



Discovery of a series of potent arylthiadiazole H₃ antagonists

Dong Xiao^{a,*}, Anandan Palani^a, Michael Sofolarides^a, Ying Huang^a, Robert Aslanian^a, Henry Vaccaro^a, Liwu Hong^a, Brian McKittrick^a, Robert E. West Jr.^b, Shirley M. Williams^b, Ren-Long Wu^b, Joyce Hwa^b, Christopher Sondey^b, Jean Lachowicz^b

^aChemical Research Department, Merck Research Laboratories, 2015 Galloping Hill Road, Kenilworth, NJ 07033, United States

^bCardiovascular/Metabolic Disease, Merck Research Laboratories, 2015 Galloping Hill Road, Kenilworth, NJ 07033, United States

ARTICLE INFO

Article history:

Received 3 September 2010

Revised 7 November 2010

Accepted 16 November 2010

Available online 21 November 2010

Keywords:

H₃ antagonists

SAR study

ABSTRACT

A series of 2-piperidinopiperidine-5-arylthiadiazoles was synthesized and subjected to a structure–activity relationship (SAR) investigation. The potency of this series was improved to the single digit nanomolar range. The key analogs were shown to be free of P450 issues, and they also maintained good ex vivo activity and brain penetration.

© 2010 Elsevier Ltd. All rights reserved.

The biogenic amine histamine is a critical neurotransmitter which mediates a wide range of neurological responses. The effect of histamine is exerted through receptors that belong to the G protein-coupled receptors (GPCRs). There are four subtypes of the histamine receptor (HR): H₁, H₂, H₃ and H₄.¹ HR antagonists have historically provided medical benefits, along with pharmaceutical blockbusters, through modulation of the H₁ and H₂ receptors, in the treatment of allergic conditions and gastric disorders, respectively. Due to the success of H₁ and H₂ therapeutics, the potential of H₃ and H₄ antagonism has attracted considerable research efforts across the industry.²

Following the pharmacological identification of the H₃ receptor in 1983,³ and its cloning in 1999,⁴ significant strides were made to identify novel, selective H₃ receptor antagonists.⁵ Although the exact therapeutic potential is unknown, H₃ antagonists lead to increased histamine levels at histaminergic neurons and subsequently may be useful in treating obesity and diabetes, as well as other CNS disorders such as cognitive disorders, Alzheimer's and Parkinson's disease, schizophrenia and sleep disorders.^{6–8}

Among the earliest H₃ antagonists investigated were imidazole based compounds;⁹ however cytochrome P450 (CYP) enzyme inhibition posed significant drug–drug interaction issues. This finding subsequently led to research in non-imidazole containing compounds.¹⁰ In our effort to discover novel H₃ antagonists we identified a high throughput screen (HTS) lead with prototype structure **1** containing a piperidine thiadiazole core. This series showed modest in vitro potency, and it is likely that the pendent urea moiety

would hinder entry into the brain due to its multiple H bond donor and acceptor characteristics. At the onset of lead optimization, based on literature precedents¹¹ and a H₃ pharmacophore model,¹² we envisioned the use of aryl groups to replace the urea, therefore increasing brain penetration¹³ (Scheme 1).

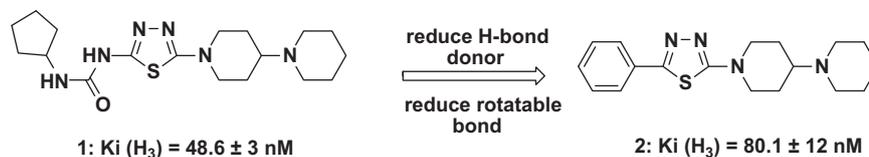
Introduction of an unsubstituted phenyl ring in place of the urea moiety afforded compound **2**. Compound **2** had comparable activity to **1** thus confirming our initial structural hypothesis. Furthermore, the caco2 permeability of **2** was improved to 104 nm/sec from 10.5 nm/sec for **1**, and the absorption was also improved to good from moderate. These results were consistent to the empirical rules that reduction of H bond donors and acceptors would improve CNS penetration.¹³ Encouraged by these findings, we undertook a full optimization of the phenyl ring. Synthesis of this phenyl series was relatively straightforward. Commercial compound **3** was converted to the dibromo intermediate **4**, which was then displaced with piperidinopiperidine. The reaction can be controlled to afford **5** exclusively, since the amino substitution deactivated further addition. The bromothiadiazole **5** was subsequently coupled under Pd catalysis conditions with aryl boron or zinc species to afford the desired products (Scheme 2).

Since H₃ activity is centrally mediated, we focused on small substitutions on the aryl group to minimize molecular weight. Table 1 shows H₃ in vitro potency¹⁴ for substitutions at the *ortho*-, *meta*- and *para*-positions. Polar substituents such as OMe, CN and acetyl groups afforded moderate improvement of potency (Table 1).

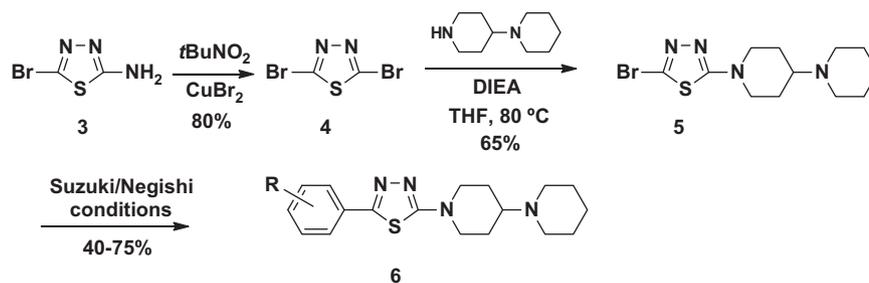
Considering these data, especially that of *meta*-substituted compounds, we speculated that an additional *meta*-substitution would provide a pseudo-symmetric interaction with the receptor,

* Corresponding author.

E-mail address: dong.xiao@merck.com (D. Xiao).

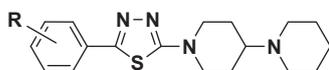


Scheme 1.



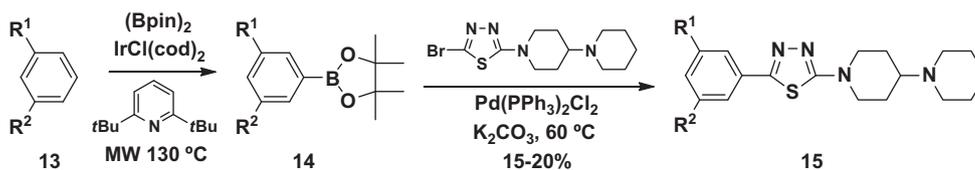
Scheme 2.

Table 1



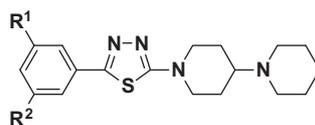
Entry	R	<i>ortho</i> -	$K_i (H_3)$ (nM)	<i>meta</i> -	<i>para</i> -
2	H	80.1 ± 12	80.1 ± 12		80.1 ± 12
7	OMe	40.0 ± 8	31.9 ± 6		40.1 ± 2
8	CN	N/A	42.6 ± 1		36.4 ± 7
9	F	36.0 ± 2	62.0 ± 11		39.0 ± 15
10	Cl	69.0 ± 1	49.0 ± 1		161 ± 69
11	Me	36.0 ± 4	60.0 ± 8		163 ± 23
12	COCH ₃	8.4 ± 2	4.0 ± 0.2		65.0 ± 4

which could further increase the binding potency. This series of analogs was investigated and synthesized using commercially available boronic acids and esters. Custom synthesis of non-commercial 3,3'-disubstituted boronic esters **14** was also developed



Scheme 3.

Table 2



Entry	15a	15b	15c	15d	15e	15f	15g
R ¹ /R ²	Cl/Cl	OMe/OMe	Cl/OMe	CN/OMe	F/OMe	F/F	CN/CN
$K_i (H_3)$ (nM)	72.0 ± 9	15.0 ± 1	12.0 ± 1	12.0 ± 3	16.0 ± 2	50.0 ± 1	23.0 ± 1

through microwave-accelerated Ir-catalyzed, direct borylation¹⁵ of the corresponding aromatics, which were then coupled with bromothiazolidine **5**. (Scheme 3).

Notable in vitro binding data are shown below. Aside from the dichloro analog **15a**, bis-*m,m'*-substitution did increase potency compared to the corresponding mono-substituted analog. Further confirming our speculation, hetero-*m,m'*-disubstitutions afforded activity in the low double digit nanomolar range, as in entries Cl/OMe, CN/OMe, and F/OMe (Table 2).

Additionally, following our initial heteroatom-containing lead, we sought to replace the phenyl group with heteroaromatic rings. Among the pyridyl substitutions, 2-pyridine showed the greatest increase in potency and was pursued further. Results are shown below (Table 3):

The 2-pyridyl analogs provided the most pronounced improvement in potency, compared to the less active pyrimidine and pyrazole analogs. Interestingly, the 3-methylpyridine and 3-methoxypyridine afforded the most potent analogs in this series, results consistent with the substituted phenyl series summarized above.

Table 3

Entry						
	16a	16b	16c	16d	16e	16f
Heteroaryl						
K_i (H_3) (nM)	30.0 ± 0.1	3.5 ± 1	6.1 ± 2	103.0 ± 22	192.0 ± 27	28 ± 7

Table 4

Entry					
	17a	17b	17c	17d	17e
R					
K_i (H_3) (nM)	40.1 ± 2	161.8 ± 2	182.3 ± 15	652.6 ± 79	600.9 ± 96

To complete the SAR study of this thiadiazole series, we also investigated the modifications at the piperidinopiperidine moiety. For example, the piperazine analogs **17b–17d** were aimed at modulating the basicity of the nitrogen. Additional analog such as **17e** was directed at reducing molecular weight. However, all these analogs showed drastic reduction of H_3 potency compared to the original analog **17a**. Indeed, only the piperidinopiperidine as the right hand side consistently afforded good potency (Table 4).

As mentioned previously, the earlier imidazole based H_3 compounds exhibited P450 inhibition issues. Therefore, in the current series, the most potent compounds **15c** and **16b** were screened against three CYP isoforms—3A4, 2D6 and 2C9. Fortunately, neither compound showed greater than 20 μ M inhibition for 3A4, 2D6 and 2C9 under pre- or co-incubation conditions. Compound **15c** was particularly noteworthy due to its ex vivo potency.¹⁶ This compound also showed high brain concentrations in mouse and

a modest AUC in rat per oral dosing, a significant improvement from the original lead **1** (Table 5).

In conclusion, a series of 2-piperidinopiperidine-5-arylthiadiazoles as centrally active H_3 antagonists was designed and was subjected to a SAR investigation. First, we replaced the urea moiety in **1** with a phenyl group, which significantly improved the absorption and permeability. We then systematically investigated the effects of *ortho*-, *meta*- and *para*-substituted phenyl rings. The use of polar groups such as OMe, CN, and COCH₃ and particularly with the *m,m'*-disubstitutions afforded improvement of potency over phenyl analog. Additionally, heteroaryl replacement of the phenyl ring was investigated. 2-Pyridyl substitutions increased potency significantly compared to the initial lead, while pyrimidine and pyrazole offered less activity. Further substitutions, 3-methylpyridine and 3-methoxypyridine offered potency in the single digit nanomolar range. The best compounds, **15c** and **16b**,

Table 5

Entry					
	K_i (H_3) (nM)	Rat AUC @10 mpk, po (ng h/ml 0–6 h)	Mouse brain level@10 mpk, po (ng/g @ 4 h)	Ex vivo @ 4 h	P450 inhib 3A4, 2D6, 2C9
	48.6 ± 3	0	ND	ND	>20 μ M
	12.0 ± 1	379	9027	90%	>20 μ M
	3.5 ± 1	0	1527	74%	>20 μ M

were devoid of P450 inhibition issues, and showed good brain concentrations and ex vivo potency. Further optimization of the pharmacokinetic profile, as well as other biological parameters, led to an orally active H₃ antagonist in our mouse obesity model. These results will be disclosed in due course.

Acknowledgments

The authors thank Dr. John J. Piwinski for supporting this research program and Dr. Pauline Ting for helpful comments on this letter.

References and notes

- Parsons, M. E.; Ganellin, C. R. *Br. J. Pharmacol.* **2006**, *147*, S127.
- Leurs, R.; Bakker, R. A.; Timmerman, H.; de Esch, I. J. P. *Nat. Rev. Drug Disc.* **2005**, *4*, 107.
- Arrang, J. M.; Garbarg, M.; Schwartz, J. C. *Nature* **1983**, *302*, 832.
- Lovenberg, T. W.; Toland, B. L.; Wilson, S. J.; Jiang, X.; Pyati, J.; Hurvar, A.; Jackson, M. R.; Erlander, M. G. *Mol. Pharmacol.* **1999**, *55*, 1101.
- For recent reviews: (a) Raddatz, R.; Tao, M.; Hudkins, R. L. *Curr. Top. Med. Chem.* **2010**, *10*, 153; (b) Gemkow, M. J.; Davenport, A. J.; Harich, S.; Ellenbroek, B. A.; Cesura, A.; Hallett, D. *Drug Discovery Today* **2009**, *14*, 509; (c) Stocking, E. M.; Letavic, M. A. *Curr. Top. Med. Chem.* **2008**, *8*, 988; (d) Esbenschade, T. A.; Browman, K. E.; Bitner, R. S.; Strakhova, M.; Cowart, M. D.; Brioni, J. D. *Br. J. Pharmacol.* **2008**, *154*, 1166; (e) Hudkins, R. L.; Raddatz, R. *Annu. Rep. Med. Chem.* **2007**, *42*, 49; (f) Berlin, M.; Boyce, C. W. *Expert Opin. Ther. Pat.* **2007**, *17*, 675; (g) Hancock, A. A. *Biochem. Pharmacol.* **2006**, *71*, 1103.
- Stark, H.; Schlicker, E.; Schunack, W. *Drugs Future* **1996**, *21*, 507.
- Kurose, Y.; Terashima, Y. *Brain Res.* **1999**, *828*, 115.
- Masaki, T.; Chiba, S.; Yoshimichi, G.; Yasuda, T.; Noguchi, H.; Kakuma, T.; Sadata, T.; Yoshimatsu, H. *Endocrinology* **2003**, *144*, 2741.
- Stark, J.; Kathmann, M.; Schlicker, E.; Schunack, W.; Schlegel, B.; Wolfgang, S. *Mini-Rev. Med. Chem.* **2004**, *9*, 965.
- Zhang, M.; Ballard, M. E.; Pan, P.; Roberts, S.; Faghieh, R.; Cowart, M. D.; Esbenschade, T. A.; Fox, G. B.; Decker, M. W.; Hancock, A. A.; Rueter, L. E. *Brain Res.* **2005**, *1045*, 142.
- For examples: (a) Medhurst, A. D.; Atkins, A. R.; Beresford, I. J.; Brackenborough, K.; Briggs, M. A.; Calver, A. R.; Cilia, J.; Cluderay, J. E.; Crook, B.; Davis, J. B.; Davis, R. K.; Davis, R. P.; Dawson, L. A.; Foley, A. G.; Gartlon, J.; Gonzalez, M. I.; Heslop, T.; Hirst, W. D.; Jennings, C.; Jones, D. N. C.; Lacroix, L. P.; Martyn, A.; Ociepa, S.; Ray, A.; Regan, C. M.; Roberts, J. C.; Schogger, J.; Southam, E.; Stean, T. O.; Trail, B. K.; Upton, N.; Wadsworth, G.; Wald, J. A.; White, T.; Witherington, J.; Woolley, M. L.; Worby, A.; Wilson, D. M. *J. Pharmacol. Exp. Ther.* **2007**, *321*, 1032; (b) Cowart, M.; Faghieh, R.; Curtis, M. P.; Gfesser, G. A.; Bennani, Y. L.; Black, L. A.; Pan, L.; Marsh, K. C.; Sullivan, J. P.; Esbenschade, T. A.; Fox, G. B.; Hancock, A. A. *J. Med. Chem.* **2005**, *48*, 38; (c) Apodaca, R.; Dvorak, C. A.; Xiao, W.; Barbier, A. J.; Boggs, J. D.; Wilson, S. J.; Lovenberg, T. W.; Carruthers, N. I. *J. Med. Chem.* **2003**, *46*, 3938.
- (a) Dastmalchi, S.; Hamzeh-Mivehroud, M.; Ghafourian, T.; Hamzeiy, H. *J. Mol. Graphics Modell.* **2008**, *26*, 834; (b) Narkhede, S. S.; Degani, M. S. *QSAR Comb. Sci.* **2007**, *26*, 744; (c) Axe, F. U.; Bembenek, S. D.; Szalma, S. J. *Mol. Graphics Modell.* **2006**, *24*, 456.
- For recent reviews on CNS penetration, see: (a) Klon, A. E. *Curr. Comput.-Aided Drug Des.* **2009**, *5*, 71; (b) Di, L.; Kerns, E. H.; Carter, G. T. *Expert Opin. Drug Discovery* **2008**, *3*, 677; (c) Obrezanova, O.; Gola, J. M. R.; Champness, E. J.; Segall, M. D. *J. Comput. Aided Mol. Des.* **2008**, *22*, 431; (d) Hitchcock, S. A.; Pennington, L. D. *J. Med. Chem.* **2006**, *49*, 7559; (e) Clark, D. E. *Annu. Rep. Med. Chem.* **2005**, *40*, 403.
- H₃ binding assay: membranes (P2 pellet) from rHu H₃-HEK cells (3 µg protein) were incubated in 200 µl 50 mM Tris-HCl, Ph 7.4, with 1 nM [³H] N-α-methylhistamine (82 Ci/mmol) and compounds at concentrations equivalent to half orders of magnitude over a five-order of magnitude range. Nonspecific binding was determined in the presence of 10⁻⁵ M thioperamide. After 30 min at 30 °C, assay mixture were filtered through 0.3% polyethylenimine soaked GF/B glass fiber filters, which were rinsed thrice with buffer, dried, impregnated with Meltilex wax scintillant and counted. K_i values were determined from curves fit to the data using GraphPad Prism nonlinear, least-squares, curve-fitting program. H₃ binding K_i values were the average of at least two independent determinations. The assay-to-assay variation was generally ±2-fold.
- Harrison, P.; Morris, J.; Marder, T. B.; Steel, P. G. *Org. Lett.* **2009**, *11*, 3586.
- H₃ ex vivo assay: Individual Imprinting Control Region mice (Taconics, nomenclature Icr/Tacr) cortexes were dissected and homogenized in ice cold assay buffer, 50 mM Na₂HPO₄-KH₂PO₄ buffer (pH 6.8). Samples were then frozen at -80 °C for at least 12 h. Protein concentration was determined by BCA Protein Assay. A 30 min room temperature incubation was performed, containing 140 µg/assay homogenized cortex sample, 0.1% BSA, 1 nM [³H] N-α-methylhistamine and 10 µM thioperamide for non-specific binding or assay buffer for total binding. Incubations were performed in quadruplet and stopped by rapid filtration on a Brandel Harvester using Unifilter-96, GF/B plates presoaked in 0.3% PEI for 30 min. The remaining radioactivity was measured on a Packard TopCount-NTX. Specific binding was calculated by subtracting the non-specific binding from the total.