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Molecular structure and spectral properties of indolenine based norsquaraines *versus* squaraines

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Novel indolenine based norsquaraine dyes, wherein the oxygen of the squaric acid bridge was substituted with a barbituric or a dicyanomethylene group, were synthesized and their molecular structure, spectral and luminescent properties were compared to those of analogous squaraine dyes. The molecular structure was investigated using X-ray analysis, NMR spectroscopy and *ab initio* DFT B3LYP/6-311G (d, p) simulations. The calculated populations of possible conformers and the barriers of internal rotations were found to be in good agreement with the experimental data. Norsquaraines absorb and emitt light within the same long-wavelength spectral range as the corresponding squaraines but due to intramolecular *H*-bonds and increased conformational rigidity they were less sensitive to solvent polarity and the presence of protein (BSA).

1. Introduction

Squaraine dyes are a subclass of cyanines that contain a 3-oxocyclobut-1-enolate (squarate) moiety in the polymethine chain [1–3]. Due to their excellent spectral characteristics such as the long-wavelength absorption and emission, high molar absorptivity, high fluorescence quantum yields, and stability, squaraines are widely used as fluorescent probes and labels for biomedical assays [4–6], clinical diagnostics [7,8], pharmaceutical research [9,10], as sensitive dyes in electronics [11], sensitizers for photovoltaic cells [12] and sunlight energy converters [13]. The molecular structure of these dyes was investigated using various experimental [14–17] and theoretical methods [18]. The spectral properties of squaraines [19], their complexes [20,21] and conjugates [22,23] with proteins were also thoroughly studied. Squaraine dyes comprising *indolenine* based terminal end groups are of most interest in particular for biomedical applications due to their increased brightness and photostability [24]. Squaraines may be substituted with various ionic and other functional groups at the indolenine nitrogens, which facilitate their solubility in hydrophobic or hydrophilic media [25,26] and reaction with biomolecules [27].

Nevertheless, the molecular structure and spectral properties of indolenine based squaraines containing hydrogen atoms at the indolenine nitrogens, so-called *norsquaraines*, have not been sufficiently investigated [28,29]. The simplest norsquaraine **nor-SqO** was synthesized and its molecular structure was simulated by the semiempirical methods AM1 and PM3 [30]. The calculations carried out in this publication predicted coexistence of very polar *cis,syn*-conformer labeled as ISQ4 (C_{2v} symmetry) as the major component and non-polar centrosymmetric *trans,anti*-conformer ISQ6 (C_{2h} symmetry) as the minor component while the ¹H NMR data evidenced the opposite conformer population. The molecular geometry of **nor-SqO** investigated by X-ray analysis was affected by the interaction with two CHCl₃ molecules existed in the unit cell, which did not allow to conclude on the anticipated dominant conformer in solutions [31].

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PIGMENTS

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The PCM-PBE0/6-31 + G(d,p) calculations of **nor-SqO** [32] predicted that the conformer ISQ6 was 0.5 kcal/mol more stable than ISQ4. Other conformers had much higher energies and had no effect on these populations. It was demonstrated also that the absorption and emission spectra of **nor-SqO** could be described as a combination of the electronic spectra of these two forms.

A few substituted norsquaraines **1a–1f** were also synthesized but their molecular structures and fluorescence properties were not investigated [33]. Importantly, to the best of our knowledge, no other norsquaraine derivatives were reported until now.

This work investigates the molecular structures and spectral properties of **nor-SqO** and its novel, first synthesized derivatives, where the squaric oxygen is substituted with a barbituric (**nor-SqB**) and dicyanomethylene (**nor-SqCN**) group, and compares these properties with those of conventional, previously reported squaraines **SqO** and **SqCN** [34] and a newly synthesized squaraine **SqB**.



2. Materials and methods

2.1. General information

Iodomethane and 2,3,3-trimethylindolenine were purchased from *Acros*; 3,4-dihydroxy-3-cyclobutene-1,2-dione (squaric acid) was from *Aldrich*; Bovine Serum Albumin (BSA, essentially fatty acid free), Silica gel 60 for column chromatography and other materials were from *Merck* and used as received. Phosphate buffer (PB) pH 7.4 (67 mM) was prepared by dissolving Na₂HPO₄·2H₂O (9.596 g) and KH₂PO₄ (1.743 g) in 1 L distilled water.

Mass spectra were measured by a *BIFLEX III MALDI-TOF* mass spectrometer using 2,5-dihydroxybenzoic acid was taken as the matrix. FAB mass spectra were recorded on a *SELMI MI-1201E* instrument using 3-nitrobenzylalcohol (NBA) or glycerol as a matrix.

The *C*, *H*, *N* elemental analysis was performed by a EuroVector Euro EA 3000 EA-IRMS elemental analyzer.

¹H NMR and ¹³C NMR spectra were measured on a Varian Mercury-VX-200 (¹H 200 MHz) spectrometer in DMSO- d_6 using signal of remaining non-deuterated solvent as an internal standard (2.50 ppm for DMSO). Selected spectra (¹H 400 MHz and ¹³C 100 MHz) were recorded in CDCl₃ using a 400 MHz Bruker Avance^{III} HD. BBI probe was equipped with Z-axis gradients coils. TMS was used as an internal standard (0.00 ppm). A full assignment was done using 2D experiments: COSY (¹H-¹H correlation), HMQC, HMBC (¹H-¹³C short and long correlation, respectfully) and J-resolved. ¹H NOESY experiments employed 128 t_1 increments with a dwell time of 83.2 µs and a mixing time of 1.5 s. A recycle delay of 2.5 s was used in all NMR experiments.

X-Ray diffraction analysis. The dark-blue crystals with metallic luster of **nor-SqO** ($C_{26}H_{24}N_2O_2$) were obtained by recrystallization from 1-propanol with 0.1% diethylamine. Crystals are monoclinic. At 293 K a = 18.1805(8), b = 6.4028(3), c = 18.7430(8) Å, $\beta = 107.185(5)^\circ$, V = 2084.4(2) Å³, $M_r = 396.28$, Z = 4, space group P2₁/n, $d_{calc} = 1.263$ g/cm³, μ (MoK_a) = 0.080 mm⁻¹, F(000) = 840. Intensities of 20723 reflections (4757 independent, $R_{int} = 0.025$) were

measured on the *Xcalibur-3* diffractometer (graphite monochromated MoK_α radiation, CCD detector, ω -scanning, $2\Theta_{max} = 55^{\circ}$). The structure was solved by direct method using SHELXTL package [35]. Positions of the hydrogen atoms were located from electron density difference maps and refined by "riding" model with $U_{iso} = nU_{eq}$ (n = 1.5 for methyl hydrogens and n = 1.2 for other hydrogen atoms) of the carrier atom. The hydrogen atoms at the indolenine nitrogens were refined using isotropic approximation. Full-matrix least-squares refinement against F^2 in anisotropic approximation for non-hydrogen atoms using 4649 reflections with $F > 4\sigma(F)$, S = 0.972). The final atomic coordinates, and crystallographic data for **nor-SqO** molecule have been deposited to the Cambridge Crystallographic Data Center, 12 Union Road, CB2 1EZ, UK (fax: +44-1223-336033; e-mail: deposit@ccdc.cam.ac.uk) and are available on request quoting the deposition numbers CCDC 998817).

Absorption spectra were recorded for the dye concentrations $c_{\text{Dye}} \sim 0.5 \,\mu\text{M}$ in phosphate buffer (PB) 7.4 and other solvent systems discussed below. All the absorption spectra were recorded in 1-cm quartz cells at 25 °C using a *PerkinElmer Lambda 35* UV/Vis spectrophotometer. Absorption maxima were determined with an accuracy of \pm 0.5 nm and rounded off.

Molar absorptivity (ε). The investigated dye (7–10 mg) was dissolved in the solvent of interest (50 mL), the stock solution was diluted to the dye concentration $c_{\rm Dye} \sim 0.5 \,\mu$ M and the absorbance (A) in the absorption band maximum was measured in a 5-cm standard quartz cell. The molar absorptivity were calculated according to the Beer—Lambert law. The molar absorptivityý of each dye was independently measured three times and the average value was taken. The reproducibility for determining the molar absorptivity was within ± 2000 M⁻¹cm⁻¹.

Emission spectra and quantum yields were measured for dye concentrations $c_{\text{Dye}} \sim 0.5 \,\mu\text{M}$ in PB pH 7.4 and other solvents listed below. The excitation wavelength was 620 nm. The fluorescence measurements were done in 1-cm standard quartz cells at 25 °C using a *Varian Cary Eclipse* spectrofluorometer. The emission spectra were corrected for wavelength-dependent instrument sensitivity. Emission maxima were determined with an accuracy of $\pm 1.0 \,\text{nm}$.

Spectral characteristics of dyes in presence of BSA (dye—BSA complexes) were measured at $c_{\text{Dye}} = 0.5 \,\mu\text{M}$ and BSA concentration $c_{\text{BSA}} = 90 \,\mu\text{M}$ (6 g/L).

Quantum chemical ab initio simulations were carried out using Density Functional Theory (DFT) and B3LYP functional [36] with 6-311G(d,p) basis sets [37] within the Gaussian 09 program [38]. The vibrational frequencies were calculated and no imaginary frequencies were found for the equilibrium geometries. The NPA charges were calculated using the NBO pro6 program [39].

Populations of the conformers (c, %) was calculated for 25 °C according to the Boltzmann distribution.

2.2. Synthesis

General procedure. A mixture of quaternized or non-quaternized indolenine (2 mmol) and squaric acid or its dicyanomethylene or barbituric derivative was refluxed in 50 mL of a butanol—toluene mixture (1:1, v/v). The solvent was removed and the residue was dried in a vacuum desiccator over P_2O_5 . The product was column purified (Silica gel 60, CHCl₃–MeOH, gradient).

2-(3,3-dimethyl-2,3-dihydro-1*H*-2-indolylidenmethyl)-4-(3,3-dimethyl-3*H*-2-indoliumylmethylene)-3-oxo-1-cyclobuten-1-olate



Scheme 1. Synthesis of squaraines and norsquaraines.

(nor-SqO): Yield 75%. Dark-blue solid. ¹H NMR (400 MHz, CDCl₃, ppm): Major conformer: δ 12.84 (2H, NH, broad s), 7.27 (2H, aromortho, d, J = 7.6 Hz), 7.25 (2H, arom-meta, m, second order), 7.13 (2H, arom-ortho,d, J = 7.8 Hz), 7.08 (2H, arom-meta. ddd, J = 7.6, 7.5, 0.8 Hz), 5.48 (2H, CH, s), 1.453 (12H, CH₃, s). Minor conformer: δ 12.54 (2H, NH, broad s), 7.27 (2H, arom-ortho, d, J = 7.6 Hz), 7.25 (2H, arom-meta, m, second order), 7.13 (2H, arom-ortho, J = 7.8 Hz), 7.08 (2H, arom-meta, ddd, J = 7.6, 7.5, 0.8 Hz), 5.52 (2H, CH, s), 1.450 (12H, CH₃, s). ¹³C NMR (100 MHz, CDCl₃, ppm): Major conformer: δ 183.42 (CO), 175.77 (CCNH), 175.14 (CHC(CO)₂), 141.96 (arom, C), 139.02 (arom, C), 128.29 (arom, CH), 123.20 (arom, CH), 122.35 (arom, CH), 111.51 (arom, CH), 85.94 (CH), 48.70 (-C(CH₃)₂), 26.65 (CH₃). Minor conformer: 185.59 (CO), 180.31 (CO), 175.91(CCNH), 175.24 (CHC(CO)₂), 141.86 (arom, C), 139.02 (arom, C), 128.24 (arom, CH), 123.26 (arom, CH), 122.48 (arom, CH), 111.36 (arom, C), 86.17 (CH), 48.70 (-C(CH3)2), 26.65 (CH3). MALDI-TOF MS, m/z calcd. for $[C_{26}H_{24}N_2O_2]^+$ 396.2, found: 397.1 $[M+H]^+$, 419.0 $[M+Na]^+$, 434.9 [M+K]⁺. Anal. calcd. (%) for C₂₆H₂₄N₂O₂: C, 78.76; H, 6.10; N, 7.07. Found: C, 78.72; H, 6.12; N, 7.12.

2-(1,3,3-trimethyl-2,3-dihydro-1*H*-2-indolylidenmethyl)-4-(1,3,3-trimethyl-3*H*-2-indoliumylmethylene)-3-oxo-1-cyclobuten-

1-olate (SqO): Yield 78%. Dark-blue solid. ¹H NMR (400 MHz, DMSO- d_6 , ppm): δ 7.51 (2H, arom. H, d, J = 7.2 Hz), 7.37–7.32 (4H, arom. H, m), 7.18–7.15 (2H, arom. H, m), 5.76 (2H, C<u>H</u>, s), 3.57 (6H, NC<u>H</u>₃, s), 1.69 (12H, (C<u>H</u>₃)₂, s). ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.33 (2H, arom. H, m), 7.16–7.13 (2H, arom. H, t, J = 7.2 Hz), 7.00 (2H, arom. H, d, J = 7.6 Hz), 5.91 (2H, C<u>H</u>, s), 3.56 (6H, NC<u>H</u>₃, s), 1.77 (12H, (C<u>H</u>₃)₂, s). ¹³C NMR (100 MHz, CDCl₃, ppm): δ 182.53 (CO), 180.31 (C<u>C</u>NCH₃), 170.94 (CH<u>C</u>(CO)₂), 143.20 (arom, C), 127.95 (arom, C), 123.91 (arom, CH), 122.24 (arom, CH), 109.31 (arom, CH), 99.23(arom, CH), 86.88 (CH), 49.38 (-C(CH₃)₂), 30.73(CH₃), 27.22 (CH₃). FAB MS, m/z calcd. for [C₂₈H₂₈N₂O₂]⁺ 424.22, found: 424.3 [M⁺], 425.3 [MH⁺]. Anal. calcd. (%) for C₂₆H₂₄N₂O₂: C, 78.76; H, 6.10; N, 7.07. Found: C, 78.72; H, 6.12; N, 7.12.

2-(3,3-dimethyl-2,3-dihydro-1*H***-2-indolylidenmethyl)-4-(3,3dimethyl-3***H***-2-indoliumylmethylene)-3-(2,4,6-trioxohexahydro-5pyrymidinylidene)-1-cyclobuten-1-olate (nor-SqB): Yield 16%. Dark-green solid. ¹H NMR (400 MHz, DMSO-** *d***₆, ppm): δ 12.19 (2H, N<u>H</u>, s) 10.27 (2H, N<u>H</u>-barb, s), 7.53 (2H, arom. H, d,** *J* **= 7.6 Hz), 7.39–7.33 (4H, arom. H, m), 7.18–7.15 (2H, arom. H, m), 6.63 (2H, C<u>H</u>, s), 1.44 (12H, (C<u>H</u>₃)₂, s). FAB MS,** *m***/***z* **calcd. for [C_{30}H_{26}N_4O_4]^+ 506.20, found: 507.1 [M+H]⁺. Anal. calcd. (%) for C_{30}H_{26}N_4O_4: C, 71.13; H, 5.17; N, 11.06. Found: C, 71.12; H, 5.13; N, 11.10.**

2-(1,3,3-trimethyl-2,3-dihydro-1*H*-2-indolylidenmethyl)-4-(1,3,3-trimethyl-3*H*-2-indoliumylmethylene)-3-(2,4,6-trioxohexahydro-5-pyrymidinylidene)-1-cyclobuten-1-olate (SqB): Yield 39%. Dark-green solid. H NMR (400 MHz, DMSO-*d*₆, ppm): δ 10.02 (2H, N<u>H</u>, s), 7.55 (2H, arom. H, d, *J* = 7.2 Hz), 7.42–7.36 (4H, arom. H, m), 7.24 (2H, arom. H, t, *J* = 6.8 Hz), 6.45 (2H, C<u>H</u>, s), 3.58 (6H, NC<u>H</u>₃, s), 1.66 (12H, (C<u>H</u>₃)₂, s). FAB MS, *m*/*z* calcd. for $[C_{32}H_{30}N_4O_4]^+$ 534.23, found: 535.2 [M+H]⁺. Anal. calcd. (%) for $C_{32}H_{30}N_4O_4$: C, 71.89; H, 5.66; N, 10.48. Found: C, 71.85; H, 5.62; N, 10.50.

3-Dicyanomethylene-2-(3,3-dimethyl-2,3-dihydro-1*H***-2-in-dolylidenmethyl)-4-(3,3-dimethyl-3***H***-2-indoliumylmethylene)-1cyclobuten-1-olate** (**nor-SqCN**): Yield 35%. Dark-green solid. ¹H NMR (400 MHz, DMSO-*d*₆, ppm): δ 12.00 (2H, N<u>H</u>, s), 7.54 (2H, arom. H, d, *J* = 7.2 Hz), 7.39–7.33 (4H, arom. H, m), 7.19 (2H, arom. H, t, *J* = 6.8 Hz), 5.69 (2H, C<u>H</u>, s), 1.44 (12H, (C<u>H</u>₃)₂, s). ¹³C NMR (100 MHz, CDCl₃, ppm): δ 178.58, 176.87, 163.00, 162.57, 141.34, 139.46, 128.56, 124.36, 122.81, 117.01, 111.95, 87.16, 49.68, 46.39, 26.45. FAB MS, *m*/*z* calcd. for $[C_{29}H_{24}N_4O]^+$ 444.20, found: 445.1 [M +H]⁺. Anal. calcd. (%) for $C_{29}H_{24}N_4O$: C, 78.36; H, 5.44; N, 12.60. Found: C, 78.32; H, 5.49; N, 12.65.

3-Dicyanomethylene-2-(1,3,3-trimethyl-2,3-dihydro-1*H***-2-in-dolylidenmethyl)-4-(1,3,3-trimethyl-3***H***-2-indoliumylmethylene)-1-cyclobuten-1-olate (SqCN):** Yield 42%. Dark-green solid. ¹H NMR (400 MHz, DMSO- d_6 , ppm): δ 7.54 (2H, arom. H, d, J = 7.4 Hz), 7.47–7.37 (4H, arom. H, m), 7.32–7.18 (2H, arom. H, m), 6.30 (2H, C<u>H</u>, s), 3.61 (6H, NC<u>H</u>₃, s), 1.68 (12H, (C<u>H</u>₃)₂, s). ¹³C NMR (100 MHz, CDCl₃, ppm): δ 173.48, 173.05, 167.94, 167.46, 142.79, 142.07, 128.25, 124.86, 122.39, 119.23, 110.12, 89.21, 49.61, 40.88, 31.82, 26.77. FAB MS, m/z calcd. for $[C_{31}H_{28}N_4O]^+$ 472.23, found: 473.2 [M +H]⁺. Anal. calcd. (%) for $C_{31}H_{28}N_4O$: C, 78.79; H, 5.97; N, 11.86. Found: C, 78.81; H, 5.99; N, 11.85.

3. Results and discussion

3.1. Synthesis

The dyes in this research were chosen in such a way to compare norsquaraines *versus* squaraines of similar structure, and to investigate the effect of substitution in the central squaraine ring on the dye structures and properties. Squaraine **SqO** and its derivatives, where the squaric oxygen is substituted with a barbituric (**SqB**) and a dicyanomethylene (**SqCN**) group, are known to be synthesized by condensation of the quaternized indolenine **2a** with the squaric acid **3a** [40,41] or squarates **3b** and **3c**, respectively [34,40,42] (Scheme 1). Analogous to squaraine **SqO**, norsquaraine **nor-SqO** was obtained starting from indolenine **2b** and squaric acid **3a** [33]. Following this approach, dyes **SqO**, **SqB**, **SqCN**, **nor-SqO** and the new norsquaraines **nor-SqB** and **nor-SqCN** were synthesized. The synthesis was carried out under reflux in a butanol—toluene mixture (1:1, v/v) as proposed in Ref. [42].



Fig. 1. Internal rotation in the squaraine and norsquaraine molecules (a) and potentially possible NH and OH prototropic forms of norsquaraines (b).

3.2. Molecular structure

Although nor-squaraines formally differ from squaraines only by the presence of a hydrogen instead of an alkyl substituent at the indolenine nitrogens, their electronic and spatial structures are quite different. Due to internal rotation around the conjugated polymethine bonds (torsion angles φ_1 , φ_1 , φ_2 , and φ_2 , Fig. 1,a) the squaraine and norsquaraine molecules can potentially exist in different conformational forms. These conformers may have different planarity and different mutual orientation of both indolenine nitrogens relatively to each other (syn- and antiforms) and to the squaric oxygens. The indolenine terminal groups can exist in the *cis*- or *trans*-form (rotation around φ_1 and $\varphi_{1'}$) relatively to the central squaric moiety. Furthermore, norsquaraines as compared to squaraines may potentially form two prototropic forms (Fig. 1,b), where the hydrogen atom is located either at the indolenine nitrogen (NH form) or the squaraine ring oxygen (OH form). In both prototropic forms, one or two seven-membered quasicycles with an intramolecular H-bond can be formed.

We investigated the molecular structures of norsquaraines **nor-SqO**, **nor-SqB** and **nor-SqCN** *versus* squaraines **SqO**, **SqB** and **SqCN** using experimental (X-ray and NMR) methods and the *ab initio* calculations (B3LYP/6-311G(d,p) method). The calculated total energies (*E*), relative energies (ΔE) (differences in the energies of conformers), populations of the conformers (*c*) at 25 °C, calculated according to the Boltzmann distribution, dipole moments (μ), and electron charges on the indolenine nitrogens (q_{N1} , q_{N2}), squaric oxygen (q_O) and/or substituted squaric oxygen (q_Z) for some typical forms are given in Table S1 (Supplementary data) while the most stable conformers are shown in Table 1 and Fig. 2.

Prototropic forms. The *ab initio* calculations show that the most stable prototropic form of the norsquaraine **nor-SqO** corresponds to the centrosymmetric NH *trans,anti*-form, where both protonated indolenine nitrogens form intermolecular *H*-bonds with two different squaraine oxygens (Table 1). This form is 11.08 kcal/mol and 26.46 kcal/mol more stable than the OH forms with one protonated squaric oxygen and one protonated indolenine nitrogen, and 27.21 kcal/mol more stable compared to the di-OH form (Table S1). Such a difference in the energies suggests that the NH form is the only prototropic form to be realized. These simulations are in good agreement with our X-ray data (Fig. 3,a) showing that **nor-SqO** in crystals exists in the theoretically predicted NH *trans,anti*-form.

X-ray analysis was used also to compare the molecular structure of **nor-SqO** and **SqO** in crystals. Both crystals did not contain solvent molecules, which could affect the molecular conformation as reported in Ref. [31]. Nevertheless, the molecular structure can be distorted by intermolecular interactions in crystals. The obtained data show

existence of two halves of nor-SqO molecules (A and B) in the asymmetric part of the unit cell, which is due to both molecules being located in the special positions relatively to the center of symmetry that coincides with the squaraine ring center. The analysis of the bond lengths in SqO [43] and nor-SqO molecules evidences their mesoionic structures (Table S2, the atom numbering is given in Fig. 3a and b). The N1-C4 bond (1.343-1.354 Å) has ordinary character, which is confirmed by comparison of its value with "standard" mean value for N (3)–Csp² bond (1.355 Å) [44]. The value of the C1–O1 bond is very close to the mean value for $Csp^2 = O$ bond (1.210 Å) in **SqO** molecule and it is slightly elongated in nor-SqO due to the formation of the N-H-O intramolecular H-bond: NH form, H-O 1.87 Å, N-H-O 158° in A and H…O 1.80 Å, N-H…O 157° in B. The C1-C2 bond has ordinary character in both dye molecules: the bond length (1.450–1.473 Å) is very close to the mean value for Csp²–Csp² bond (1.455 Å). Pronounced delocalization of the electron density is observed within the C2-C3-C4 chain in both dyes: the C2-C3 and C3-C4 bond lengths (1.371-1.405 Å) are in between the mean values for the $Csp^{2} = Csp^{2}$ (1.326 Å) and $Csp^{2}-Csp^{2}$ (1.455 Å) bonds.

The analysis of *non-bonded* interactions in the **SqO** crystals reveals a moderate *intramolecular* steric repulsion between the indolenine methyl groups and squaric moiety: the H…C van der Waals radii sum [45] is 2.87 Å as compared to the shortened intramolecular contacts 2.54–2.87 Å (Fig. S1, Table S2). This repulsion is compensated by a slight twisting of the indolenine and squaric moieties relatively to each other (Fig. 3,b). Two parts of **SqO** molecule are nonequivalent due to *intermolecular* interactions in crystals causing also a moderate *non-planarity* of the molecules (the torsion angles are up to 17.1° and the dihedral angles are up to 25.3°). The participation of the methyl group of part **B** in the C–H… π hydrogen bonding with squaric moiety of the neighboring molecule (symmetry operation is 2–x, –y, 1–z; H…C 2.62 Å, C–H…C 133°) causes a more pronounced twisting as compared to part **A** (Fig. 3,b and S1).

In contrast to squaraine **SqO**, the *H*-bonding in norsquaraine **nor-SqO** stabilizes the *planar* geometry (the torsion angle do not exceed 4.9° and the dihedral angles 7.7°). It also causes the elongation of the C1–O1 bond and equalization the C2–C3 and C3–C4 bonds. Thus, the intramolecular and intermolecular steric interactions affect the planar conformation and π -conjugation of squaraine **SqO** [43] in a greater degree than that for norsquaraine **nor-SqO**.

NMR study. The ¹H NMR spectrum of **SqO** measured in DMSO- d_6 (Fig. S2,a) and CDCl₃ (Fig. S2,b) and the ¹H NMR spectrum of **nor-SqO** in DMSO- d_6 (Fig. 4,a) exhibit only one set of peaks. In contrast, the room temperature ¹H NMR and ¹³C NMR spectra of **nor-SqO** in CDCl₃ shows two sets of peaks (Fig. 4,b and c). A change in the solvents ratio (CDCl₃ : DMSO- d_6) causes a change in the relative intensities of these

Table 1

The *ab initio* calculated total energies (*E*), relative energies (ΔE), populations of the conformers (*c*) at 25 °C, dipole moments (μ), and electron charges on the indolenine nitrogens (q_{N1} , q_{N2}), squaric oxygen (q_O) and/or substituted squaric oxygen atom (q_Z). The most stable conformers are underlined.

| Dye/Form | Conformation | <i>E</i> , a.u. | ΔE , kcal/mol (c, %) | μ, D | $q_{ m N1},q_{ m N2}$ | <i>q</i> z, <i>q</i> o | Torsion angle ϕ_1, ϕ_2 $\phi_{1'}, \phi_{2'}$ |
|---|---------------|---------------------|------------------------------|-------|-----------------------|------------------------|---|
| nor-SqO NH cis,syn | | - 1265.48611 | 1.35 (9.0%) | 4.18 | -0.544 -0.544 | - 0.745 - 0.745 | 0.0, 0.0, 0.0, 0.0 |
| <u>nor-SqO</u> <u>NH</u> <u>trans,anti</u> | A HA | <u>– 1265.48827</u> | 0.00 (91.0%) | 0.00 | -0.543 -0.543 | -0.674 -0.674 | 0.0, 0.0, 0.0, 0.0 |
| SqO cis,syn(2) | AT THE | -1344.09204 | 0.90 (16.75%) | 0.49 | -0.383 -0.383 | - 0.635 - 0.690 | 0.0, 0.0, 0.0, 0.0 |
| SqO <u>trans,anti(1)</u> | AN AN | <u>-1344.09348</u> | <u>0.00 (79.04%)</u> | 0.00 | -0.383 -0.383 | -0.662 -0.662 | 0.0, 0.0, 0.0, 0.0 |
| SqO trans,syn | to the second | - 1344.09074 | 1.72 (4.08%) | 1.39 | -0.383 -0.377 | -0.660 -0.662 | 0.1, 0.1, 0.8, 0.3 |
| <u>nor-SqB</u> <u>NH cis.syn</u> | the state | <u>– 1679.23895</u> | <u>0.00 (100%)</u> | 7.85 | -0.538 -0.538 | - 0.360 - 0.707 | -1.1, 0.3, -1.1, 0.3 |
| SqB <u>cis.syn(2)</u> | | <u>– 1757.84196</u> | <u>0.00 (95%)</u> | 5.43 | -0.373 -0.373 | -0.384 -0.641 | -3.1, 6.6, -3.1, 6.7 |
| SqB cis,anti | | - 1757.83912 | 1.78 (5%) | 7.04 | -0.373 -0.373 | -0.381 -0.640 | -23.3, -24.3, 1.2, 4.8 |
| <u>nor-SqCN</u> <u>NH cis.syn</u> | | <u>– 1414.07885</u> | <u>0.00 (100%)</u> | 10.21 | -0.536 -0.536 | -0.396 -0.713 | 0.0, 0.0, 0.0, 0.0 |
| <u>SqCN</u> <u>cis,syn</u> | Anx o xn | <u>– 1492.67819</u> | <u>0.00 (100%)</u> | 10.12 | -0.372 -0.372 | -0.420 -0.650 | -14.6, -20.9, -14.6, -20.9 |

two sets (Fig. 4,d). However, the addition of an equimolar amount of one solvent to a dye dissolved in the other solvent has almost no effect on the spectra evidencing that the above phenomenon is not connected with the specific interaction between the dye and the solvent molecules but is just due to the general solvent effect.

The two sets of peaks (Fig. 4,b) in the ¹H NMR spectrum of **nor-SqO** in CDCl₃ belong to two conformers at a ratio of 2:1. Both conformers have a symmetrical structure as evidenced by the presence of only one set of peaks for the methine CH, indolenine NH and 3-(CH₃)₂ hydrogens. These conformers have intramolecular hydrogen bonds with a low field chemical shift of the NH hydrogen at $\delta = 12.84$ ppm for the *trans,anti*-form and $\delta = 12.54$ ppm for the *cis,syn*-form. Only two structures of the four possible symmetrical forms shown in Fig. 5, **nor-SqO** NH *trans, anti* and **nor-SqO** NH *cis,syn*, have intramolecular hydrogen bonds.

Also the ${}^{1}H{-}^{1}H$ NOESY supports the coexistence of these two conformers, confirming that the hydrogens of the 3-(CH₃)₂ and the methine CH groups in both conformers are in close proximity to each other and therefore cross peaks are seen in the 2D-NOESY experiment. Nor-SqO

NH *trans, anti* has a cross peak between $\delta = 1.45$ ppm (3-(CH₃)₂) and $\delta = 5.48$ ppm (C<u>*H*</u>) and **nor-SqO** NH *cis,syn* has a cross peak between $\delta = 1.45$ ppm (3-(CH₃)₂ hydrogens) and $\delta = 5.52$ ppm (C<u>*H*</u>) (Fig. S3,a).

The two conformers can be distinguished using ¹³C NMR spectroscopy (Fig. 4,c and S4). The **nor-SqO** NH *trans, anti* has C_2 symmetry (Fig. 5); each squarate oxygen forms one intra-molecular H-bond with the NH hydrogen and the chemical shift of the carbon is 183.42 ppm. In contrast, **nor-SqO** NH *cis,syn* has a σ_2 symmetry and thus the two oxygens are different. One oxygen forms two intramolecular *H*-bonds and the chemical shift of the carbon is 185.59 ppm while the other oxygen is lacking a *H*-bond and the chemical shift of the carbon is observed at a higher field, $\delta = 180.31$ ppm.

The one bond coupling constants (${}^{1}J_{N-H}$) between the ${}^{15}N$ and ${}^{1}H$ of the two conformers were extracted from the 2D experiment HMBC ${}^{15}N^{-1}H$ (Fig. S3,b) and their values are almost the same: 92 Hz and 94 Hz for the major and the minor conformers, respectively. These values confirm that in solution the hydrogen atom is also bound to the nitrogen atom as predicted by *ab initio* simulation.

When the ¹H NMR spectrum of **nor-SqO** in CDCl₃ (Fig. 4,b), is



Fig. 2. The most stable conformers of squaraine and norsquaraine molecules according to the *ab initio* simulations.

compared to the spectrum in DMSO- d_6 , at room temperature, the latter one shows only one set of broad peaks (Fig. 4,a). A series of ¹H NMR experiments at high temperatures (330–370 K) displays a narrowing of some of the peaks as we increase the temperature. For example, the line width of the methine hydrogen (CH) is decreasing from 4.86 Hz at 300 K to 0.8 Hz at 370 K (Fig. S5). These observations suggest that the two conformers, NH *trans, anti* and NH *cis,syn,* are both present in DMSO- d_6 and the equilibrium is rapid in the NMR time scale at room temperature to be above the coalesce but not fully narrow. Further, most of the ¹³C signals are broad at room temperature (Fig. S4) and the carbonyl signal is very broad, so it disappears in the spectrum noise. This equilibrium is obviously slow in the NMR time scale when the compound is dissolved in CDCl₃ to yield two sets of peaks.

Conformations. Ab initio simulations indicate that norsquaraine **nor-SqO** exists mostly in the centrosymmetric *trans, anti-*form with an

insufficient contribution of *cis,syn*-form, which is 1.35 kcal/mol less stable (Table 1 and S1, Fig. 5). According to the Boltzmann distribution the ratio between these forms at 25 °C is about 91% : 9%. This is in good agreement with data reported in Ref. [32], where *trans,anti*-form was found to be 0.5 kcal/mol less stable compared to *cis,syn*-form. The most stable *trans,anti*-form of **nor-SqO** is realized in crystals (Fig. 3,a, Table 1). However, due to the ΔE between these conformers is not pronounced it is anticipated that *trans,anti*- and *cis,syn*-forms of **nor-SqO** and **SqO** can coexist in solutions.

Similar to **nor-SqO**, the squaraine molecule **SqO** can also exist in different conformations (Table 1 and S1). The most stable conformer of **SqO** is also centrosymmetric; it has a *trans*-orientation of indolenine moieties and *anti*-orientation of indolenine nitrogens (Fig. 2). However, the calculated energy of this form is only about 0.90 kcal/mol lower compared to that of the *cis,syn*-form and 1.72 kcal/mol lower than that



Fig. 3. The nor-SqO (a) and SqO (b) structures according to the X-ray data.

of the *trans,syn*-form (Table 1). According to the Boltzmann distribution the ratio between these forms at 25 °C is about 79%: 17%: 4%, respectively. The contribution of other conformers is only minor (< 0.2%). The most stable theoretically predicted *trans, anti*(1) conformer is realized in crystals (Fig. 3,b) [43].

Planarity. Ab initio calculations show that both **SqO** and **nor-SqO** molecules are flat (φ_1 , φ_2 , φ_1 , φ_2 ; $\approx 0^\circ$). The planarity of **nor-SqO** was found to be in good agreement with X-ray data (Fig. 3,a). However the planarity of **SqO** in crystals (Fig. 3,b) is somewhat disrupted ($\varphi_1 = +2.8^\circ$, $\varphi_2 = +7.0^\circ$, $\varphi_{1'} = -7.8^\circ$, $\varphi_{2'} = -17.1^\circ$, torsion angles are designated in Fig. 1) [43]. The non-planarity of **SqO** in crystals is rather due to the above mentioned *inter*-molecular interactions in the crystal phase (Fig. S1, Table S2) than of *intra*-molecular sterical hindrance caused by the indolenine N-methyl groups.

Substitution of squaric oxygen. Similar to **nor-SqO**, barbituric (**nor-SqB**) and dicyanomethylene (**nor-SqCN**) norsquaraines according to *ab initio* calculations also exist entirely in the **NH** forms (Table 1 and **S1**). Substitution of the squaric acid oxygen with these rather bulky groups has a strong impact on the conformations of the squaraine and

norsquaraine molecules. The calculations show that **SqB**, **nor-SqB**, **SqCN**, and **nor-SqCN** exceptionally have a *cis*-orientation of indolenine moieties and a *syn*-orientation of the indolenine nitrogens (Fig. 2). These simulations are in good agreement with the ¹H NMR data exhibiting only one set of peaks for these dyes. The nitrogen atoms are located on the same side as the squaric oxygen. The exception is squaraine **SqB**, where the nitrogen atoms are on the same side as the barbituric moiety.

In contrast to oxosquaraine **SqO**, barbituric (**SqB**) and especially dicyanomethylene (**SqCN**) squaraines are less planar (see torsion angles in Table 1). At the same time, norsquaraines **nor-SqB** and **nor-SqCN** as well as **nor-SqO** are very flat.

Similar to **nor-SqO**, the intramolecular *H*-bonds between the indolenine NH hydrogens and squaric acid oxygen are realized also in the **nor-SqB** and **nor-SqCN** molecules. However, in contrast to **nor-SqO**, these *H*-bonds are formed with the same squarate oxygen atom, so that the molecules have a mirror-symmetrical structure. In the **SqB** and **nor-SqB** the barbituric oxygens form additional *H*-bonds with the methine hydrogens.



Fig. 4. ¹H NMR (*a,b,d*) and ¹³C NMR (*c*) spectra of norsquaraine nor-SqO in DMSO-*d*₆ (*a*), CDCl₃ (*b*, *c*) and CDCl₃: DMSO-*d*₆ 1:1 (*d*).



nor-SqO, NH trans.anti(2)

Fig. 5. Symmetrical forms of nor-SqO.

The intramolecular H-bonds facilitate flattening but also increase the conformational rigidity of the norsquaraine molecules (Fig. 3,a) as compared to squaraines (Fig. 3,b). Thus, the calculations show that the rotation barrier associated with torsion angle φ_1 in the **nor-SqO** molecule is about 26.1 kcal/mol, whereas in the SqO molecule it is substantially less pronounced, 18.3 kcal/mol. The rotation barriers associated with torsion angle ϕ_2 are higher (33.6 kcal/mol and 24.0 kcal/ mol. respectively).

Electron density distribution. Both squaraine and norsquaraine chromophores are neutral and have a zwitter-ionic structure. The electronic charge in all these molecules is delocalized mostly within the polymethine chain (Table S3). The calculations exhibit a negative charge on the squaric oxygen (q_0) and a less pronounced charge on the indolenine nitrogens (q_{N1}, q_{N2}) and on the carbon of the substituted squaric oxygens (q_7) (Table 1). The positive charge is mostly on the squaric "carbonyl" carbons and on the indolenine carbons at the position 2.

The most stable conformers of squaraine SqO and norsquaraine nor-**SqO** are centrosymmetric; $q_0 = q_Z$, $q_{N1} = q_{N2}$ and therefore the molecular dipole moments (μ) are about zero. While the negative charges on the oxygens (q_0, q_z) in **SqO** and **nor-SqO** are almost equal (-0.66 and -0.67, respectively), the charges on the **nor-SqO** nitrogens (q_{N1} , q_{N2}) are greatly increased as compared to SqO (-0.54 vs. -0.38).

Substitution of the oxygen atom in SqO and nor-SqO with barbituric (SqB and nor-SqB) and dicyanomethylene (SqCN and nor-SqCN) groups causes a noticeable increase in the calculated dipole moments (μ) which increases in the order of SqO (0 D) = nor-SqO (0 D) \ll SqB (5.43 D) < nor-SqB (7.85 D) < SqCN (10.12 D) \approx nor-SqCN (10.21 D).

Molecular structure of norsquaraines versus squaraines. Squaraines and norsquaraines have similar molecular structure, electronic density distribution and dipole moments but norsquaraines have an increased "local" polarity expressed by the increased electron charges on the indolenine nitrogens as compared to those of squaraines. In addition, norsquaraine molecules contain intramolecular H-bonds and therefore they are conformationally more rigid and more planar.

3.3. Spectral properties

3.3.1. Dyes free in solutions

The absorption and emission maxima (λ_{max}), molar absorptivities (ε), quantum yields ($\Phi_{\rm F}$), Stokes shifts ($\Delta v_{\rm st}$), and spectral band halfwidths ($\Delta v_{1/2}$) of norsquaraine dyes measured free in solutions and after non-covalent binding to protein (BSA), are summarized in Table 2.

The absorption spectra of norsquaraine nor-SqO and squaraine SqO, measured in CHCl₃ consist of only one long-wavelength band with a molar absorptivity $\hat{\epsilon} = 218,000 \text{ M}^{-1} \text{cm}^{-1}$ and $219,000 \text{ M}^{-1} \text{cm}^{-1}$, respectively.

Substitution of squaric oxygen in both SqO and nor-SqO molecules

with the barbituric moiety (SqB, nor-SqB) causes a 6-27 nm red-shift of the absorption and emission bands (Table 2). Introduction of the dicyanomethylene group (SqCN, nor-SqCN) results in a more pronounced red-shift (20-60 nm). Dyes SqB, nor-SqB, SqCN, nor-SqCN as compared to SqO and nor-SqO exhibit an additional, short-wavelength absorption band at 427-441 nm (barbituric) and 381-388 nm (dicyanomethylene) with molar absorptivity (ϵ) of up to 35,000 M⁻¹cm⁻¹, which makes them suitable for excitation within the blue/UV spectral range (Fig. 6).

The absorption and emission bands of norsquaraines are in the long-wavelength range of 654-694 nm and 669-712 nm, respectively. These bands are 11-28 nm red-shifted as compared to the corresponding squaraines SqO, SqB and SqCN (Table 2), which is more likely due to their more planar molecular structure. The main absorption spectra of squaraines and norsquaraines consist of an intense, longwavelength band and a less intense shoulder on the shorter-wavelength slope of the main band (Fig. 6). This sholder is typical for cyanines and squaraines and known to belong to a vibronic mode [19].

While the absorption bands of the investigated squaraines and norsquaraines measured in CHCl₃ do not exhibit any sign of aggregation, these dyes do strongly aggregate in aqueous media. The extinction coefficients measured in CHCl₃ significantly decrease in PB pH 7.4 -DMSO (5:1, v/v) and a pronounced aggregation band appears on the shorter-wavelength slope of the absorption band (Fig. 7,a-f). The aggregation band of cyanines and squaraines is known to be located very close to the vibronic shoulder [46]. Barbituric, dicyanomethylene squaraines and norsquaraines exhibit more pronounced aggregation than SqO and nor-SqO. The level of aggregation for the dye pairs SqO/ nor-SqO and SqCN/nor-SqCN seems very similar while for nor-SqB it is higher compared to SqB.

It is worth mentioning that the investigated dyes have reduced solubility not only in aqueous media but also in CHCl₃ and DMSO. As a result, we were unable to measure ¹³C NMR spectra of SqB and nor-SqB in CDCl₃ and DMSO.

The Stokes shifts (Δv_{st}) for norsquaratines and squaratines in CHCl₃ increase in the order: oxo < dicyanomethylene < barbituric (Table 2), while the molecular polarity represented by the dipole moment increases in the order: oxo < barbituric < dicyanomethylene. The increased Δv_{st} of barbituric and dicyanomethylene derivatives as compared to SqO and nor-SqO is more likely due to their more polar structure. The substantially increased Δv_{st} of SqB and nor-SqB as compared to other derivatives is supposedly due to the change in the intramolecular H-bonds between barbituric oxygens and methine hydrogens in the excited state. The intramolecular H-bonds cause a decrease in the molar absorptivities of barbituric derivatives compared to other dyes. Interestingly, the Stokes shifts (Δv_{st}) and the spectral band half-widths ($\Delta v_{1/2}$) for **nor-SqO** in CHCl₃ are higher than those of **SqO**

Table 2

Spectral characteristics of free dyes ($c_{\rm Dye} \sim 0.5 \,\mu\text{M}$) and their complexes with 90 μ M BSA at $T = 25^{\circ}\text{C}$.

| Dye | Media | λ_{\max} (Ab), nm | λ_{max} (Em), nm | ε , M ⁻¹ cm ⁻¹ | $\Phi_{\rm F}$, % | $\Delta\nu_{sb}~cm^{-1}$ | $\Delta v_{1/2}$ (Ab), cm ⁻¹ | $\Delta\nu_{1/2}$ (Em), cm $^{-1}$ |
|-----------------------------|------------------------------|---------------------------|--------------------------|--|--------------------|--------------------------|---|------------------------------------|
| | CHCl ₃ | 654 | 669 | 218,000 | 45 | 340 | 780 | 730 |
| | MeOH | 647 | 662 | 200,000 | 44 | 350 | 830 | 800 |
| н ₆₆ н | DMF | 657 | 673 | 206,000 | 45 | 360 | 820 | 770 |
| 101-340 | DMSO | 659 | 678 | 219,000 | 43 | 430 | 840 | 780 |
| | DMSO—CHCl ₃ (1:1) | 658 | 675 | 215,000 | 47 | 380 | 790 | 770 |
| | DMF-water (1:1) | 647 | 666 | n/d | 29 | 440 | 870 | 790 |
| | PB pH 7.4—DMSO (5:1) | 639 | 659 | 92,000 | 13 | 470 | 960 | 850 |
| | BSA-Complex PB pH 7.4 | 655 | 669 | 200,000 | 36 | 320 | 870 | 890 |
| | CHCl ₃ | 633 | 642 | 219,000 | 25 | 220 | 690 | 690 |
| | MeOH | 625 | 635 | n/d | 4 | 250 | 640 | 690 |
| I 69 I | DMSO | 641 | 652 | n/d | 15 | 260 | 660 | 650 |
| SqO | PB pH 7.4—DMSO (5:1) | 624 | 635 | n/d | 3 | 280 | 750 | 740 |
| | BSA-Complex PB pH 7.4 | 639 | 645 | 203,000 | 44 | 150 | 670 | 670 |
| Î | CHCl ₃ | 669 | 693 | 63,000 | 48 | 520 | 980 | 830 |
| HN NH | | 441 | | 16,000 | | | | |
| $A^{\circ} I^{\circ} X_{A}$ | PB pH 7.4—DMSO (5:1) | 673 | n/a ^a | n/a ^a | < 1 | n/a ^a | n/a ^a | n/a ^a |
| | BSA-Complex PB pH 7.4 | 662 | 683 | 23,000 | 9 | 460 | 2270 | 980 |
| nor-SaB | | 460 | | 15,000 | | | | |
| o o | CHCl | 641 | 669 | 103 000 | 4 | 650 | 1110 | 1380 |
| HN NH | Grog | 427 | 00) | 27 000 | | 000 | 1110 | 1000 |
| at of the | PB pH 7 4—DMSO (5.1) | 618 | 646 | 86,000 | < 1 | 700 | 1410 | 1620 |
| | | 426 | 010 | 23,000 | ~ 1 | 700 | 1110 | 1020 |
| i d⊎ i SaD | BSA-Complex PB pH 7 4 | 633 | 657 | 91,000 | 34 | 580 | 1180 | 1540 |
| зчь | bon complex i b pii / | 422 | 007 | 13,000 | 01 | 500 | 1100 | 1010 |
| NC_CN | CHCl | 694 | 712 | 120.000 | 28 | 360 | 750 | 730 |
| ALXA | Grog | 388 | /12 | 33,000 | 20 | 500 | 700 | 700 |
| | PB pH 7 4—DMSO (5.1) | 698 | n/a ^a | 28,000 | < 1 | n/a ^a | n/a ^a | n/a ^a |
| nor-SqCN | | 389 | ii/u | 19,000 | ~ 1 | ii/ u | ii) u | ii/ u |
| • | BSA-Complex PB pH 7 4 | 690 | 704 | 63,000 | 13 | 290 | 1730 | 830 |
| | borr dompren i b pri / i i | 394 | , | 32,000 | 10 | 200 | 1,00 | 000 |
| NC_CN | CHCla | 683 | 701 | 196 000 | 37 | 380 | 800 | 830 |
| $\gamma = 1 - 1 \gamma$ | 3 | 381 | | 35.000 | 57 | | | |
| | PB pH 7 4—DMSO (5:1) | 660 | 678 | 70,000 | < 1 | 400 | 2030 | 1510 |
| SqCN | 12 pri / 1 Dillo (0.1) | 384 | 0,0 | 16,000 | ~ 1 | | 2000 | 1010 |
| - | BSA-Complex PB pH 7 4 | 682 | 702 | 162 000 | 50 | 420 | 830 | 850 |
| | Dort complex 1 D pi1 7.4 | 380 | 704 | 36,000 | 50 | 120 | 000 | 000 |
| | | 560 | | 50,000 | | | | |

^a Not available because of the low solubility and/or high aggregation.



Fig. 6. Absorption and emission spectra of nor-SqO, nor-SqB and nor-SqCN measured in CHCl₃.

while for the dye pairs **nor-SqB/SqB** and **nor-SqCN/SqCN** an opposite correlation is observed. For norsquaraines $\Delta \nu_{1/2}(Ab) > \Delta \nu_{1/2}(Em)$, while for **SqO** they are almost the same and for **SqB** and **SqCN** $\Delta \nu_{1/2}(Ab) < \Delta \nu_{1/2}(Em)$. Both $\Delta \nu_{st}$ and $\Delta \nu_{1/2}$ increase, when CHCl₃ is replaced with water, which is typical for polar dyes.

The quantum yields (Φ_F) of norsquaraines nor-SqO and nor-SqB measured in CHCl₃ are 1.8 and 12 fold higher compared to SqO and SqB while for nor-SqCN the Φ_F is surprisingly 1.3 times lower.

The quantum yields of *squaraine* dyes are known to be strongly affected by the solvent polarity [47,48]. The Φ_F of **SqO**, **SqB** and **SqCN** substantially increases, when a polar solvent such as water, alcohol or

DMSO is substituted with a less polar solvent such as CHCl₃ (Table 2). For instance, the quantum yield of **SqO** in CHCl₃ ($\Phi_F = 25\%$) is 8 fold decreased in PB pH 7.4 — DMSO (5:1) ($\Phi_F = 3\%$) and 6 fold in MeOH ($\Phi_F = 4\%$).

In contrast, the solvent polarity has a marginal effect on the quantum yield of norsquaraines. The quantum yield of nor-SqO in DMSO ($\Phi_F = 43\%$), DMF ($\Phi_F = 45\%$) and MeOH ($\Phi_F = 44\%$) is almost the same as in the less polar solvent $CHCl_3$ ($\Phi_F = 45\%$) and DMSO—CHCl₃ (1:1, v/v) ($\Phi_F = 47\%$) (Table 2). At the same time, the $\Phi_{\rm F}$ of **nor-SqO** measured in DMSO or DMF decreases upon dilution with water. Thus, the $\Phi_{\rm F}$ is 43% in DMSO and 45% in DMF while it is only 29% in water-DMSO or water-DMF (1:1, v/v) and 13% in PB pH 7.4—DMSO (5:1). The quantum yield decrease in aqueous media can be attributed to the aggregation of nor-SqO molecules. This is evidenced by the increase in the aggregation band at the short-wavelength slope of the main absorption band (around 600 nm) and the decrease in the main band (around 650 nm) (Fig. 8,a). Furthermore, upon increasing the water ratio to more then about 40:1 (v/v), an additional aggregation band appears at the long-wavelength slope of the main absorption band (at about 700 nm).

In case of **SqCN**, **nor-SqCN** and especially **SqB** and **nor-SqB**, the aggregation noticeably increases causing a dramatic decrease in the quantum yields: $\Phi_F < 1\%$ in PB pH 7.4 — DMSO (5:1, v/v) (Table 2).

As shown above, the ¹H NMR and ¹³C NMR spectra of **nor-SqO**, measured in CDCl₃ as compared to DMSO-*d*₆, contain two groups of signals, which is due to coexistence of two conformers. However, the absorption spectra measured in CHCl₃, MeOH, DMF, and DMSO contain only one band (Fig. 8,b). This suggests that the above conformers have similar spectral properties. The solvent causes a slight red-shift in the



Fig. 7. Absorption spectra of norsquaraines and squaraines in CHCl3 and PB pH 7.4 in presence of DMSO (5:1, v/v) and 90 µM BSA.



Fig. 8. Absorption spectra of nor-SqO ($c_{dye} = 0.5 \,\mu$ M) in PB 7.4 — DMSO mixtures (*a*) and in different solvents (*b*).

nor-SqO spectra and this shift increases in the order of PB–DMSO (5:1) < MeOH < $CHCl_3$ < DMF < DMSO while the molar absorptivity remain almost unchanged (Table 2).

3.3.2. Non-covalent complexation with BSA

The absorption and emission spectra and the quantum yields of *squaraines* measured in aqueous buffers are known to be substantially affected by the presence of proteins in particular bovine serum albumin (BSA), which is due to the formation of dye—BSA complexes: there is a moderate red-shift and noticeable increase in the quantum yield [42,49]. These effects have been explained by decrease in the environment polarity [46,50] and increase in the conformational rigidity [51] of dye molecules, when they move from water to protein phase.

We found that the spectral properties of *norsquaraines* also do change in the presence of BSA (Fig. S6,a,b). For both investigated dye

classes, squaraines and norsquaraines, measured at $c_{\rm Dye} = 0.5 \,\mu$ M these characteristics reach "saturation" when the concentration of BSA reaches $c_{\rm BSA} \sim 90 \,\mu$ M and then do not change anymore. Therefore, the spectral characteristics of the dye—BSA complexes presented in Table 2 were measured at $c_{\rm BSA} = 90 \,\mu$ M.

When compared to PB pH 7.4 upon complexation with BSA squaraines SqO and SqB — DMSO (5:1, v/v) exhibit an about 15/10 nm redshift of the absorption/emission bands and SqCN shows a 22/24 nm red-shift. Norsquaraine nor-SqO in presence of BSA also shows about the same red-shift (16/10 nm) as SqO while nor-SqB and nor-SqCN exhibit ~10 nm blue-shift, which is more likely due to their higher polarity.

In general, the Stokes shifts of squaraines and norsquaraines (except **SqCN**) in aqueous BSA are shorter compared to CHCl₃, which is most likely due to the more viscous protein environment.

The fluorescence intensities of *squaraines* measured in aqueous media (PB pH 7.4 — DMSO, 5:1, v/v) dramatically increase in presence of BSA (Fig. S6,b). The quantum yields (Φ_F) increase by factors of ~15, > 34, and > 50, respectively (Table 2). It was difficult to determine the accurate values of these increases for **SqB** and **SqCN** because of the extremely low Φ_F in PB pH 7.4 — DMSO (5:1, v/v). Importantly, the Φ_F of *norsquaraine* **nor-SqO** increases only by factor of 2.8 (as compared to 15 fold for **SqO**), while in the absence of BSA it is higher compared to **SqO** (Table 2, Fig. S6,a). Thus, **nor-SqO** is less sensitive to BSA than **SqO**, which can be attributed to its more rigid structure. **Nor-SqB** and **nor-SqCN** demonstrate a substantially more pronounced increases in the Φ_F (> 9 and > 13), which is most likely due to a higher polar structure of these molecules (higher dipole moments, Table 1).

The increase in the quantum yields of the investigated squaraines and norsquaraines in the presence of BSA is supposedly not only due to the decrease in the environment *polarity* and increase in the conformational *rigidity* of these dye molecules but also because of the decrease in their *aggregation*. The decrease in aggregation can be seen from the decrease of the aggregation band (Fig. 7a–f) and increase in the extinction coefficients (Table 2).

The quantum yields of BSA complexes of norsquaraines are lower than those for squaraines: 36/44% for **nor-SqO/SqO**, 9/34% for **nor-SqB/SqB**, and 13/50% for **nor-SqCN/SqCN**. This correlation is also in good agreement with the aggregation ability of these dyes: While **nor-SqO** and **SqO** in BSA exhibit an insufficient tendency to aggregate (Fig. 7a and b), **nor-SqCN** (Fig. 7,e) and especially **nor-SqB** (Fig. 7,c) demonstrate a much more pronounced aggregation tendency compared to **SqCN** (Fig. 7,f) and **SqB** (Fig. 7,d).

4. Conclusions

In summary, we have synthesized the indolenine based norsquaraine dye nor-SqO and its novel barbituric (nor-SqB) and dicyanomethylene (nor-SqCN) derivatives; investigated their molecular structures, spectral properties and complexation with BSA versus those of conventional squaraines of similar structures (SqO, SqB and SqCN). Due to the intramolecular H-bonds, norsquaraine molecules are more planar and rigid compared to squaraines, and have therefore higher fluorescence quantum yields in solutions. In solution norsquaraine nor-SqO exists as a mixture of two conformers, where the non-polar centrosymmetric one is dominant. In contrast, barbituric and dicyanomethylene squaraines and norsquaraines are highly polar. Due to the more rigid structure and lower polarity, the quantum yield of nor-SqO is less sensitive to the presence of protein (BSA) than SqO. This suggests that nor-SqO might be useful for biomedical assays based on a specific interaction of the dye-biomolecule conjugates with biological counterparts (antibody-antigen, biotin-avidin, complimentary oligonucleotides, and receptor-ligand interactions) in heterogeneous biological media, where the presence of non-targeting proteins and other large-molecular-weight biomolecules could affect the assay. At the same time, highly polar norsquaraines **nor-SqB** and **nor-SqCN** as well as squaraines SqO, SqB and SqCN might be promising fluorescent probes for sensing applications in proteins and other high-molecularweight species.

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Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.dyepig.2018.12.007.

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