described; for comparison purposes their spectroscopic and physical data are also included.

General procedures for the scission-oxidation-alkylation sequence

Method A. To a solution of the starting amino acid or peptide (0.2 mmol) in dry dichloromethane (8 mL) were added iodine (15 mg, 0.06 mmol, 0.3 equiv) and (diacetoxyiodo)benzene (DIB) (97 mg, 0.3 mmol, 1.5 equiv). The reaction mixture was stirred at 25–26 °C for 4 h, under irradiation with visible light. Then the solution was cooled to 0 °C, and 2-methyl-1-(trimethylsilyloxy)-1-propene (110 μL, 86 mg, 0.6 mmol, 3 equiv) or vinyloxytrimethylsilane (89 µL, 70 mg, 0.6 mmol, 3 equiv) or ethyl vinyl ether (86 µL, 65 mg, 0.6 mmol, 3.0 equiv) was injected, followed by dropwise addition of BF₃·OEt₂ (51 µL, 57 mg, 0.4 mmol, 2 equiv). The mixture was allowed to reach room temperature and stirred for 3 h; then it was poured into 10% aqueous Na₂S₂O₃-saturated aqueous NaHCO₃ (1:1, 10 mL) and extracted with CH₂Cl₂. The organic layer was dried over sodium sulfate, filtered and evaporated under reduced pressure. The residue was purified by chromatography on silica gel (hexanes-EtOAc) to give the products.

Method B. As in Method A but using trimethylsilyl triflate (TMSOTf) (72 µL, 89 mg, 0.4 mmol, 2 equiv) as the Lewis acid.

N-Benzoyl-α,α-dimethyl-DL-β-homophenylalaninal

(8). Obtained from N-benzoyl-DL-phenylalanine (7) according to Method A, using 2-methyl-1-(trimethylsilyloxy)-1-propene as the nucleophile. The reaction mixture was purified by rotatory chromatography (hexanes-EtOAc, 90:10), giving the aldehyde (±)-8 (61%) as a crystalline solid; Mp 133-134 °C (EtOAc*n*-hexane); $v_{\text{max}}/\text{cm}^{-1}$ 3439, 3083, 3067, 1724, 1660; δ_{H} (500 MHz, CDCl₃) 1.25 (3H, s, 2-Me_a), 1.31 (3H, s, 2-Me_b), 2.75 (1H, dd, J = 11.2, 14.6 Hz, 4-H_a), 3.10 (1H, dd, J = 4.1,

14.2 Hz, 4-H_b), 4.69 (1H, ddd, J = 4.1, 10.2, 10.8 Hz, 3-H), 6.38 (1H, d, J = 9.8 Hz, NH), 7.18 (1H, dd, J = 6.8, 6.8 Hz, Ar), 7.23–7.28 (4H, m, Ar), 7.36 (2H, dd, J = 7.5, 7.8 Hz, Ar), 7.45 (1H, dd, J = 7.1, 7.8 Hz, Ar), 7.52 (2H, d, J = 7.1 Hz, Ar), 9.59(1H, s, CHO); $\delta_{\rm C}$ (125.7 MHz, CDCl₃) 19.5 (CH₃), 20.1 (CH₃), 36.7 (CH₂), 50.5 (C), 54.7 (CH), 126.7 (3 \times CH), 128.5 (4 \times CH), 128.9 (2 × CH), 131.3 (CH), 134.6 (C), 137.8 (C), 167.4 (C), 205.3 (CH); m/z 295 (M⁺, <1%), 105 (100, [PhCO]⁺); HRMS calcd for C₁₉H₂₁NO₂ 295.1572, found 295.1580; calcd for C₇H₅O 105.0340, found 105.0343. C₁₉H₂₁NO₂ requires C, 77.26; H, 7.17; N, 4.74%. Found: C, 77.18; H, 7.43; N, 4.74.

(9).9 N-Benzoyl-DL-N-Benzoyl-dl-β-homophenylalaninal phenylalanine (7) was treated as in Method A, using ethyl vinyl ether as the nucleophile. The reaction mixture was purified by chromatography (hexanes-EtOAc, 60:40), giving the aldehyde (±)-9 (60%) as a syrup; $v_{\text{max}}/\text{cm}^{-1}$ 3437, 3065, 1723, 1657; $\delta_{\rm H}$ (500 MHz, CDCl₃) 2.73 (1H, ddd, J = 1.6, 6.0, 17.3 Hz, 2- H_a), 2.77 (1H, ddd, J = 1.3, 5.4, 17.7 Hz, 2- H_b), 2.97 (1H, dd, J = 7.6, 13.6 Hz, 4-H_a), 3.09 (1H, dd, J = 6.9, 13.6 Hz, 4-H_b), 4.76 (1H, m, 3-H), 6.58 (1H, d, J = 8.2 Hz, NH), 7.21–7.25 (3H, m, Ar), 7.32 (2H, dd, J = 7.3, 7.6 Hz, Ar), 7.40 (2H, dd, J = 7.6, 7.6 Hz, Ar), 7.48 (1H, dd, J = 7.3, 7.6 Hz, Ar), 7.68 (2H, d, J = 7.6 Hz, Ar), 9.77 (1H, dd, J = 0.6, 0.9 Hz, CHO); $\delta_{\rm C}$ (125.7 MHz, CDCl₃) 40.1 (CH₂), 46.7 (CH₂), 46.9 (CH), 126.9 $(3 \times CH)$, 128.6 $(2 \times CH)$, 128.8 $(2 \times CH)$, 129.2 $(2 \times CH)$, 131.6 (CH), 134.3 (C), 137.3 (C), 167.0 (C), 201.2 (CH); *m/z* 268 (M⁺ + H, 12%), 176 (69, M⁺ - PhCH₂), 105 (100, [PhCO]⁺), 91 (37, [PhCH₂]⁺), 77 (81, [Ph]⁺); HRMS calcd for C₁₇H₁₈NO₂ 268.1338, found 268.1327; calcd for C₁₀H₁₀NO₂ 176.0712, found 176.0708; calcd for C₇H₅O 105.0340, found 105.0341; calcd for C₇H₇ 91.0548, found 91.0549; calcd for C₆H₅ 77.0391, found 77.0388. C₁₇H₁₇NO₂ requires C, 76.38; H, 6.41; N, 5.24%. Found: C, 76.38; H, 6.41; N, 5.03.

N-Benzoyl-α,α-dimethyl-DL-β-homoalaninal (14). Obtained from N-benzoyl-DL-alanine (10) according to Method A, using 2-methyl-1-(trimethylsilyloxy)-1-propene as the nucleophile. The reaction mixture was purified by chromatography (hexanes-EtOAc, 70:30), giving the aldehyde (\pm)-14 (69%) as a syrup; $v_{\text{max}}/\text{cm}^{-1}$ 3439, 3081, 1725, 1650, 1519, 1487; δ_{H} (500 MHz, CDCl₃) 1.14 (3H, s, 2-Me_a), 1.18 (3H, s, 2-Me_b), 1.23 (3H, d, J = 6.9 Hz, 3-Me), 4.43 (1H, m, 3-H), 6.58 (1H, d, J = 8.8 Hz, NH), 7.41 (2H, dd, J = 7.3, 7.6 Hz, Ar), 7.48 (1H, dd, J = 7.3, 7.6 Hz, Ar), 7.73 (2H, d, J = 6.9 Hz, Ar), 9.54 (1H, s, CHO); $\delta_{\rm C}$ (125.7 MHz, CDCl₃) 16.3 (CH₃), 18.9 (CH₃), 20.0 (CH₃), 49.4 (CH), 49.9 (C), 126.8 (2 × CH), 128.6 (2 × CH), 131.5 (CH), 134.5 (C), 166.8 (C), 205.9 (CH); m/z 220 (M⁺ + H, 6%), 219 $(M^+, <1\%)$, 148 (32, $M^+ - H - Me₂CCHO)$, 105 (100, [PhCO]⁺), 77 (36, [Ph]⁺). HRMS calcd for C₁₃H₁₈NO₂ 220.1338, found 220.1333; calcd for C₁₃H₁₇NO₂ 219.1259, found 219.1270; calcd for C₉H₁₀NO 148.0762, found 148.0764; calcd for C₇H₅O 105.0340, found 105.0339; calcd for C₆H₅ 77.0391, found 77.0394. C₁₃H₁₇NO₂ requires C, 71.21; H, 7.81; N, 6.39%. Found: C, 71.10; H, 8.04; N, 6.51.

N-Benzoyl-DL-β-homoalaninal (15). 11a,b N-Benzoyl DL-alanine (10) was treated as in Method A, using ethyl vinyl ether as the nucleophile. The reaction mixture was purified by chromatography (hexanes-EtOAc, 50:50), giving the aldehyde (\pm)-15 (79%) as an oil; $v_{\text{max}}/\text{cm}^{-1}$ 3440, 3063, 1724, 1656, 1517; $\delta_{\rm H}$ (500 MHz, CDCl₃) 1.36 (3H, d, J = 6.8 Hz, 3-Me), 2.74 (1H, dd, J = 5.7, 17.3 Hz, 2-H₂), 2.80 (1H, ddd, J = 1.9, 5.7, 17.3 Hz, 2-H_b), 4.61 (1H, m, 3-H), 6.56 (1H, br b, NH), 7.40 (2H, dd, J = 7.3, 7.9 Hz, Ar), 7.48 (1H, dd, J = 7.6 Hz, Ar), 7.73 (2H, d, J = 8.2 Hz, Ar), 9.81 (1H, s, CHO); $\delta_{\rm C}$ (125.7 MHz, CDCl₃) 20.5 (CH₃), 41.6 (CH), 49.6 (CH₂), 126.9 (2 × CH), 128.5 (2 × CH), 131.5 (CH), 134.3 (C), 166.8 (C), 201.1 (CH); m/z 191 $(M^+, <1\%)$, 163 (2, $M^+ - CO$), 121 (4, $[PhCONH_2]^+$), 105 (100, $[PhCO]^{+}$), 77 (75, $[Ph]^{+}$); HRMS calcd for $C_{11}H_{13}NO_{2}$ 191.0946, found 191.0941; calcd for C₁₀H₁₃NO 163.0997, found 163.1004; calcd for C₇H₇NO 121.0528, found 121.0524; calcd for C₇H₅O 105.0340, found 105.0344; calcd for C₆H₅ 77.0391, found 77.0389. C₁₁H₁₃NO₂ requires C, 69.09; H, 6.85; N, 7.32%. Found: C, 69.23; H, 7.02; N, 7.43.

N-Benzovl-α,α-dimethyl-DL-β-homoleucinal (16). Obtained from N-benzoyl DL-leucine (11) according to Method A, using 2-methyl-1-(trimethylsilyloxy)-1-propene as the nucleophile. The reaction mixture was purified by chromatography (hexanes-EtOAc, 9:1), giving the aldehyde (\pm)-16 (60%) as an oil; $v_{\text{max}}/\text{cm}^{-1}$ 3440, 3081, 3065, 1725, 1706, 1657, 1519; δ_{H} $(500 \text{ MHz}, \text{CDCl}_3) 0.91 (3\text{H}, d, J = 6.7 \text{ Hz}, 5\text{-Me}_a), 0.95 (3\text{H}, d)$ d, J = 6.7 Hz, 5-Me_b), 1.13 (3H, s, 2-Me_a), 1.17 (3H, s, 2-Me_b), 1.30 (1H, ddd, J = 2.6, 9.8, 14.4 Hz, 4-H_a), 1.46 (1H, ddd, J = 3.6, 11.4, 14.0 Hz, 4-H_b), 1.64 (1H, m, 5-H), 4.44 (1H, ddd, J = 2.6, 9.8, 11.0 Hz, 3-H), 6.29 (1H, br d, J = 9.3 Hz, NH), 7.42 (2H, dd, J = 7.3, 7.9 Hz, Ar), 7.49 (1H, dd, J = 7.2, 7.8 Hz, Ar), 7.74 (2H, d, J = 7.2 Hz, Ar), 9.54 (1H, s, CHO); $\delta_{\rm C}$ (125.7 MHz, CDCl₃) 19.2 (CH₃), 19.6 (CH₃), 21.4 (CH₃), 23.8 (CH₃), 25.2 (CH), 39.8 (CH₂), 50.5 (C), 51.9 (CH), 126.8 (2 × CH), 128.6 (2 × CH), 131.5 (CH), 134.5 (C), 167.3 (C), 205.8 (CH); m/z 262 (M⁺ + H, 3%), 190 (31, M⁺ - H - Me_2CCHO), 105 (100, $[PhCO]^+$). HRMS calcd for $C_{16}H_{24}NO_2$ 262.1807, found 262.1801; calcd for C₁₂H₁₆NO 190.1232, found 190.1223; calcd for C₇H₅O 105.0340, found 105.0344. C₁₆H₂₃NO₂ requires C, 73.53; H, 8.87; N, 5.36%. Found: C, 73.43; H, 9.09; N, 5.27.

N-Benzoyl-DL-β-homoleucinal (17). 11c N-Benzoyl DL-leucine (11) was treated as in Method A, using ethyl vinyl ether as the nucleophile. The reaction mixture was purified by chromatography (hexanes-EtOAc, 70:30), giving the aldehyde (±)-17 (56%) as a oil; $v_{\text{max}}/\text{cm}^{-1}$ 3437, 3065, 1704, 1655, 1518, 1486; $\delta_{\rm H}$ (500 MHz, CDCl₃) 0.94 (3H, d, J = 6.6 Hz, 5-Me_a), 0.95 $(3H, d, J = 6.9 Hz, 5-Me_b), 1.42 (1H, ddd, J = 5.0, 10.1, 15.1)$ Hz, 4-H_a), 1.67 (1H, m, 4-H_b), 1.68 (1H, m, 5-H), 2.72 (1H, ddd, J = 2.2, 5.9, 17.3 Hz, 2-H_a), 2.78 (1H, br dd, J = 5.5, 17.3 Hz, 2-H_b), 4.61 (1H, m, 3-H), 6.50 (1H, br d, J = 8.8 Hz, NH), 7.40 (2H, dd, J = 7.3, 7.9 Hz, Ar), 7.48 (1H, dd, J = 7.6, 7.8 Hz, Ar), 7.73 (2H, d, J = 7.0 Hz, Ar), 9.80 (1H, s, CHO); $\delta_{\rm C}$ (125.7 MHz, CDCl₃) 22.0 (CH₃), 22.9 (CH₃), 25.2 (CH), 43.7 (CH₂), 44.0 (CH), 48.8 (CH₂), 126.9 (2 × CH), 128.6 (2 × CH), 131.5 (CH), 134.4 (C), 167.0 (C), 201.4 (CH); m/z 234 (M⁺ + H, 3%), 233 (M $^+$, <1%), 190 (3, M $^+$ – CH₂CHO), 105 (100, $[PhCO]^{+}$), 77 (53, $[Ph]^{+}$). HRMS calcd for $C_{14}H_{20}NO_{2}$ 234.1494, found 234.1502; calcd for C₁₄H₁₉NO₂ 233.1416, found 233.1419; calcd for C₁₂H₁₆NO 190.1232, found 190.1228; calcd for C₇H₅O 105.0340, found 105.0345; calcd for

C₆H₅ 77.0391, found 77.0394. C₁₄H₁₉NO₂ requires C, 72.07; H, 8.21; N, 6.00%. Found: C, 72.23; H, 8.24; N, 6.01.

Ethyl 4-benzamido-5,5-dimethyl-6-oxo hexanoate (18). Obtained from Bz-Glu(OEt)-OH (12) according to Method A, using 2methyl-1-(trimethylsilyloxy)-1-propene as the nucleophile. The reaction mixture was purified by rotary chromatography (hexanes-EtOAc, 85:15), giving the aldehyde (\pm)-18 (66%) as an oil; $v_{\text{max}}/\text{cm}^{-1}$ 3428, 1725, 1658, 1519; δ_{H} (500 MHz, CDCl₃) note: the usual numbering (CHO as C-1) is used, not the one given by IUPAC nomenclature: 1.13 (3H, dd, J = 6.7, 7.0Hz, Et), 1.15 (3H, s, 2-Me_a), 1.20 (3H, s, 2-Me_b), 1.81 (1H, m, $4-H_a$), 1.96 (1H, dddd, J = 3.2, 6.9, 6.9, 13.9 Hz, $4-H_b$), 2.30-2.45 (2H, m, 5-H₂), 3.94-4.08 (2H, m, Et), 4.35 (1H, ddd, J = 3.2, 9.8, 12.9 Hz, 3-H), 6.55 (1H, d, <math>J = 9.8 Hz, NH), 7.41(2H, dd, J = 7.3, 7.6 Hz, Ar), 7.48 (1H, dd, J = 7.3, 7.6 Hz, Ar),7.74 (2H, d, J = 7.5 Hz, Ar), 9.53 (1H, s, CHO); $\delta_{\rm C}$ (125.7 MHz, CDCl₃) 14.1 (CH₃), 19.1 (CH₃), 19.8 (CH₃), 25.0 (CH₂), 31.3 (CH₂), 50.3 (C), 53.6 (CH), 60.6 (CH₂), 126.9 (2 × CH), 128.6 (2 × CH), 131.7 (CH), 133.9 (C), 167.3 (C), 173.6 (C), 205.4 (CH); m/z 306 (M⁺ + H, 1%), 234 (43, M⁺ -Me₂CCHO), 105 (100, [PhCO]⁺), 77 (48, [Ph]⁺). HRMS calcd for C₁₇H₂₄NO₄ 306.1705, found 306.1714; calcd for C₁₃H₁₆NO₃ 234.1130, found 234.1131; calcd for C₇H₅O 105.0340, found 105.0343; calcd for C₆H₅ 77.0391, found 77.0388. C₁₇H₂₃NO₄ requires C, 66.86; H, 7.59; N, 4.59%. Found: C, 66.63; H, 7.77; N, 4.58.

Ethyl 4-benzamido-6-oxohexanoate (19). Obtained from Bz-Glu(OEt)-OH (12) according to Method A, using vinyloxytrimethylsilane as the nucleophile. The reaction mixture was purified by chromatography (hexanes-EtOAc, 50:50), giving the aldehyde (±)-**19** (51%) as a oil; $v_{\text{max}}/\text{cm}^{-1}$ 3434, 1724, 1658, 1518; $\delta_{\rm H}$ (500 MHz, CDCl₃) 1.18 (3H, dd, J = 6.9, 7.3 Hz, Et), 1.97 (1H, m, 4-H_a), 2.06 (1H, m, 4-H_b), 2.38–2.51 (2H, m, 5-H₂), 2.75 (1H, dd, J = 5.7, 17.1 Hz, 2-H_a), 2.83 (1H, ddd, $J = 1.9, 5.7, 17.3 \text{ Hz}, 2-H_b$, 4.01-4.11 (2H, m, Et), 4.51 (1H, m, 3-H), 6.94 (1H, d, J = 8.2 Hz, NH), 7.40 (2H, dd, J = 7.8, 7.9 Hz, Ar), 7.47 (1H, dd, J = 7.3, 7.6 Hz, Ar), 7.75 (2H, d, J = 7.3 Hz, Ar), 9.79 (1H, s, CHO); $\delta_{\rm C}$ (125.7 MHz, CDCl₃) 14.0 (CH₃), 28.9 (CH₂), 31.2 (CH₂), 45.7 (CH), 48.5 (CH₂), 60.7 (CH₂), 126.9 (2 × CH), 128.5 (2 × CH), 131.6 (CH), 133.9 (C), 167.0 (C), 173.7 (C), 200.8 (CH); m/z 278 (M⁺ + H, 6%), 234 (12, M^+ – CH₂CHO), 172 (99, M^+ – PhCO), 144 (60, M^+ - PhCO - CHO), 105 (100, [PhCO]⁺), 77 (99, [Ph]⁺); HRMS calcd for C₁₅H₂₀NO₄ 278.1392, found 278.1395; calcd for C₁₃H₁₆NO₃ 234.1130, found 234.1123; calcd for C₈H₁₄NO₃ 172.0974, found 172.0976; calcd for C₇H₁₄NO₂ 144.1025, found 144.1020; calcd for C₇H₅O 105.0340, found 105.0344; calcd for C₆H₅ 77.0391, found 77.0393. C₁₅H₁₉NO₄ requires C, 64.97; H, 6.91; N, 5.05%. Found: C, 64.98; H, 6.96; N, 5.19.

N-Methoxycarbonyl-α,α-dimethyl-pl-β-homoprolinal

(21). Obtained from the ornithine derivative 13 according to Method A, using 2-methyl-1-(trimethylsilyloxy)-1-propene as the nucleophile. The reaction mixture was purified by chromatography (hexanes-EtOAc, 60:40), giving the aldehyde (±)-21 (83%) as a syrup; $v_{\text{max}}/\text{cm}^{-1}$ 1719, 1690, 1454, 1386; δ_{H} (500 MHz, CDCl₃, 70 °C) 1.00 (3H, s, 2-Me_a), 1.01 (3H, s, 2- Me_b), 1.74–1.90 (3H, m, 4- H_a + 5- H_2), 2.00 (1H, m, 4- H_b), 3.19

(1H, m, 6-H_a), 3.65 (3H, s, OMe), 3.71 (1H, m, 6-H_b), 4.14 (1H, m, 3-H), 9.53 (1H, s, CHO); $\delta_{\rm C}$ (125.7 MHz, CDCl₃, 70 °C) 16.3 (CH₃), 20.0 (CH₃), 24.4 (CH₂), 27.1 (CH₂), 47.9 (CH₂), 50.5 (C), 52.2 (CH₃), 62.0 (CH), 156.4 (C), 203.4 (CH); m/z 200 (M^+ + H, <1%), 199 (M^+ , <1%), 128 (100, M^+ -Me₂CCHO). HRMS calcd for C₁₀H₁₈NO₃ 200.1287, found 200.1284; calcd for C₁₀H₁₇NO₃ 199.1208, found 199.1199; calcd for C₆H₁₀NO₂ 128.0712, found 128.0717. C₁₀H₁₇NO₃ requires C, 60.28; H, 8.60; N, 7.03%. Found: C, 59.99; H, 8.61; N, 7.36.

N-Benzyloxycarbonyl-α,α-dimethyl-DL-β-homoprolinal Obtained from DL-proline benzyl carbamate (22) according to Method A, using 2-methyl-1-(trimethylsilyloxy)-1-propene as the nucleophile. The reaction mixture was purified by chromatography (hexanes-EtOAc, 70:30), giving the aldehyde (±)-24 (86%) as a syrup; $v_{\text{max}}/\text{cm}^{-1}$ 1689, 1451, 1416; δ_{H} (500 MHz, CDCl₃, 70 °C) 1.01 (3H, s, 2-Me), 1.02 (3H, s, 2-Me), 1.74-1.90 (3H, m, $3-H_a + 4-H_2$), 1.98 (1H, m, $3-H_b$), 3.24 (1H, m, 5-H_a), 3.75 (1H, m, 5-H_b), 4.18 (1H, m, 2-H), 5.06 (1H, d, J = 12.4 Hz, OCH_aPh), 5.12 (1H, d, J = 12.3 Hz, OCH_bPh), 7.25–7.36 (5H, m, Ar), 9.54 (1H, s, CHO); $\delta_{\rm C}$ (125.7 MHz, CDCl₃, 70 °C) 16.5 (CH₃), 20.1 (CH₃), 22.4 (CH₂), 27.1 (CH₂), 47.9 (CH₂), 50.5 (C), 62.1 (CH), 67.2 (CH₂), 128.0 (2 × CH), 128.1 (CH), 128.5 (2 × CH), 136.9 (C), 155.9 (C), 203.5 (CH); m/z 275 (M⁺, <1%), 204 (80, M⁺ - Me₂CCHO), 91 (100, $[PhCH_2]^+$). HRMS calcd for $C_{16}H_{21}NO_3$ 275.1521, found 275.1512; calcd for C₁₂H₁₄NO₂ 204.1025, found 204.1026; calcd for C₇H₇ 91.0548, found 91.0548. C₁₆H₂₁NO₃ requires C, 69.79; H, 7.69; N, 5.09%. Found: C, 69.62; H, 7.75; N, 5.13.

N-Benzyloxycarbonyl-DL-β-homoprolinal (25). 15a-d Obtained from the proline derivative 22 according to Method A, using 2ethyl vinyl ether as the nucleophile. The reaction mixture was purified by chromatography (hexanes-EtOAc, 50:50), giving the aldehyde (\pm)-25 (85%) as a syrup; $v_{\text{max}}/\text{cm}^{-1}$ 1717, 1692, 1420; $\delta_{\rm H}$ (500 MHz, CDCl₃, 70 °C) 1.69 (1H, m, 3-H_a), 1.81-1.92 (2H, m, 4-H₂), 2.14 (1H, m, 3-H_b), 2.50 (1H, ddd, $J = 1.9, 7.9, 16.4 \text{ Hz}, 1'-H_a$, 2.90 (1H, br b, 1'-H_b), 3.43 (1H, ddd, J = 5.4, 6.9, 10.7 Hz, 5-H_a), 3.49 (1H, m, 5-H_b), 4.30 (1H, m, 2-H), 5.13 (2H, s, OCH₂Ph), 7.28-7.35 (5H, m, Ar), 9.75 (1H, br s, CHO); $\delta_{\rm C}$ (125.7 MHz, CDCl₃, 70 °C) 23.5 (CH₂), 31.5 (CH₂), 46.6 (CH₂), 48.9 (CH₂), 53.0 (CH), 66.9 (CH₂), 128.0 (2 × CH), 128.5 (3 × CH), 137.0 (C), 154.8 (C), 200.0 (CH); m/z 247 (M⁺, 2%), 204 (27, M⁺ – CH₂CHO), 91 (100, $[PhCH_2]^+$). HRMS calcd for $C_{14}H_{17}NO_3$ 247.1208, found 247.1205; calcd for C₁₂H₁₄NO₂ 204.1025, found 204.1027; calcd for C₇H₇ 91.0548, found 91.0546. C₁₄H₁₇NO₃ requires C, 68.00; H, 6.93; N, 5.66%. Found: C, 68.35; H, 7.14; N, 5.30.

(2R,4R)-Benzyl 4-(tert-butyldimethylsilyloxy)-2-(2-methyl-1oxopropan-2-yl)pyrrolidine-1-carboxylate (26) and (2S,4R)benzyl 4-(tert-butyldimethylsilyloxy)-2-(2-methyl-1-oxo propan-2-yl)pyrrolidine-1-carboxylate (27). Obtained from hydroxyproline derivative 23 according to Method A, using 2-methyl-1-(trimethylsilyloxy)-1-propene as the nucleophile. The reaction mixture was purified by rotatory chromatography (hexanes-EtOAc, 90:10), giving the 2,4-cis product 26 (63%) and the 2,4-trans isomer 27 (19%).

Product 26. Syrup; $[\alpha]_D$ +8 (c 1.05 in CHCl₃); $v_{\text{max}}/\text{cm}^{-1}$ 1724, 1691, 1471; $\delta_{\rm H}$ (500 MHz, CDCl₃, 70 °C) note: the usual numbering (CHO as C-1) is used, not the one given by IUPAC nomenclature: 0.07 (3H, s, SiMe_a), 0.08 (3H, s, SiMe_b), 0.91 (9H, s, tBu), 1.00 (3H, s, 2-Me_a), 1.07 (3H, s, 2-Me_b), 1.72 (1H, ddd, J = 7.6, 7.6, 13.2 Hz, 4-H_a), 2.27 (1H, ddd, J = 7.5, 7.5, 13.2 Hz, 4-H_b), 2.96 (1H, dd, J = 7.9, 11.3 Hz, 6-H_a), 3.99 (1H, dd, J = 7.3, 11.1 Hz, 6-H_b), 4.19 (1H, dd, J = 8.0, 8.1 Hz, 3-H), 4.24 (1H, dddd, J = 7.4, 7.5, 7.5, 7.6 Hz, 5-H), 5.04 (1H, br d, J = 12.6 Hz, OCH_aPh), 5.13 (1H, d, J = 12.3 Hz, OCH_bPh), 7.30–7.35 (5H, m, Ar), 9.56 (1H, br b, CHO); $\delta_{\rm C}$ (125.7 MHz, $CDCl_3$, 70 °C) -4.8 (2 × CH_3), 16.0 (CH_3), 18.0 (C), 20.2 (CH_3) , 25.8 (3 × CH_3), 36.3 (CH_2) , 50.2 (C), 54.6 (CH_2) , 60.7 (CH), 67.3 (CH₂), 69.7 (CH), 128.1 (2 × CH), 128.2 (CH), 128.5 (2 × CH), 136.7 (C), 155.7 (C), 203.2 (CH); m/z 334 (14, M^+ - Me₂CCHO), 290 (41, M^+ - Me₃CSi(Me)₂), 91 (100, $[PhCH_2]^+$). HRMS calcd for $C_{18}H_{28}NO_3Si$ 334.1838, found 334.1823; calcd for C₁₆H₂₀NO₄ 290.1392, found 290.1378; calcd for C₇H₇ 91.0548, found 91.0545. C₂₂H₃₅NO₄Si requires C, 65.15; H, 8.70; N, 3.45%. Found: C, 65.02; H, 8.82; N, 3.56.

Product 27. Syrup; $[\alpha]_D$ -37 (c 0.18 in CHCl₃); v_{max}/cm^{-1} 1722, 1694, 1469, 1415; $\delta_{\rm H}$ (500 MHz, CDCl₃, 70 °C) 0.04 (3H, s, SiMe_a), 0.06 (3H, s, SiMe_b), 0.86 (9H, s, tBu), 1.00 (6H, s, 2-Me₂), 1.86 (1H, ddd, J = 3.5, 7.9, 13 Hz, 4-H_a), 1.97 (1H, m, 4-H_b), 3.20 (1H, dd, J = 3.6, 11.9 Hz, 6-H_a), 3.77 (1H, m, 6- H_b), 4.33 (1H, m, 5-H), 4.38 (1H, br dd, J = 7.6, 7.9 Hz, 3-H), 5.06 (1H, m, OCH_aPh), 5.14 (1H, m, OCH_bPh), 7.28–7.34 (5H, m, Ar), 9.56 (1H, br b, CHO); $\delta_{\rm C}$ (125.7 MHz, CDCl₃, 70 °C) $-4.8 (2 \times CH_3), 15.9 (CH_3), 18.0 (C), 20.0 (CH_3), 25.7 (3 \times$ CH₃), 37.0 (CH₂), 50.0 (C), 56.8 (CH₂), 61.0 (CH), 67.3 (CH₂), 70.8 (CH), 128.0 (2 × CH), 128.1 (CH), 128.5 (2 × CH), 136.9 (C), 156.8 (C), 203.5 (CH); m/z 334 (5, M^+ – Me₂CCHO), 290 $(20, M^+ - Me_3CSi(Me)_2), 91 (100, [PhCH_2]^+)$. HRMS calcd for C₁₈H₂₈NO₃Si 334.1838, found 334.1850; calcd for C₁₆H₂₀NO₄ 290.1392, found 290.1400; calcd for C₇H₇ 91.0548, found 91.0548. C₂₂H₃₅NO₄Si requires C, 65.15; H, 8.70; N, 3.45%. Found: C, 65.45; H, 8.88; N, 3.67.

(2R,4R)-Benzyl 4-(tert-butyldimethylsilyloxy)-2-(2-oxoethyl) pyrrolidine-1-carboxylate (28) and (2S,4R)-benzyl 4-(tert-butyldimethylsilyloxy)-2-(2-oxoethyl)pyrrolidine-1-carboxylate (29). Obtained from hydroxyproline derivative 23 according to Method A, using vinyloxytrimethylsilane as the nucleophile. The reaction mixture was purified by rotatory chromatography (hexanes-EtOAc, 90:10), giving the 2,4-cis product 28 (34%) and the 2,4-*trans* isomer **29** (20%).

Product 28. Colorless oil; $\lceil \alpha \rceil_D$ +5 (c 0.63 in CHCl₃); v_{max} cm $^{-1}$ 1694, 1417; $\delta_{\rm H}$ (500 MHz, CDCl₃, 70 °C) note: the usual numbering (CHO as C-1) is used, not the one given by IUPAC nomenclature: 0.08 (6H, s, SiMe₂), 0.90 (9H, s, tBu), 1.78 (1H, br d, J = 13.2 Hz, 4-H_a), 2.24 (1H, ddd, J = 5.2, 8.4, 13.5 Hz, 4-H_b), 2.85 (1H, m, 2-H_a), 3.09 (1H, m, 2-H_b), 3.37 (1H, br d, $J = 12.6 \text{ Hz}, 6-\text{H}_a$), 3.64 (1H, m, 6-H_b), 4.37 (1H, m, 3-H), 4.39 (1H, m, 5-H), 5.13 (1H, d, J = 12 Hz, OCH_aPh), 5.16 (1H, d, J = 11.5 Hz, OCH_bPh), 7.30–7.36 (5H, m, Ar), 9.77 (1H, br b, CHO); $\delta_{\rm C}$ (100.6 MHz, CDCl₃, 25 °C) A mixture of rotamers was observed: -5.0 (CH₃), -4.9 (CH₃), 17.9 (C), 25.7 (3 × CH₃), 39.7/40.4 (CH₂), 49.2/49.9 (CH₂), 51.9/52.5 (CH), 55.2/ 55.7 (CH₂), 66.9/67.1 (CH₂), 70.6/71.3 (CH), 127.9 (2 × CH),

128.1 (CH), 128.5 (2 × CH), 136.5/136.7 (C), 154.5/154.8 (C), 201.1/201.2 (CH); m/z 320 (M⁺ – Me₃C, 9%), 292 (11, M⁺ – $Me_3C - CO)$, 276 (15, $M^+ - Me_3C - CH_2CHO)$, 91 (100, $[PhCH_2]^+$). HRMS calcd for $C_{16}H_{22}NO_4Si$ 320.1318, found 320.1320; calcd for C₁₅H₂₂NO₃Si 292.1369, found 292.1363; calcd for C₇H₇ 91.0548, found 91.0544. C₂₀H₃₁NO₄Si requires C, 63.63; H, 8.28; N, 3.71%. Found: C, 63.59; H, 8.15; N, 3.78.

Product 29. Colorless oil; $[\alpha]_D$ -35 (c 0.26 in CHCl₃); $v_{\text{max}}/\text{cm}^{-1}$ 1694, 1416, 1357; δ_{H} (500 MHz, CDCl₃, 70 °C) 0.06 (3H, s, SiMe_a), 0.07 (3H, s, SiMe_b), 0.88 (9H, s, tBu), 1.77 (1H, ddd, $J = 4.7, 7.3, 12.0 \text{ Hz}, 4-\text{H}_a$), 2.17 (1H, m, 4-H_b), 2.58 (1H, br dd, J = 6.9, 16.0 Hz, 2-H_a), 2.96 (1H, m, 2-H_b), 3.46 (1H, dd, J = 4.4, 11.4 Hz, 6-H_a), 3.50 (1H, m, 6-H_b), 4.37 (1H, m, 5-H), 4.41 (1H, m, 3-H), 5.12-5.18 (2H, m, OCH₂Ph), 7.29-7.35 (5H, m, Ar), 9.75 (1H, br b, CHO); $\delta_{\rm C}$ (100.6 MHz, CDCl₃, 25 °C) A mixture of rotamers was observed: -4.9 (CH₃), -4.8 (CH₃), 17.9 (C), 25.7 (3 \times CH₃), 40.9/41.8 (CH₂), 48.5/49.4 (CH₂), 51.3/51.9 (CH₂), 55.0/55.3 (CH), 66.8/67.0 (CH₂), 69.5/70.0 (CH), 127.8 (2 × CH), 128.0 (CH), 128.5 (2 × CH), 136.8 (C), 155.4 (C), 200.3/200.5 (CH); m/z 292 (M⁺ – Me₃C – CO, 33%), 91 (100, $[PhCH_2]^+$). HRMS calcd for $C_{15}H_{22}NO_3Si$ 292.1369, found 292.1364; calcd for C₇H₇ 91.0548, found 91.0544. C₂₀H₃₁NO₄Si requires C, 63.63; H, 8.28; N, 3.71%. Found: C, 63.61; H, 8.28; N, 3.70.

N-(N-Benzoyl-1-aminocyclohexanecarbonyl)-α,α-dimethyl-DLβ-homoleucinal (37). Obtained from the dipeptide 30 according to Method B, using 2-methyl-1-(trimethylsilyloxy)-1-propene as the nucleophile. The reaction mixture was purified by rotatory chromatography (hexanes-EtOAc, 70:30), giving product 37 (50%) as a syrup; $v_{\text{max}}/\text{cm}^{-1}$ 3433, 1724, 1662, 1515; $\delta_{\rm H}$ (500 MHz, CDCl₃) 0.86 (3H, d, J = 6.6 Hz, 5-Me), 0.88 (3H, d, J = 6.3 Hz, 5-Me), 1.02 (3H, s, 2-Me), 1.05 (3H, s, 2-Me), 1.15 (1H, ddd, J = 2.5, 11.8, 13.3 Hz, 4-H_a), 1.33 (1H, ddd, $J = 3.2, 11.7, 14.1 \text{ Hz}, 4-H_b$, 1.38 (1H, m, 5'-H_a), 1.46 (2H, m, $4'-H_a + 6'-H_a$), 1.56 (1H, m, 5-H), 1.66 (1H, m, 5'-H_b), 1.70 $(2H, m, 4'-H_b + 6'-H_b), 1.96 (2H, m, 3'-H_a + 7'-H_a), 2.25 (2H, m, 3'-H_a)$ dd, J = 13.2, 13.2 Hz, 3'-H_b + 7'-H_b), 4.23 (1H, ddd, J = 2.2, 9.8, 11.8 Hz, 3-H), 6.05 (1H, s, NH), 7.45 (2H, dd, J = 7.3, 7.9 Hz, Ar), 7.50 (1H, d, J = 7.6 Hz, NH), 7.53 (1H, dd, J = 7.3, 7.6 Hz, Ar), 7.73 (2H, d, J = 7.0 Hz, Ar), 9.46 (1H, s, CHO); $\delta_{\rm C}$ (125.7 MHz, CDCl₃) 18.0 (CH₃), 18.6 (CH₃), 21.4 (CH₃), 21.6 (CH₂), 21.7 (CH₂), 23.8 (CH₃), 25.0 (CH), 25.2 (CH₂), 32.2 (CH₂), 32.5 (CH₂), 39.4 (CH₂), 50.6 (CH), 50.8 (C), 61.1 (C), 126.8 (2 × CH), 128.8 (2 × CH), 131.9 (CH), 134.8 (C), 168.5 (C), 173.8 (C), 205.1 (CH); m/z 387 (M⁺ + H, <1%), 230 $(21, M^+ - NH-CH(CH_2CHMe_2)-C(Me)_2-CHO), 202 (73, M^+ - M^+)$ $CONH-CH(CH_2CHMe_2)-C(Me)_2-CHO)$, 105 (100, $[PhCO]^+$). HRMS calcd for C₂₃H₃₅N₂O₃ 387.2648, found 387.2641; calcd for C₁₄H₁₆NO₂ 230.1181, found 230.1183; calcd for C₁₃H₁₆NO 202.1232, found 202.1235; calcd for C₇H₅O 105.0340, found 105.0341. C₂₃H₃₄N₂O₃ requires C, 71.47; H, 8.87; N, 7.25%. Found: C, 71.36; H, 8.97; N, 6.93.

N-(N-Benzoyl-2-methylalanyl)-α,α-dimethyl-β-α,α-dimethylβ-homophenylalaninal (38). Obtained from the dipeptide 31 according to Method B, using 2-methyl-1-(trimethylsilyloxy)-1propene as the nucleophile. The reaction mixture was purified by chromatography (hexanes-EtOAc, 60:40), giving product 38 (61%) as a crystalline solid; Mp 138–139 °C (EtOAc–n-hexane); $v_{\text{max}}/\text{cm}^{-1}$ 3439, 3088, 1722, 1671, 1512, 1484; δ_{H} (500 MHz, CDCl₃) 1.19 (3H, s, 2-Me or 2'-Me), 1.21 (3H, s, 2-Me or 2'-Me), 1.37 (3H, s, 2-Me or 2'-Me), 1.41 (3H, s, 2-Me or 2'-Me), 2.59 (1H, dd, J = 11.7, 14.2 Hz, 4-H_a), 2.98 (1H, dd, J = 4.1, 14.2 Hz, 4-H_b), 4.48 (1H, ddd, J = 4.1, 9.8, 11.7 Hz, 3-H), 6.62 (1H, br s, NH), 7.07 (1H, br d, J = 9.8 Hz, NH), 7.13–7.20 (5H, m, Ar), 7.42 (2H, dd, J = 7.3, 7.9 Hz, Ar), 7.50 (1H, dd, J = 7.6, 7.8 Hz, Ar), 7.70 (2H, d, J = 6.9 Hz, Ar), 9.50 (1H, s, CHO); $\delta_{\rm C}$ (125.7 MHz, CDCl₃) 19.1 (CH₃), 19.5 (CH₃), 24.6 (CH₃), 25.3 (CH₃), 36.4 (CH₂), 50.5 (C), 54.2 (CH), 57.8 (C), 126.5 (CH), 126.9 (2 × CH), 128.3 (2 × CH), 128.6 (2 × CH), 129.0 (2 × CH), 131.6 (CH), 134.7 (C), 137.9 (C), 167.5 (C), 173.9 (C), 204.8 (C); m/z 381 (M⁺ + H, 5%), 309 (6, M⁺ - Me₂CCHO), 190 (64, M^+ – NHCH(CH₂Ph)-C(Me)₂-CHO), 162 (70, M^+ – CONHCH(CH₂Ph)-C(Me)₂-CHO), 105 (100, [PhCO]⁺). HRMS calcd for C₂₃H₂₉N₂O₃ 381.2178, found 381.2170; calcd for $C_{19}H_{21}N_2O_2$ 309.1603, found 309.1616; calcd for $C_{11}H_{12}NO_2$ 190.0868, found 190.0861; calcd for C₁₀H₁₂NO 162.0919, found 162.0925; calcd for C₇H₅O 105.0340, found 105.0343. C₂₃H₂₈N₂O₃ requires C, 72.61; H, 7.42; N, 7.36%. Found: C, 72.77; H, 7.68; N, 7.07.

N-(N-Benzoyl-L-phenylalanyl)-α,α-dimethyl-L-β-homoalaninal (39) and N-(N-benzoyl-L-phenylalanyl)-α,α-dimethyl-D-β-homoalaninal (40). Obtained from the dipeptide 32 according to Method B, but using DIB (0.4 mmol) and iodine (0.1 mmol) in the scission step, and 2-methyl-1-(trimethylsilyloxy)-1-propene as the nucleophile in the addition step. The reaction mixture was purified by chromatography (hexanes-EtOAc, 75:25), giving the diastereomeric products 39 (49%) and 40 (37%), in 86% overall yield.

Product 39. Crystalline solid; Mp 133-134 °C (EtOAc*n*-hexane); $[\alpha]_D - 8$ (c 0.24 in CHCl₃); v_{max}/cm^{-1} 3423, 3310, 1723, 1653, 1512; $\delta_{\rm H}$ (500 MHz, CDCl₃) 0.90 (3H, s, 2-Me_a), 0.94 (3H, s, 2-Me_b), 1.02 (3H, d, J = 6.9 Hz, 3-Me), 3.12 (1H, dd, J = 7.9, 13.9 Hz, 3'-H_a), 3.21 (1H, dd, J = 6.6, 13.9 Hz, 3'-H_b), 4.17 (1H, m, 3-H), 4.86 (1H, ddd, J = 6.6, 7.6, 7.9 Hz, 2-H), 6.43 (1H, d, J = 9.5 Hz, NH), 6.91 (1H, d, J = 7.6 Hz, NH), 7.21-7.30 (5H, m, Ar), 7.40 (2H, dd, J = 7.3, 7.9 Hz, Ar), 7.49 (1H, dd, J = 7.3, 7.6 Hz, Ar), 7.71 (2H, d, J = 7.3 Hz, Ar), 9.35 (1H, s, CHO); $\delta_{\rm C}$ (125.7 MHz, CDCl₃) 16.1 (CH₃), 18.4 (CH₃), 19.0 (CH₃), 38.3 (CH₂), 49.0 (CH), 49.6 (C), 55.2 (CH), 127.0 (2 × CH), 127.1 (CH), 128.6 (2 × CH), 128.8 (2 × CH), 129.3 (2 × CH), 131.8 (CH), 133.7 (C), 136.5 (C), 167.4 (C), 170.5 (C), 204.8 (CH); m/z 366 (M⁺, 1%), 224 (17, M⁺ - $CONHCH(Me)-C(Me)_2-CHO)$, 105 (100, $[PhCO]^+$). HRMS calcd for C₂₂H₂₆N₂O₃ 366.1943, found 366.1944; calcd for C₁₅H₁₄NO 224.1075, found 224.1074; calcd for C₇H₅O 105.0340, found 105.0338. C₂₂H₂₆N₂O₃ requires C, 72.11; H, 7.15; N, 7.64%. Found: C, 72.24; H, 7.35; N, 7.76.

Product **40**. Syrup; $[\alpha]_D$ -13 (*c* 0.21 in CHCl₃); v_{max}/cm^{-1} 3423, 3310, 1723, 1648, 1510; $\delta_{\rm H}$ (500 MHz, CDCl₃) 0.90 (3H, d, J = 6.9 Hz, 3-Me), 0.93 (3H, s, 2-Me_a), 0.95 (3H, s, 2-Me_b), 3.12 (1H, dd, J = 8.2, 13.6 Hz, $3'-H_a$), 3.24 (1H, dd, J = 6.3, 13.6, 3'-H_b), 4.15 (1H, m, 3-H), 4.88 (1H, ddd, J = 6.9, 7.9, 7.9 Hz, 2'-H), 6.59 (1H, br b, NH), 7.16 (1H, m, Ar), 7.20–7.30 (5H, m, Ar + NH), 7.39 (2H, dd, J = 6.8, 7.3 Hz, Ar), 7.49 (1H, Theorem 1)dd, J = 7.3, 7.6 Hz, Ar), 7.72 (2H, d, J = 8.1 Hz, Ar), 9.32 (1H,

br b, CHO); $\delta_{\rm C}$ (125.7 MHz, CDCl₃) 15.6 (CH₃), 18.0 (CH₃), 18.7 (CH₃), 38.3 (CH₂), 48.5 (CH), 49.6 (C), 55.2 (CH), 127.0 (CH), 127.1 (2 × CH), 128.5 (2 × CH), 128.7 (2 × CH), 129.3 (2 × CH), 131.8 (CH), 133.7 (C), 136.7 (C), 167.6 (C), 170.6 (C), 204.3 (CH); m/z 366 (M⁺, 5%), 224 (18, M⁺ – CONHCH(Me)-C(Me)₂-CHO), 105 (100, [PhCO]⁺). HRMS calcd for C₂₂H₂₆N₂O₃ 366.1943, found 366.1948; calcd for C₇H₅O 105.0340, found 105.0343. C₂₂H₂₆N₂O₃ requires C, 72.11; H, 7.15; N, 7.64%. Found: C, 72.23; H, 6.89; N, 7.77.

N-(N-Benzoyl-L-leucyl)-α,α-dimethyl-L-β-(41) N-(Nbenzovl-L-leucvl)-α,α-dimethyl-p-β-homoalaninal (42). The products were generated from dipeptide 33 according to Method B, using 2-methyl-1-(trimethylsilyloxy)-1-propene as the nucleophile. The reaction mixture was purified by rotatory chromatography (hexanes-EtOAc, 80:20), affording compounds 41 (57%) and 42 (29%).

Compound 41. Crystalline solid; Mp 184-185 °C (EtOAc*n*-hexane); $[\alpha]_D$ -22 (c 0.43 in CHCl₃); v_{max}/cm^{-1} 3428, 1723, 1656, 1517; $\delta_{\rm H}$ (500 MHz, CDCl₃) 0.95 (6H, d, J = 6.3 Hz, 4'-Me₂), 1.05 (3H, d, J = 8.2 Hz, 3-Me), 1.06 (3H, s, 2-Me_a), dddd, J = 6.8, 6.8, 6.8, 9.4 Hz, 3-H), 4.67 (1H, ddd, J = 7.5, 7.5, 7.5 Hz, 2'-H), 6.87 (1H, br b, NH), 6.94 (1H, d, J = 7.9 Hz, NH), 7.40 (2H, dd, J = 7.7, 7.7 Hz, Ar), 7.49 (1H, dd, J = 7.3, 7.6 Hz, Ar), 7.78 (2H, d, J = 7.5 Hz, Ar), 9.45 (1H, s, CHO); $\delta_{\rm C}$ (100.6 MHz, CDCl₃) 15.8 (CH₃), 17.9 (CH₃), 19.2 (CH₃), 22.2 (CH₃), 22.8 (CH₃), 25.0 (CH), 41.4 (CH₂), 48.6 (CH), 50.0 (C), 52.5 (CH), 127.1 (2 × CH), 128.6 (2 × CH), 131.8 (CH), 133.8 (C), 167.6 (C), 171.9 (C), 204.7 (CH); m/z 333 (M⁺ + H, 1%), 190 (100, M^+ – CONHCH(Me)CMe₂CHO), 105 (91, [PhCO]⁺); HRMS calcd for C₁₉H₂₉N₂O₃ 333.2178, found 333.2181; calcd for C₁₂H₁₆NO 190.1232, found 190.1239. C₁₉H₂₈N₂O₃ requires C, 68.65; H, 8.49; N, 8.43%. Found: C, 68.90; H, 8.35; N, 8.14.

Compound 42. Crystalline solid; Mp 152–153 °C (net); $[\alpha]_D$ -28 (c 0.60 in CHCl₃); $v_{\text{max}}/\text{cm}^{-1}$ 3428, 1721, 1656, 1517; δ_{H} $(500 \text{ MHz}, \text{CDCl}_3) \ 0.96 \ (3\text{H}, d, J = 6.5 \text{ Hz}, 4'-\text{Me}_a), \ 0.97 \ (3\text{H}, d)$ d, J = 6.3 Hz, 4'-Me_b), 1.00 (3H, s, 2-Me_a), 1.03 (3H, s, 2-Me_b), 1.13 (3H, d, J = 6.9 Hz, 3-Me), 1.63–1.82 (3H, m, 3'-H₂ + 4'-H), 4.24 (1H, dddd, J = 6.9, 6.9, 6.9, 9.5 Hz, 3-H), 4.61 (1H, ddd, J = 6.0, 6.3, 8.2 Hz, 2'-H), 6.67 (1H, d, <math>J = 8.2 Hz, NH),6.75 (1H, d, J = 9.8 Hz, NH), 7.43 (2H, dd, J = 7.4, 7.4 Hz, Ar), 7.50 (1H, dd, J = 7.3, 7.6 Hz, Ar), 7.78 (2H, d, J = 6.9 Hz, Ar), 9.43 (1H, s, CHO); δ_C (100.6 MHz, CDCl₃) 16.0 (CH₃), 17.9 (CH₃), 19.0 (CH₃), 22.3 (CH₃), 22.9 (CH₃), 25.0 (CH), 40.4 (CH₂), 48.4 (CH), 50.1 (C), 52.2 (CH), 127.1 (2 × CH), 128.6 (2 × CH), 131.8 (CH), 133.9 (C), 167.9 (C), 171.3 (C), 204.8 (CH); m/z 333 (M⁺ + H, 2%), 190 (86, M⁺ - CONHCH(Me) CMe_2CHO), 105 (100, $[PhCO]^+$); HRMS calcd $C_{19}H_{29}N_2O_3$ 333.2178, found 333.2177; calcd for C₇H₅O 105.0340, found 105.0340. C₁₉H₂₈N₂O₃ requires C, 68.65; H, 8.49; N, 8.43%. Found: C, 68.80; H, 8.32; N, 8.10.

N-(N-Benzoyl-L-prolyl)-α,α-dimethyl-L-β-homoleucinal (43) and N-(N-benzoyl-L-prolyl)-α,α-dimethyl-p-β-homoleucinal (44). The products were generated from dipeptide 34 according to Method B, using 2-methyl-1-(trimethylsilyloxy)-1-propene as the nucleophile. The reaction mixture was purified by rotatory

chromatography (hexanes-EtOAc, 80:20), affording compounds 43 (47%) and 44 (22%).

Compound 43. Syrup; $[\alpha]_D$ -85 (c 0.65 in CHCl₃); $v_{\text{max}}/\text{cm}^{-1}$ 3300, 1723, 1674, 1610, 1536; $\delta_{\rm H}$ (500 MHz, CDCl₃) 0.91 (3H, d, J = 6.6 Hz, 5-Me_a), 0.92 (3H, d, J = 6.6 Hz, 5-Me_b), 0.96 $(3H, s, 2-Me_a), 1.05 (3H, s, 2-Me_b), 1.18 (1H, ddd, J = 2.2, 9.8,$ 12.6 Hz, 4-H_a), 1.32 (1H, ddd, J = 3.5, 11.3, 13.0 Hz, 4-H_b), 1.65 (1H, m, 5-H), 1.81 (1H, m, 4'-H_a), 1.91–2.04 (2H, m, 4'-H_b) + 3'-H_a), 2.50 (1H, m, 3'-H_b), 3.37 (1H, m, 5'-H_a), 3.50 (1H, m, 5'-H_b), 4.26 (1H, dd, J = 10, 11.7 Hz, 3-H), 4.82 (1H, m, 2'-H), 7.26 (1H, d, J = 9.8 Hz, NH), 7.41 (3H, m, Ar), 7.54 (2H, m, Ar), 9.47 (1H, s, CHO); $\delta_{\rm C}$ (125.7 MHz, CDCl₃) 17.0 (CH₃), 18.9 (CH₃), 21.4 (CH₃), 23.8 (CH₃), 25.0 (CH), 25.2 (CH₂), 26.3 (CH₂), 38.6 (CH₂), 50.0 (CH₂), 50.5 (CH), 51.0 (C), 59.5 (CH), 126.8 (2 × CH), 128.5 (2 × CH), 130.1 (CH), 136.3 (C), 170.7 (C), 171.5 (C), 204. 7 (CH); *m/z* 358 (M⁺, 3%), 202 (50, M^{+} - NHCH(CH₂CHMe₂)-C(Me)₂-CHO), 174 (95, M^{+} -CONHCH(CH₂CHMe₂)-C(Me)₂-CHO), 105 (100, [PhCO]⁺). HRMS calcd for $C_{21}H_{30}N_2O_3$ 358.2256, found 358.2262; calcd for C₁₂H₁₂NO₂ 202.0868, found 202.0858; calcd for C₁₁H₁₂NO 174.0919, found 174.0921; calcd for C₇H₅O 105.0340, found 105.0341. C₂₁H₃₀N₂O₃ requires C, 70.36; H, 8.44; N, 7.81%. Found: C, 70.48; H, 8.42; N, 8.10.

Compound 44. Syrup; $[\alpha]_D$ –97 (c 0.34 in CHCl₃); $v_{\text{max}}/\text{cm}^{-1}$ 3424, 3303, 1725, 1674, 1660, 1602, 1420; $\delta_{\rm H}$ (500 MHz, $CDCl_3$) 0.83 (3H, d, J = 6.9 Hz, 5-Me_a), 0.90 (3H, d, J = 6.6Hz, 5-Me_b), 1.07 (3H, s, 2-Me_a), 1.08 (3H, s, 2-Me_b), 1.16 (1H, m, 4-H_a), 1.31 (1H, m, 4-H_b), 1.56 (1H, m, 5-H), 1.83 (1H, m, $4'-H_a$), 1.99–2.08 (2H, m, $4'-H_b + 3'-H_a$), 2.43 (1H, m, $3'-H_b$), 3.45 (1H, m, 5'-H_a), 3.53 (1H, m, 5'-H_b), 4.24 (1H, dd, J = 9.5, 9.8 Hz, 3-H), 4.76 (1H, dd, J = 4.8, 7.3 Hz, 2'-H), 7.11 (1H, br d, J = 9.8 Hz, NH), 7.40-7.50 (5H, m, Ar), 9.49 (1H, s, CHO); $\delta_{\rm C}$ (125.7 MHz, CDCl₃) 18.4 (CH₃), 18.7 (CH₃), 21.5 (CH₃), 23.8 (CH₃), 25.2 (CH), 25.4 (CH₂), 27.1 (CH₂), 39.6 (CH₂), 50.4 (CH₂), 50.5 (C), 51.0 (CH), 59.9 (CH), 126.9 (2 × CH), 128.5 (2 × CH), 130.3 (CH), 136.2 (C), 170.9 (C), 171.5 (C), 205.2 (CH); m/z 358 (M⁺, 2%), 202 (27, M⁺ – NHCH $(CH_2CHMe_2)-C(Me)_2-CHO)$, 174 (74, M^+ – CONHCH (CH₂CHMe₂)-C(Me)₂-CHO), 105 (100, [PhCO]⁺). HRMS calcd for C₂₁H₃₀N₂O₃ 358.2256, found 358.2251; calcd for $C_{12}H_{12}NO_2$ 202.0868, found 202.0863; calcd for $C_{11}H_{12}NO$ 174.0919, found 174.0925; calcd for C₇H₅O 105.0340, found 105.0336. C₂₁H₃₀N₂O₃ requires C, 70.36; H, 8.44; N, 7.81%. Found: C, 70.19; H, 8.43; N, 7.76.

N-(N-Benzoyl-L-alanyl)-α,α-dimethyl-DL-β-homoleucinal (45). The products were generated from dipeptide 35 according to Method B, using 2-methyl-1-(trimethylsilyloxy)-1-propene as the nucleophile. The reaction mixture was purified by rotatory chromatography (hexanes-EtOAc, 75:25), affording compound **45** (78%) as a 9:6 mixture of diastereomers; $v_{\text{max}}/\text{cm}^{-1}$ 3422, 3307, 1724, 1676, 1645, 1514; $\delta_{\rm H}$ (500 MHz, CDCl₃) major isomer: 0.90 (3H, d, J = 6.8 Hz, 5-Me_a), 0.91 (3H, d, J = 6.6Hz, 5-Me_b), 0.98 (3H, s, 2-Me_a), 1.01 (3H, s, 2-Me_b), 1.14 (1H, m, 4-H_a), 1.39 (1H, m, 4-H_b), 1.49 (3H, d, J = 6.9 Hz, 2'-Me), 1.61 (1H, m, 5-H), 4.24 (1H, m, 3-H), 4.81 (1H, m, 2'-H), 7.07 (1H, br d, J = 10.4 Hz, NH), 7.22 (1H, br d, J = 7.6 Hz, NH), 7.40 (2H, dd, J = 7.8, 7.8 Hz, Ar), 7.49 (1H, dd, J = 7.3, 7.5 Hz, Ar), 7.79 (2H, d, J = 8.1 Hz, Ar), 9.46 (1H, s, CHO); minor

isomer: 0.75 (3H, d, J = 6.7 Hz, 5-Me_a), 0.84 (3H, d, J = 6.5Hz, 5-Me_b), 1.08 (3H, s, 2-Me_a), 1.09 (3H, s, 2-Me_b), 1.12 (1H, m, 4-H_a), 1.34 (1H, m, 4-H_b), 1.50 (3H, d, J = 6.9 Hz, 2'-Me), 1.51 (1H, m, 5-H), 4.24 (1H, m, 3-H), 4.81 (1H, m, 2'-H), 7.04 (1H, br d, J = 10.4 Hz, NH), 7.24 (1H, br d, J = 8.2 Hz, NH), 7.40 (2H, dd, J = 7.8, 7.8 Hz, Ar), 7.49 (1H, dd, J = 7.3, 7.5 Hz, Ar), 7.76 (2H, d, J = 7.5 Hz, Ar), 9.51 (1H, s, CHO); $\delta_{\rm C}$ (125.7 MHz, CDCl₃) major isomer: 18.2 (CH₃), 18.4 (CH₃), 18.6 (CH₃), 21.2 (CH₃), 23.8 (CH₃), 25.1 (CH), 39.1 (CH₂), 49.4 (CH), 50.7 (C), 50.9 (CH), 127.1 (2 \times CH), 128.5 (2 \times CH), 131.8 (CH), 133.7 (C), 167.6 (C), 172.5 (C), 204.9 (CH); minor isomer: 18.0 (CH₃), 18.4 (CH₃), 19.1 (CH₃), 21.2 (CH₃), 23.6 (CH₃), 24.9 (CH), 38.8 (CH₂), 49.4 (CH), 50.7 (C), 50.9 (CH), 127.1 (2 × CH), 128.5 (2 × CH), 131.8 (CH), 133.8 (C), 167.4 (C), 172.5 (C), 204.8 (CH); m/z 333 (M⁺ + H, 1%), 176 $(31, M^+ - NHCH(CH_2CHMe_2)-C(Me)_2-CHO), 148 (55, M^+ - M^+)$ CONHCH(CH₂CHMe₂)-C(Me)₂-CHO), 105 (100, [PhCO]⁺). HRMS calcd for C₁₉H₂₉N₂O₃ 333.2178, found 333.2166; calcd for C₁₀H₁₀NO₂ 176.0712, found 176.0706; calcd for C₉H₁₀NO 148.0762, found 148.0763; calcd for C₇H₅O 105.0340, found 105.0345. C₁₉H₂₈N₂O₃ requires C, 68.65; H, 8.49; N, 8.43%. Found: C, 68.50; H, 8.61; N, 8.69.

N-(N-Benzyloxycarbonyl-L-valyl)-α,α-dimethyl-DL-β-homoleucinal (46). The products were generated from dipeptide 36 according to Method B, using 2-methyl-1-(trimethylsilyloxy)-1propene as the nucleophile. The reaction mixture was purified by rotatory chromatography (hexanes-EtOAc, 85:15), affording compound 46 (76%) as a 10:7 mixture of diastereomers: syrup; $v_{\text{max}}/\text{cm}^{-1}$ 3426, 1714, 1672, 1506, 1469; δ_{H} (500 MHz, $CDCl_3$) major isomer: 0.86–0.90 (9H, m, 5-Me₂ + 3'-Me_a), 0.95 $(3H, d, J = 6.6 Hz, 3'-Me_b), 1.01 (3H, s, 2-Me_a), 1.06 (3H, s, s, s)$ 2-Me_b), 1.16 (1H, m, 4-H_a), 1.32 (1H, m, 4-H_b), 1.52 (1H, m, 5-H), 2.14 (1H, m, 3'-H), 3.92 (1H, ddd, J = 8.8, 8.8, 8.9 Hz, 2'-H), 4.21 (1H, br dd, J = 12.0, 12.0 Hz, 3-H), 5.08 (1H, d, J = 12.0 Hz, OCH_aPh), 5.11 (1H, d, J = 12.5 Hz, OCH_bPh), 5.37 (1H, br b, NH), 6.11 (1H, br d, J = 9.8 Hz, NH), 7.30–7.35 (5H, m, Ar), 9.44 (1H, s, CHO); minor isomer: 0.85 (3H, d, $J = 6.6 \text{ Hz}, 5\text{-Me}_a$, $0.86-0.90 \text{ (6H, m, } 5\text{-Me}_b + 3'\text{-Me}_a$), 0.952-Me_b), 1.15 (1H, m, 4-H_a), 1.32 (1H, m, 4-H_b), 1.52 (1H, m, 5-H), 2.14 (1H, m, 3'-H), 3.92 (1H, ddd, J = 8.8, 8.8, 8.9 Hz, 2'-H), 4.21 (1H, br dd, J = 12.0, 12.0 Hz, 3-H), 5.09–5.15 (2H, m, OCH₂Ph), 5.38 (1H, br b, NH), 6.14 (1H, br d, J = 12.0 Hz, NH), 7.30-7.35 (5H, m, Ar), 9.45 (1H, s, CHO); major isomer: $\delta_{\rm C}$ (125.7 MHz, CDCl₃) 17.7 (CH₃), 18.7 (CH₃), 19.0 (CH₃), 19.4 (CH₃), 21.1 (CH₃), 23.8 (CH₃), 24.9 (CH), 30.3 (CH), 39.3 (CH₂), 50.3 (C), 51.0 (CH), 61.0 (CH), 67.1 (CH₂), 128.0 (2 × CH), 128.2 (CH), 128.5 (2 × CH), 136.2 (C), 156.5 (C), 171.2 (C), 205.0 (CH); minor isomer: $\delta_{\rm C}$ (125.7 MHz, CDCl₃) 17.7 (CH₃), 18.5 (CH₃), 18.7 (CH₃), 19.4 (CH₃), 21.2 (CH₃), 23.7 (CH₃), 25.0 (CH), 30.1 (CH), 39.4 (CH₂), 50.4 (C), 51.2 (CH), 61.0 (CH), 67.1 (CH₂), 128.0 (2 × CH), 128.2 (CH), 128.5 (2 × CH), 136.2 (C), 156.6 (C), 171.2 (C), 204.9 (CH); m/z 391 (M⁺ + H, <1%), 390 (M⁺, <1%), 319 (8, M⁺ - C(Me)₂-CHO), 234 $(12, M^+ - NHCH(CH_2CHMe_2)-C(Me)_2-CHO), 91 (100,$ $[PhCH_2]^+$). HRMS calcd for $C_{22}H_{34}N_2O_4$ 390.2518, found 390.2519; calcd for C₁₈H₂₇N₂O₃ 319.2022, found 319.2027; calcd for C₁₃H₁₆NO₃ 234.1130, found 234.1122; calcd for C₇H₇

91.0548, found 91.0552. C₂₂H₃₄N₂O₄ requires C, 67.66; H, 8.78; N, 7.17%. Found: C, 67.57; H, 8.93; N, 7.11.

General procedure for the reduction of the amino aldehydes to the amino alcohols

To a solution of the starting β-peptide aldehydes (0.4 mmol) in dry ⁱPrOH (12 mL) cooled at 0 °C, was added LiBH₄ (17 mg, 0.8 mmol, 2.0 equiv). The reaction mixture was stirred at 0 °C for 40 min. Then it was poured into saturated brine and extracted with EtOAc. The organic layer was dried over anhydrous sodium sulfate, filtered and evaporated under vacuum. The residue was purified by chromatography on silica gel (hexanes-EtOAc) giving the γ -amino alcohol derivatives.

N-(N-Benzoyl-L-phenylalanyl)-α,α-dimethyl-L-β-homoalaninol (47). Obtained by reduction of the peptide aldehyde 39 according to the General Procedure. The reaction mixture was purified by rotatory chromatography (hexanes–EtOAc, 70:30), affording compound 47 (78%) as a crystalline solid: Mp 187-188 °C (EtOAc-n-hexane); $[\alpha]_D$ -17 (c 0.27 in CHCl₃); v_{max}/cm^{-1} 3423, 1650, 1509, 1484; $\delta_{\rm H}$ (500 MHz, CDCl₃) 0.47 (3H, s, 2-Me_a), 0.90 (3H, s, 2-Me_b), 1.01 (3H, d, J = 6.9 Hz, 3-Me), 2.91 (1H, d, J = 11.7 Hz, 1-H_a), 3.10 (1H, d, J = 11.7 Hz, 1-H_b), 3.15 (1H, dd, J = 8.5, 13.9 Hz, 3'-H_a), 3.25 (1H, dd, J = 6.3, 13.9 Hz, 3'-H_b), 3.93 (1H, dddd, J = 6.8, 6.8, 6.9, 8.7 Hz, 3-H), 4.92 (1H, ddd, J = 6.3, 7.9, 8.2 Hz, 2'-H), 6.34 (1H, br d, J = 9.0 Hz, NH), 6.85 (1H, brd, J = 7.3 Hz, NH), 7.24 (1H, m, Ar), 7.29-7.34 (4H, m, Ar), 7.42 (2H, dd, J = 7.3, 7.9 Hz, Ar), 7.51 (1H, dd, J = 7.3, 7.6 Hz, Ar), 7.72 (2H, d, J = 8.2 Hz, Ar); $\delta_{\rm C}$ (125.7 MHz, CDCl₃) 15.3 (CH₃), 18.3 (CH₃), 23.0 (CH₃), 38.5 (CH₂ + C), 49.6 (CH), 55.2 (CH), 69.6 (CH₂), 127.0 (2 \times CH), 127.2 (CH), 128.6 (2 \times CH), 128.9 (2 \times CH), 129.3 (2 \times CH), 131.9 (CH), 133.6 (C), 136.4 (C), 167.4 (C), 171.3 (C); m/z 368 (M⁺, 1%), 350 (3, M⁺ – H₂O), 252 (26, M⁺ – NHCH $(Me)C(Me)_2CH_2OH)$, 224 (21, M^+ – CONHCH(Me)C-(Me)₂CH₂OH), 105 (100, [PhCO]⁺); HRMS calcd for C₂₂H₂₈N₂O₃ 368.2100, found 368.2107; calcd for C₂₂H₂₆N₂O₂ 350.1994, found 350.1985; calcd for C₁₆H₁₄NO₂ 252.1025, found 252.1019; calcd for C₁₅H₁₄NO 224.1075, found 224.1070; calcd for C₇H₅O 105.0340, found 105.0343. C₂₂H₂₈N₂O₃ requires C, 71.71; H, 7.66; N, 7.60%. Found: C, 71.79; H, 7.68; N, 7.68.

N-(N-Benzoyl-L-phenylalanyl)-α,α-dimethyl-D-β-homoalaninol (48). Obtained by reduction of the peptide aldehyde 40 according to the General Procedure. The reaction mixture was purified by rotatory chromatography (hexanes-EtOAc, 75:25), affording compound 48 (69%) as a crystalline solid: Mp 134-135 °C (EtOAc–*n*-hexane); $[\alpha]_D$ –11 (*c* 0.19 in CHCl₃); $v_{\text{max}}/\text{cm}^{-1}$ 3423, 1731, 1650, 1510; $\delta_{\rm H}$ (500 MHz, CDCl₃) 0.54 (3H, s, 2- Me_a), 0.86 (3H, d, J = 6.9 Hz, 3-Me), 0.91 (3H, s, 2-Me_b), 2.92 (1H, d, J = 11.7 Hz, 1-H_a), 3.14 (1H, dd, J = 7.9, 13.2 Hz, 3'- H_a), 3.21 (1H, dd, J = 6.6, 13.3 Hz, 3'- H_b), 3.25 (1H, d, J = 12Hz, 1-H_b), 3.87 (1H, dddd, J = 6.6, 6.6, 6.7, 9.1 Hz, 3-H), 4.02 (1H, OH), 4.99 (1H, ddd, J = 6.6, 8.2, 8.2 Hz, 2'-H), 6.95 (1H, br d, J = 9.5 Hz, NH), 7.19 (1H, m, Ar), 7.21–7.25 (4H, m, Ar), 7.35 (1H, brb, NH), 7.37 (2H, dd, J = 7.6, 7.8 Hz, Ar), 7.49 (1H, dd, J = 7.3, 7.6 Hz, Ar), 7.70 (2H, d, J = 8.2 Hz, Ar);

 $\delta_{\rm C}$ (100.6 MHz, CDCl₃) 14.8 (CH₃), 18.1 (CH₃), 22.9 (CH₃), 38.6 (C), 38.8 (CH₂), 49.0 (CH), 55.2 (CH), 69.7 (CH₂), 126.9 (CH), 127.1 (2 × CH), 128.5 (4 × CH), 129.4 (2 × CH), 131.8 (CH), 133.6 (C), 136.6 (C), 167.6 (C), 171.8 (C); m/z 368 (M⁺, 1%), 350 (1, M^+ – H_2O), 252 (58, M^+ – NHCH(Me)C- $(Me)_2CH_2OH)$, 224 (44, M^+ – CONHCH(Me)C(Me)_2CH_2OH), 105 (100, [PhCO]⁺); HRMS calcd for C₂₂H₂₈N₂O₃ 368.2100, found 368.2104; calcd for C₂₂H₂₆N₂O₂ 350.1994, found 350.1984; calcd for C₁₆H₁₄NO₂ 252.1025, found 252.1014; calcd for C₁₅H₁₄NO 224.1075, found 224.1072; calcd for C₇H₅O 105.0340, found 105.0341. C₂₂H₂₈N₂O₃ requires C, 71.71; H, 7.66; N, 7.60%. Found: C, 71.64; H, 7.48; N, 7.62.

N-(N-Benzovl-L-leucyl)-α,α-dimethyl-L-β-homoalaninol (49). Obtained by reduction of the peptide aldehyde 41 according to the General Procedure. The reaction mixture was purified by rotatory chromatography (hexanes-EtOAc, 60:40), affording compounds 49 (76%) as a crystalline solid; Mp 186-187 °C (EtOAc-n-hexane); $[\alpha]_D$ -35 (c 0.62 in CHCl₃); v_{max}/cm^{-1} 3427, 1651, 1516; $\delta_{\rm H}$ (500 MHz, CDCl₃) 0.69 (3H, s, 2-Me_a), 0.95 (6H, d, J = 6.6 Hz, $2 \times Me$), 0.96 (3H, s, 2-Me_b), 0.97 (3H, d, J = 6.3 Hz, Me), 1.71 (1H, m, 3'-H_a), 1.77 (1H, m, 4'-H), 1.82 (1H, m, 3'-H_b), 3.03 (1H, brdd, J = 9.1, 11.0 Hz, 1-H_a), 3.33 (1H, brd, J = 11.4 Hz, 1-H_b), 3.92 (1H, dddd, J = 6.6, 6.9, 6.9, 8.8 Hz, 3-H), 4.09 (1H, brb, OH), 4.89 (1H, ddd, J = 5.1, 5.4, 8.8 Hz, 2'-H), 7.34 (1H, dd, J = 7.3, 8.2 Hz, Ar), 7.46 (1H, dd, J = 7.3, 7.6 Hz, Ar), 7.44–7.50 (2H, brb, 2 × NH), 7.81 (1H, d, J = 8.0 Hz, Ar); δ_C (100.6 MHz, CDCl₃) 14.9 (CH₃), 18.4 (CH₃), 22.1 (CH₃), 22.9 (CH₃), 23.0 (CH₃), 25.0 (CH), 38.9 (C), 41.8 (CH₂), 49.0 (CH), 52.5 (CH), 69.8 (CH₂), 127.2 (2 × CH), 128.4 (2 × CH), 131.7 (CH), 133.8 (C), 167.6 (C), 173.4 (C); m/z 335 (M⁺ + H, <1%), 261 (5, M⁺ - C(Me)₂CH₂OH), 218 $(66, M^{+} - NHCH(Me)C(Me)_{2}CH_{2}OH), 190 (57, M^{+} - M^{+})$ $CONHCH(Me)C(Me)_2CH_2OH)$, 105 (100, $[PhCO]^+$); HRMS calcd for C₁₉H₃₁N₂O₃ 335.2335, found 335.2339; calcd for C₁₅H₂₁N₂O₂ 261.1603, found 261.1592; calcd for C₁₃H₁₆NO₂ 218.1181, found 218.1180; calcd for C₁₂H₁₆NO 190.1232, found 190.1227; calcd for C₇H₅O 105.0340, found 105.0340. C₁₉H₃₁N₂O₃ requires C, 68.23; H, 9.04; N, 8.38%. Found: C, 68.59; H, 8.95; N, 8.48.

N-(N-Benzoyl-L-leucyl)-α,α-dimethyl-p-β-homoalaninol (50). Obtained by reduction of the peptide aldehyde 42 according to the General Procedure. The reaction mixture was purified by rotatory chromatography (hexanes-EtOAc, 60:40), affording compounds 50 (79%) as a crystalline solid; Mp 163-164 °C (EtOAc–n-hexane); $[\alpha]_D$ –39 (c 0.66 in CHCl₃); v_{max}/cm^{-1} 3425, 1654, 1510; $\delta_{\rm H}$ (500 MHz, CDCl₃) 0.62 (3H, s, 2-Me_a), $0.95 \text{ (3H, d, } J = 6.9 \text{ Hz, Me)}, 0.96 \text{ (3H, s, 2-Me}_b), 0.97 \text{ (3H, d, me)}$ J = 6.9 Hz, Me), 1.12 (3H, d, J = 6.9 Hz, Me), 1.67–1.75 (2H, m, 3'-H_a + 4'-H), 1.77 (1H, m, 3'-H_b), 2.94 (1H, d, J = 11.7 Hz, $1-H_a$), 3.25 (1H, d, J = 11.9 Hz, $1-H_b$), 3.98 (1H, dddd, J = 6.6, 6.8, 6.9, 9.4 Hz, 3-H), 4.69 (1H, ddd, J = 6.0, 6.3, 7.9 Hz, 2'-H),6.95 (1H, brd, J = 8.2 Hz, NH), 7.03 (1H, br d, J = 9.5 Hz, NH), 7.41 (2H, dd, J = 7.3, 7.9 Hz, Ar), 7.50 (1H, dd, J = 7.3, 7.6 Hz, Ar), 7.76 (2H, d, J = 8.0 Hz, Ar); $\delta_{\rm C}$ (125.7 MHz, CDCl₃) 15.3 (CH₃), 18.3 (CH₃), 22.4 (CH₃), 22.8 (CH₃), 23.1 (CH₃), 25.0 (CH), 38.9 (C), 40.7 (CH₂), 49.1 (CH), 52.5 (CH), 69.8 (CH₂), 127.1 (2 × CH), 128.6 (2 × CH), 131.9 (CH), 133.8 (C), 167.9

(C), 172.7 (C); m/z 335 (M⁺ + H, 1%), 261 (9, M⁺ - C $(Me)_2CH_2OH)$, 218 (97, M^+ – NHCH(Me)C(Me)₂CH₂OH), 190 $(92, M^+ - CONHCH(Me)C(Me)_2CH_2OH), 105 (100, [PhCO]^+);$ HRMS calcd for C₁₉H₃₁N₂O₃ 335.2335, found 335.2343; calcd for C₁₅H₂₀N₂O₂ 260.1525, found 260.1515; calcd for C₁₃H₁₆NO₂ 218.1181, found 218.1177; calcd for C₁₂H₁₆NO 190.1232, found 190.1229; calcd for C₇H₅O 105.0340, found 105.0342. C₁₉H₃₁N₂O₃ requires C, 68.23; H, 9.04; N, 8.38%. Found: C, 68.06; H, 8.88; N, 8.41.

N-(N-Benzoyl-L-prolyl)-α,α-dimethyl-L-β-homoleucinol Obtained by reduction of the peptide aldehyde 43 according to the General Procedure. The reaction mixture was purified by rotatory chromatography (hexanes-EtOAc, 1:1), affording compound 51 (77%) as a syrup; $[\alpha]_D$ -92 (c 0.41 in CHCl₃); v_{max} cm⁻¹ 3426, 1656, 1613; $\delta_{\rm H}$ (500 MHz, CDCl₃) 0.64 (3H, s, 2- Me_a), 0.89 (3H, d, J = 6.6 Hz, 5- Me_a), 0.94 (3H, d, J = 6.6 Hz, 5-Me_b), 1.00 (3H, s, 2-Me_b), 1.28-1.33 (2H, m, 4-H₂), 1.64 (1H, m, 5-H), 1.84 (1H, m, 4'-H_a), 2.03 (1H, m, 3'-H_a), 2.08 (1H, m, 4'- H_b), 2.48 (1H, m, 3'- H_b), 2.93 (1H, brb, OH), 3.01 (1H, d, J =11.7 Hz, 1-H_a), 3.30 (1H, d, J = 11.9 Hz, 1-H_b), 3.46 (1H, m, 5'- H_a), 3.55 (1H, m, 5'- H_b), 3.92 (1H, ddd, J = 3.5, 10.2, 10.7 Hz, 3-H), 4.80 (1H, m, 2'-H), 7.02 (1H, brd, J = 9.5 Hz, NH), 7.40–7.48 (5H, m, Ar); $\delta_{\rm C}$ (125.7 MHz, CDCl₃) 18.3 (CH₃), 21.3 (CH₃), 23.1 (CH₃), 23.9 (CH₃), 25.4 (CH), 25.5 (CH₂), 27.0 (CH₂), 38.1 (CH₂), 39.0 (C), 50.3 (CH₂), 51.2 (CH), 59.7 (CH), 70.3 (CH₂), 126.8 (2 × CH), 128.5 (2 × CH), 130.3 (CH), 136.0 (C), 171.2 (C), 172.4 (C); m/z 360 (M⁺, 1%), 287 (73, M⁺ - C(Me)₂CH₂OH), 202 (51, M⁺ - NHCH(CH₂CHMe₂)C- $(Me)_2CH_2OH)$, 174 (54, M^+ – $CONHCH(CH_2CHMe_2)C$ - $(Me)_2CH_2OH)$, 105 (100, $[PhCO]^+$); HRMS calcd for C₂₁H₃₂N₂O₃ 360.2413, found 360.2404; calcd for C₁₇H₂₃N₂O₂ 287.1760, found 287.1752; calcd for C₁₂H₁₂NO₂ 202.0868, found 202.0864; calcd for C₁₁H₁₂NO 174.0919, found 174.0918; calcd for C₇H₅O 105.0340, found 105.0345. C₂₁H₃₂N₂O₃ requires C, 69.97; H, 8.95; N, 7.77%. Found: C, 70.13; H, 9.01; N, 7.83.

N-(N-Benzoyl-L-prolyl)-α,α-dimethyl-p-β-homoleucinol (52). Obtained by reduction of the peptide aldehyde 44 according to the General Procedure. The reaction mixture was purified by rotatory chromatography (hexanes-EtOAc, 1:1), affording compound **51** (79%) as a syrup; $[\alpha]_D$ –130 (c 0.13 in CHCl₃); v_{max} cm⁻¹ 3449, 1656, 1615; $\delta_{\rm H}$ (500 MHz, CDCl₃) 0.68 (3H, s, 2- Me_a), 0.82 (3H, d, J = 6.6 Hz, 5- Me_a), 0.87 (3H, d, J = 6.6 Hz, 5-Me_b), 1.02 (3H, s, 2-Me_b), 1.26–1.32 (2H, m, 4-H₂), 1.52 (1H, m, 5-H), 1.85 (1H, m, 4'-H_a), 1.99–2.09 (2H, m, 3'-H_a + 4'-H_b), 2.50 (1H, m, 3'-H_b), 3.02 (1H, d, J = 11.7 Hz, 1-H_a), 3.38 (1H, d, J = 11.6 Hz, 1-H_b), 3.47 (1H, m, 5'-H_a), 3.53 (1H, m, 5'-H_b), 3.89 (1H, ddd, J = 4.4, 9.4, 9.5 Hz, 3-H), 4.88 (1H, dd, J = 4.4, 7.3 Hz, 2'-H), 7.12 (1H, brd, J = 9.5 Hz, NH), 7.41–7.48 (5H, m, Ar); $\delta_{\rm C}$ (125.7 MHz, CDCl₃) 18.4 (CH₃), 21.3 (CH₃), 23.0 (CH₃), 23.8 (CH₃), 25.4 (CH₂), 25.5 (CH), 26.9 (CH₂), 38.3 (CH₂), 38.9 (C), 50.4 (CH₂), 51.5 (CH), 59.7 (CH), 70.3 (CH₂), 126.9 (2 × CH), 128.5 (2 × CH), 130.4 (CH), 136.0 (C), 171.7 (C), 172.3 (C); m/z 360 (M⁺, 1%), 287 (7, M⁺ - C- $(Me)_2CH_2OH)$, 202 (36, M^+ – NHCH(CH₂CHMe₂)C- $(Me)_2CH_2OH)$, 174 (55, M^+ – $CONHCH(CH_2CHMe_2)C$ - $(Me)_2CH_2OH)$, 105 $(100, [PhCO]^+)$; HRMS calcd for C₂₁H₃₂N₂O₃ 360.2413, found 360.2415; calcd for C₁₂H₁₂NO₂ 202.0868, found 202.0875; calcd for C₁₁H₁₂NO 174.0919, found 174.0924; calcd for C₇H₅O 105.0340, found 105.0344. C₂₁H₃₂N₂O₃ requires C, 69.97; H, 8.95; N, 7.77%. Found: C, 69.76; H, 8.87; N, 7.68.

N-(N-Benzoyl-L-alanyl)-α,α-dimethyl-L-β-homoleucinol (53)and N-(N-benzovl-L-alanyl)-α.α-dimethyl-p-β-homoleucinol (54). Obtained by reduction of the peptide aldehyde 45 according to the General Procedure. The reaction mixture was purified by rotatory chromatography (hexanes-EtOAc, 60:40), affording compounds 53 (41%) and 54 (29%).

Product 53. Syrup; $[\alpha]_D$ +1 (c 0.29 in CHCl₃); $v_{\text{max}}/\text{cm}^{-1}$ 3421, 1653, 1647, 1512; $\delta_{\rm H}$ (500 MHz, CDCl₃) 0.59 (3H, s, 2- Me_a), 0.87 (3H, d, J = 6.3 Hz, 5- Me_a), 0.90 (3H, d, J = 6.6 Hz, 5-Me_b), 0.98 (3H, s, 2-Me_b), 1.26 (1H, ddd, J = 2.5, 10.5, 13.3 Hz, 4-H_a), 1.41 (1H, ddd, J = 3.5, 11.3, 13.9 Hz, 4-H_b), 1.51 (3H, d, J = 6.9 Hz, 2'-Me), 1.62 (1H, m, 5-H), 2.92 (1H, d, J =11.7 Hz, 1-H_a), 3.26 (1H, d, J = 11.7 Hz, 1-H_b), 3.90 (1H, ddd, J = 2.5, 9.5, 12.0 Hz, 3-H), 4.88 (1H, dddd, J = 6.9, 6.9, 7.3, 7.3Hz, 2'-H), 7.28 (1H, d, J = 9.5 Hz, NH), 7.38 (1H, d, J = 7.6Hz, NH), 7.41 (2H, dd, J = 7.3, 7.9 Hz, Ar), 7.51 (1H, dd, J =7.3, 7.6 Hz, Ar), 7.79 (2H, d, J = 7.8 Hz, Ar); $\delta_{\rm C}$ (125.7 MHz, CDCl₃) 18.3 (CH₃), 18.6 (CH₃), 21.0 (CH₃), 23.0 (CH₃), 23.9 (CH₃), 25.4 (CH), 37.8 (CH₂), 38.9 (C), 49.4 (CH), 51.4 (CH), 70.1 (CH₂), 127.1 (2 × CH), 128.6 (2 × CH), 131.9 (CH), 133.5 (C), 167.6 (C), 174.0 (C); m/z 335 (M⁺ + H, <1%), 316 (2, M⁺ - H₂O), 176 (34, [BzNH-CH(Me)CO), 148 (45, [BzNH=CH $(Me)^+$, 105 (96, $[PhCO]^+$), 86 (100, $[NH_2=CHCH_2CH$ $(Me)_2$]⁺), 77 (32, [Ph]⁺); HRMS calcd for $C_{19}H_{31}N_2O_3$ 335.2335, found 335.2351; calcd for C₁₉H₂₈N₂O₂ 316.2151, found 316.2141; calcd for C₁₀H₁₀NO₂ 176.0712, found 176.0707; calcd for C₉H₁₀NO 148.0762, found 148.0760; calcd for C₇H₅O 105.0340, found 105.0338; calcd for C₅H₁₂N 86.0970, found 86.0973; calcd for C₆H₅ 77.0391, found 77.0392. C₁₉H₃₀N₂O₃ requires C, 68.23; H, 9.04; N, 8.38%. Found: C, 68.44; H, 8.96; N, 8.30.

Product 54. Crystalline solid; Mp 158–159 °C (EtOAc*n*-hexane); $[\alpha]_D$ -40 (c 0.36 in CHCl₃); v_{max}/cm^{-1} 3421, 1655, 1649, 1512, 1484; $\delta_{\rm H}$ (500 MHz, CDCl₃) 0.71 (3H, s, 2-Me_a), 0.76 (3H, d, J = 6.6 Hz, 5-Me_a), 0.79 (3H, d, J = 6.6 Hz, 5-Me_b), 1.02 (3H, s, 2-Me_b), 1.25 (1H, m, 4-H_a), 1.31 (1H, ddd, J = 3.5, 11.0, 14.5 Hz, 4-H_b), 1.48 (1H, m, 5-H), 1.55 (3H, d, J = 6.9 Hz, 2'-Me), 3.05 (1H, d, J = 11.7 Hz, 1-H_a), 3.35 (1H, d, $J = 11.9 \text{ Hz}, 1\text{-H}_b$, 3.90 (1H, ddd, J = 2.5, 9.5, 11.7 Hz, 3-H), 4.88 (1H, dddd, J = 6.9, 6.9, 7.3, 7.3 Hz, 2'-H), 7.01 (1H, br d, J = 9.1 Hz, NH), 7.13 (1H, br d, J = 7.3 Hz, NH), 7.43 (2H, dd, J = 7.6, 7.8 Hz, Ar), 7.52 (1H, dd, J = 7.3, 7.6 Hz, Ar), 7.78 (2H, d, J = 8.2 Hz, Ar); $\delta_{\rm C}$ (125.7 MHz, CDCl₃) 18.6 (CH₃), 19.1 (CH₃), 21.1 (CH₃), 23.0 (CH₃), 23.7 (CH₃), 25.2 (CH), 37.9 (CH₂), 39.0 (C), 49.3 (CH), 51.5 (CH), 70.2 (CH₂), 127.0 (2 × CH), 128.6 (2 × CH), 131.9 (CH), 133.6 (C), 167.4 (C), 173.7 (C); m/z 335 (M⁺ + H, <1%), 316 (1, M⁺ - H₂O), 176 (23, [PhCONH-CH(Me)CO]), 148 (37, [PhCONH=CH(Me)]⁺), 105 (100, $[PhCO]^+$), 86 (100, $[NH_2=CHCH_2CH(Me)_2]^+$), 77 $(31, [Ph]^+)$; HRMS calcd for $C_{19}H_{31}N_2O_3$ 335.2335, found 335.2344; calcd for $C_{19}H_{28}N_2O_2$ 316.2151, found 316.2156; calcd for $C_{10}H_{10}NO_2$ 176.0712, found 176.0711; calcd for C₉H₁₀NO 148.0762, found 148.0760; calcd for C₇H₅O

105.0340, found 105.0336; calcd for C₅H₁₂N 86.0970, found 86.0968; calcd for C_6H_5 77.0391, found 77.0391. $C_{19}H_{30}N_2O_3$ requires C, 68.23; H, 9.04; N, 8.38%. Found: C, 68.58; H, 9.10; N, 8.22.

N-(N-Benzovloxycarbonyl-L-valyl)-α,α-dimethyl-L-β-homoleucinol (55) and N-(N-benzoyloxycarbonyl-L-valyl)-α,α-dimethyl-D-β-homoleucinol (56). Obtained by reduction of the peptide aldehyde 46 according to the General Procedure. The reaction mixture was purified by rotatory chromatography (hexanes-EtOAc, 85:15), affording compounds 55 (46%) and 56 (32%).

Product 55. Amorphous solid; $[\alpha]_D$ +2 (c 0.21 in CHCl₃); $v_{\text{max}}/\text{cm}^{-1}$ 3425, 1718, 1653, 1506; δ_{H} (500 MHz, CDCl₃) 0.61 (3H, s, 2-Me_a), 0.86 (3H, d, J = 6.3 Hz, 5-Me_a), 0.91 (3H, d,J = 6.6 Hz, 5-Me_b), 0.94 (3H, d, J = 6.9 Hz, 3'-Me_a), 0.97 (3H, d, J = 6.9 Hz, 3'-H_b), 0.98 (3H, s, 2-Me_b), 1.24–1.30 (2H, m, 4-H₂), 1.55 (1H, m, 5-H), 2.11 (1H, m, 3'-H), 2.95 (1H, d, $J = 11.9 \text{ Hz}, 1-H_a$), 3.23 (1H, d, $J = 11.7 \text{ Hz}, 1-H_b$), 3.87–3.95 (2H, m, 3-H + 2'-H), 5.07 (1H, d, J = 12.0 Hz, OCH_aPh), 5.12 $(1H, d, J = 12.3 \text{ Hz}, OCH_bPh), 5.40 (1H, d, J = 8.8 \text{ Hz}, NH),$ 6.10 (1H, d, J = 9.5 Hz, NH), 7.31–7.37 (5H, m, Ar); $\delta_{\rm C}$ (125.7 MHz, CDCl₃) 17.9 (CH₃), 18.5 (CH₃), 19.4 (CH₃), 21.0 (CH₃), 23.1 (CH₃), 24.0 (CH₃), 25.3 (CH), 30.2 (CH), 38.2 (CH₂), 38.9 (C), 51.5 (CH), 61.1 (CH), 67.1 (CH₂), 70.0 (CH₂), 128.0 (2 × CH), 128.2 (CH), 128.6 (2 × CH), 136.1 (C), 156.6 (C), 172.8 (C); m/z 393 (M⁺ + H, 1%), 319 (9, M⁺ - C(Me)₂-CHO), 234 (14, M⁺ - NHCH(CH₂CHMe₂)-C(Me)₂-CH₂OH), 108 (18, [PhCH₂OH]⁺), 91 (100, [PhCH₂]⁺), 86 (29, $[NH_2=CHCH_2CH(Me)_2]^+$). HRMS calcd for $C_{22}H_{37}N_2O_4$ 393.2753, found 393.2749; calcd for C₁₈H₂₇N₂O₃ 319.2022, found 319.2015; calcd for C₁₃H₁₆NO₃ 234.1130, found 234.1130; calcd for C₇H₈O 108.0575, found 108.0574; calcd for C₇H₇ 91.0548, found 91.0549; calcd for C₅H₁₂N 86.0970, found 86.0967. C₂₂H₃₆N₂O₄ requires C, 67.32; H, 9.24; N, 7.14%. Found: C, 67.34; H, 8.98; N, 7.29.

Product **56**. Amorphous solid; $[\alpha]_D$ -30 (c 0.26 in CHCl₃); $v_{\text{max}}/\text{cm}^{-1}$ 3422, 1715, 1653, 1520, 1506; δ_{H} (500 MHz, $CDCl_3$) 0.63 (3H, s, 2-Me_a), 0.83 (3H, d, J = 6.6 Hz, 5-Me_a), 0.89 (3H, d, J = 6.6 Hz, 5-Me_b), 0.92 (3H, d, J = 6.6 Hz, 3'-Me_a), 0.99 (3H, d, J = 6.9 Hz, 3'-H_b), 1.00 (3H, s, 2-Me_b), 1.23–1.30 (2H, m, 4-H₂), 1.50 (1H, m, 5-H), 2.21 (1H, m, 3'-H), 3.01 (1H, br d, J = 11.5 Hz, 1-H_a), 3.34 (1H, br d, J = 12.0 Hz, 1-H_b), 3.89 (1H, m, 3-H), 3.96 (1H, m, 2'-H), 5.08 (1H, d, J = 12.3 Hz, OCH_aPh), 5.13 (1H, d, J = 12.3 Hz, OCH_bPh), 5.27 (1H, br b, NH), 6.03 (1H, br d, J = 8.8 Hz, NH), 7.32–7.38 (5H, m, Ar); $\delta_{\rm C}$ (125.7 MHz, CDCl₃) 17.7 (CH₃), 18.7 (CH₃), 19.5 (CH₃), 21.1 (CH₃), 23.1 (CH₃), 23.9 (CH₃), 25.2 (CH), 30.0 (CH), 38.3 (CH₂), 38.8 (C), 51.6 (CH), 61.0 (CH), 67.2 (CH₂), 70.0 (CH₂), 128.1 (2 \times CH), 128.3 (CH), 128.6 (2 \times CH), 136.0 (C), 156.5 (C), 172.4 (C); m/z 393 (M⁺ + H, 1%), 319 (12, M^+ – C(Me)₂-CHO), 234 (20, M^+ – NHCH $(CH_2CHMe_2)-C(Me)_2-CH_2OH)$, 91 (100, $[PhCH_2]^+$), 86 (21, $[NH_2=CHCH_2CH(Me)_2]^+$). HRMS calcd for $C_{22}H_{37}N_2O_4$ 393.2753, found 393.2743; calcd for C₁₃H₁₆NO₃ 234.1130, found 234.1138; calcd for C₇H₇ 91.0548, found 91.0544; calcd for C₅H₁₂N 86.0970, found 86.0967. C₂₂H₃₆N₂O₄ requires C, 67.32; H, 9.24; N, 7.14%. Found: C, 67.37; H, 9.13; N, 7.18.

General procedure for the reduction of the \beta-amino esters to the **β**-amino aldehydes

A solution of the β-amino ester (0.2 mmol) in methanol (8 mL), was cooled to -78 °C and DIBAL-H was added (1 M solution in dichloromethane, 2 mL, 2 mmol). The reaction mixture was stirred for 6 h; then the solution was poured into a saturated solution of Rochelle's salt and extracted with CH₂Cl₂. The organic layer was dried on anhydrous sodium sulfate, filtered and evaporated under vacuum. The residue was purified by column chromatography (hexanes–EtOAc) to give the β-amino aldehydes.

General procedure for the reduction of the \beta-amino esters to the γ-amino alcohols

A solution of the β-amino ester (0.2 mmol) in methanol (8 mL), was cooled to 0 °C; then DIBAL-H was added (1 M solution in dichloromethane, 2 mL, 2 mmol; except in the case of substrate 57, where 0.6 mmol of the reducing agent were used). The reaction mixture was stirred for 3 h at 0 °C, and then was extracted, evaporated and purified as before to give the γ -amino alcohols.

Acknowledgements

This work was supported by the Research Projects CTQ2006-14260/PPQ and CTQ2009-07109 of the Plan Nacional de Investigación Científica, Desarrollo e Innovación Tecnológica, Ministerio de Educación y Ciencia (MEC) and Ministerio de Ciencia e Innovación (MICINN), Spain. We also acknowledge financial support from FEDER funds. C.J.S. thanks the Gobierno de Canarias for a predoctoral fellowship.

Notes and references

- 1 For a review on the subject, see: A. Moulin, J. Martinez and J. A. Fehrentz, J. Pept. Sci., 2007, 13, 1-15.
- 2 For peptidyl α -aldehydes, see: (a) D. Balamurugan and K. M. Muraleedharan, Tetrahedron, 2009, 65, 10074–10082; (b) M. A. Jones, J. D. Morton, J. M. Coxon, S. B. McNabb, H. Y.-Y. Lee, S. G. Aitken, J. M. Mehrtens, L. J. G. Robertson, A. T. Neffe, S. Miyamoto, R. Bickerstaffe, K. Gately, J. M. Wood and A. D. Abell, Bioorg. Med. Chem., 2008, 16, 6911-6923; (c) A. D. Abell, M. A. Jones, A. T. Neffe, S. G. Aitken, T. P. Cain, R. J. Payne, S. B. McNabb, J. M. Coxon, B. G. Stuart, D. Pearson, H. Y.-Y. Lee and J. D. Morton, Med. Chem., 2007, 50, 2916-2920; (d) E. Isabel, R. Aspiotis, C. Black, J. Colucci, R. Fortin, A. Giroux, E. L. Grimm, Y. Han, C. Mellon, D. W. Nicholson, D. M. Rasper, J. Renaud, S. Roy, J. Tam, P. Tawa, J. P. Vaillancourt, S. Xanthoudakis and R. J. Zamboni, Bioorg. Med. Chem. Lett., 2007, 17, 1671-1674; (e) S. I. Al-Gharabli, S. T. A. Shah, S. Weik, M. F. Schmidt, J. R. Mesters, D. Kuhn, G. Klebe, R. Hilgenfeld and J. Rademann, ChemBioChem, 2006, 7, 1048–1055; (f) R. J. Payne, K. M. Brown, J. M. Coxon, J. D. Morton, H. Y.-Y. Lee and A. D. Abell, Aust. J. Chem., 2004, 57, 877-884; (g) N. J. Ede, S. N. Eagle, G. Wickham, A. M. Bray, B. Warne, K. Shoemaker and S. Rosenberg, J. Pept. Sci., 2000, 6, 11-18; (h) G. W. Bemis, J. M. C. Golec, D. J. Lauffer, M. D. Mullican and M. A. Murcko, Patent US6025147, 1995.
- 3 For bioactive peptidyl β-aldehydes, see: (a) B. Yin, R. Dhal, V. Maisonneuve and G. Dujardin, Eur. J. Org. Chem., 2006, 3309-3313; (b) A. Richard and L. Bourel-Bonnet, Chem.-Eur. J., 2005, 11, 7315-7321; (c) Y. Choe, L. S. Brinen, M. S. Price, J. C. Engel, M. Lange, C. Grisostomi, S. G. Weston, P. V. Pallai, H. Cheng, L. W. Hardy, D. S. Hartsough, M. McMakin, R. F. Tilton, C. M. Baldino and C. S. Craik, Bioorg. Med. Chem., 2005, 13, 2141-2156; (d) J. Nkemgu-Njinkeng,

- V. Rosenkranz, M. Wink and D. Steverding, Antimicrob. Agents Chemother., 2002, 46, 2038-2040; (e) J. F. Lynas, P. Harriott, A. Healy and M. A. McKervey, Bioorg. Med. Chem. Lett., 1998, 8, 373-378; (f) M. Iqbal, P. A. Messina, B. Freed, M. Das, S. Chatterjee, R. Tripathy, M. Tao, K. A. Josef, B. Dembofsky, D. Dunn, E. Griffith, R. Siman, S. E. Senadhi, W. Biazzo, D. Bozyczko-Coyne, S. L. Meyer, M. A. Ator and R. Bihovsky, Bioorg. Med. Chem. Lett., 1997, 7, 539-544.
- 4 For the synthesis and use of α -aminoglyoxals, see: (a) A. El-Dahshan, S. Weik and J. Rademann, Org. Lett., 2007, 9, 949-952; (b) M. Groarke, M. A. McKervey and M. Nieuwenhuyzen, Tetrahedron Lett., 2000, 41, 1275-1278; (c) P. Darkins, M. Groarke, M. A. McKervey, H. M. Moncrieff, N. McCarthy and M. Nieuwenhuyzen, J. Chem. Soc., Perkin Trans. 1, 2000, 381-389; (d) J. F. Lynas, S. L. Martin and B. Walker, J. Pharm. Pharmacol., 2001, 53, 473–480.
- 5 (a) M. F. Schmidt, A. El-Dahshan, S. Keller and J. Rademann, Angew. Chem., Int. Ed., 2009, 48, 6346-6349; (b) J. F. Lynas, S. J. Hawthorne and B. Walker, Bioorg. Med. Chem. Lett., 2000, 10, 1771-1773.
- 6 (a) S. Bajusz, I. Véghelyi, K. Németh, E. Barabás, A. Juhász, J. Langó, E. Lavich, Z. Mohai, I. Moravcsik, Z. Taschler, G. Tóth, M. Patthy, J. Salát, A. Szeker and K. Makk, Patent WO9849189, 1998; (b) S. Bajusz, I. Fauszt, K. Németh, E. Barabás, A. Juhász and M. Patthy, Bioorg. Med. Chem. Lett., 1998, 8, 1477-1482; (c) H. Yang and R. G. Carter, J. Org. Chem., 2009, 74, 2246-2249; (d) M. Terada and Y. Toda, J. Am. Chem. Soc., 2009, 131, 6354-6355; (e) H. Zhang, Y. Chuan, Z. Li and Y. Peng, Adv. Synth. Catal., 2009, 351, 2288–2294; (f) F. A. Davis and M. Song, Org. Lett., 2007, 9, 2413-2416; (g) A. Labonne, L. Zani, L. Hintermann and C. Bolm, J. Org. Chem., 2007, 72, 5704-5708; (h) Y. Chi, E. P. English, W. C. Pomerantz, W. S. Horne, L. A. Joyce, L. R. Alexander, W. S. Fleming, E. A. Hopkins and S. Gellman, J. Am. Chem. Soc., 2007, 129, 6050-6055; (i) J. W. Yang, M. Stadler and B. List, Angew. Chem., Int. Ed., 2007, 46, 609–611; (j) T. Kano, Y. Yamaguchi, O. Tokuda and K. Maruoka, J. Am.Chem. Soc., 2005, 127, 16408-16409; (k) P. Gizecki, R. Dhal, C. Poulard, P. Gosselin and P. Dujardin, J. Org. Chem., 2003, 68, 4338-4344; (1) A. Mann, L. Quaranta, G. Reginato and M. Taddei, Tetrahedron Lett., 1996, 37, 2651-2654.
- 7 For the addition of nucleophiles to iminium ions, see: (a) A. Yazici and S. G. Pyne, Synthesis, 2009, 339-368 (part I); A. Yazici and S. G. Pyne, Synthesis, 2009, 513-541 (part 2) (b) D. Ferraris, Tetrahedron, 2007, 63, 9581-9597; (c) G. K. Friestad and A. K. Mathies, Tetrahedron, 2007, 63, 2541-2569; (d) S. E. Schaus and A. Ting, Eur. J. Org. Chem., 2007, 5797–5815; (e) M. Petrini and E. Torregiani, Synthesis, 2007, 159–186; (f) M. Petrini, Chem. Rev., 2005, 105, 3949-3977; (g) J. Royer, M. Bonin and L. Micouin, *Chem. Rev.*, 2004, **104**, 2311–2352; (h) B. E. Maryanoff, H. C. Zhang, J. H. Cohen, I. J. Turchi and C. A. Maryanoff, Chem. Rev., 2004, 104, 1431–1628.
- 8 (a) For a preliminary communication on this work, see: A. Boto, R. Hernández and C. J. Saavedra, Synlett, 2010, 659-663. For other related work from our group, see: (b) A. Boto and I. Romero-Estudillo, Org. Lett., 2011, 13, 3426-3429; (c) A. Boto, D. Hernández and R. Hernández, Eur. J. Org. Chem., 2010, 6633-6642; (d) A. Boto, D. Hernández and R. Hernández, Eur. J. Org. Chem., 2010, 3847-3857; (e) C. Saavedra, R. Hernandez, A. Boto and E. Alvarez, J. Org. Chem., 2009, 74, 4655-4665; (f) A. Boto, D. Hernández, R. Hernández and E. Alvarez, Eur. J. Org. Chem., 2009, 3853-3857; (g) A. Boto, D. Hernández and R. Hernández, Tetrahedron Lett., 2008, 49, 455-458; (h) A. Boto, D. Hernández, R. Hernández, A. Montoya and E. Suárez, Eur. J. Org. Chem., 2007, 325-334; (i) A. Boto, D. Hernández and R. Hernández, *Org. Lett.*, 2007, **9**, 1721–1724; (*j*) A. Boto, D. Hernández, R. Hernández and E. Álvarez, *J. Org. Chem.*, 2007, **72**, 9523-9532; (k) A. Boto, J. A. Gallardo, D. Hernández and R. Hernández, J. Org. Chem., 2007, 72, 7260-7269; (I) A. Boto, R. Hernández, Y. León, J. R. Murguía and A. Rodríguez-Afonso, Eur. J. Org. Chem., 2005, 673-682; (m) For a review on the modification of amino acids and carbohydrates through radical chemistry, see: S. G. Hansen and T. Skrydstrup, Top. Curr. Chem., 2006, 264, 135-162; (n) For a review covering radical chemistry with hypervalent iodine reagents, see: V. V. Zhdankin and P. J. Stang, Chem. Rev., 2008, 108, 5299-5358.
- 9 For the β-amino aldehyde 9, see: (a) D. S. Laitar, J. W. Kramer, B. T. Whiting, E. B. Lobkovsky and G. W. Coates, Chem. Commun., 2009, 5704-5706; (b) P. Gizecki, R. Dhal, C. Poulard, P. Gosselin and G. Dujardin, J. Org. Chem., 2003, 68, 4338-4344; (c) W. J. Greenlee, P. L. Allibone, D. S. Perlow, A. A. Patchett, E. H. Ulm and T. C. Vassil, J. Med. Chem., 1985, 28, 434-442.

- 10 (a) For substrates 10-12, see ref. 8e (b) For substrate 13, see: A. Boto, R. Hernández and E. Suárez, J. Org. Chem., 2000, 65, 4930-4937.
- (a) For the β -amino aldehyde 15, see ref. 9a and also: C. N. C. Drey, R. J. Ridge and E. Mtetwa, J. Chem. Soc., Perkin Trans. 1, 1980, 378-380; (b) B. Weiner, A. Baeza, T. Jerphagnon and B. L. Feringa, J. Am. Chem. Soc., 2009, 131, 9473–9474; (c) For the β-amino aldehyde 17, see ref. 9a; (d) For the β-amino aldehyde 21, see: T. Shono, Y. Matsumura and T. Kenji, J. Am. Chem. Soc., 1981, 103, 1172-1176.
- 12 A. Boto, J. A. Gallardo, R. Hernández and C. J. Saavedra, Tetrahedron Lett., 2005, 46, 7807-7811.
- 13 For proline benzyl carbamate 22, see: (a) K. Hattori, H. Sajiki and K. Hirota, Tetrahedron, 2000, 56, 8433-8442; (b) S. E. Denmark, J. P. Edwards, T. Weber and D. W. Piotrowski, Tetrahedron: Asymmetry, 2010, 21, 1278-1302
- 14 For the hydroxyproline derivative 23, see: (a) S. C. Mayers, J. Ramanjulu, M. D. Vera, A. J. Pfizenmayer and M. M. Jouille, J. Org. Chem., 1994, 59, 5192-5205; (b) J. C. Jung and M. A. Avery, Tetrahedron: Asymmetry, 2006, 17, 2479-2486.
- 15 For the β-amino aldehyde 25, see: (a) A. Fleurant, J. P. Celerier and G. Lhommet, Tetrahedron: Asymmetry, 1992, 3, 695-696; (b) Y. Sugano and J. A. Katzenellenbogen, Bioorg. Med. Chem. Lett., 1996, 6, 361-366; (c) H. Konno, S. Kusumoto, S. Kanai, Y. Yamahana, K. Nosaka and K. Akaji, Heterocycles, 2006, 68, 2579-2585; (d) E. C. Carlson, L. K. Rathbone, H. Yang, N. D. Collett and R. G. Carter, J. Org. Chem., 2008, 73, 5155-5158. For compounds closely related to products 28 and

- 29, see: (e) M. Sunagawa, H. Matsumura, T. Inoue, M. Fukasawa and M. Kato, Patent US4962103, 1990.
- 16 (a) C. H. Larsen, B. H. Ridgway, J. T. Shaw and K. A. Woerpel, J. Am. Chem. Soc., 1999, 121, 12208-12209; (b) D. M. Smith, M. B. Tran and K. A. Woerpel, *J. Am. Chem. Soc.*, 2003, **125**, 14149–14152; (c) C. H. Larsen, B. H. Ridgway, J. T. Shaw, D. M. Smith and K. A. Woerpel, J. Am. Chem. Soc., 2005, 127, 10879-0884; (d) D. Smith and K. A. Woerpel, Org. Biomol. Chem., 2006, 4, 1195-1201; (e) K. M. Bonger, T. Wennekes, D. V. Filippov, G. Lodder, G. A. van der Marel and H. S. Overkleeft, Eur. J. Org. Chem., 2008, 3678-3688; (f) For preliminary studies of the application of this effect in the scission-addition reaction, see: A. Boto, D. Hernández and R. Hernández, Tetrahedron Lett., 2009, 50, 3974-3977.
- 17 (a) For substrates 30-35, see ref. 8e (b) For substrate 36, see: A. Bodanszky, M. Bodanszky, N. Chandramouli, J. Z. Kwei, J. Martinez and J. C. Tolle, J. Org. Chem., 1980, 45, 72-76; (c) U. Schmidt, R. Utz, A. Lieberknecht, H. Griesser, B. Potzolli, J. Bahr, K. Wagner and P. Fischer, Synthesis, 1987, 236-241; (d) Y. Okada, S. Iguchi, T. Hirai, Y. Goto, M. Yagyu and H. Yajima, Chem. Pharm. Bull., 1984, 32, 4608-4615.
- 18 Handbook of Biologically Active Peptides, ed. A. J. Kastin, Academic Press, San Diego, 2006, Ch. 14.
- 19 H. Itokawa, K. Kondo, Y. Hitotsuyanagi, M. Isomura and K. Takeya, Chem. Pharm. Bull., 1993, 41, 1402-1410.
- 20 W. L. F. Armarego and C. L. L. Chai, Purification of Laboratory Chemicals, Elsevier, UK, 5th edn, 2002.