

S-Ribosylhomocysteine analogues with the carbon-5 and sulfur atoms replaced by a vinyl or (fluoro)vinyl unit

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Abstract—Treatment of the protected ribose or xylose 5-aldehyde with sulfonyl-stabilized fluorophosphonate gave (fluoro)vinyl sulfones. Stannyldesulfonylation followed by iododestannylation afforded 5,6-dideoxy-6-fluoro-6-iodo-D-ribo or xylo-hex-5-enofuranoses. Coupling of the hexenofuranoses with alkylzinc bromides gave 10-carbon ribosyl- and xylosylhomocysteine analogues incorporating a fluoroalkene. The fluoroalkenyl and alkenyl analogues were evaluated for inhibition of *Bacillus subtilis* S-ribosylhomocysteinase (LuxS). One of the compounds, 3,5,6-trideoxy-6-fluoro-D-erythro-hex-5-enofuranose, acted as a competitive inhibitor of moderate potency ($K_i = 96 \mu\text{M}$).

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1. Introduction

S-Adenosyl-L-homocysteine (SAH) is a byproduct of many methyltransferase reactions and a potent inhibitor of the methyltransferases. In eukaryotes and some bacteria, detoxification of SAH is mediated by SAH hydrolase (EC 3.3.1.1), which effects hydrolytic cleavage of SAH to L-homocysteine (Hcy) and adenosine (Fig. 1).¹ Hcy appears to be a risk factor for coronary artery diseases.² Alternatively, most bacteria utilize enzyme 5'-methylthioadenosine (MTA)/SAH nucleosidase (EC 3.2.2.9) to irreversibly cleave SAH yielding adenine and S-ribosyl-L-homocysteine (SRH).³ The SRH is then converted to Hcy and 2,4-dihydroxy-2,3-pentadione (DPD) by a metalloenzyme S-ribosylhomocysteinase (LuxS).⁴ DPD⁵ spontaneously cyclizes and complexes with borate to form a furanosyl borate diester, which acts as a type 2 autoinducer for bacterial interspecies quorum sensing.⁶ Since quorum sensing regulates many bacterial behaviors such as virulence and biofilm formation, LuxS and other proteins involved in quorum sensing have been proposed as attractive targets for novel antibacterial drug design.⁷

Several substrate analogues of SRH (e.g., **1** and **2**) showed submicromolar inhibition of LuxS.^{4e,h}

We have previously observed that SAH hydrolase is capable of the addition of water across 5',6' isolated double bond of adenosine analogues **3** and **4** (Fig. 2).^{1c,8} The resulting adduct (or its derivative) caused covalent modification and inactivation^{8b} of the enzyme, a process which required the catalytic activity of the enzyme. Since LuxS catalyzes a similar reaction as SAH hydrolase (i.e., overall elimination of Hcy), we designed analogues of SRH with the vinyl or halovinyl moieties incorporated in place of the carbon-5 and sulfur atoms (e.g., **5**). We envisaged that these ribosyl analogues might serve as mechanistic probes to study the mechanism of action of LuxS and evaluate the similarities between SAH hydrolase and LuxS. As mentioned above, LuxS inhibitors may provide a novel class of antibacterial agents. We now describe the syntheses of SRH analogues with the carbon-5 and sulfur atoms replaced by vinyl or (6-fluoro)vinyl motifs and discuss their interactions with LuxS enzyme.

2. Chemistry

Our initial plan to prepare compound **5** and its congeners is illustrated in Scheme 1. Treatment of the diacetone

Keywords: LuxS enzyme, Negishi coupling; S-Ribosylhomocysteine; Vinyl fluorides; Vinyl stannanes.

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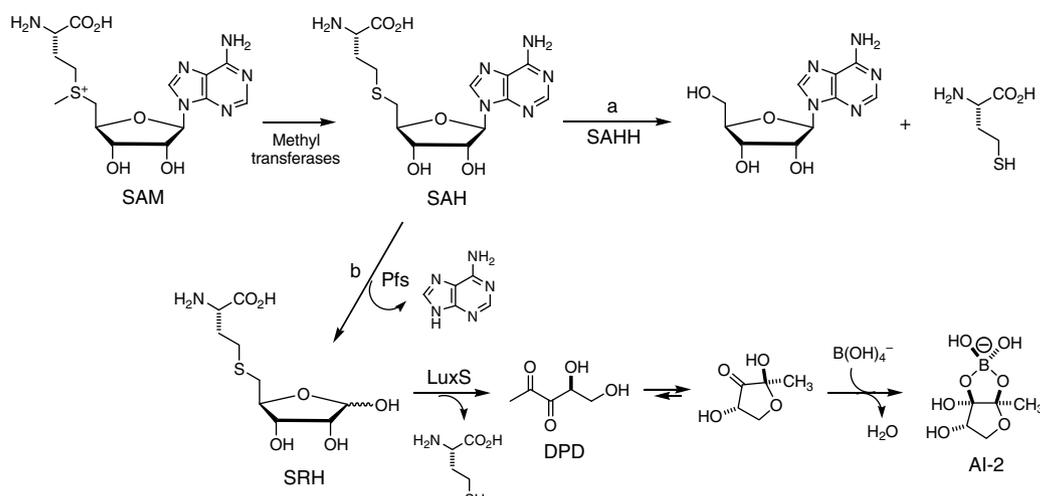


Figure 1. Reaction pathways for SAH detoxification in eukaryotes (a) and the majority of bacteria (b). The latter is also utilized by the bacteria to produce the type 2 autoinducer.

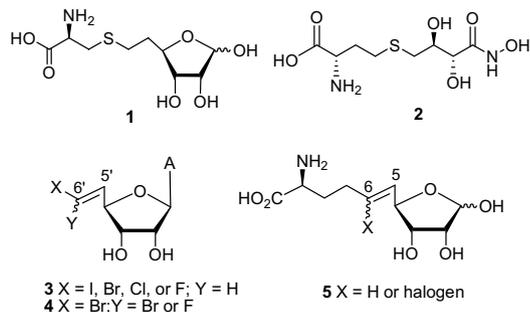
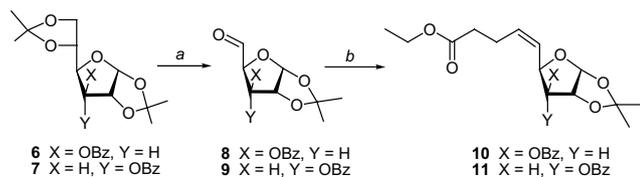


Figure 2. Inhibitors of LuxS enzyme (**1** and **2**).^{4c,h} 5'-Deoxy-5'-(halomethylene)adenosine analogues (**3** and **4**) which serve as suicide substrates for SAH hydrolase⁸ and targeted SRH analogues (**5**) in which the sulfur and C5 atoms are replaced by a vinyl unit.

3-*O*-benzoylglucose **6** or allose **7** with periodic acid selectively removed the 5,6-*O*-isopropylidene group. Subsequent oxidative cleavage of the exposed vicinal diol^{9a} gave the corresponding 5-aldehydes **8** and **9**, respectively, in high yields (Scheme 1). Wittig olefination of aldehyde **8** with the ylide derived from commercially available [4-ethoxy-4-oxobutyltriphenylphosphonium bromide provided a complex mixture of products. Column chromatography yielded protected 5,6,7,8-tetra-deoxy- α -D-xylo-non-5(*Z*)-enofuranuronate **10** (18% yield). The stereochemistry was assigned as *Z*, based on the magnitude of the coupling constants for olefinic protons (³*J*_{5,6} = 11.1 Hz), and literature precedence for the Wittig condensations of aliphatic aldehydes with

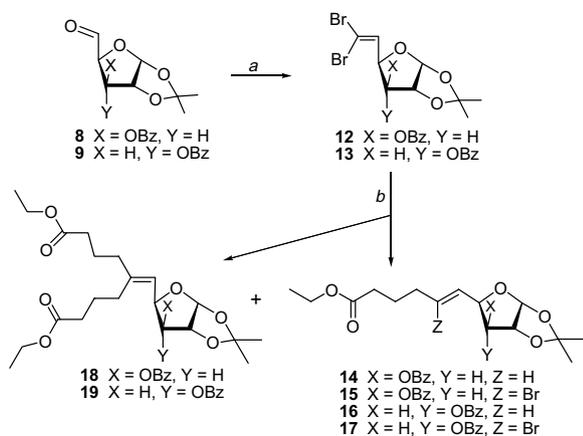


Scheme 1. Reagent: (a) H₅IO₆/EtOAc; (b) Ph₃PCH₂CH₂CH₂CO₂Et/HMDS/THF.

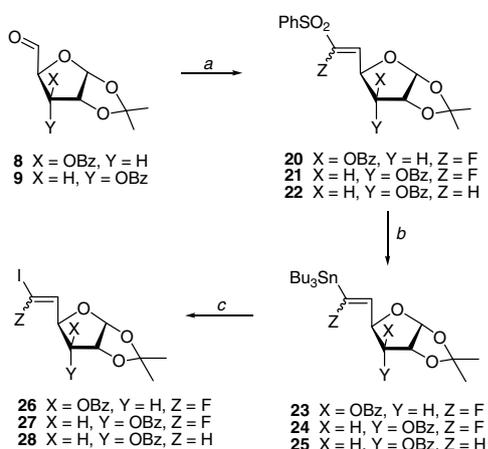
the non-stabilized ylides.^{9b} Similarly, Wittig-treatment of *ribo* 5-aldehyde **9** gave **11**; a nine-carbon analogue of SRH. Unfortunately, our attempts to add bromine (CH₂Cl₂/0 °C) across the double bond of **10** or **11** (as well as **16**) produced a complex mixture which did not give the desired SRH analogues of type **5** bearing a (6-bromo)vinyl unit when treated with DBU.¹⁰

In an alternative approach, we attempted a synthesis of 6-bromoalkenyl analogues **5** (X = Br) via Pd-catalyzed monoalkylation^{11–13} of the readily available (*gem*-dibromo)vinyl sugar precursors (e.g., **12** and **13**) with the corresponding alkylzinc reagents. Thus, dibromolefination of *xylo* 5-aldehyde **8** by the Corey–Fuchs procedure¹⁴ gave 5-(dibromomethylene)-5-deoxyxylose **12** (81% from **6**; Scheme 2). Analogous treatment of the *ribo* 5-aldehyde **9** afforded **13**.¹⁵ Treatment of **12** with 3 equiv of 4-ethoxy-4-oxobutylzinc bromide in the presence of Pd(PPh₃)₄ at 55 °C gave monoalkylated 5,6,7,8,9-penta-deoxy- α -D-xylo-dec-5(*E*)-enofuranuronate **14** (18%, ³*J*_{5,6} = 15.4 Hz) and dialkylated **18** (48%) products, but did not yield the desired 6-bromoalkenyl product **15**. Analogous Negishi coupling of 5-(dibromomethylene)-5-deoxyribose **13** afforded only dialkylated product **19** (54%). Changing catalyst [(Pd₂(dba)₃], solvent (THF), reaction temperature (rt to 60 °C) as well as adding additives (CuI, tricyclohexylphosphine) did not lead to the formation of **15** or **17** but instead produced dialkylated byproducts **18** and **19** (3–49%) in agreement with a recent report.^{13b}

We next explored stereoselective coupling of the *gem*-dihalovinyl sugars containing two different halogens. We chose 5-deoxy-5-(fluoroiodomethylene) hexenofuranoses **26** and **27** because the iodo and fluoro substituents are known to have quite different reactivity towards oxidative-addition in Pd-mediated couplings.^{11b,16,17} The precursors **26** and **27** were prepared employing McCarthy's stannyldesulfonylation methodology.^{18,19} Thus, treatment of the *xylo* aldehyde **8** with the enolate generated from the sulfonyl-stabilized fluorophosphonate²⁰

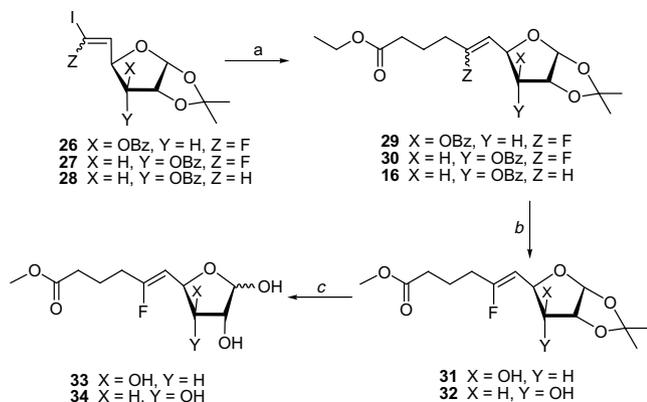


Scheme 2. Reagents: (a) $\text{PPh}_3/\text{CBr}_4$; (b) $\text{BrZn}(\text{CH}_2)_3\text{CO}_2\text{Et}/\text{Pd}(\text{PPh}_3)_4/\text{benzene}/\Delta$.



Scheme 3. Reagents and conditions: (a) $\text{PhSO}_2\text{CHFPO}(\text{OEt})_2$ or $\text{PhSO}_2\text{CH}_2\text{PO}(\text{OEt})_2/\text{LHMDS}/\text{THF}/-78^\circ\text{C}$; (b) $\text{Bu}_3\text{SnH}/\text{AIBN}/\text{toluene}/85^\circ\text{C}$; (c) $\text{NIS}/\text{CH}_2\text{Cl}_2$.

gave (fluoro)vinyl sulfones **20** (*E/Z*, 7:3; 76%; **Scheme 3**). The stereoselective radical-mediated stannyldesulfonylation of **20** with Bu_3SnH produced (fluoro)vinyl stannanes **23** (*E/Z*, 7:3; 95%). Iodostannylation of **23** with



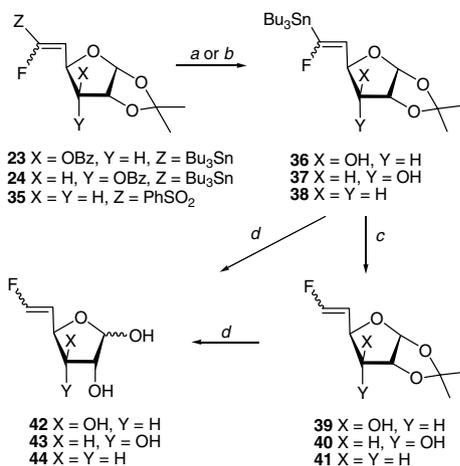
Scheme 4. Reagents: (a) $\text{BrZn}(\text{CH}_2)_3\text{CO}_2\text{Et}/\text{Pd}(\text{PPh}_3)_4/\text{benzene}/\Delta$; (b) NH_3/MeOH ; (c) $\text{TFA}/\text{H}_2\text{O}$.

N-iodosuccinimide (NIS) quantitatively afforded 6-fluoro-6-iodo-xylo-hex-5-enofuranoses **26** with retention of the *E/Z* configuration. The *ribo* analogue **27** (*E/Z*, 3:2; 57% overall yield from **9**) was similarly prepared. The isomeric ratio for the fluorinated sugars could be distinguished by the magnitude of the $^3J_{\text{F-H5}}$ in the NMR spectra.

Pd-mediated cross-coupling of the *xylo* analogue **26** (*E/Z*, 4:1) with 2 equiv of 4-ethoxy-4-oxobutylzinc bromide resulted in selective consumption of (*E*)-**26** to afford (*Z*)-**29** in 61% isolated yield or 76% based on consumption of (*E*)-**26** (**Scheme 4**). A small amount of (*E*)-**29** was also isolated, although monocoupling with *gem*-dihalovinyl substrates is considered to be *trans* selective.^{12,13b,16} Similar monoalkylation of the *ribo* analogue **27** (*E/Z*, 3:2) with $\text{BrZn}(\text{CH}_2)_3\text{COOEt}$ yielded (*Z*)-**30** [54%, 90% based on the conversion of (*E*)-**27**]²¹ and (*E*)-**30** [12%, 30% from (*Z*)-**27**]. Coupling of the (iodo)vinyl (*E*)-**28**, prepared as depicted in **Scheme 3** (**9** → **22** → **25** → **28**), with $\text{BrZn}(\text{CH}_2)_3\text{COOEt}$ gave the unfluorinated analogue (*E*)-**16** (56%) with the retention of configuration. Treatment of (*Z*)-**29** with NH_3/MeOH removed the benzoyl group and converted the ethyl ester into a methyl ester (*Z*)-**31** (74%). Subsequent removal of the isopropylidene group with aqueous trifluoroacetic acid (TFA) at 0°C gave (*Z*)-**33** (61%; α/β , 1:1). Successive treatment of (*Z*)-**30** with NH_3/MeOH followed by $\text{TFA}/\text{H}_2\text{O}$ gave (*Z*)-**34** (52% overall yield; α/β , 3:7); a 10-carbon 6-fluoroalkenyl analogue of SRH.

The 5,6-dideoxy-6-fluorohex-5-enofuranoses **42** and **43**, deprotected analogues of **3** ($\text{X} = \text{F}$), were synthesized by protiodestannylation of the (fluoro)vinyl stannanes **23** and **24**. Thus, treatment of **23** (*E/Z*, 7:3) with NH_3/MeOH at 25°C resulted in the removal of 3-*O*-benzoyl group to give **36** (**Scheme 5**). However, prolonged heating of **36** (or **23**) with NH_3/MeOH at 65°C for 48 h effected protiodestannylation to yield a separable mixture of (*E*)-**39** (29%) and (*Z*)-**39** (48%). Treatment of (*E*)-**39** with $\text{TFA}/\text{H}_2\text{O}$ at 0°C gave (*E*)-**42** (α/β , ~1:1). Analogous debenzoylation and protiodestannylation of **24** (*E/Z*, 1:1) with NH_3/MeOH yielded (*E*)-**40** (32%) and (*Z*)-**40** (26%). Acid-catalyzed removal of the isopropylidene group in (*E*)-**40** gave 5,6-dideoxy-6-fluoro-*D*-ribo-hex-5-enofuranose (*E*)-**43** (67%; α/β , ~1:4). Alternatively, concomitant protiodestannylation and removal of acetone unit in **36** or **37** with TFA also afforded **42** and **43**.

The 3,5,6-trideoxy 6-fluorohex-5-enofuranose **44**, which lacks a hydroxyl group at C3 and therefore cannot participate in the second enolization step of the LuxS-catalyzed reaction,^{4b} was also prepared. Thus, oxidation of the diacetone 3-deoxyglucose²² with H_5IO_6 ^{9a} and in situ treatment of the rather unstable 3-deoxyribose 5-aldehyde with the enolate generated from the sulfonyl-stabilized fluorophosphonate²⁰ gave the (fluoro)vinyl sulfones **35** (48%; *E/Z*, 2:1). Subjection of **35** to the stannyldesulfonylation/protiodestannylation^{18b} sequence afforded 3-deoxy (6-fluoro)vinyl sugar **41**, which was deprotected to yield **44** (12% from **35**). Alter-



Scheme 5. Reagents and conditions: (a) NH₃/MeOH/25 °C; (b) Bu₃SnH/AIBN/toluene/85 °C; (c) NH₃/MeOH/65 °C or NH₃/MeOH/CsF/65 °C; (d) TFA/H₂O.

natively, treatment of vinyl stannanes **38** with TFA effected simultaneous protiodestannylation and removal of the acetone unit to give **44** (23% from **35**; *E/Z*, ~1:3, α/β ~1:4).

3. Inhibition of LuxS

The (6-fluoro)vinyl *xylo*- (**42**) and *ribo*-hexofuranoses (**43**) and their 3-deoxy analogue **44** as well as (6-fluoro)vinyl *xylo*- and *ribo*-decofuranoses (**33** and **34**) were evaluated^{4h} as potential inhibitors of *Bacillus subtilis* *S*-ribosylhomocysteine (LuxS). Compound **44** exhibited competitive inhibition of moderate potency, with a K_I value of $96 \pm 3 \mu\text{M}$ (Fig. 3). None of the other compounds showed significant inhibition under the assay conditions.

4. Summary and conclusions

We have developed synthesis of six-, nine-, and 10-carbon analogues of ribosyl- and xylosylhomocysteines in which the carbon-5 and sulfur atoms are replaced by a vinyl or (fluoro)vinyl unit. These fluoroalkenyl and alkenyl analogues of SRH were synthesized employing either the Wittig reaction or Pd-catalyzed coupling routes. They were evaluated against *B. subtilis* *S*-ribosylhomocysteine (LuxS). Only 3,5,6-trideoxy-6-fluoro-*D*-erythro-hex-5-enofuranose acted as competitive inhibitor of moderate potency with $K_I = 96 \mu\text{M}$.

5. Experimental

¹H (Me₄Si) NMR spectra were determined with solution in CDCl₃ at 400 or 600 MHz, ¹³C (Me₄Si) at 100.6 MHz and ¹⁹F (CFCl₃) at 376.5 MHz unless otherwise noted. Mass spectra (MS) were obtained by atmospheric pressure chemical ionization (APCI) and HRMS by electron impact techniques unless otherwise noted. Reagent grade chemicals were used as received. Solvents were

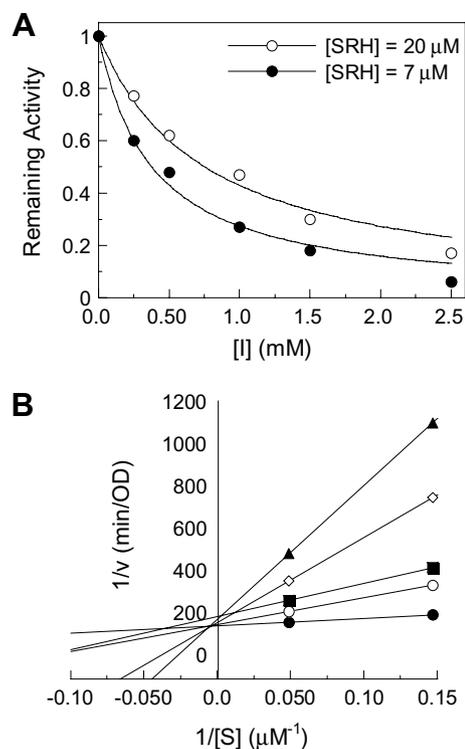


Figure 3. Inhibition of Co²⁺-substituted *B. subtilis* LuxS by compound **44**. (A) Plot of remaining LuxS activity (relative to that in the absence of inhibitor) as a function of [I]. (B) Lineweaver–Burke plot of data from part A to show the competitive inhibition mode.

dried by reflux over and distillation from CaH₂ under an argon atmosphere except THF (K/benzophenone). TLC was performed on Merck kieselgel 60-F₂₅₄ with MeOH/CHCl₃ (1:9) and EtOAc/MeOH (95:5) as developing systems, and products were detected with 254 nm light or by visualization with Ce(SO₄)₂/(NH₄)₆Mo₇O₂₄·4H₂O/H₂SO₄/H₂O reagent. Merck kieselgel 60 (230–400 mesh) was used for column chromatography. Elemental analyses were determined by Galbraith Laboratories, Knoxville, TN.

5.1. Ethyl 3-*O*-benzoyl-5,6,7,8-tetradeoxy-1,2-*O*-isopropylidene- α -*D*-xylo-non-5(*Z*)-enofuranuronate (**10**)

Step (a). H₅IO₆ (150 mg, 0.66 mmol) was added to a stirred solution of **6** (200 mg, 0.55 mmol) in dried EtOAc at ambient temperature. A precipitate appeared within the first 5 min and the resulting solution was stirred for 90 min. The precipitate was filtered off and was washed with EtOAc (2 × 5 mL). The combined organic layer was washed with NaHCO₃/H₂O (10 mL), NaCl/H₂O (10 mL), dried (Na₂SO₄), and evaporated to yield 3-*O*-benzoyl-1,2-*O*-isopropylidene- α -*D*-xylo-pentodialdo-1,4-furanose (**8**; 160 mg, 95%; approximately 90% pure based on ¹H NMR): ¹H NMR δ 1.35 and 1.48 (2 × s, 2 × 3, 2 × CH₃), 4.76 (d, $J_{4-3} = 3.2$ Hz, 1, H₄), 4.88 (d, $J_{2-1} = 3.1$ Hz, 1, H₂), 5.77 (d, $J_{1-2} = 3.1$ Hz, 1, H₁), 6.18 (d, $J_{3-4} = 3.3$ Hz, 1, H₃), 7.42–8.01 (m, 5, Ar), 9.78 (s, 1, H₅). **Step (b).** LHMDS (1 M/THF; 0.69 mL, 0.69 mmol) was added dropwise to a stirred solution of Ph₃PCH₂CH₂CH₂CO₂Et/Br (314 mg,

0.69 mmol) in anhydrous THF (4 mL) in a flame-dried flask under N₂ at ambient temperature. After 15 min, a solution of the crude, preferentially freshly prepared, aldehyde **8** (160 mg of the material from Step (a)) in THF (2 mL) was added via syringe and stirring was continued overnight. EtOAc (30 mL) and NaHCO₃/H₂O (10 mL) were added and the separated organic was washed with NaCl/H₂O (10 mL), dried (Na₂SO₄), and evaporated. Column chromatography (10 → 30% hexanes/EtOAc) gave **10** (39 mg, 18%) as an oil: ¹H NMR δ 1.24 (t, *J* = 7.2 Hz, 3, CH₃), 1.37 and 1.62 (2× s, 2× 3, 2× CH₃), 2.41 (t, *J*_{8-7/7'} = 6.9 Hz, 2, H8/8'), 2.50 ('q', *J*_{7-6/8/8'} = 7.3 Hz, 2, H7/7'), 4.15 (q, *J* = 7.1 Hz, 2, CH₂), 4.71 (d, *J*₂₋₁ = 3.8 Hz, 1, H2), 5.22 (dd, *J*₄₋₅ = 7.7 Hz, *J*₄₋₃ = 2.8 Hz, 1, H4), 5.46 (d, *J*₃₋₄ = 2.8 Hz, H3), 5.58 (dd, *J*₅₋₆ = 11.1 Hz, *J*₅₋₄ = 7.9 Hz, 1, H5), 5.68 (dt, *J*₆₋₅ = 11.1 Hz, *J*_{6-7/7'} = 7.1 Hz, 1, H6), 6.05 (d, *J*₁₋₂ = 3.7 Hz, 1, H1), 7.48–8.04, (m, 5, Ar); ¹³C NMR δ 14.62 (CH₃), 24.08 (C7), 26.62 and 27.19 (CMe₂), 34.23 (C8), 60.90 (CH₂), 75.54 (C2), 78.59 (C3), 84.20 (C4), 105.02 (C1), 112.47 (CMe₂), 123.90 (C6), 128.91 (Bz), 129.78 (Bz), 130.15 (Bz), 133.85 (Bz), 134.39 (C5), 165.64 (Bz), 173.09 (C9); MS *m/z* 391 (100, MH⁺), HRMS (AP-ESI) *m/z* calcd for C₂₁H₂₆O₇Li (MLi⁺) 397.1839; found: 397.1833.

5.2. Ethyl 3-*O*-benzoyl-5,6,7,8-tetra-deoxy-1,2-*O*-isopropylidene- α -*D*-ribo-non-5(*Z*)-enofuranuronate (**11**)

Step (a). Oxidation of **7** (200 mg, 0.55 mmol) with H₅IO₆ (150 mg, 0.66 mmol) as described for **10**, gave 3-*O*-benzoyl-1,2-*O*-isopropylidene- α -*D*-ribo-pentodialdo-1,4-furanose (**9**; 145 mg, 85%; approximately 90% pure). ¹H NMR δ 1.39 and 1.61 (2× s, 2× 3, 2× CH₃), 4.64 (dd, *J*₄₋₅ = 2.2 Hz, *J*₄₋₃ = 9.2 Hz, 1, H4), 5.01 (t, *J*_{2-1/3} = 4.2 Hz, 1, H2), 5.13 (dd, *J*₃₋₄ = 9.2 Hz, *J*₃₋₂ = 4.6 Hz, 1, H3), 6.02 (d, *J*₁₋₂ = 3.4 Hz, 1, H1), 7.48–8.03 (m, 5, Ar), 9.77 (d, *J*₅₋₄ = 2.2 Hz, 1, H5). Step (b). Treatment of the crude **9** (145 mg) with Ph₃P(CH₂)₃CO₂Et/Br (275 mg, 0.60 mmol) and LHDMS (1 M/THF; 0.60 mmol, 0.60 mL) as described for **10**, gave **11** (18 mg, 12%): ¹H NMR δ 1.26 (t, *J* = 7.1 Hz, 3, CH₃), 1.36 and 1.62 (2× s, 2× 3, 2× CH₃), 2.38 (t, *J*_{8-7/7'} = 8.2 Hz, 2, H8/8'), 2.46–2.55 (m, 1, H7), 2.55–2.67 (m, 1, H7'), 4.15 (q, *J* = 7.1 Hz, 2, CH₂), 4.74 (dd, *J*₃₋₄ = 9.1 Hz, *J*₃₋₂ = 4.8 Hz, 1, H3), 4.96 ('t', *J*_{2-1/3} = 4.3 Hz, 1, H2), 5.20 (t, *J*_{4-3/5} = 8.7 Hz, 1, H4), 5.50 (ddt, *J*₅₋₆ = 10.9 Hz, *J*₅₋₄ = 8.7 Hz, *J*_{5-7/7'} = 1.0 Hz, 1, H5), 5.72 (dt, *J*₆₋₅ = 10.9 Hz, *J*_{6-7/7'} = 7.1 Hz, 1, H6), 5.93 (d, *J*₁₋₂ = 3.8 Hz, 1, H1), 7.48–8.04, (m, 5, Ar); ¹³C NMR δ 14.30 (CH₃), 24.01 (C7), 26.99 and 27.04 (CMe₂), 34.53 (C8), 60.89 (CH₂), 73.39 (C4), 77.34 (C2), 77.56 (C3), 104.61 (C1), 113.40 (CMe₂), 127.26 (C5), 128.90 (Bz), 129.80 (Bz), 130.28 (Bz), 133.80 (Bz), 135.14 (C6), 166.25 (Bz), 173.06 (C9). HRMS (AP-ESI) *m/z* calcd for C₂₁H₂₆O₇Li (MLi⁺) 397.1839; found: 397.1828.

5.3. 3-*O*-Benzoyl-5,6-dideoxy-6,6-dibromo-1,2-*O*-isopropylidene- α -*D*-xylo-hex-5-enofuranose (**12**)

(Dibromomethylene)triphenylphosphorane [generated in situ by stirring CBr₄ (8.09 g, 24.5 mmol), Ph₃P (6.46 g, 24.5 mmol) and activated Zn (dust; 1.60 g,

24.5 mmol) in dried CH₂Cl₂ (100 mL) at 0 °C (ice-bath) for 30 min followed by stirring at ambient temperature under N₂ for 3 h] was added to the solution of freshly prepared aldehyde **8** [prepared as described for **10** (Step (a)) from **6** (4.68 g, 12.9 mmol) and dried for 1 h under vacuum prior to use] in CH₂Cl₂ (75 mL). After stirring for 14 h at ambient temperature, the reaction mixture was partitioned (NaHCO₃/H₂O/CHCl₃), and the organic layer was washed (H₂O, brine), dried (MgSO₄), and the volatiles were evaporated. Column chromatography (15 → 25% EtOAc/hexane) gave **12** (4.68 g, 81% overall from **6**) as a solidifying viscous oil: ¹H NMR δ 1.34 and 1.58 (2× s, 2× 3, 2× CH₃), 4.68 (d, *J*₂₋₁ = 3.7 Hz, 1, H2), 5.04 (dd, *J*₄₋₅ = 7.6 Hz, *J*₄₋₃ = 3.0 Hz, 1, H4), 5.54 (d, *J*₃₋₄ = 3.0 Hz, 1, H3), 6.00 (d, *J*₁₋₂ = 3.7 Hz, 1, H1), 6.60 (d, *J*₅₋₄ = 7.6 Hz, 1, H5), 7.48 (t, *J* = 7.6 Hz, 2 Ar), 7.60 (tt, *J* = 1.3, 7.6 Hz, 1 Ar), 8.02 ('dd', *J* = 1.4, 7.7 Hz, 2 Ar); ¹³C NMR δ 24.96 and 25.51 (CMe₂), 75.71 (C3), 78.18 (C4), 82.04 (C2), 93.26 (C6), 103.30 (C1), 111.26 (CMe₂), 127.31 (Bz), 127.73 (Bz), 128.44 (Bz), 130.49 (C5), 132.37 (Bz), 163.81 (Bz); MS *m/z* 451 (5, MH⁺ [⁸¹Br₂]), 449 (10, MH⁺ [^{81/79}Br₂]), 447 (5, MH⁺ [⁷⁹Br₂]).

5.4. Ethyl 3-*O*-benzoyl-5,6,7,8,9-pentadeoxy-1,2-*O*-isopropylidene- α -*D*-xylo-dec-5(*E*)-enofuranuronate (**14**) and ethyl 3-*O*-benzoyl-6-[3-(ethoxycarbonyl)propyl]-5,6,7,8,9-pentadeoxy-1,2-*O*-isopropylidene- α -*D*-xylo-dec-5-enofuranuronate (**18**)

Pd[P(Ph)₃]₄ (22 mg, 0.014 mmol) was added to a stirred solution of **12** (42 mg, 0.094 mmol) in anhydrous benzene (3 mL) in a flame-dried flask under N₂ at ambient temperature. After 2 min, 4-ethoxy-4-oxobutylzinc bromide (0.5 M/THF; 0.56 mL, 129 mg, 0.28 mmol) was added and the resulting mixture was heated at 55 °C for 6 h. The reaction mixture was cooled down to ambient temperature and was partitioned between EtOAc (30 mL) and NaHCO₃/H₂O (10 mL). The separated organic layer was washed with H₂O (10 mL), NaCl/H₂O (10 mL), dried (Na₂SO₄), and evaporated. Column chromatography (10 → 30% EtOAc/hexanes) gave recovered **12** (7 mg, 13%), **14** (7 mg, 18%), and **18** (19 mg, 48%). Compound **14** had: ¹H NMR δ 1.23 (t, *J* = 7.1 Hz, 3, CH₃), 1.28 and 1.58 (2× s, 2× 3, 2× CH₃), 1.68 (quint, *J*_{8-7/7'/9/9'} = 7.5 Hz, 2, H8/8'), 2.07 ('q', *J*_{7-6/8/8'} = 7.0 Hz, 2, H7/7'), 2.23 (t, *J*_{9-8/8'} = 7.4 Hz, 2, H9/9'), 4.15 (q, *J* = 7.1 Hz, 2, CH₂), 4.70 (d, *J*₂₋₁ = 3.7 Hz, 1, H2), 4.86 (dd, *J*₄₋₅ = 7.1 Hz, *J*₄₋₃ = 2.8 Hz, 1, H4), 5.42 (d, *J*₃₋₄ = 2.7 Hz, 1, H3), 5.56 (dd, *J*₅₋₆ = 15.4 Hz, *J*₅₋₄ = 7.3 Hz, 1, H5), 5.92 (dt, *J*₆₋₅ = 15.4 Hz, *J*_{6-7/7'} = 6.9 Hz, 1, H6), 6.03 (d, *J*₁₋₂ = 3.7 Hz, 1, H1), 7.46–8.05, (m, 5, Ar); ¹³C NMR δ 14.59 (CH₃), 23.34 (C8), 26.69 and 27.16 (CMe₂), 30.10 (C7), 34.05 (C9), 60.74 (CH₂), 75.90 (C2), 81.58 (C3), 85.44 (C4), 105.00 (C1), 112.52 (CMe₂), 128.35 (C5), 130.17 (Bz), 130.65 (Bz), 132.47 (Bz), 133.60 (Bz), 135.50 (C6), 165.89 (Bz), 173.74 (C10); MS *m/z* 405 (100, MH⁺). Compound **18** had: ¹H NMR δ 1.21 (t, *J* = 7.1 Hz, 3, CH₃), 1.29 (t, *J* = 7.1 Hz, 3, CH₃), 1.36 and 1.61 (2× s, 2× 3, 2× CH₃), 1.69 (quint, *J* = 7.5 Hz, 2H), 1.71–1.80 (m, 2H), 2.05 (t, *J* = 7.5 Hz, 2H), 2.10–2.26 (m, 4H), 2.33 (t, *J* = 7.2 Hz, 2H), 4.09 (q, *J* = 7.1 Hz, 2, CH₂), 4.18 (q, *J* = 7.1 Hz, 2, CH₂), 4.70 (d, *J*₂₋₁ = 3.8 Hz, 1, H2), 5.14 (dd, *J*₄₋₅ =

8.7 Hz, $J_{4-3} = 2.9$ Hz, 1, H4), 5.35 (d, $J_{5-4} = 8.4$ Hz, 1, H5), 5.41 (d, $J_{3-4} = 2.8$ Hz, 1, H3), 6.02 (d, $J_{1-2} = 3.8$ Hz, 1, H1), 7.47–8.06, (m, 5, Ar); ^{13}C NMR δ 14.60 (CH₃), 14.65 (CH₃), 26.65 and 27.15 (CMe₂), 23.32 and 23.84 (C8/8'), 30.53 and 36.22 (C7/7'), 34.04 and 34.08 (C9/9'), 60.59 (CH₂), 60.75 (CH₂), 75.90 (C4), 78.71 (C3), 84.24 (C2), 104.84 (C1), 112.36 (CMe₂), 119.09 (C5), 146.90 (C6), 128.92 (Bz), 128.92 (Bz), 129.81 (Bz), 130.17 (Bz), 133.83 (Bz), 165.70 (Bz), 173.62 and 173.75 (C10/10'); MS m/z 519 (100, MH⁺).

5.5. Ethyl 3-*O*-benzoyl-5,6,7,8,9-pentadeoxy-1,2-*O*-isopropylidene- α -D-ribo-dec-5(*E*)-enofuranuronate (16)

Treatment (55 °C, 3 h) of **28** (*E*; 20 mg, 0.048 mmol) with 4-ethoxy-4-oxobutylzinc bromide (0.5 M/THF; 0.19 mL, 65 mg, 0.096 mmol) as described for **14/18** gave **16** (11 mg, 56%): ^1H NMR δ 1.24 (t, $J = 7.1$ Hz, 3, CH₃), 1.35 and 1.59 (2 \times s, 2 \times 3, 2 \times CH₃), 1.72 (quint, $J_{8-7/7/9/9'} = 7.4$ Hz, 2, H8/8'), 2.13 (q', $J_{7-6/8/8'} = 7.2$ Hz, 2, H7/7'), 2.28 (t, $J_{9-8/8'} = 7.6$ Hz, 2, H9/9'), 4.10 (q, $J = 7.1$ Hz, 2, CH₂), 4.65 (dd, $J_{4-5} = 7.6$ Hz, $J_{4-3} = 8.9$ Hz, 1, H4), 4.74 (dd, $J_{3-4} = 9.2$ Hz, $J_{3-2} = 4.6$ Hz, 1, H3), 4.96 (t, $J_{2-1/3} = 4.3$ Hz, 1, H2), 5.53 (dd, $J_{5-6} = 15.4$ Hz, $J_{5-4} = 7.3$ Hz, 1, H5), 5.89 (d, $J_{1-2} = 4.0$ Hz, 1, H1), 5.91 (dt, $J_{6-5} = 15.8$ Hz, $J_{6-7/7'} = 6.8$ Hz, 1, H6), 7.46–8.05, (m, 5, Ar); ^{13}C NMR δ 14.62 (CH₃), 24.40 (C8), 26.92 and 27.00 (CMe₂), 31.99 (C7), 33.89 (C9), 60.65 (CH₂), 76.92 (C2), 77.65 (C3), 78.60 (C4), 104.37 (C1), 113.35 (CMe₂), 127.04 (C5), 128.85 (Bz), 129.83 (Bz), 130.28 (Bz), 133.76 (Bz), 136.28 (C6), 166.29 (Bz), 173.84 (C10); MS m/z 405 (100, MH⁺).

5.6. Ethyl 3-*O*-benzoyl-5,6,7,8,9-pentadeoxy-6-[3-(ethoxycarbonyl)propyl]-1,2-*O*-isopropylidene- α -D-ribo-dec-5-enofuranuronate (19)

Treatment of **13**¹⁵ (42 mg, 0.094 mmol) with Pd[P(Ph)₃]₄ (22 mg, 0.014 mmol) and 4-ethoxy-4-oxobutylzinc bromide (0.56 mL, 129 mg, 0.28 mmol) as described for **14/18** gave **19** (26 mg, 54%): ^1H NMR δ 1.22 (t, $J = 7.1$ Hz, 6, 2 \times CH₃), 1.37 and 1.62 (2 \times s, 2 \times 3, 2 \times CH₃), 1.75 (quint, $J = 7.5$ Hz, 4H), 2.11 (t, $J = 7.0$ Hz, 2H), 2.24 (t, $J = 9.0$ Hz, 2H), 2.26 (t, $J = 9.0$ Hz, 2H), 2.32 (t, $J = 7.2$ Hz, 2H), 4.10 (q, $J = 7.1$ Hz, 2, CH₂), 4.16 (q, $J = 7.1$ Hz, 2, CH₂), 4.74 (dd, $J_{3-4} = 9.0$ Hz, $J_{3-2} = 4.8$ Hz, 1, H3), 4.95 (t, $J_{2-3} = 4.3$ Hz, 1, H2), 5.01 (t, $J_{4-5} = 9.0$ Hz, 1, H4), 5.26 (d, $J_{5-4} = 8.9$ Hz, 1, H5), 5.89 (d, $J_{1-2} = 3.9$ Hz, 1, H1), 7.40–8.10 (m, 5, Ar); ^{13}C NMR δ 14.55 (CH₃), 14.61 (CH₃), 21.48 (C8/8'), 23.11 and 27.03 (CMe₂), 30.08 and 30.12 (C7/7'), 32.34 and 34.04 (C9/9'), 60.73 (CH₂), 60.83 (CH₂), 73.86 (C4), 78.71 (C3), 84.24 (C2), 104.37 (C1), 113.28 (CMe₂), 122.65 (C5), 128.92 (Bz), 128.92 (Bz), 129.81 (Bz), 130.17 (Bz), 133.83 (Bz), 143.90 (C6), 165.70 (Bz), 170.57 and 171.62 (C10/10'); MS m/z 519 (100, MH⁺).

5.7. (*E/Z*)-3-*O*-benzoyl-5,6-dideoxy-6-fluoro-1,2-*O*-isopropylidene-6-phenylsulfonyl- α -D-xylo-hex-5-enofuranose (20)

LHMDS (0.84 mL, 140 mg, 0.84 mmol) was added dropwise to a solution of diethyl fluoro(phenylsulfo-

nyl)methylphosphonate²⁰ (260 mg, 0.84 mmol) in anhydrous THF (8 mL) in a flame-dried flask under N₂ at –78 °C. After 30 min, a solution of **8** (265 mg, 0.82 mmol) in THF (4 mL) was added and stirring was continued for 1.5 h. EtOAc (30 mL) and NH₄Cl/H₂O (10 mL) were added and the reaction mixture was allowed to warm to ambient temperature. The separated organic layer was washed with NaHCO₃/H₂O (10 mL), NaCl/H₂O (10 mL), dried (Na₂SO₄), and evaporated. Column chromatography (10 \rightarrow 30% EtOAc/hexanes) gave **20** (166 mg, 76%; *E/Z*, 7:3) as inseparable mixture of isomers: HRMS (AP-ESI) m/z : calcd for C₂₂H₂₂FO₇S (MH⁺) 449.1065; found: 449.1071. ^{19}F NMR δ –110.25 (d, $J_{\text{F-H5}} = 18.8$ Hz, 0.30F, *Z*), –119.30 (d, $J_{\text{F-H5}} = 32.1$ Hz, 0.70F, *E*). Compound (*E*)-**20** had: ^1H NMR δ 1.35 and 1.56 (2 \times s, 2 \times 3, 2 \times CH₃), 4.73 (d, $J_{2-1} = 3.7$ Hz, 1, H2), 5.23 (dt, $J_{4-5} = 7.4$ Hz, $J_{4-3/\text{F}} = 2.3$ Hz, 1, H4), 5.49 (d, $J_{3-4} = 3.1$ Hz, 1, H3), 6.03–6.05 (m, 1, H1), 6.43 (dd, $J_{5-\text{F}} = 32.4$ Hz, $J_{5-4} = 7.2$ Hz, 1, H5), 7.48–8.03 (m, 10, Ar); ^{13}C NMR δ 26.55 and 27.10 (CMe₂), 73.78 (C4), 78.21 (C3), 83.77 (C2), 105.31 (C1), 113.13 (CMe₂), 112.10 (d, $^2J_{5-\text{F}} = 3.3$ Hz, C5), 128.99 (Ph), 129.08 (Bz), 129.13 (Bz), 129.79 (Ph), 129.94 (Ph), 130.12 (Ph), 130.17 (Bz), 134.25 (Bz), 135.02 (Ph), 156.00 (d, $^1J_{6-\text{F}} = 300.0$ Hz, C6), 165.36 (Bz). Compound (*Z*)-**20** had: ^1H NMR δ 1.38 and 1.69 (2 \times s, 2 \times 3, 2 \times CH₃), 4.76 (d, $J_{2-1} = 3.7$ Hz, 1, H2), 5.68 (d, $J_{3-4} = 2.8$ Hz, 1, H3), 5.99 (dd, $J_{5-\text{F}} = 19.3$ Hz, $J_{5-4} = 8.6$ Hz, 1, H5), 6.05–6.07 (m, 1, H1), 6.07–6.09 (m, 1, H4), 7.48–8.03 (m, 10, Ar); ^{13}C NMR δ 26.98 and 27.32 (CMe₂), 73.15 (d, $^3J_{4-\text{F}} = 10.14$ Hz, C4), 79.06 (C3), 83.93 (C2), 105.37 (C1), 113.35 (CMe₂), 114.10 (d, $^2J_{5-\text{F}} = 15.0$ Hz, C5), 128.99 (Ph), 129.08 (Bz), 129.13 (Bz), 129.79 (Ph), 129.94 (Ph), 130.12 (Ph), 130.17 (Bz), 134.25 (Bz), 135.02 (Ph), 155.58 (d, $^1J_{6-\text{F}} = 292.3$ Hz, C6), 165.36 (Bz). *Note*: Freshly prepared aldehyde **8**, dried under vacuum for 2 h at ambient temperature prior to the use, gave the best results.

5.8. (*E/Z*)-3-*O*-Benzoyl-5,6-dideoxy-6-fluoro-1,2-*O*-isopropylidene-6-phenylsulfonyl- α -D-ribo-hex-5-enofuranose (21)

Treatment of **9** (200 mg, 0.68 mmol) with diethyl fluoro(phenylsulfonyl)methylphosphonate²⁰ (212 mg, 0.68 mmol) and LHMDS (0.68 mL, 114 mg, 0.68 mmol) as described for **20** gave **21** (216 mg, 71%; *E/Z*, 6:4): HRMS (AP-ESI) m/z : calcd for C₂₂H₂₂FO₇S (MH⁺) 449.1065; found: 449.1069. ^{19}F NMR δ –108.98 (d, $J_{\text{F-H5}} = 22.6$ Hz, 0.40F, *Z*), –121.25 (d, $J_{\text{F-H5}} = 30.1$ Hz, 0.60F, *E*). Compound (*E*)-**21** had: ^1H NMR δ 1.28 and 1.38 (2 \times s, 2 \times 3, 2 \times CH₃), 4.85 (dd, $J_{3-4} = 9.0$ Hz, $J_{3-2} = 4.7$ Hz, 1, H3), 5.00 (t', $J_{2-1/3} = 4.5$ Hz, 1, H2), 5.10 (t, $J_{4-3/5} = 8.2$ Hz, 1, H4), 5.94 (d, $J_{1-2} = 3.7$ Hz, 1, H1), 6.37 (dd, $J_{5-\text{F}} = 31.3$ Hz, $J_{5-4} = 8.3$ Hz, 1, H5), 7.44–8.20 (m, 10, Ar); ^{13}C NMR δ 26.86 and 26.93 (CMe₂), 71.36 (d, $^3J_{4-\text{F}} = 2.2$ Hz, C4), 76.64 (d, $^4J_{3-\text{F}} = 1.8$ Hz, C3), 77.54 (C2), 104.96 (C1), 114.10 (CMe₂), 114.22 (d, $^2J_{5-\text{F}} = 3.1$ Hz, C5), 128.88 (Ph), 129.13 (Bz), 129.18 (Bz), 129.88 (Ph), 130.28 (Ph), 133.98 (Bz), 135.10 (Ph), 136.96 (Ph), 156.91 (d, $^1J_{6-\text{F}} = 306.0$ Hz, C6), 165.96 (Bz). Compound (*Z*)-**21** had: ^1H NMR δ 1.28 and 1.38 (2 \times s, 2 \times 3, 2 \times CH₃), 4.84 (dd,

$J_{3-4} = 9.2$ Hz, $J_{3-2} = 4.6$ Hz, 1, H3), 5.01 (t, $J_{2-1/3} = 4.6$ Hz, 1, H2), 5.86 (dd, $J_{5-F} = 19.8$ Hz, $J_{5-4} = 9.9$ Hz, H5), 5.96 (d, $J_{1-2} = 3.7$ Hz, 1, H1), 6.07 (t, $J_{4-3/5} = 10.4$ Hz, 1, H4), 7.44–8.20 (m, 10, Ar); ^{13}C NMR δ 27.17 and 27.39 (CMe₂), 70.71 (d, $^3J_{4-F} = 8.5$ Hz, C4), 77.17 (C3), 77.86 (C2), 104.82 (C1), 114.37 (CMe₂), 116.23 (d, $^2J_{5-F} = 16.2$ Hz, C5), 128.95 (Ph), 129.22 (Bz), 129.46 (Bz), 129.80 (Ph), 130.02 (Ph), 133.61 (Bz), 134.00 (Ph), 135.19 (Ph), 156.62 (d, $^1J_{6-F} = 296.3$ Hz, C6), 166.30 (Bz).

5.9. (*E*)-3-*O*-Benzoyl-5,6-dideoxy-1,2-*O*-isopropylidene-6-phenylsulfonyl- α -D-ribo-hex-5-enofuranose (**22**)

Treatment of **9** (150 mg, 0.50 mmol) with diethyl (phenylsulfonyl)methylphosphonate²⁰ (146 mg, 0.50 mmol) and LHMDS (0.50 mL, 84 mg, 0.50 mmol) as described for **20** gave **22** (166 mg, 82%): ^1H NMR δ 1.33 and 1.55 (2 \times s, 2 \times 3, 2 \times CH₃), 4.76 (dd, $J_{3-4} = 9.5$ Hz, $J_{3-2} = 4.6$ Hz, 1, H3), 4.92 (ddd, $J_{4-5} = 3.7$ Hz, $J_{4-6} = 1.7$ Hz, $J_{4-3} = 9.5$ Hz, 1, H4), 5.01 (t, $J_{2-3/1} = 4.2$ Hz, 1, H2), 5.90 (d, $J_{1-2} = 3.7$ Hz, 1, H1), 6.79 (dd, $J_{6-4} = 1.8$ Hz, $J_{6-5} = 15.0$ Hz, 1, H6), 7.09 (dd, $J_{5-6} = 15.0$ Hz, $J_{5-4} = 3.8$ Hz, 1, H5), 7.50–8.05 (m, 10, Ar); MS *m/z* 431 (100, MH⁺).

5.10. (*E/Z*)-3-*O*-Benzoyl-5,6-dideoxy-6-fluoro-1,2-*O*-isopropylidene-6-tributylstannyl- α -D-xylo-hex-5-enofuranose (**23**)

Bu₃SnH (407 mg, 0.38 mL, 1.4 mmol) was added dropwise to a degassed solution of **20** (300 mg, 0.70 mmol; *E/Z*, 7:3) in anhydrous toluene (5 mL) in a flame-dried flask under N₂ at ambient temperature. After an additional 10 min of degassing with N₂, AIBN (86 mg, 0.53 mmol) was added and the reaction mixture was refluxed at 110 °C with stirring for 5 h. The volatiles were evaporated and the residue was partitioned between EtOAc (50 mL) and NaHCO₃/H₂O (30 mL). The organic layer was washed with NaCl/H₂O (30 mL), dried (Na₂SO₄), and evaporated. Column chromatography (hexanes \rightarrow 10% EtOAc/hexanes) gave **23** (794 mg, 95%; *E/Z*, 7:3) as an inseparable mixture: MS *m/z* 599 (89, MH⁺, ^{120}Sn), 597 (63, MH⁺, ^{118}Sn), 595 (33, MH⁺, ^{116}Sn), 541 (100, M-57, ^{120}Sn), 539 (78, M-57, ^{118}Sn), 537 (42, M-57, ^{116}Sn); ^{19}F NMR δ -87.67 (d, $J_{F-H5} = 34.3$ Hz, 84% of 0.30F, Z), -87.67 (dd, $J_{F-Sn} = 229.5$ Hz, $J_{F-H5} = 34.8$ Hz, 16% of 0.30F, Z), -92.73 (d, $J_{F-H5} = 52.7$ Hz, 84% of 0.70F, E), -92.73 (ddd, $J_{F-Sn} = 213.1$ Hz, $J_{F-H5} = 52.7$ Hz, $J_{F-H4} = 4.9$ Hz, 16% of 0.70F, E). Compound (*E*)-**23** had: ^1H NMR δ 0.90–1.60 (m, 27, 3 \times Bu), 1.34 and 1.36 (2 \times s, 2 \times 3, 2 \times CH₃), 4.71 (d, $J_{2-1} = 3.8$ Hz, 1, H2), 5.10 (dd, $J_{5-F} = 52.6$ Hz, $J_{5-4} = 7.4$ Hz, 1, H5), 5.32 (d, $J_{3-4} = 3.0$ Hz, 1, H3), 5.47–5.49 (m, 1, H4), 6.02 (d, $J_{1-2} = 3.8$ Hz, 1, H1), 7.47–8.06 (m, 5, Ar); ^{13}C NMR δ 10.33 (Bu), 11.15 (Bu), 17.90 (Bu), 27.42 and 27.54 (CMe₂), 28.24 (Bu), 70.56 (d, $-3J_{4-F} = 17.6$ Hz, C4), 77.21 (C3), 77.52 (C2), 104.31 (C1), 113.07 (CMe₂), 120.53 (d, $^2J_{5-F} = 3.9$ Hz, C5), 128.75 (Bz), 129.83 (Bz), 130.25 (Bz), 133.61 (Bz), 166.16 (Bz), 177.14 (d, $^2J_{6-F} = 262.0$ Hz, C6). Compound (*Z*)-**23** had: ^1H NMR δ 0.90–1.60 (m, 27, 3 \times Bu), 1.38 and 1.69 (2 \times s, 2 \times 3, 2 \times CH₃), 4.69 (d, $J_{1-2} = 3.9$ Hz, 1, H2), 4.75 (d, $J_{3-4} = 7.9$ Hz, 1, H3), 5.47–5.49

(m, 1, H4), 5.98 (d, $J_{1-2} = 3.8$ Hz, 1, H1), 6.02 (dd, $J_{5-F} = 34.3$ Hz, $J_{5-4} = 9.2$ Hz, 1, H5), 7.47–8.06 (m, 5, Ar); ^{13}C NMR δ 10.33 (Bu), 11.15 (Bu), 17.90 (Bu), 27.42 and 27.54 (CMe₂), 28.24 (Bu), 74.73 (d, $^3J_{4-F} = 22.2$ Hz, C4), 77.38 (d, $^4J_{3-F} = 1.4$ Hz, C3), 77.52 (C2), 104.53 (C1), 113.47 (CMe₂), 121.24 (d, $^2J_{5-F} = 9.5$ Hz, C5), 128.75 (Bz), 129.88 (Bz), 130.34 (Bz), 133.73 (Bz), 166.35 (Bz), 180.03 (d, $^2J_{6-F} = 254.3$ Hz, C6).

5.11. (*E/Z*)-3-*O*-Benzoyl-5,6-dideoxy-6-fluoro-1,2-*O*-isopropylidene-6-tributylstannyl- α -D-ribo-hex-5-enofuranose (**24**)

Treatment of **21** (300 mg, 0.70 mmol; *E/Z*, 3:2) with Bu₃SnH (407 mg, 0.38 mL, 1.4 mmol) and AIBN (86 mg, 0.53 mmol) as described for **23** gave **24** (397 mg, 95%; *E/Z*, 3:2): MS *m/z* 599 (89, MH⁺, ^{120}Sn), 597 (63, MH⁺, ^{118}Sn), 595 (33, MH⁺, ^{116}Sn), 541 (100, M-57, ^{120}Sn), 539 (78, M-57, ^{118}Sn), 537 (42, M-57, ^{116}Sn); ^{19}F NMR δ -87.58 (d, $J_{F-H5} = 33.1$ Hz, 84% of 0.40F, Z), -87.58 (ddd, $J_{F-Sn} = 226.7$ Hz, $J_{F-H5} = 32.8$ Hz, $J_{F-H4} = 4.1$ Hz, 16% of 0.40F), -94.80 (d, $J_{F-H5} = 51.1$ Hz, 84% of 0.60F, E), -94.80 (ddd, $J_{F-Sn} = 213.9$ Hz, $J_{F-H5} = 50.8$ Hz, $J_{F-H4} = 4.5$ Hz, 16% of 0.60F, E). Compound (*E*)-**24** had: ^1H NMR δ 0.70–1.70 (m, 27, 3 \times Bu), 1.24 and 1.26 (2 \times s, 2 \times 3, 2 \times CH₃), 4.62 (dd, $J_{3-4} = 9.3$ Hz, $J_{3-2} = 4.7$ Hz, 1, H3), 4.86–4.87 (m, 1, H2), 4.90 (dd, $J_{5-F} = 51.0$ Hz, $J_{5-4} = 8.4$ Hz, 1, H5), 5.28 (t, $J_{4-3/5} = 8.9$ Hz, 1, H4), 5.79 (d, $J_{1-2} = 3.9$ Hz, 1, H1), 7.57–8.06 (m, 5, Ar); ^{13}C NMR δ 11.15 (Bu), 14.03 (Bu), 29.96 (Bu), 27.48 and 27.59 (CMe₂), 29.23 (Bu), 70.55 (d, $^3J_{4-F} = 18.1$ Hz, C4), 77.36 (C3), 77.48 (C2), 104.50 (C1), 113.54 (CMe₂), 120.59 (d, $^2J_{5-F} = 3.8$ Hz, C5), 128.77 (Bz), 129.81 (Bz), 130.38 (Bz), 133.69 (Bz), 166.44 (Bz), 176.10 (d, $^1J_{6-F} = 260.0$ Hz, C6). Compound (*Z*)-**24** had: ^1H NMR δ 0.70–1.70 (m, 27, 3 \times Bu), 1.28 and 1.30 (2 \times s, 2 \times 3, 2 \times CH₃), 4.47 (t, $J_{4-3/5} = 9.3$ Hz, 1, H4), 4.71 (dd, $J_{3-4} = 9.0$ Hz, $J_{3-2} = 4.8$ Hz, 1, H3), 4.85–4.86 (m, 1, H2), 5.81 (d, $J_{1-2} = 3.8$ Hz, 1, H1), 5.83 (dd, $J_{5-F} = 33.7$ Hz, $J_{5-4} = 9.5$ Hz, 1, H5), 7.57–8.06 (m, 5, Ar); ^{13}C NMR δ 11.15 (Bu), 14.03 (Bu), 29.96 (Bu), 27.48 and 27.59 (CMe₂), 29.23 (Bu), 74.72 (d, $^3J_{4-F} = 22.0$ Hz, C4), 77.16 (C3), 77.67 (C2), 104.28 (C1), 113.13 (CMe₂), 121.18 (d, $^2J_{5-F} = 9.9$ Hz, C5), 128.77 (Bz), 129.76 (Bz), 130.28 (Bz), 130.80 (Bz), 166.24 (Bz), 177.00 (d, $^1J_{6-F} = 255.0$ Hz, C6).

5.12. (*E*)-3-*O*-Benzoyl-5,6-dideoxy-1,2-*O*-isopropylidene-6-tributylstannyl- α -D-ribo-hex-5-enofuranose (**25**)

Treatment of **22** (*E*; 300 mg, 0.70 mmol) with Bu₃SnH (407 mg, 0.38 mL, 1.4 mmol) and AIBN (86 mg, 0.53 mmol) as described for **23** gave **25** (385 mg, 95%): ^1H NMR δ 1.51–1.90 (m, 33, 3 \times Bu and 2 \times CH₃), 4.66 (dd, $J_{4-5} = 6.5$ Hz, $J_{4-3} = 8.4$ Hz, 1, H4), 4.77 (dd, $J_{3-4} = 9.2$ Hz, $J_{3-2} = 4.7$ Hz, 1, H3), 4.97 (t, $J_{2-1/3} = 3.6$ Hz, 1, H2), 5.93 (d, $J_{1-2} = 3.8$ Hz, 1, H1), 6.10 (dd, $J_{5-6} = 19.1$ Hz, $J_{5-4} = 6.5$ Hz, 1, H5), 6.50 (dd, $J_{6-5} = 19.1$ Hz, $J_{6-4} = 0.9$ Hz, 1, H6), 7.57–8.06 (m, 5, Ar); MS *m/z* 581 (89, MH⁺, ^{120}Sn), 579 (63, MH⁺, ^{118}Sn), 577 (33, MH⁺, ^{116}Sn), 523 (100, M-57, ^{120}Sn), 521 (78, M-57, ^{118}Sn), 519 (42, M-57, ^{116}Sn).

5.13. (*E/Z*)-3-*O*-Benzoyl-5,6-dideoxy-6-fluoro-6-iodo-1,2-*O*-isopropylidene- α -D-xylo-hex-5-enofuranose (26**)**

A solution of NIS (50 mg, 0.23 mmol) in anhydrous CH₂Cl₂ (3 mL) was added dropwise to a stirred solution of **23** (90 mg, 0.15 mmol; *E/Z*, 7:3) in CH₂Cl₂ (5 mL) under N₂ at –20 °C. After 1 h, CHCl₃ (30 mL) and diluted NaHSO₃/H₂O (10 mL) were added. The separated organic layer was washed with NaHCO₃/H₂O (10 mL), NaCl/H₂O (10 mL), dried (Na₂SO₄), and evaporated. Column chromatography (hexanes → 30% EtOAc/hexanes) gave **26** (155 mg, 83%; *E/Z*, 8:2) as an inseparable mixture: MS *m/z* 435 (100, MH⁺); ¹⁹F NMR δ –56.54 (d, J_{F-H5} = 15.8 Hz, 0.20F, Z), –60.98 (d, J_{F-H5} = 33.1 Hz, 0.80F, E). Compound (*E*)-**26** had: ¹H NMR δ 1.35 and 1.38 (2× s, 2× 3, 2× CH₃), 4.69 (d, J_{2-1} = 3.8 Hz, 1, H2), 5.28 (ddd, J_{4-5} = 9.8 Hz, J_{4-3} = 2.9 Hz, J_{4-6} = 1.6 Hz, 1, H4), 5.45 (d, J_{3-4} = 3.0 Hz, 1, H3), 5.58 (dd, J_{5-F} = 33.0 Hz, J_{5-4} = 8.7 Hz, 1, H5), 5.99 (d, J_{1-2} = 3.7 Hz, 1, H1), 7.47–8.06 (m, 5, Ar); ¹³C NMR δ 26.60 and 27.12 (CMe₂), 74.04 (d, $^3J_{4-F}$ = 4.4 Hz, C4), 77.88 (C3), 83.73 (C2), 104.74 (C1), 107.89 (d, $^1J_{6-F}$ = 338.7 Hz, C6), 112.85 (CMe₂), 116.80 (d, $^2J_{5-F}$ = 5.4 Hz, C5), 129.04 (Bz), 129.50 (Bz), 130.18 (Bz), 134.09 (Bz), 165.54 (Bz). Compound (*Z*)-**26** had: ¹H NMR δ 1.35 and 1.68 (2× s, 2× 3, 2× CH₃), 4.71 (d, J_{1-2} = 3.7 Hz, 1, H2), 4.84 (dd, J_{4-5} = 8.7 Hz, J_{4-3} = 2.7 Hz, 1, H4), 5.48 (d, J_{3-4} = 3.0 Hz, 1, H3), 5.77 (dd, J_{5-F} = 15.4 Hz, J_{5-4} = 8.8 Hz, 1, H5), 6.03 (d, J_{1-2} = 3.7 Hz, 1, H1), 7.47–8.06 (m, 5, Ar); ¹³C NMR δ 26.78 and 27.27 (CMe₂), 77.88 (C3), 80.06 (d, $^3J_{4-F}$ = 8.2 Hz, C4), 83.84 (C2), 105.04 (C1), 112.94 (d, $^2J_{5-F}$ = 16.9 Hz, C5), 112.95 (CMe₂), 114.78 (d, $^1J_{6-F}$ = 332.0 Hz, C6), 129.04 (Bz), 129.40 (Bz), 130.18 (Bz), 134.14 (Bz), 165.54 (Bz).

5.14. (*E/Z*)-3-*O*-Benzoyl-5,6-dideoxy-6-fluoro-6-iodo-1,2-*O*-isopropylidene- α -D-ribo-hex-5-enofuranose (27**)**

Treatment of **24** (250 mg, 0.42 mmol; *E/Z*, 3:2) with NIS (142 mg, 0.63 mmol) as described for **26** gave **27** (155 mg, 85%; *E/Z*, 3:2) as an inseparable mixture: HRMS (AP-FAB) *m/z*: calcd for C₁₆H₁₆FIO₅Li (MLi⁺) 441.0181; found: 441.0192. ¹⁹F NMR δ –56.42 (d, J_{F-H5} = 15.1 Hz, 0.40F, Z), –63.30 (d, J_{F-H5} = 33.5 Hz, 0.60F, E). Compound (*E*)-**27** had: ¹H NMR δ 1.36 and 1.60 (2× s, 2× 3, 2× CH₃), 4.75 (dd, J_{3-4} = 9.2 Hz, J_{3-2} = 4.7 Hz, 1, H3), 4.98 (t, $J_{2-1/3}$ = 4.5 Hz, 1, H2), 5.16 (t, $J_{4-3/5}$ = 9.0 Hz, 1, H4), 5.49 (dd, J_{5-F} = 32.7 Hz, J_{5-4} = 8.7 Hz, 1, H5), 5.89 (d, J_{1-2} = 3.8 Hz, 1, H1), 7.50–8.10 (m, 5, Ar); ¹³C NMR δ 26.96 and 27.11 (CMe₂), 72.45 (d, $^3J_{4-F}$ = 4.3 Hz, C4), 76.74 (d, $^4J_{3-F}$ = 2.1 Hz, C3), 77.32 (C2), 104.39 (C1), 113.78 (CMe₂), 114.96 (d, $^1J_{6-F}$ = 331.5 Hz, C6), 119.90 (d, $^2J_{5-F}$ = 5.5 Hz, C5), 128.92 (Bz), 129.46 (Bz), 130.50 (Bz), 133.94 (Bz), 166.21 (Bz). Compound (*Z*)-**27** had: ¹H NMR δ 1.38 and 1.64 (2× s, 2× 3, 2× CH₃), 4.72–4.75 (m, 1, H4), 4.84 (dd, J_{3-4} = 9.2 Hz, J_{3-2} = 4.6 Hz, 1, H3), 4.98 (t, $J_{2-3/1}$ = 4.5 Hz, 1, H2), 5.68 (dd, J_{5-F} = 15.3 Hz, J_{5-4} = 8.9 Hz, 1, H5), 5.91 (d, J_{1-2} = 3.8 Hz, 1, H1), 7.50–8.14 (m, 5, Ar); ¹³C NMR δ 26.96 and 27.11 (CMe₂), 76.85 (d, $^4J_{3-F}$ = 2.1 Hz, C3), 77.63 (C2), 78.13 (d, $^3J_{4-F}$ = 8.3 Hz, C4), 104.54 (C1), 108.75 (d, $^1J_{6-F}$ = 339.4 Hz, C6), 113.90 (CMe₂), 115.77 (d, $^2J_{5-F}$ =

16.2 Hz, C5), 128.91 (Bz), 129.49 (Bz), 130.37 (Bz), 133.95 (Bz), 166.21 (Bz).

5.15. (*E*)-3-*O*-Benzoyl-5,6-dideoxy-6-iodo-1,2-*O*-isopropylidene- α -D-ribo-hex-5-enofuranose (28**)**

Treatment of **25** (150 mg, 0.25 mmol) with NIS (85 mg, 0.38 mmol) as described for **26** gave **28** (93 mg, 87%): ¹H NMR δ 1.35 and 1.62 (2× s, 2× 3, 2× CH₃), 4.67 (dd, J_{2-3} = 9.2 Hz, J_{2-1} = 4.6 Hz, 1, H2), 4.77 (dd, J_{3-4} = 3.4 Hz, J_{3-2} = 9.2 Hz, 1, H3), 4.97 (t, $J_{4-3/5}$ = 4.2 Hz, 1, H4), 5.91 (d, J_{1-2} = 3.8 Hz, 1, H1), 6.61–6.70 (m, 2, H5/6), 7.49–8.08 (m, 5, Ar); ¹³C NMR δ 26.95 and 26.96 (CMe₂), 76.37 (C4), 77.59 (C3), 79.61 (C2), 81.32 (C6), 104.43 (C1), 113.68 (CMe₂), 128.89 (Bz), 129.60 (Bz), 130.94 (Bz), 133.89 (Bz), 141.58 (C5), 166.08 (Bz); MS *m/z* 417 (100, MH⁺).

5.16. Ethyl 3-*O*-benzoyl-5,6,7,8,9-pentadeoxy-6-fluoro-1,2-*O*-isopropylidene- α -D-xylo-dec-5-(*E/Z*)-enofuranuronate (29**)**

Pd[P(Ph)₃]₄ (5 mg, 0.004 mmol) was added to a stirred solution of **26** (30 mg, 0.07 mmol; *E/Z*, 4:1) in anhydrous benzene (3 mL) under N₂ at ambient temperature. After 2 min, 4-ethoxy-4-oxobutylzinc bromide (0.5 M/THF; 0.28 mL, 65 mg, 0.14 mmol) was added and the resulting mixture was heated at 55 °C for 5 h. EtOAc (30 mL) and NaHCO₃/H₂O (10 mL) were added and the separated organic layer was washed with H₂O (10 mL), NaCl/H₂O (10 mL), dried (Na₂SO₄), and then was evaporated. Column chromatography (10 → 30% EtOAc/hexanes) gave (*Z*)-**29** (18 mg, 61%, 76% based on the conversion of the *E*-isomer), (*E*)-**29** (2 mg, 7%, 35% based on the conversion of the *Z*-isomer) and more polar byproduct tentatively assigned as 3-*O*-debenzoylated-(*Z*)-**26** [~5%, TLC; ¹⁹F NMR δ –57.21 (J_{F-H5} = 16.4 Hz)]. Compound (*Z*)-**29** had: ¹H NMR δ 1.22 (t, J = 7.1 Hz, 3, CH₃), 1.36 and 1.61 (2× s, 2× 3, 2× CH₃), 1.81 ('quint', $J_{8-7/7'/9/9'}$ = 7.4 Hz, 2, H8/8'), 2.26 (dt, J_{7-F} = 18.1 Hz, $J_{7-8/8'}$ = 7.4 Hz, 2, H7/7'), 2.30 (t, $J_{9-8/8'}$ = 7.4 Hz, 2, H9/9'), 4.09 (q, J = 7.1 Hz, 2, CH₂), 4.69 (d, J_{2-1} = 3.7 Hz, 1, H2), 4.84 (dd, J_{5-F} = 35.8 Hz, J_{5-4} = 8.4 Hz, H5), 5.33 (dd, J_{4-5} = 8.5 Hz, J_{4-3} = 2.8 Hz, 1, H4), 5.45 (d, J_{3-4} = 2.8 Hz, 1, H3), 6.00 (d, J_{1-2} = 3.7 Hz, 1, H1), 7.48–8.04 (m, 5, Ar); ¹³C NMR δ 14.59 (CH₃), 21.49 (C8), 26.64 and 27.11 (CMe₂), 31.61 (d, $^2J_{7-F}$ = 25.4 Hz, C7), 33.34 (C9), 60.75 (CH₂), 73.37 (d, $^3J_{4-F}$ = 6.6 Hz, C4), 77.63 (C2), 78.33 (C3), 100.03 (d, $^2J_{5-F}$ = 10.9 Hz, C5), 104.78 (C1), 112.60 (CMe₂), 128.97 (Bz), 129.70 (Bz), 130.16 (Bz), 133.93 (Bz), 162.94 (d, $^1J_{6-F}$ = 260.7 Hz, C6), 165.61 (Bz), 173.32 (C10); ¹⁹F NMR δ –99.93 (dt, J_{F-H5} = 35.8 Hz, J = 18.0 Hz); HRMS (AP-FAB⁺) *m/z* calcd for C₂₂H₂₇FO₇Li (MLi⁺) 429.1910; found: 429.1900. Compound (*E*)-**29** had: ¹H NMR δ 1.28 (t, J = 7.1 Hz, 3, CH₃), 1.35 and 1.61 (2× s, 2× 3, 2× CH₃), 1.85–1.95 (m, 2, H8/8'), 2.38 (t, $J_{9-8/8'}$ = 7.2 Hz, 2, H9/9'), 2.39–2.50 (m, 2, H7/7'), 4.15 (q, J = 7.1 Hz, 2, CH₂), 4.69 (d, J_{2-1} = 3.8 Hz, 1, H2), 4.93 ('dt', J_{4-5} = 9.3 Hz, $J_{4-F/3}$ = 2.5 Hz, 1, H4), 5.30 (dd, J_{5-F} = 18.6 Hz, J_{5-4} = 9.4 Hz, H5), 5.38 (d, J_{3-4} = 2.9 Hz, 1, H3), 6.00 (d, J_{1-2} = 3.8 Hz, 1, H1), 7.48–8.04 (m, 5, Ar); ¹⁹F NMR δ

–94.53 ('q', $J = 22.1$ Hz). HRMS (AP-FAB⁺) m/z calcd for C₂₂H₂₇FO₇Li (MLi⁺) 429.1910; found: 429.1903.

5.17. Ethyl 3-*O*-benzoyl-5,6,7,8,9-pentadeoxy-6-fluoro-1,2-*O*-isopropylidene- α -*D*-ribo-dec-5(*E/Z*)-enofuranuronate (**30**)

Treatment of **27** (42 mg, 0.097 mmol; *E/Z*, 3:2) with Pd[P(Ph)₃]₄ (22 mg, 0.01 mmol) and 4-ethoxy-4-oxobutylzinc bromide (0.5 M/THF; 0.30 mmol, 0.60 mL) as described for **29** followed by column chromatography (10 → 40% EtOAc/hexanes) gave (*Z*)-**30** (22 mg, 54%; 90% based on the conversion of *E*-isomer), (*E*)-**30** (5 mg, 12%; 30% based on the conversion of the *Z*-isomer), and more polar 3-*O*-debenzoylated-(*Z*)-**27** (3 mg, 10%). Compound (*Z*)-**30** had: ¹H NMR δ 1.24 (t, $J = 7.1$ Hz, 3, CH₃), 1.37 and 1.60 (2× s, 2× 3, 2× CH₃), 1.84 ('quint', $J_{8-7/7'/9/9'} = 7.4$ Hz, 2, H8/8'), 2.25 (dt, $J_{7-F} = 17.6$ Hz, $J_{7-8/8'} = 7.5$ Hz, 2, H7/7'), 2.32 (t, $J_{9-8/8'} = 7.4$ Hz, 2, H9/9'), 4.09 (q, $J = 7.1$ Hz, 2, CH₂), 4.72 (dd, $J_{3-4} = 9.2$ Hz, $J_{3-2} = 4.7$ Hz, 1, H3), 4.75 (dd, $J_{5-F} = 35.0$ Hz, $J_{5-4} = 8.9$ Hz, 1, H5), 4.95 (t, $J_{2-1/3} = 4.3$ Hz, 1, H2), 5.19 (t, $J_{4-3/5} = 9.1$ Hz, 1, H4), 5.89 (d, $J_{1-2} = 3.8$ Hz, 1, H1), 7.48–8.09 (m, 5, Ar); ¹³C NMR δ 14.60 (CH₃), 21.45 (C8), 26.95 and 27.00 (CMe₂), 31.63 (d, $^2J_{7-F} = 26.5$ Hz, C7), 33.32 (C9), 60.79 (CH₂), 71.39 (d, $^3J_{4-F} = 6.3$ Hz, C4), 77.54 (C2), 73.63 (C3), 103.40 (d, $^2J_{5-F} = 11.8$ Hz, C5), 104.39 (C1), 113.52 (CMe₂), 128.84 (Bz), 129.71 (Bz), 130.33 (Bz), 133.77 (Bz), 164.13 (d, $^1J_{6-F} = 272.7$ Hz, C6), 165.33 (Bz), 173.30 (C10); ¹⁹F NMR δ –102.14 (dt, $J_{F-H5} = 34.1$ Hz, $J_{F-H7/7'} = 17.6$ Hz). HRMS (AP-ESI) m/z calcd for C₂₂H₂₈FO₇ (MH⁺) 423.1814; found 423.1815. Compound (*E*)-**30** had: ¹H NMR δ 1.26 (t, $J = 7.1$ Hz, 3, CH₃), 1.36 and 1.61 (2× s, 2× 3, 2× CH₃), 1.90 ('quint', $J_{8-7/7'/9/9'} = 6.9$ Hz, 2, H8/8'), 2.38 (t, $J_{9-8/8'} = 6.9$ Hz, 2, H9/9'), 2.50 (dt, $J_{7-F} = 23.0$ Hz, $J_{7-8/8'} = 7.3$ Hz, 2, H7/7'), 4.14 (q, $J = 7.1$ Hz, 2, CH₂), 4.75–4.80 (m, 2, H3/4), 4.95 (t, $J_{2-1/3} = 4.0$ Hz, 1, H2), 5.17 (dd, $J_{5-F} = 19.2$ Hz, $J_{5-4} = 8.7$ Hz, 1, H5), 5.88 (d, $J_{1-2} = 3.8$ Hz, 1, H1), 7.48–8.09 (m, 5, Ar); ¹³C NMR δ 14.63 (CH₃), 21.99 (C8), 26.92 and 27.00 (CMe₂), 28.49 (d, $^2J_{7-F} = 27.0$ Hz, C7), 33.67 (C9), 60.78 (CH₂), 73.65 (d, $^3J_{4-F} = 14.7$ Hz, C4), 77.50 (C2), 77.61 (C3), 104.58 (d, $^2J_{5-F} = 25.8$ Hz, C5), 104.37 (C1), 113.39 (CMe₂), 128.88 (Bz), 129.64 (Bz), 130.25 (Bz), 133.83 (Bz), 165.15 (d, $^1J_{6-F} = 256.5$ Hz, C6), 165.99 (Bz), 173.21 (C10); ¹⁹F NMR δ –94.73 ('q', $J_{F-H5/7/7'} = 22.8$ Hz). HRMS (AP-ESI) m/z calcd for C₂₂H₂₈FO₇ (MH⁺) 423.1814; found: 423.1819. The 3-*O*-debenzoylated-(*Z*)-**27** had: ¹H NMR δ 1.27 and 1.58 (2× s, 2× 3, 2× CH₃), 3.82–3.84 (m, 1, H3), 4.18 (t, $J_{4-3/5} = 8.8$ Hz, 1, H4), 4.60–4.62 (m, 1, H2), 5.61 (dd, $J_{5-F} = 15.1$ Hz, $J_{5-4} = 8.9$ Hz, 1, H5), 5.83 (d, $J_{1-2} = 3.7$ Hz, 1, H1); ¹³C NMR δ 26.99 and 27.07 (CMe₂), 76.63 (C3), 78.56 (C2), 80.47 (d, $^3J_{4-F} = 8.0$ Hz, C4), 104.16 (C1), 115.85 (d, $^1J_{6-F} = 344.1$ Hz, C6), 113.70 (CMe₂), 115.85 (d, $^2J_{5-F} = 15.7$ Hz, C5); ¹⁹F NMR δ –56.50 (d, $J_{F-H5} = 15.1$ Hz); MS m/z 331 (100, MH⁺).

5.18. Methyl 5,6,7,8,9-pentadeoxy-6-fluoro-1,2-*O*-isopropylidene- α -*D*-xylo-dec-5(*Z*)-enofuranuronate (**31**)

Compound (*Z*)-**29** (26 mg, 0.062 mmol) was dissolved in MeOH (6 mL) and the saturated NH₃/MeOH (3 mL)

was added at 0 °C (ice bath). The resulting mixture was stirred for 48 h (0 °C → ambient temperature). The volatiles were evaporated and the residue was column chromatographed (15 → 50% EtOAc/hexanes) to give **31** (14 mg, 74%): ¹H NMR δ 1.35 and 1.55 (2× s, 2× 3, 2× CH₃), 1.90 (quint, $J_{8-7/7'/9/9'} = 7.2$ Hz, 2, H8/8'), 2.23–2.40 (m, 2, H7/7'), 2.41 (t, $J_{9-8/8'} = 7.2$ Hz, 2, H9/9'), 3.70 (s, 3, CH₃), 4.17 (d, $J_{3-4} = 2.6$ Hz, 1, H3); 4.59 (d, $J_{2-1} = 3.7$ Hz, 1, H2), 4.84 (dd, $J_{5-F} = 37.6$ Hz, $J_{5-4} = 7.7$ Hz, 1, H5), 5.08 ('dm', $J_{4-5} = 7.6$ Hz, 1, H4), 5.95 (d, $J_{1-2} = 3.7$ Hz, 1, H1); ¹³C NMR δ 21.40 (C8), 26.59 and 27.12 (CMe₂), 31.65 (d, $^2J_{7-F} = 26.6$ Hz, C7), 33.29 (C9), 52.08 (CH₃), 75.29 (d, $^3J_{4-F} = 4.9$ Hz, C4), 76.74 (d, $^4J_{3-F} = 1.0$ Hz, C3), 85.51 (C2), 101.15 (d, $^2J_{5-F} = 11.0$ Hz, C5), 104.74 (C1), 112.08 (CMe₂), 161.86 (d, $^1J_{6-F} = 261.8$ Hz, C6), 174.02 (C10); ¹⁹F NMR δ –100.23 (dt, $J_{F-H5} = 38.0$ Hz, $J_{F-H7} = 18.0$ Hz); MS m/z 305 (100, MH⁺).

5.19. Methyl 5,6,7,8,9-pentadeoxy-6-fluoro-1,2-*O*-isopropylidene- α -*D*-ribo-dec-5(*Z*)-enofuranuronate (**32**)

Saturated NH₃/MeOH (3 mL) was added to a solution of (*Z*)-**30** (26 mg, 0.062 mmol) in MeOH (3 mL) and the mixture was stirred at 0 °C for 48 h to ambient temperature. The volatiles were evaporated and the residue was column chromatographed (15 → 60% EtOAc/hexanes) to give **32** (13 mg, 69%): ¹H NMR δ 1.39 and 1.62 (2× s, 2× 3, 2× CH₃), 1.90 (quint, $J_{8-7/7'/9/9'} = 7.3$ Hz, 2, H8/8'), 2.31 (dt, $J_{7-F} = 17.6$ Hz, $J_{7-8/8'} = 7.4$ Hz, 2, H7/7'), 2.40 (t, $J_{9-8/8'} = 6.9$ Hz, 2, H9/9'), 3.75 (s, 3, CH₃), 4.56–4.72 (m, 4, H2/3/4/5), 5.82 (d, $J_{1-2} = 3.9$ Hz, 1, H1); ¹³C NMR δ 21.44 (C8), 26.81 and 26.94 (CMe₂), 31.74 (d, $^2J_{7-F} = 26.1$ Hz, C7), 33.20 (C9), 52.07 (CH₃), 73.94 (d, $^3J_{4-F} = 5.1$ Hz, C4), 76.80 (C2), 78.62 (C3), 103.63 (d, $^2J_{5-F} = 11.6$ Hz, C5), 104.00 (C1), 113.07 (CMe₂), 163.91 (d, $^1J_{6-F} = 262.70$ Hz, C6), 173.90 (C10); ¹⁹F NMR δ –100.23 (dt, $J_{F-H5} = 37.1$ Hz, $J_{F-H7/7'} = 18.1$ Hz). HRMS (AP-ESI) m/z calcd for C₁₄H₂₂FO₆ (MH⁺) 305.1395; found: 305.1396.

5.20. Methyl 5,6,7,8,9-pentadeoxy-6-fluoro- α/β -*D*-xylo-dec-5(*Z*)-enofuranuronate (**33**)

A solution of **31** (17 mg, 0.056 mmol) in TFA/H₂O (9:1, 3 mL) was stirred for 45 min at 0 °C and was evaporated and coevaporated [toluene (3×), CH₃CN (2×)]. The residue was dissolved in H₂O and the aqueous layer was extracted with ether (2×). The water layer was evaporated to give **33** (9 mg, 61%; α/β , 1:1): ¹H NMR (MeOH-*d*₄) δ 1.81–1.94 (m, 2, H8/8'), 2.27–2.38 (m, 2, H7/7'), 2.38–2.45 (m, 2, H9/9'), 3.67 (m, 3, CH₃), 3.92 (dd, $J_{3-4} = 3.8$ Hz, $J_{3-2} = 1.8$ Hz, 0.5, H3), 3.97 (dd, $J_{3-4} = 3.9$ Hz, $J_{3-2} = 2.6$ Hz, 0.5, H3), 4.00–4.04 (m, 1, H2), 4.91 (d, $J_{4-5} = 8.9$ Hz, 0.5, H4), 4.99 (d, $J_{4-5} = 9.1$ Hz, 0.5, H4), 5.04–5.12 (m, 1, H5), 5.08 (s, 0.5, H1 β), 5.37 (d, $J_{1-2} = 4.0$ Hz, 0.5, H1 α); ¹⁹F NMR δ –106.29 (dt, $J_{F-H5} = 37.2$ Hz, $J_{F-H7} = 17.6$ Hz, 0.5F), –106.87 (dt, $J_{F-H5} = 37.8$ Hz, $J_{F-H7} = 18.2$ Hz, 0.5F). HRMS (AP-ESI) m/z calcd for C₁₁H₁₈FO₆ (MH⁺) 265.1082; found: 265.1088.

5.21. Methyl 5,6,7,8,9-pentadeoxy-6-fluoro- α/β -D-ribo-dec-5(Z)-enofuranuronate (**34**)

A solution of **32** (12 mg, 0.04 mmol) in TFA/H₂O (9:1, 3 mL) was stirred for 30 min at 0 °C and was evaporated and coevaporated [toluene (3 \times)]. The residue was dissolved in H₂O and the aqueous layer was extracted with ether (2 \times). The water layer was evaporated to give **34** (8 mg, 76%; α/β , 3:7): ¹H NMR (D₂O) δ 1.72–1.76 (m, 2, H8/8'), 2.20 (dt, J_{7-F} = 18.1 Hz, $J_{7-8/8'}$ = 8.4 Hz, 2, H7/7'), 2.31–2.38 (m, 2, H9/9'), 3.59 (d, J_{2-3} = 2.4 Hz, 0.7, H2), 3.82–3.85 (m, 0.3, H2), 3.90–3.93 (s, 0.7, H3), 3.90–4.06 (m, 4.3, H3/H4/CH₃), 4.64–4.77 (m, 1, H5), 5.12 (s, 0.7, H1), 5.28 (d, J_{1-2} = 3.7 Hz, 0.3, H1); ¹⁹F NMR δ –104.06 (dt, J_{F-H5} = 36.4 Hz, $J_{F-H7/7'}$ = 17.8 Hz, 0.3, α); –105.04 (dt, J_{F-H5} = 35.8 Hz, $J_{F-H7/7'}$ = 18.8 Hz, 0.7F, β). HRMS (AP-ESI) m/z calcd for C₁₁H₁₈FO₆ (MH⁺) 265.1082; found: 265.1090.

5.22. (E/Z)-3,5,6-Trideoxy-6-fluoro-1,2-O-isopropylidene-6-phenylsulfonyl- α -D-erythro-hex-5-enofuranose (**35**)

Step (a). Treatment of diacetone 3-deoxyglucose²² (204 mg, 0.83 mmol) with H₅IO₆ (228 mg, 1.00 mmol) as described for **10** (Step (a), except no aqueous workup was performed) gave 3-deoxy-1,2-O-isopropylidene- α -D-erythro-pentodialdo-1,4-furanose [\sim 85% pure; ¹H NMR δ 9.75 (d, J_{5-4} = 4.8 Hz, H5)] was directly used in the next step. *Step (b)*. Treatment of the crude aldehyde with diethyl fluoro(phenylsulfonyl)methylphosphonate²⁰ (297 mg, 0.96 mmol) and LHMDs (0.96 mL, 0.96 mmol) as described for **20**, gave **35** (150 mg, 48%; E/Z, 2:1) as an inseparable mixture of isomers. Compound (E)-**35** had: ¹H NMR δ 1.30 and 1.47 (2 \times s, 2 \times 3, 2 \times CH₃), 1.71 (ddd, J_{3-4} = 10.9 Hz, $J_{3-3'}$ = 15.5 Hz, J_{3-2} = 4.6 Hz, 1, H3), 2.28 (dd, $J_{3'-4}$ = 4.5 Hz, $J_{3'-3}$ = 13.4 Hz, 1, H3'), 4.77 (t, $J_{2-1/3}$ = 4.0 Hz, 1, H2), 4.93–4.97 (m, 1, H4), 5.85 (d, J_{1-2} = 3.6 Hz, 1, H1), 6.31 (dd, J_{5-F} = 32.4 Hz, J_{5-4} = 7.5 Hz, 1, H5), 7.56–7.97 (m, 5, Ar); ¹³C NMR δ 26.42 and 27.00 (CMe₂), 39.25 (d, J_{3-F} = 2.0 Hz, C3), 71.26 (d, J_{4-F} = 2.4 Hz, C4), 80.69 (C2), 105.90 (C1), 112.01 (CMe₂), 116.77 (d, J_{5-F} = 3.7 Hz, C5), 129.17 (Ph), 129.91 (ph), 135.08 (Ph), 137.22 (Ph), 155.54 (d, J_{6-F} = 301.9 Hz, C6); ¹⁹F NMR δ –122.72 (d, J_{F-H5} = 32.5 Hz, 0.66). Compound (Z)-**35** had: ¹H NMR δ 1.34 and 1.60 (2 \times s, 2 \times 3, 2 \times CH₃), 1.68 (ddd, J_{3-4} = 10.5 Hz, $J_{3-3'}$ = 15.2 Hz, J_{3-2} = 4.7 Hz, 1, H3), 2.46 (ddd, $J_{3'-4}$ = 4.6 Hz, $J_{3'-3}$ = 13.2 Hz, 1, H3'), 4.79 (t, $J_{2-1/3}$ = 3.9 Hz, 1, H2), 5.71 (ddd, J_{4-5} = 8.7 Hz, J_{4-3} = 10.6 Hz, $J_{4-3'}$ = 4.5 Hz, 1, H4), 5.84–5.85 (m, 1, H1), 5.86 (dd, J_{5-F} = 20.1 Hz, J_{5-4} = 8.6 Hz, H5), 7.56–7.97 (m, 5, Ar); ¹⁹F NMR δ –114.04 (d, J_{F-H5} = 20.0 Hz, 0.33); MS m/z 329 (100, MH⁺).

5.23. (E/Z)-3,5,6-Trideoxy-6-fluoro-1,2-O-isopropylidene-6-tributylstannyl- α -D-erythro-hex-5-enofuranose (**38**)

Treatment of **35** (128 mg, 0.39 mmol) with Bu₃SnH (0.21 mL, 228 mg, 0.78 mmol) and AIBN (481 mg, 0.29 mmol) as described for **23**, gave **38** (83 mg, 44%; E/Z, 1:1): ¹⁹F NMR δ –92.83 (d, J_{F-H5} = 33.9 Hz, 84% of 0.50F, Z), –92.83 (ddd, J_{F-Sn} = 230.4 Hz, J_{F-H5} = 34.7 Hz, J_{F-H4} = 5.2 Hz 16% of 0.50F, Z), –96.75 (d,

J_{F-H5} = 52.7 Hz, 84% of 0.50F, E), –96.75 (ddd, J_{F-Sn} = 221.1 Hz, J_{F-H5} = 52.7 Hz, J_{F-H4} = 4.9 Hz, 16% of 0.50F, E); MS m/z 479 (100, MH⁺, ¹²⁰Sn), 477 (73, MH⁺, ¹¹⁸Sn), 475 (48, MH⁺, ¹¹⁶Sn). Compound (E)-**38** had: ¹H NMR δ 0.98–1.70 (m, 34, 3 \times Bu/2 \times CH₃/H3), 2.26 (dd, $J_{3'-4}$ = 4.3 Hz, $J_{3'-3}$ = 13.4 Hz, 1, H3), 4.45–4.55 (m, 1, H4), 4.75 (t, $J_{2-1/3}$ = 4.2 Hz, 1, H2), 4.96 (dd, J_{5-F} = 52.9 Hz, J_{5-4} = 7.5 Hz, 1, H5), 5.83 (d, J_{1-2} = 3.8 Hz, 1, H1); ¹³C NMR δ 10.31 (Bu), 11.00 (Bu), 13.97 (Bu), 27.04 and 27.08 (CMe₂), 27.50 (Bu), 40.19 (d, J_{3-F} = 1.6 Hz, C3), 71.25 (d, J_{4-F} = 17.3 Hz, C4), 81.06 (C2), 105.47 (C1), 111.41 (CMe₂), 123.42 (d, J_{5-F} = 3.7 Hz, C5), 174.29 (d, J_{5-F} = 323.5 Hz, C6). Compound (Z)-**38** had: ¹H NMR δ 0.98–1.70 (m, 34, 3 \times Bu/2 \times CH₃/H3), 2.11 (dd, $J_{3'-4}$ = 4.3 Hz, $J_{3'-3}$ = 13.4 Hz, 1, H3'), 4.73 (t, $J_{2-1/3}$ = 4.2 Hz, 1, H2), 5.21 (ddd, J_{4-5} = 7.5 Hz, J_{4-3} = 4.4 Hz, $J_{4-3'}$ = 15.2 Hz, 1, H4), 5.81 (d, J_{1-2} = 3.7 Hz, 1, H1), 5.84 (dd, J_{5-F} = 34.2 Hz, J_{5-4} = 9.2 Hz, 1, H5); ¹³C NMR δ 10.31 (Bu), 11.00 (Bu), 13.97 (Bu), 27.04 and 27.08 (CMe₂), 27.50 (Bu), 41.00 (d, J_{3-F} = 1.8 Hz, C3), 74.73 (d, J_{4-F} = 21.7 Hz, C4), 80.83 (C2), 105.69 (C1), 111.17 (CMe₂), 123.26 (d, J_{5-F} = 8.1 Hz, C5), 177.16 (d, J_{6-F} = 316.5 Hz, C6).

5.24. (E/Z)-5,6-Dideoxy-6-fluoro-1,2-O-isopropylidene- α -D-xyllo-hex-5-enofuranose (**39**)

Step (a). Compound **23** (200 mg, 0.34 mmol; E/Z, 7:3) was dissolved in saturated NH₃/MeOH (20 mL) and the resulting solution was stirred overnight at ambient temperature. The volatiles were evaporated to give **36** in quantitative yield of sufficient purity for use in the subsequent reaction. *Step (b)*. Compound **36** (crude from Step (a), 0.34 mmol) was dissolved in NH₃/MeOH (20 mL) and the resulting mixture was heated in a pressure Ace tube at 65 °C for 18 h. The volatiles were evaporated and the residue was column chromatographed (hexanes/EtOAc, 8:2 \rightarrow 3:7) to give (E)-**39** (20 mg, 29% from **23**) and (Z)-**39** (33 mg, 48% from **23**). Compound (E)-**39** had: ¹H NMR δ 1.35 and 1.58 (2 \times s, 2 \times 3, 2 \times CH₃), 1.78 (br s, 1, OH3), 4.32 (d, J_{3-4} = 2.5 Hz, 1, H3), 4.59 (d, J_{2-1} = 3.7 Hz, 1, H2), 4.69 ('dm', J_{4-5} = 7.0 Hz, 1, H4), 5.53 (ddd, J_{5-F} = 18.1 Hz, J_{5-6} = 11.2 Hz, J_{5-4} = 7.1 Hz, 1, H5), 5.94 (d, J_{1-2} = 3.7 Hz, 1, H1), 6.86 (ddd, J_{6-F} = 82.9 Hz, J_{6-5} = 11.2 Hz, J_{6-4} = 1.0 Hz, 1, H6); ¹³C NMR δ 26.47 and 27.06 (CMe₂), 76.48 (d, J_{3-F} = 2.0 Hz, C3), 76.80 (d, J_{4-F} = 12.6 Hz, C4), 85.31 (C2), 104.87 (C1), 106.20 (d, J_{5-F} = 13.7 Hz, C5), 112.25 (CMe₂), 153.79 (d, J_{6-F} = 262.6 Hz, C6); ¹⁹F NMR δ –122.18 (dd, J_{F-H5} = 17.8 Hz, J_{F-H6} = 82.9 Hz). Compound (Z)-**39** had: 1.35 and 1.54 (2 \times s, 2 \times 3, 2 \times CH₃), 1.81 (br s, 1, OH3), 4.22 (br s, 1, H3), 4.58 (d, J_{2-1} = 3.7 Hz, 1, H2), 5.07 (ddd, J_{5-F} = 40.1 Hz, J_{5-6} = 4.9 Hz, J_{5-4} = 7.5 Hz, 1, H5), 5.12–5.15 (m, 1, H4), 5.96 (d, J_{1-2} = 3.7 Hz, 1, H1), 6.63 (dd, J_{6-F} = 82.7 Hz, J_{6-5} = 4.8 Hz, J_{6-4} = 1.2 Hz, 1, H6); ¹³C NMR δ 26.57 and 27.10 (CMe₂), 74.49 (d, J_{4-F} = 5.1 Hz, C4), 76.74 (d, J_{3-F} = 1.9 Hz, C3), 85.47 (C2), 104.80 (C1), 106.24 (C5), 112.24 (CMe₂), 150.20 (d, J_{6-F} = 265.2 Hz, C6); ¹⁹F NMR δ –121.02 (dd, J_{F-H5} = 41.1 Hz, J_{F-H6} = 83.3 Hz). MS (APCI⁺) m/z 205 (100, MH⁺). Anal. Calcd for C₉H₁₃FO₄ (204.19): C, 52.94; H, 6.42. Found: C, 53.19; H, 6.63.

5.25. (*E/Z*)-5,6-Dideoxy-6-fluoro-1,2-*O*-isopropylidene- α -*D*-ribo-hex-5-enofuranose (**40**)

Step (a). Compound **24** (200 mg, 0.34 mmol; *E/Z*, 1:1) was dissolved in NH₃/MeOH (20 mL) and stirred overnight at ambient temperature. The volatiles were evaporated to give **37** in quantitative yield of sufficient purity for use in the subsequent step. *Step (b)*. Treatment of **37** (crude, 0.34 mmol) with NH₃/MeOH (20 mL) at 65 °C, as described for **39**, gave unchanged **37** (17 mg, 10% from **24**; *E/Z*, 2:3) and **40** as separable isomers (*E*; 22 mg, 32% from **24**) and (*Z*; 18 mg, 26% from **24**). Compound (*E*)-**40** had: ¹H NMR δ 1.40 and 1.60 (2 \times s, 2 \times 3, 2 \times CH₃), 2.37 (d, $J_{\text{OH3-3}}$ = 10.0 Hz, 1, OH3), 3.70–3.73 (m, 1, H3), 4.12 (t, $J_{4-5/3}$ = 8.3 Hz, 1, H4), 4.60 (t, $J_{2-1/3}$ = 4.6 Hz, 1, H2), 5.53 (ddd, J_{5-F} = 17.1 Hz, J_{5-6} = 11.2 Hz, J_{5-4} = 7.8 Hz, 1, H5), 5.84 (d, J_{1-2} = 3.9 Hz, 1, H1), 6.82 (ddd, J_{6-F} = 82.4 Hz, J_{6-5} = 11.1 Hz, J_{6-4} = 0.7 Hz, 1, H6); ¹³C NMR δ 26.75 and 26.84 (CMe₂), 76.55 (d, $^3J_{4-F}$ = 17.0 Hz, C4), 76.49 (C3), 78.57 (C2), 104.08 (C1), 109.44 (d, $^2J_{5-F}$ = 12.9 Hz, C5), 113.11 (CMe₂), 152.81 (d, $^1J_{6-F}$ = 262.1 Hz, C6); ¹⁹F NMR δ -123.67 (dd, J_{F-H5} = 17.1 Hz, J_{F-H6} = 82.5 Hz). Compound (*Z*)-**40** had: 1.40 and 1.60 (2 \times s, 2 \times 3, 2 \times CH₃), 2.39 (d, $J_{\text{OH3-3}}$ = 11.0 Hz, 1, OH3), 3.74 (ddd, $J_{3-\text{OH3}}$ = 10.9 Hz, J_{3-4} = 8.9 Hz, J_{3-2} = 5.1 Hz, 1, H3), 4.60 (t, $J_{2-1/3}$ = 4.5 Hz, 1, H2), 4.70 (t, $J_{4-3/5}$ = 8.8 Hz, 1, H4), 4.89 (ddd, J_{5-F} = 40.0 Hz, J_{5-6} = 4.9 Hz, J_{5-4} = 8.9 Hz, 1, H5), 5.84 (d, J_{1-2} = 3.9 Hz, 1, H1), 6.69 (ddd, J_{6-F} = 82.7 Hz, J_{6-5} = 4.9 Hz, J_{6-4} = 0.8 Hz, 1, H6); ¹³C NMR δ 26.79 and 26.92 (CMe₂), 73.18 (d, $^3J_{4-F}$ = 5.1 Hz, C4), 76.74 (d, $^4J_{3-F}$ = 2.0 Hz, C3), 78.58 (C2), 104.20 (C1), 108.69 ($^2J_{5-F}$ = 1.9 Hz, C5), 113.17 (CMe₂), 153.71 (d, $^1J_{6-F}$ = 265.8 Hz, C6); ¹⁹F NMR δ -123.90 (dd, J_{F-H5} = 40.1 Hz, J_{F-H6} = 82.6 Hz); MS (APCI⁺) *m/z* 205 (100, MH⁺). Anal. Calcd for C₉H₁₃FO₄ (204.19): C, 52.94; H, 6.42. Found: C, 53.07; H, 6.67.

5.26. (*E/Z*)-3,5,6-Trideoxy-6-fluoro-1,2-*O*-isopropylidene- α -*D*-erythro-hex-5-enofuranose (**41**)

Treatment of **38** (100 mg, 0.21 mmol; *E/Z*, 1:1) with NH₃/MeOH (15 mL) and CsF (51 mg, 0.33 mmol) at 65 °C for 4 h, as described for **39** (Step (b)), gave **41** (16 mg, 40%; *E/Z*, ~45:55): ¹⁹F NMR δ -124.79 (dd, J_{F-H5} = 41.8 Hz, J_{F-H6} = 83.0 Hz, 0.55F), -125.95 (dd, J_{F-H5} = 16.7 Hz, J_{F-H6} = 82.9 Hz, 0.45F); MS (APCI⁺) *m/z* 189 (100, MH⁺). Compound (*E*)-**41** had: ¹H NMR δ 1.35 and 1.55 (2 \times s, 2 \times 3, 2 \times CH₃), 1.55–1.68 (m, 1, H3), 2.20 (dd, $J_{3'-3}$ = 13.5 Hz, $J_{3'-4}$ = 4.3 Hz, 1, H3'), 4.62 (ddd, J_{4-3} = 11.5 Hz, J_{4-5} = 8.2 Hz, $J_{4-3'}$ = 4.3 Hz, 1, H4), 4.75–4.79 (m, 1, H2), 5.42 (ddd, J_{5-F} = 16.8 Hz, J_{5-6} = 11.2 Hz, J_{5-4} = 8.3 Hz, 1, H5), 5.82–5.84 (m, 1, H1), 6.80 (dd, J_{6-F} = 82.7 Hz, J_{6-5} = 11.2 Hz, 1, H6). Compound (*Z*)-**41** had: 1.28 and 1.57 (2 \times s, 2 \times 3, 2 \times CH₃), 1.55–1.68 (m, 1, H3), 2.28 (dd, $J_{3'-3}$ = 13.5 Hz, $J_{3'-4}$ = 4.3 Hz, 1, H3'), 4.75–4.79 (m, 1, H2), 4.92 (ddd, J_{5-F} = 41.3 Hz, J_{5-4} = 8.1 Hz, J_{5-6} = 4.8 Hz, 1, H5), 5.13 (ddd, J_{4-3} = 11.5 Hz, J_{4-5} = 8.0 Hz, $J_{4-3'}$ = 4.1 Hz, 1, H4), 5.82–5.84 (m, 1, H1), 6.53 (dd, J_{6-F} = 82.8 Hz, J_{6-5} = 4.8 Hz, 1, H6).

5.27. (*E*)-5,6-Dideoxy-6-fluoro- α/β -*D*-xylo-hex-5-enofuranose (**42**)

A solution of (*E*)-**39** (13 mg, 0.064 mmol) in TFA/H₂O (9:1; 3 mL) was stirred for 50 min at 0 °C (ice bath). The volatiles were evaporated, coevaporated [toluene (3 \times) and MeCN (2 \times)], and the residue was flash column chromatographed (50 \rightarrow 95% EtOAc/hexanes) to give **42** (4 mg, 38%; α/β , 1:1): ¹H NMR (MeOH-*d*₄) δ 3.91–4.03 (m, 2 H_{2/3}), 4.56–4.63 (m, 1, H4), 5.07 (s, 0.5, H1 β), 5.37 (d, J_{1-2} = 4.0 Hz, 0.5, H1 α), 5.51 (ddd, J_{5-F} = 17.8 Hz, J_{5-6} = 11.2 Hz, J_{5-4} = 8.9 Hz, 0.5, H5), 5.65 (ddd, J_{5-F} = 17.9 Hz, J_{5-6} = 11.1 Hz, J_{5-4} = 8.9 Hz, 0.5, H5), 6.87 (dd, J_{6-F} = 84.0 Hz, J_{6-5} = 11.0 Hz, 0.5, H6), 6.90 (dd, J_{6-F} = 83.7 Hz, J_{6-5} = 11.0 Hz, 0.5, H6); ¹³C NMR (MeOH-*d*₄) δ 75.29 (d, $^3J_{4-F}$ = 13.8 Hz, C4), 77.00 and 77.20 (C3), 77.73 (C2), 78.17 (d, $^3J_{4-F}$ = 13.7 Hz, C4), 81.26 (C2), 96.72 (C1 α), 103.17 (C1 β), 108.71 (d, $^2J_{5-F}$ = 12.0 Hz, C5), 109.32 (d, $^2J_{5-F}$ = 11.7 Hz, C5), 152.14 (d, $^1J_{6-F}$ = 258.7 Hz, C6), 152.24 (d, $^1J_{6-F}$ = 259.2 Hz, C6); ¹⁹F NMR (MeOH-*d*₄) δ -126.55 (dd, J_{F-H5} = 17.8 Hz, J_{F-H6} = 83.9 Hz, 0.5F), -126.85 (dd, J_{F-H5} = 18.1 Hz, J_{F-H6} = 84.0 Hz, 0.5F); MS (APCI⁻) *m/z* 163 (100, MH⁻).

Analogous treatment of **39** (*E/Z*, 1:1; 20 mg, 0.040 mmol) gave **42** (5 mg, 76%) as a mixture (*E/Z*, ~1:1; α/β , ~1:1). Compound (*E/Z*)-**37** had: ¹⁹F NMR (MeOH-*d*₄) δ -126.55 (dd, J_{F-H5} = 17.3 Hz, J_{F-H6} = 84.0 Hz; *E*, 0.25, α), -126.85 (dd, $^3J_{F-H5}$ = 17.5 Hz, $^2J_{F-H6}$ = 84.0 Hz; *E*, 0.25, β), -127.66 (dd, J_{F-H5} = 41.5 Hz, J_{F-H6} = 84.9 Hz; *Z*, 0.25, α), -128.34 (dd, J_{F-H5} = 42.2 Hz, J_{F-H6} = 84.8 Hz; *Z*, 0.25, β); MS (APCI⁻) *m/z* 163 (100, MH⁻).

Treatment of the crude **36** [from Step (a) for the preparation of **39**] with TFA/H₂O (1 h, 0 °C) followed by evaporation, coevaporation [toluene (2 \times) and MeCN (1 \times)], partition (H₂O/ethyl ether), and evaporation of the aqueous layer also gave **42** (55% from **23**, α/β , 1:1).

5.28. (*E*)-5,6-Dideoxy-6-fluoro- α/β -*D*-ribo-hex-5-enofuranose (**43**)

Treatment of (*E*)-**40** (13 mg, 0.064 mmol) with TFA/H₂O (9:1, 3 mL) as described for **42**, gave **43** (7 mg, 67%; α/β , 1:4): ¹H NMR (MeOH-*d*₄) δ 3.77 (t, $J_{3-2/4}$ = 6.1 Hz, 0.2, H3), 3.87 (d, J_{2-1} = 4.5 Hz, 0.8, H2), 4.01–4.05 (m, 1, H2 α and H3 β), 4.20 (t, $J_{4-3/5}$ = 8.0 Hz, 0.8, H4), 4.30 (dd, J_{4-5} = 8.2 Hz, J_{4-3} = 6.4 Hz, 0.2, H4), 5.12 (br s, 0.8, H1), 5.28 (d, J_{1-2} = 4.1 Hz, 0.2, H1), 5.42 (ddd, J_{5-F} = 17.5 Hz, J_{5-6} = 11.1 Hz, J_{5-4} = 8.3 Hz, 0.2, H5), 5.49 (ddd, J_{5-F} = 17.6 Hz, J_{5-6} = 11.1 Hz, J_{5-4} = 8.5 Hz, 0.8, H5), 6.86 (dd, J_{6-F} = 83.9 Hz, J_{6-5} = 11.1 Hz, 0.8, H6), 6.87 (dd, J_{6-F} = 83.7 Hz, J_{6-5} = 11.0 Hz, 0.2, H6); ¹³C NMR (MeOH-*d*₄) δ 70.90 (C2 α), 75.32 (d, $^4J_{3-F}$ = 2.6 Hz, C3 α), 75.5 (d, $^4J_{3-F}$ = 2.5 Hz, C3 β), 76.13 (C2 β), 77.55 (d, $^3J_{4-F}$ = 13.6 Hz, C4 α), 77.88 (d, $^3J_{4-F}$ = 13.7 Hz, C4 β), 96.63 (C1 α), 102.09 (C1 β), 111.19 (d, $^2J_{5-F}$ = 11.4 Hz, C5 α), 112.65 (d, $^2J_{5-F}$ = 10.6 Hz, C5 β), 151.98 (d, $^1J_{6-F}$ = 258.6 Hz, C6 β), 152.17 (d, $^1J_{6-F}$ = 258.7 Hz, C6 α); ¹⁹F NMR (MeOH-

d_4) δ -128.55 (dd, $J_{F-H5} = 17.4$ Hz, $J_{F-H6} = 83.5$ Hz, $0.2F$, α), -129.00 (dd, $J_{F-H5} = 17.3$ Hz, $J_{F-H6} = 83.7$ Hz, $0.8F$, β); MS (APCI⁻) m/z 163 (100, MH⁻).

Analogous treatment of **40** (E/Z , 1:1; 16 mg, 0.032 mmol) gave **43** (3 mg, 57%) as a mixture (E/Z , $\sim 3:1$; α/β , $\sim 1:4$ for E isomer and α/β , $\sim 1:15$ for Z isomer): ¹⁹F NMR (MeOH- d_4) δ -128.01 (dd, $J_{F-H5} = 41.3$ Hz, $J_{F-H6} = 83.4$ Hz; Z , 0.02F, α) -128.55 (dd, $J_{F-H5} = 17.6$ Hz, $J_{F-H6} = 84.2$ Hz; E , 0.14F, α), -129.00 (dd, $J_{F-H5} = 17.5$ Hz, $J_{F-H6} = 83.8$ Hz; E , 0.60F, β), -129.69 (dd, $J_{F-H5} = 40.8$ Hz, $J_{F-H6} = 84.4$ Hz; Z , 0.24F, β); MS (APCI⁻) m/z 163 (100, MH⁻).

Treatment of the crude **37** [from Step (a) for the preparation of **40**] with TFA/H₂O (1 h, 0 °C) followed by evaporation, coevaporation [toluene (2 \times) and MeCN (1 \times)], partition (H₂O/ethyl ether), and evaporation of the aqueous layer also gave **43** (45% from **24**, α/β , 1:3).

5.29. (E/Z)-3,5,6-Trideoxy-6-fluoro- α -D-erythro-hex-5-enofuranose (**44**)

Treatment of **38** (62 mg, 0.13 mmol; E/Z , 3:2) with TFA/H₂O (9:1, 1 mL; 1 h, 0 °C) followed by evaporation, coevaporation [toluene (2 \times) and MeCN (1 \times)], partition (H₂O/ethyl ether), and evaporation of the aqueous layer gave **44** (10 mg, 52%; E/Z $\sim 1:3$, α/β , $\sim 1:4$): ¹H NMR (D₂O) δ 1.85–2.10 (m, 2, H3,3'), 4.05–4.25 (m, 1, H2), 4.58–4.75 (m, 1, H4), 4.86 (ddd, $J_{5-F} = 41.7$ Hz, $J_{5-4} = 8.9$ Hz, $J_{5-6} = 4.7$ Hz, 0.15, H5), 4.92 (ddd, $J_{5-F} = 41.6$ Hz, $J_{5-4} = 8.9$ Hz, $J_{5-6} = 4.7$ Hz, 0.6, H5), 5.12 (s, 0.15, H1 β), 5.13 (s, 0.6, H1 β), 5.24 (d, $J_{1-2} = 3.5$ Hz, 0.05, H1 α), 5.25 (d, $J_{1-2} = 3.8$ Hz, 0.2, H1 α), 5.39 (ddd, $J_{5-F} = 17.4$ Hz, $J_{5-6} = 11.2$ Hz, $J_{5-4} = 9.3$ Hz, 0.2, H5), 5.45 (ddd, $J_{5-F} = 17.6$ Hz, $J_{5-6} = 11.1$ Hz, $J_{5-4} = 9.1$ Hz, 0.05, H5), 6.52 (ddd, $J_{6-F} = 83.7$ Hz, $J_{6-5} = 4.7$ Hz, $J_{6-4} = 1.0$ Hz, 0.15, H6), 6.55 (dd, $J_{6-F} = 83.7$ Hz, $J_{6-5} = 4.7$ Hz, $J_{6-4} = 1.0$, 0.60, H6), 6.77 (dd, $J_{6-F} = 83.9$ Hz, $J_{6-5} = 10.7$ Hz, 0.05, H6), 6.78 (dd, $J_{6-F} = 83.9$ Hz, $J_{6-5} = 11.0$ Hz, 0.20, H6); ¹⁹F NMR (D₂O) δ -126.35 (dd, $J_{F-H5} = 17.3$ Hz, $J_{F-H6} = 83.6$ Hz; E , 0.20F, β), -126.45 (dd, $J_{F-H5} = 17.2$ Hz, $J_{F-H6} = 82.8$ Hz; E , 0.05F, α), -126.81 (dd, $J_{F-H5} = 42.0$ Hz, $J_{F-H6} = 83.7$ Hz; Z , 0.15F, α), -127.55 (dd, $J_{F-H5} = 42.0$ Hz, $J_{F-H6} = 83.8$ Hz; Z , 0.60F, β); HRMS (LCT-ESI) m/z : calcd for C₆H₉FO₃ [M+Na]⁺ 171.0433; found: 171.0434.

Analogous treatment of **41** (10 mg, 0.053 mmol; E/Z , $\sim 45:55$) with TFA/H₂O gave **44** (5 mg, 64%; E/Z $\sim 1:2$, α/β $\sim 1:3$).

5.30. Enzymatic assay

Inhibition assays were performed in a buffer containing 50 mM Hepes (pH 7.0), 150 mM NaCl, 150 μ M 5,5'-dithio-bis-(2-nitrobenzoic acid),²³ and various concentrations of SRH (0–55 μ M) and inhibitors (0–1 mM). The reactions were initiated by the addition of Co²⁺-substituted LuxS from *B. subtilis* (final concentration 0.4–0.5 μ M) and monitored continuously at 412 nm ($\epsilon = 14,150$ M⁻¹ cm⁻¹) in a Perkin-Elmer λ 25 UV–vis

spectrophotometer at room temperature. The initial rates recorded from the early regions of the progress curves were fitted into the Lineweaver–Burk equation $1/V = K_M'/(k_{cat} [E]_0) \times 1/[S] + 1/(k_{cat} [E]_0)$ and the Michaelis–Menten equation $V = k_{cat} [E]_0 [S]/(K_M' + [S])$ using KaleidaGraph 3.5 to determine the inhibition pattern. K_I values were calculated from the equation $K_M' = K_M \times (1 + [I]/K_I)$, where $K_M = 2.2$ μ M.

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