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Letter

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## **Optimization of 3-Pyrimidin-4-yl-oxazolidin-2-ones as Orally Bioavailable and Brain Penetrant Mutant IDH1 Inhibitors**

Qian Zhao,\*<sup>,†</sup> James R. Manning,<sup>\*,†</sup> James Sutton,<sup>†,‡</sup> Abran Costales,<sup>†,§</sup> Martin Sendzik,<sup>†,⊥</sup> Cynthia M. Shafer,<sup>†</sup> Julian R. Levell,<sup>⊥,∥</sup> Gang Liu,<sup>⊥</sup> Thomas Caferro,<sup>⊥</sup> Young Shin Cho,<sup>⊥</sup> Mark Palermo,<sup>⊥</sup> Gregg Chenail,<sup>⊥</sup> Julia Dooley,<sup>⊥</sup> Brian Villalba,<sup>⊥</sup> Ali Farsidjani,<sup>⊥</sup> Jinyun Chen,<sup>⊥</sup> Stephanie Dodd,<sup>⊥</sup> Ty Gould,<sup>⊥</sup> Guiqing Liang,<sup>⊥</sup> Kelly Slocum,<sup>⊥</sup> Minying Pu,<sup>⊥</sup> Brant Firestone,<sup>⊥</sup> Joseph Growney,<sup>⊥</sup> Tycho Heimbach,<sup>⊥</sup> and Raymond Pagliarini<sup>⊥</sup>

<sup>†</sup>Novartis Institutes for BioMedical Research, 5300 Chiron Way, Emeryville, California 94608, United States

<sup>1</sup>Novartis Institutes for BioMedical Research, 250 Massachusetts Avenue, Cambridge, Massachusetts 02139, United States

KEYWORDS mutant IDH1, inhibition of 2-HG production, in vivo anticancer activity, brain penetration, clinical candidate

**ABSTRACT:** Mutant isocitrate dehydrogenase 1 (IDH1) is an attractive therapeutic target for the treatment of various cancers such as AML, glioma and glioblastoma. We have evaluated 3-pyrimidin-4-yl-oxazolidin-2-ones as mutant IDH1 inhibitors that bind to an allosteric, induced pocket of IDH1<sup>R132H</sup>. This letter describes SAR exploration focused on improving both the *in vitro* and *in vivo* metabolic stability of the compounds, leading to the identification of **19** as a potent and selective mutant IDH1 inhibitor that has demonstrated brain penetration and excellent oral bioavailability in rodents. In a preclinical patient-derived IDH1 mutant xenograft tumor model study, **19** efficiently inhibited the production of the biomarker, 2-HG.

As an isocitrate dehydrogenase, NADP<sup>+</sup>-dependent IDH1 catalyzes the oxidative decarboxylation of isocitrate to produce  $\alpha$ ketoglutarate ( $\alpha$ -KG) as part of the citric acid cycle in aerobic metabolism,<sup>1-5</sup> while NADP<sup>+</sup> is reduced to NADPH.<sup>2-4</sup> Heterozygous mutations in human IDH1 at Arg<sup>132</sup> (R132\*) have been identified in multiple cancer types, including acute myeloid leukemia (AML), glioblastoma, glioma, chondrosarcoma and cholangiocarcinoma. The gain-of-function mutant loses normal enzymatic function and reduces a-KG to 2hydroxyglutarate (2-HG), resulting in elevated intracellular levels of this metabolite.<sup>6</sup> 2-HG has been found to inhibit enzymatic function of many  $\alpha$ -KG-dependent dioxygenases, including histone and DNA demethylases, causing widespread alterations in histone and DNA methylation which may potentially contribute to tumorigenesis.<sup>7</sup> Recently, a first-in-class inhibitor of mutant IDH1 has shown efficacy against IDH1mutant glioma and cholangiocarcinoma in clinical studies.<sup>8,9</sup> A number of preclinical studies also provided evidence to support targeting mutant IDH1 tumors with small molecule inhibitors.10-15

We recently reported the identification of clinical candidate **IDH305** (3) as a brain penetrant, potent and selective mutant IDH1 inhibitor (Figure 1).<sup>16</sup> A major part of the lead optimization efforts focused on improving intrinsic clearance without compromising physicochemical properties (e.g. solubility). In this letter, we describe the rational design of fluoromethyl and fluoroethyl (as in 3) substituted oxazolidinones to enhance metabolic stability.



Figure 1. SAR progression from hit-to-lead to clinical candidate

As reported previously, our hit-to-lead optimization efforts from 1 produced a potent and selective lead compound, **IDH889** (2).<sup>17</sup> Oral dosing of 2 in a murine IDH1 mutant tumor xenograft model resulted in robust reduction of tumorderived 2-HG, a PD biomarker of mutant IDH1<sup>R132\*</sup> activity. However, in addition to low aqueous solubility (39 µM at pH 6.8). 2 also displayed poor rodent PK due to its high *in vivo* clearance (Table 1). This correlated with high in vitro intrinsic clearance  $(CL_{int})$ across different species (rat/mouse/dog/human 588/143/548/205 µL/min/mg). Thus, improving the CL<sub>int</sub> was identified as a top priority for the series.

*In silico* and *in vivo* metabolic studies had identified debenzylation resulting from benzylic oxidation as a major metabolic pathway for **1** (Scheme 1). Common strategies to improve metabolic stability focus on: reducing the lipophilicity of the molecule to affect binding to cytochrome P450 proteins (CYPs), and blocking metabolically labile sites.<sup>19</sup>

#### Scheme 1. Major Metabolite of 1 in Rat Liver Microsomes



Our early optimization efforts focused on the exploration of 5membered heterocyclic aromatic rings on the amine side chain, which were expected to reduce benzylic oxidation<sup>2</sup> through the inductive effect of the electronegative heteroatoms and reduction of the overall lipophilicity of the molecule. We elected to maintain the isopropyl group on the oxazolidinone ring throughout our initial studies because it enhanced potency relative to compounds lacking this moiety. This is likely due both to its interaction with the hydrophobic protein pocket, and, more importantly, its van der Waals contact with the aryl group of the amine side chain. Indeed, both the bound and small molecule X-ray structures of 1 overlay closely, suggesting a prearrangement of the molecule into the active binding conformation that is driven by hydrophobic collapse.<sup>17,18</sup> We believe this ligand preorganization is critical for maintaining mutant IDH1 potency. Additionally, a co-crystal structure of  $2^{17}$  with the homodimer of IDH1<sup>R132H</sup> showed a hydrogen bond between Ser<sup>278</sup> and one of the ortho nitrogen atoms in the pyrimidine ring. Therefore, in our optimization efforts, at least one hydrogen bond acceptor, either a nitrogen or oxygen atom, was maintained in ring A (Figure 1).

Scheme 2. Major Metabolites of 4 in Rat Liver Microsomes



Rat liver microsomal CL<sub>int</sub> 924 µL/min/mg

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Most of the heterocyclic analogs thus prepared did indeed attenuate the aforementioned benzylic oxidation observed in Met ID studies; however, the rat *in vitro* microsomal  $CL_{int}$  remained high. For example, benzylic oxidation of oxadiazole **4** was reduced during incubation with rat liver microsomes (Scheme 2), but the major observed metabolites now resulted from oxazolidinone ring opening and oxidation. The change in the site of metabolism observed with **4** was not predicted by *in silico* modeling, but we were nonetheless encouraged by the reduced benzylic oxidation observed with **4** and *para*-chloro analog **5**, which is more potent and has improved  $CL_{int}$ . The tetra-methyl substituted analog of **5** (**6**) (Figure 2) was designed and synthesized to block oxazolidinone ring oxidation. Unfortunately, this resulted in a 20-fold loss in potency, and only a slight improvement in  $CL_{int}$  was observed.



Figure 2. Biochemical Potency and Rat Liver Microsomal  $CL_{int}$  for 5 and 6.

In parallel to the work on the heteroaromatic analogs, compounds that displayed acceptable CL<sub>int</sub> (although were insufficiently potent) were evaluated to understand what structural characteristics were important for metabolic stability. For instance, unsubstituted oxazolidinone 7 (Figure 3) had low clearance, while the corresponding isopropyl analog 8 displayed high clearance. It seemed counterintuitive that analogs containing sterically encumbered oxazolidinone rings would undergo ring metabolism more readily than their unsubstituted congeners. We hypothesized that the substituents on the oxazolidinone might play a role in CYP recognition and binding that could be important for the metabolic stability of the molecules.



Figure 3. Biochemical Potency and Rat Liver Microsomal  $CL_{int}$  for 7 and 8.

The co-crystal structures of both **IDH125** and **IDH889**<sup>17</sup> showed van der Waals interactions between the isopropyl group and ring A. Thus, the decreased potency of **7** relative to isopropyl analog **8** was not surprising because of the ligand preorganization resulting from the aforementioned hydrophobic collapse.<sup>17</sup> However, the relative importance of the isopropyl group to inducing the hydrophobic collapse was unclear. Therefore, the unsubstituted oxazolidinone was viewed as providing a starting point for building in potency while maintaining good metabolic stability. In addition to high clearance, isopropyl analogs also generally displayed low aqueous solubility and high plasma protein binding across species. Substituents that would reduce lipophilicity as compared to isopropyl would likely improve the overall physicochemical profile of the series and lead to better drug-like properties.

Toward this goal, an effort was initiated to evaluate various substituents on both the oxazolidinone and the biaryl amine side chain to identify a suitable combination of metabolic stability and potency. A number of 5,6- and 6,6-membered biaryl analogs were synthesized, and the imidazole analog 9 was identified as a promising lead, with a cellular  $IC_{50}$  of 0.5  $\mu M$ and much improved in vitro CLint across species (mouse/rat/human 73/34/59 µL/min/mg). The addition of a methyl group onto the oxazolidinone ring (10) increased cellular potency 10-fold, while only moderately increasing the rat  $CL_{int}$  (103 µL/min/mg) as compared to 2 and other isopropyl analogs (Table 1). Compound 10 also displayed significantly lower in vivo clearance vs 2 (17 vs 41 mL/min/kg) during pharmacokinetic profiling in rat (IV/PO 1/10 mg/kg). This translated to a 35-fold increase in oral AUC and a 17-fold improvement in bioavailability vs 2 at the same oral dosage.

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Table 1. Cellular Potency, *in vitro* CL<sub>int</sub> and Rat PK Data of 9, 10 and 2



	9	10	2
cellular HCT116-IDH1 <sup>R132H/+</sup> (IC <sub>50</sub> / $\mu$ M)	0.56	0.047	0.014
<i>in vitro</i> liver microsomal CL <sub>int</sub> (µL/min/mg)	34	101	588
<i>in vivo</i> plasma clearance (mL/min/kg)	n.d. <sup>a</sup>	17	41
oral AUC ( $\mu M \cdot h$ )	n.d.	14	0.4
oral bioavailability (%)	n.d.	67	4

<sup>&</sup>lt;sup>a.</sup> Not determined

Given these encouraging results, we continued to explore the oxazolidinone substitutions to further improve the stability and potency of the imidazole series. We anticipated that the oxazolidinone SAR would translate beyond the imidazole series described here and that the findings could prove useful for the optimization of the other biaryl series as well. The data summarized in Table 2 highlights both steric and electronic effects of the substituents on cellular IC50 and microsomal CLint. Ethyl analog 11 was about 3-fold more potent than methyl analog 10; however, the CL<sub>int</sub> of 11 was significantly higher. Introduction of a fluorine atom generated a pair of fluoroethyl diastereomers, 12 and 13. While both isomers demonstrated a trend towards improved stability over ethyl analog 11, Sisomer 13 was shown to be the more potent diastereomer by 10-fold over 12. A significant improvement in stability was obtained with fluoromethyl analog 14 with maintenance of cellular potency.

Throughout our medicinal chemistry campaign, both the 5fluoro and 5-hydrogen pyrimidine cores had been explored, and the 5-substituent was identified as a handle for the finetuning of potency, lipophilicity and physicochemical properties. As observed in the other biaryl series, isopropyl analog **15** displayed good potency and high  $CL_{int}$ . The (*S*)-fluoroethyl analog of **15** (**16**) demonstrated an improvement in potency (4fold) and stability vs **15**. Once again, lower  $CL_{int}$  was observed with fluoromethyl analog **17** relative to **16**. However, unlike with the 5-fluoro pyrimidines (**13-14**), the cellular potency dropped over 7-fold from **16** to **17**.

Encouraged by the improvement in  $CL_{int}$  with the fluoromethyl and (*S*)-1-fluoroethyl oxazolidinones, we decided to modify the phenyl substitutions of the biaryl amine side chain and identified a number of potent compounds with low to medium  $CL_{int}$ , exemplified by **18** and **19**.

Table 2. Cellular Potency and  $\ensuremath{CL_{int}}$  in Rat liver Microsomes



Cmpd	Х	Y	R	Cellular HCT116- IDH1 <sup>R132H</sup>	Rat liver microsomal CL <sub>int</sub>
				/+ (IC <sub>50</sub> /μM)	(µL/min/m g)
10	Cl	F	Me	0.047	103
11	Cl	F	Et	0.013	448
12	Cl	F	( <i>R</i> )-1- F-Et	0.24	237
13	Cl	F	( <i>S</i> )-1- F-Et	0.022	272
14	Cl	F	F-Me	0.024	28
15	Cl	Н	<i>i</i> -Pr	0.015	489
16	Cl	Н	( <i>S</i> )-1- F-Et	0.004	148
17	Cl	Н	F-Me	0.030	75
18	CF <sub>2</sub> H	Н	( <i>S</i> )-1- F-Et	0.015	72
19	CF <sub>3</sub>	Н	F-Me	0.039	7

Both 18 and 19 demonstrated a good balance of cellular potency, *in vitro* clearance, permeability and aqueous solubility (Table 3). Although 19 was about 2-fold less potent than 18, its superior microsomal CL<sub>int</sub> correlated with significantly improved *in vivo* clearance, AUC and oral bioavailability in rat vs 18. When rat plasma protein binding data were used to approximate unbound plasma AUC, 10 mg/kg oral dosing of 18 and 19 provided cellular IC<sub>50</sub> coverage for roughly the same amount of time (Figure SI-1). The AUC brain to plasma ratio of 19 in rat at 10 mg/kg was 0.17, suggesting 19 is brain penetrant. The limited exposure of 18 in rat brain is presumably due to the p-glycoprotein driven efflux mechanism (MDR1-MDCK efflux ratio >9).

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Table 3. Physicochemical Properties and Rodent PK Data for 18 and 19 (IV/PO 1/10 mg/kg for rat and mouse)

	10	10
	18	19
cellular HCT116-IDH1 <sup>R132H/+</sup> IC <sub>50</sub> (μM)	0.015	0.039
solubility at pH 6.8 (µM)	422	124
Caco-2 Papp (AB/BA)	21/31	20/24
MDR1-MDCK (AB/BA)	6/57	11/39
plasma protein binding (rat/mouse)	95/92	98/98
<i>in vitro</i> liver microsomal CL <sub>int</sub> (µL/min/mg) (rat/mouse)	72/63	7.7/11
<i>in vivo</i> plasma clearance (mL/min/kg) (rat/mouse)	20/n.d. <sup>a</sup>	2.6/5
oral AUC ( $\mu$ M·h) (rat/mouse)	15/n.d.	180/172
oral bioavailability(%) (rat/mouse)	82/n.d.	127/ 149
brain to plasma (total) AUC ratio (rat)	< 0.1	0.17

Not determined

Compound 19 was evaluated for specificity against mutant IDH1 relative to wild-type (WT) IDH1 in a biochemical assay measuring consumption (mutant) or production (WT) of NADPH.<sup>21</sup> Compound **19** exhibited over 50 to 100-fold selectivity for mutant IDH1 isoforms vs WT (IDH1<sup>R132H</sup> IC<sub>50</sub> 0.021  $\mu$ M; IDH1<sup>R132C</sup> IC<sub>50</sub> 0.045  $\mu$ M; IDH1<sup>WT</sup> IC<sub>50</sub> 2.52  $\mu$ M). Compound **19** was also profiled in a patient-derived IDH1 mutant HCT116-IDH1<sup>R132H/+</sup> mechanistic xenograft tumor model in mice to evaluate in vivo inhibition of 2-HG production. Compound 19, dosed orally at 150 mg/kg, inhibited new 2-HG production. Given the high baseline tissue concentration of 2-HG, it takes several hours for it to be cleared from tissues, as observed previously in the pharmacodynamic (PD) study with **2**.<sup>17</sup> Unbound plasma concentration of **19**, approximated from mouse plasma protein binding of 98%, stayed above the HCT116-IDH1<sup>R132H/+</sup> cellular IC<sub>50</sub> for 24 hours. This resulted in reduction of tumor 2-HG concentration (Figure 4), with maximal 2-HG inhibition of over 90% relative to the vehicletreated tumors at 16-24 hours post-treatment. As a comparison, when 3 was dosed orally at 200 mg/kg in the same PK/PD study,<sup>16</sup> 2-HG levels began to rebound after 16 hours posttreatment as the free concentration of 3 dropped below the in vitro cellular IC50.



Figure 4. Percent 2-HG inhibition in HCT116-IDH1<sup>R132H/+</sup> xenograft tumor tissue in correlation with total and calculated unbound plasma concentration following (a) 150 mg/kg dose of 19 and (b) 200 mg/kg dose of 3.

The synthesis of **19** is described in Scheme 3. Condensation of (S)-2-methylpropane-2-sulfinamide<sup>22</sup> and *tert*-butyl 4-formyl-1H-imidazole-1-carboxylate provided 20 in 74% yield. The of methylmagnesium bromide to addition tertbutanesulfinimine 20,<sup>23</sup> followed by Boc deprotection gave 21 in 86% yield over two steps. Palladium-catalyzed cross $coupling^{24}$  of **21** with 1-bromo-4-(trifluoromethyl)benzene followed by removal of the tert-butanesulfinyl group, provided amine 22 efficiently in 81% yield over two steps. Silyl protection of (S)-4-(hydroxymethyl)oxazolidin-2-one (23) produced 24 in 86% yield, and the subsequent  $S_NAr$  reaction of 24 with 2,4-difluoropyrimidine afforded compound 25 in 81% yield. The removal of the silvl protecting group and subsequent conversion to fluoromethyl compound 26 was achieved efficiently in one-pot using perfluorobutanesulfonyl fluoride (PFBSF) and triethylamine trihydrofluoride.<sup>24</sup> Coupling of 22 and 26 in DMSO then provided 19 in 74% yield.

Scheme 3. Synthesis of 19

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Examples can be found in the literature demonstrating that metabolism can be reduced for some molecules when a metabolically labile C-H is replaced by C-F. These include the cholesterol-absorption inhibitor Ezetimibe<sup>26</sup> and the cathepsin K inhibitor Odanacatib<sup>27</sup> for the treatment of osteoporosis. Multiple factors may contribute to the improvement of CL<sub>int</sub> of the fluoroalkyl-substituted analogs (12-14, 16-19), including a reduction in potential for oxazolidinone ring oxidation by an inductive effect of the electronegative fluorine atom. Another factor could be the effect of fluorine on molecular lipophilicity. Examples of a hydrogen-to-fluorine substitution decreasing lipophilicity have been reported by Roche scientists.<sup>28</sup> Those compounds contained a common motif whereby an oxygen atom was located either two or three carbons away from a fluorine. The decreased lipophilicity was attributed to the polarization of the oxygen atom by the neighboring fluorine. Interestingly, this structural pattern is also observed in our fluoroalkyl analogs. It is noted that some modifications to the biaryl amine side chain modestly improved CL<sub>int</sub> of the isopropyl-substituted analogs, but not as substantially as the replacement of the isopropyl group with methyl, fluoromethyl or fluoroethyl groups. This suggests that the compounds may be recognized differently within the CYP enzyme active site, and that the substituents on the oxazolidinone ring can affect the binding of the compounds in the active site.

In conclusion, our optimization of lead compound 2 with a focus on improving *in vitro* CL<sub>int</sub> led to the discoveries of both the imidazole subseries and fluoroalkyl-substituted oxazolidinones that demonstrated improved metabolic stability. Potent, orally bioavailable and brain-penetrant mutant IDH1 inhibitor **19** effectively inhibited 2-HG production in a preclinical xenograft tumor model. The fluoroalkyl oxazolidinone SAR was also applied to other biaryl series and a number of compounds with good potency and desirable *in vivo* properties, including the clinical candidate **IDH305**, were subsequently identified.

#### ASSOCIATED CONTENT

#### **Supporting Information**

Synthetic procedures, analytical data, assay protocols, PK, PK/PD and efficacy study protocols and data (PDF). This material is available free of charge on the ACS Publications website.

#### AUTHOR INFORMATION

#### **Corresponding Authors**

\* Email: qianz53705@gmail.com.

\* Tel: (510) 879-9429. Email: james.manning@novartis.com.

#### Present Addresses

<sup>‡</sup> (J.S.) Ideaya Biosciences, 280 Utah Avenue, Suite 250, South San Francisco, CA 94080

<sup>§</sup>(A.C.) Chevron Oronite LLC, 100 Chevron Way, Richmond, CA 24802

[(J.R.L.) Constellation Pharmaceuticals, 215 First Street, Suite 200, Cambridge, MA 02142

#### Notes

The authors declare the following competing financial interest(s): The authors are all current or former employees of Novartis and some own shares in Novartis.

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Cell 2-HG IC<sub>50</sub> 0.014 μM Rat liver microsomal CL<sub>int</sub> 588 μL/min/mg Rat 10 mg/kg oral AUC 0.4 μM·h

Cell 2-HG IC<sub>50</sub> 0.039 μM Rat liver microsomal CL<sub>int</sub> 7 μL/min/mg Rat 10 mg/kg oral AUC 180 μM·h