



Contents lists available at ScienceDirect

Bioorganic & Medicinal Chemistry Letters

journal homepage: www.elsevier.com/locate/bmcl

Design, synthesis, and characterization of novel, nonquaternary reactivators of GF-inhibited human acetylcholinesterase



Stanton F. McHardy^{a,*}, Jonathan A. Bohmann^a, Michael R. Corbett^a, Bismarck Campos^a, Michael W. Tidwell^a, Paul Marty Thompson^a, Chris J. Bembien^a, Tony A. Menchaca^a, Tony E. Reeves^a, William R. Cantrell Jr.^a, William E. Bauta^a, Ambrosio Lopez^a, Donald M. Maxwell^b, Karen M. Brecht^b, Richard E. Sweeney^b, John McDonough^b

^a Southwest Research Institute, Chemistry and Chemical Engineering Division, Department of Medicinal and Process Chemistry, 6220 Culebra Road, San Antonio, TX 78266, USA

^b USAMRICD Research Division, Pharmacology Branch, 3100 Ricketts Point Road, APG, MD 21010, USA

ARTICLE INFO

Article history:

Received 10 December 2013

Revised 13 February 2014

Accepted 18 February 2014

Available online 28 February 2014

Keywords:

Acetylcholinesterase

Reactivation

Cyclosarin (GF)

GF-inhibited hAChE

Structure–activity relationship

Heteroaryl keto-oximes

ABSTRACT

The goal of this research was to identify structurally novel, non-quaternarypyridinium reactivators of GF (cyclosarin)-inhibited hAChE that possess the capacity to mediate in vitro reactivation of GF-inhibited human acetylcholinesterase (hAChE). New compounds were designed, synthesized and assessed in GF-inhibited hAChE assays. Structure activity relationships for AChE binding and reactivation of GF-inhibited hAChE were developed. Lead compounds from two different chemical series, represented by compounds **17** and **38**, displayed proficient in vitro reactivation of GF-inhibited hAChE, while also possessing low inhibition of native enzyme.

© 2014 Elsevier Ltd. All rights reserved.

Exposure to highly toxic organophosphorus compounds in the form of pesticides and their metabolites (e.g., paraoxon, malaoxon) or nerve agents (VX, Sarin, Soman or Cyclosarin) pose substantial threats to both military and civilian populations. Nerve agents of this type have been studied extensively and there is significant evidence showing their toxic effects are a result of phosphorylation of the active site serine residue in acetylcholinesterase (AChE).¹ Since AChE is responsible for hydrolyzing acetylcholine to choline, this irreversible inhibition of the AChE enzyme causes an accumulation of acetylcholine in both peripheral and central synapses, ultimately leading to deleterious effects, such as ocular pain, diminished vision, tightness of the chest, wheezing, bronchial constriction and secretion, and ultimately respiratory failure and death. Of significant relevance to our research efforts is the fact that nerve agents also penetrate the blood–brain barrier, inhibiting central AChE, thus causing a number of CNS related effects, such as confusion, ataxia, convulsions, slurred speech, and coma. Furthermore, studies have suggested that exposure to sub-lethal doses of nerve agent

can dramatically impact CNS mediated behavioral attributes such as decreased ability to perform tasks.² Other studies have shown that the detrimental effects of a single dose of agent can be prolonged³ and most of the human data, which is focused on organophosphate pesticides, suggest long-term CNS effects such as irritability, impaired memory, insomnia or anxiety.⁴

Pyridinium oxime compounds have been postulated to act as reactivators by binding to nerve agent-inhibited AChE and hydrolyzing the phosphonate group from the serine residue via nucleophilic attack of the oxime moiety on the phosphorus atom. The resulting reactivation of AChE allows the enzyme to continue converting acetylcholine to choline, helping maintain functional levels of acetylcholine.⁵ Quaternary pyridinium aldoxime compounds, such as 2-PAM (Fig. 1) are limited to peripheral reactivation,⁶ however, results with uncharged dihydropyridine 'pro-drugs' of 2-PAM have supported central protection.⁷ Historically,⁸ studies on non-pyridinium based AChE oxime reactivators have received little attention, with the exception of the simple acetyl oximes such as MINA and DAM.⁹ During the course of our investigation reported here, a small number of non-pyridinium based AChE reactivators were reported, represented by the dihydroisoquinoline **1**,¹⁰ amidine **2**,¹¹ phenyl ketooxime **3**,¹² and the amide oxime RS41A¹³ (Fig. 1).

* Corresponding author at present address: University of Texas San Antonio, Department of Chemistry, Center for Innovative Drug Discovery, One UTSA Circle, San Antonio, TX 78249, USA. Tel.: +1 210 458 8676; fax: +1 210 458 7428.

E-mail address: stanton.mchardy@utsa.edu (S.F. McHardy).

Figure 2. Structure of prepared amide oxime analogs

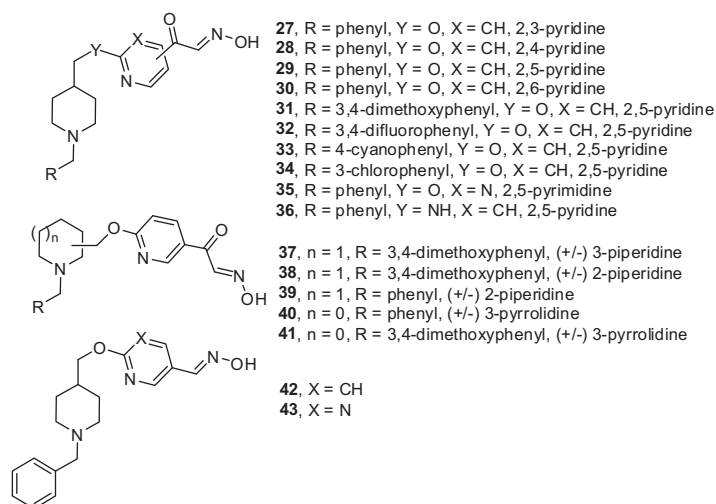
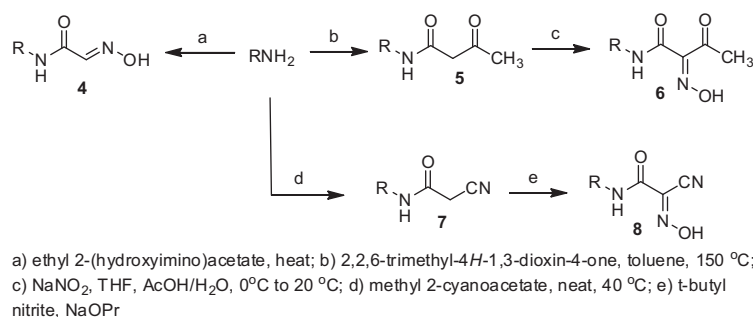
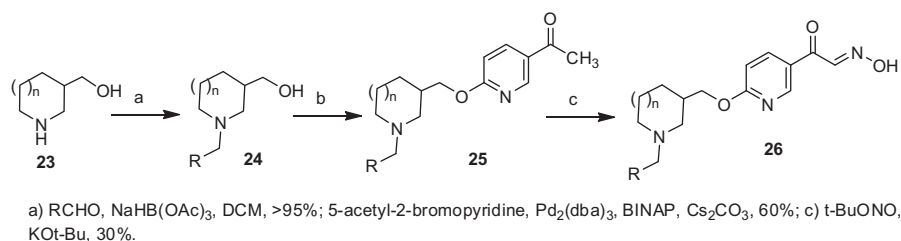


Figure 3. Structure of heteroaryl keto-oximes.



Scheme 1. Synthesis of amide oxime analogs.



Scheme 2. Synthesis of heteroaryl keto-oxime 26.

substantial effects on activity. The initial analog in this series, compound **29**, which possessed the 2,5-substitution pattern on the pyridine, provided 23% reactivation and a K_i of 31 μ M. Compound **30**, the 2,6-substituted pyridine, as compared to **29** showed a decrease in the affinity for native hAChE (199 μ M vs 31 μ M) as well as the corrected reactivation (7% vs 23%). Moving the keto-oxime group to the 2,4-position **28** decreased both reactivation (17%) and inhibition (144 μ M) relative to **29**. However, the 2,3-substitution pattern in **27** displayed a marked improvement in reactivation (45%), while possessing low inhibition (180 μ M). Based on these SAR findings the order of reactivation activity for substitution on the pyridine ring is 2,3- > 2,5- > 2,4- > 2,6. Changing the piperidine sidechain from benzyl **29** to 3,4-dimethoxybenzyl **31** produced equivalent reactivation activity, but showed a >60-fold decrease in affinity for native enzyme. Additional modifications to the *N*-benzyl side chain, as represented by compounds **32–34**, led to moderate improvements in reactivation activity with the 3,4-difluoro analog **32**. Installment of the 2,5-substituted pyrimi-

dine ring **35**, produced compounds with very similar activity to the corresponding pyridine **29**. The ether linkage in **29** can also be replaced by an amine, resulting in a compound **36** with equivalent reactivation activity and a 9-fold decrease in affinity for native enzyme.

Finally, we attempted to translate SAR trends from the 4-substituted piperidines to the 2- and 3-substituted piperidines, as well as pyrrolidines, an effort which produced lead compounds with good activity. Compound **38** produced a significant increase in reactivation activity (65%) as compared to the corresponding 3-substituted piperidine **37** (38%) or the 4-substituted analog **29**. The 3-substituted pyrrolidines **40** and **41** showed similar reactivation activity, however, as observed before, the 3,4-dimethoxybenzyl side chain displayed a decrease in affinity for native enzyme. Analogs **42** and **43**, which possess the oxime directly attached to the pyridine or pyrimidine ring, produced a significant decrease in reactivation relative to the corresponding keto-oximes **29** and **35**.²⁵

Table 1
In vitro GF-Inhibited hAChE reactivation data^a

Compound	Reactivation ^b	K _i ^c	Compound	Reactivation ^b	K _i ^c
MINA	7.7	0	28	17.2	144
9	7.5	2430	29	23.0	31.2
10	7.7	3178	30	7.3	199
11	11.8	234	31	25.3	1930
12	23.7	2300	32	30.8	159
13	0.1	732	33	21.4	1608
14	4.8	29938	34	18.9	298
15	22.7	1625	35	23.8	55.2
16	2.7	595	36	22.9	285
17	36.5	1926	37	38.2	345
18	28.9	2671	38	64.9	233
19	7.2	6.2	39	33.9	551
20	2.4	1.5	40	25.1	137
21	6.9	3.1	41	24.7	2517
22	0.8	0.59	42	0.4	270000
27	45.3	180	43	2.0	198

^a Mean and STD data available in [Supplementary information](#).

^b % Corrected cumulative reactivation of GF-inhibited hAChE by 20 uM oxime at 4 h.²⁴

^c K_i inhibition constant for native AChE (uM).

In conclusion, two new chemical series of non-pyridinium based reactivators of hAChE have been discovered, both of which possess a range of reactivation activities for GF-inhibited hAChE and a varying degrees of selectivity over affinity for native enzyme. Lead compounds from these studies, such as **15**, **17**, **27**, **29** and **38**, which possess substantial reactivation activity, low affinity for native enzyme, and favorable physiochemical properties, have been advanced to blood brain barrier permeability, multiple in vivo reactivation, and agent-challenge studies. The results of these studies will be reported in due course.

Acknowledgments

Support for this work by Defense Threat Reduction Agency (HDTRA1-10-C-0041) is gratefully acknowledged.

Supplementary data

Supplementary data associated with this article can be found, in the online version, at <http://dx.doi.org/10.1016/j.bmcl.2014.02.049>.

References and notes

- Marrs, T. C.; Maynard, R. L.; Sidell, F. R. *In Chemical Warfare Agents, Toxicology and Treatment*, 2nd ed.; Wiley & Sons: Ltd, 2007.

- DiGiovanni, C. *Am. J. Psychiatr.* **1999**, *156*, 1500; D'Mello, G. D. *Human Exp. Toxicol.* **1993**, *12*, 3.
- McDonough, J. H. et al *Behav. Toxicol. Teratol.* **1986**, *8*, 179.
- Jamal, G. A. *Adv. Drug React. Toxicol. Rev.* **1995**, *14*, 85.
- Eyer, P. A.; Worek, F. *In Chemical Warfare Agents, Toxicology and Treatment*, 2nd ed.; Marrs, T. C.; Maynard, R. L.; Sidell, F. R., Eds.; Wiley & Sons Ltd: **2007**; Chapter 15, pp 305–330.
- Taylor, P. (2011) 'Chapter 8. Anticholinesterase Agents' (Chapter). Brunton, L. L., Lazo, J. S., Parker, K. L.: Goodman & Gillman's The Pharmacological Basis of Therapeutics.
- (a) Bodor, N.; Shek, E.; Higuchi, T. *Science* **1975**, *190*, 155–156; (b) DeMar, J. C.; Clarkson, E. D.; Ratcliffe, R. H.; Campbell, A. J.; Thangavelu, S. G.; Herdman, C. A.; Leader, H.; Schulz, S. M.; Marek, E.; Medynets, M. A.; Ku, T. C.; Evans, S. A.; Khan, F. A.; Owens, R. R.; Nambiar, M. P.; Gordon, R. K. *Chem. Biol. Interact.* **2010**, *187*, 191.
- (a) Kassa, J.; Kuca, K.; Bartosova, L.; Kunesova, G. *Curr. Org. Chem.* **2007**, *11*, 267; (b) Mercey, G.; Verdet, T.; Renou, J.; Kliachyna, M.; Baati, R.; Nachon, F.; Jean, L.; Renard, P. Y. *Acc. Chem. Res.* **2012**, *45*, 756.
- (a) Shih, T. M.; Duniho, S. M.; McDonough, J. H. *Toxicol. Appl. Pharmacol.* **2003**, *188*, 69; (b) Shih, T. M.; Skovira, J. W.; O'Donnell, J. C.; McDonough, J. H. *J. Mol. Neurosci.* **2010**, *40*, 63; (c) Skovira, J. W.; O'Donnell, J. C.; Koplovitz, I.; Kan, R. K.; McDonough, J. H.; Shih, T. M. *Chem. Biol. Interact.* **2010**, *187*, 318.
- (a) Mercey, G.; Verdet, T.; Saint-Andre, G.; Gillon, E.; Wagner, A.; Baati, R.; Ludovic, J.; Nachon, F.; Renard, P. Y. *Chem. Commun.* **2011**, *47*, 5295; (b) Mercey, G.; Renou, J.; Verdet, T.; Kliachyna, M.; Baati, R.; Gillon, E.; Arboles, M.; Loidice, M.; Nachon, F.; Jean, L.; Renard, P. Y. *J. Med. Chem.* **2012**, *55*, 10791.
- Kalisiak, J.; Ralph, E. C.; Cashman, J. R. *J. Med. Chem.* **2012**, *55*, 465.
- De Koning, M. C.; Van Grol, M.; Noort, D. *Toxicol. Lett.* **2011**, *206*, 54.
- (a) Kovarik, Z.; Macek, N.; Sit, R. K.; Radic, Z.; Fokin, V. V.; Sharpless, K. B.; Taylor, P. *Chem. Biol. Interact.* **2013**, *203*, 77; (b) Radic, Z.; Sit, R. K.; Kovarik, Z.; Berend, S.; Garcia, E.; Zhang, L.; Amitai, G.; Green, C.; Radic, B.; Fokin, V. V.; Sharpless, K. B.; Taylor, P. *J. Biol. Chem.* **2012**, *287*, 11798; (c) Sit, R. K.; Radic, Z.; Gerardi, V.; Zhang, L.; Garcia, E.; Katalinic, M.; Amitai, G.; Kovarik, Z.; Fokin, V. V.; Sharpless, K. B.; Taylor, P. *Chem. Biol. Interact.* **2011**, *286*, 19422.
- McHardy, S. F.; Corbett, R. M.; Maxwell, D. M.; Tidwell, M. W.; Campos, B.; Bembien, C. J. US Patent Application 20130035351, **2013**.
- Kuca, K.; Jun, D.; Musilek, K. *Mini-Rev. Med. Chem.* **2006**, *6*, 269. and reference cited within.
- (a) Lipinski, C. A.; Lombardo, F.; Dominy, B. W.; Feeney, P. J. *Adv. Drug Delivery Rev.* **2001**, *46*, 3–26; (b) Wager, T. T.; Hou, X.; Verhoest, P. R.; Villalobos, A. *ACS Chem. Neurosci.* **2010**, *6*, 435.
- Kumar, P.; Dhawan, K. N.; Kishor, K.; Bhargava, K. P.; Satsangi, R. K. *J. Heterocycl. Chem.* **1982**, *19*, 677.
- Carlier, P. R.; Du, D. M.; Han, Y.; Liu, J.; Pang, Y. P. *Bioorg. Med. Chem. Lett.* **1999**, *9*, 2335.
- Abdel-Magid, A. F.; Carson, K. G.; Harris, B. D.; Maryanoff, C. A.; Shah, R. D. *J. Org. Chem.* **1996**, *61*, 3849.
- Gowrisankar, S.; Sergeev, A. G.; Anbarasan, P.; Spannenberg, A.; Neumann, H.; Beller, M. *J. Am. Chem. Soc.* **2010**, *132*, 11592.
- Yonishi, S.; Aoki, S.; Matsushima, Y.; Akahane, A., US Patent 20050113387, **2005**.
- Mohammed, A. H. A.; Nagendrappa, G. *Tetrahedron Lett.* **2003**, *44*, 2753.
- Ellman, G. L.; Courtney, K. D.; Andres, V.; Featherstone, R. M. *Biochem. Pharmacol.* **1961**, *7*, 88.
- Reactivation at the 4 h time point was initially assessed due to slow reactivation rates observed with early analogs. Future work will involve testing the more advanced analogs at therapeutically relevant time points.
- pK_a's of various analogs were assessed via a capillary electrophoresis method (Analiza, Inc.) across a 24-point titration curve. pK_a's for oximes **42** and **43** were higher (11.2 and 10.7, respectively) than the corresponding keto-oxime derivatives **29** and **35** (7.8 and 7.6, respectively).