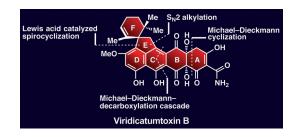


Communications



Total Synthesis

Total Synthesis and Structural Revision of Viridicatumtoxin B



Will the real viridicatumtoxin B please stand up: The total synthesis of viridicatumtoxin B resulted in its structural revision and opens the way for analogue construction and biological evaluation of this complex tetracycline-like antibiotic. The highly convergent strategy employed allows for swift construction of the entire carbocyclic framework of the molecule.



Total Synthesis

DOI: 10.1002/anie.201304691

Total Synthesis and Structural Revision of Viridicatumtoxin B**

K. C. Nicolaou,* Christian Nilewski, Christopher R. H. Hale, Heraklidia A. Ioannidou, Abdelatif ElMarrouni, and Lizanne G. Koch

The discovery and development of antibiotics in the first half of the 20th century revolutionized medicine and led to a dramatic enhancement of life expectancy.[1] However, the emergence of drug-resistant bacterial strains caused by the extensive use of antibiotics in both humans and livestock risks rendering the world's arsenal of antibiotics ineffective. [2] The development of novel antibacterial agents is, therefore, of high priority.

The nefarious molecular architectures of tetracyclines have challenged synthetic chemists for over six decades,[3] with impressive results being reported most recently from the Myers research group.^[4] Structurally related to the tetracycline antibiotics are the viridicatumtoxins (1-3, Figure 1), which constitute an intriguing subclass of naturally occurring potent antibacterial agents. Originally isolated in 1973 from a Penicillium species, [5] viridicatumtoxin A (1) was structurally elucidated through the aid of X-ray crystallographic analysis in 1976. [6] In 2008, Kim and co-workers reported the re-isolation of viridicatumtoxin A (1) as well as the isolation of viridicatumtoxin B, whose structure was assigned as 2 (Figure 1).^[7] Finally, spirohexaline (3) was reported in early 2013.[8] Viridicatumtoxins A (1) and B (2) were shown to exhibit antibacterial activity against a variety of Grampositive bacterial strains, including methicillin-resistant (MRSA) Staphylococcus aureus MIC = 0.25

[*] Prof. Dr. K. C. Nicolaou, Dr. C. Nilewski, [+] C. R. H. Hale, [+] Dr. H. A. Ioannidou, Dr. A. ElMarrouni, L. G. Koch Department of Chemistry and The Skaggs Institute for Chemical Biology, The Scripps Research Institute 10550 North Torrey Pines Road, La Jolla, CA 92037 (USA) and

Department of Chemistry and Biochemistry, University of California San Diego 9500 Gilman Drive, La Jolla, CA 92093 (USA)

Department of Chemistry, BioScience Research Collaborative Rice University

6100 Main Street, Houston, TX 77005 (USA) E-mail: kcn@rice.edu

- [+] These authors contributed equally to this work.
- [**] Financial support was provided by the National Institutes of Health (USA) (grant AI055475-11) and the Skaggs Institute of Research. Fellowships to C.N. (Feodor Lynen Research Fellowship, Alexander von Humboldt Foundation), C.R.H.H. (NSF Graduate Research Fellowship), and A.E. (Fundación Alfonso Martín Escudero) are gratefully appreciated. We thank Dr. R. K. Chadha for X-ray crystallographic assistance and Dr. D. H. Huang and Dr. L. Pasternack for NMR spectroscopic assistance. We thank Prof. W.-G. Kim for graciously providing scanned NMR spectra of natural viridicatumtoxin B.



Supporting information for this article is available on the WWW under http://dx.doi.org/10.1002/anie.201304691.

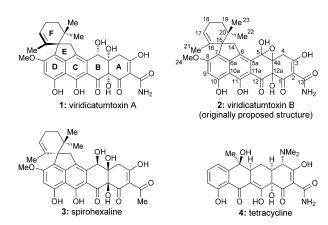


Figure 1. Molecular structures of viridicatumtoxin A (1), B (2), spirohexaline (3), and tetracycline (4).

0.5 μg mL⁻¹, respectively; compare tetracycline (4, Figure 1) and vancomycin: MIC = 8 and $0.25-1 \,\mu g \, mL^{-1}$, respectively].^[7] Their mode of action was implied to involve disruption of bacterial peptidoglycan biosynthesis through inhibition of UPP synthase. [9] Despite their impressive antibacterial activities, no synthetic approaches toward the viridicatumtoxins have been reported to date.

Inspired by the structural complexity, the incomplete and curious structural assignment containing an epoxy hemiacetal structural motif, and potent antibiotic properties of viridicatumtoxin B (2), we embarked on its total synthesis. Our objectives were to fully elucidate the structure of viridicatumtoxin B (2) and establish the foundation for the synthesis and biological evaluation of designed analogues within this family of antibiotics. A number of distinct structural features add significantly to the challenge of synthesizing viridicatumtoxin B (2) compared to the tetracyclines (e.g. 4, Figure 1). These features include the EF spirosystem of viridicatumtoxin B, its hindered quaternary carbon center at C15, its highly oxidized AB ring system, and the extensive substitution on the ABCD tetracyclic core of the molecule. On the basis of biosynthetic considerations^[10] and the known structure of viridicatumtoxin A (1),^[6] we tentatively assigned the relative configuration of viridicatumtoxin B (2) as shown in Figure 2. Our highly convergent strategy toward 2 was derived from the retrosynthetic analysis shown in Figure 2. Thus, disconnection of the indicated six carbon-carbon bonds through the reactions shown revealed four key building blocks: allylic bromide 5, cyclic anhydride 6, quinone monoketal 7 (known),[11] and isoxazole phenyl ester 8. Their construction and assimilation into the targeted molecule would involve three cyclizations (as indicated in Figure 2), the extrusion of

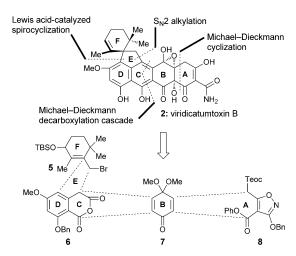


Figure 2. Retrosynthetic analysis of viridicatumtoxin B (2). Bn = benzyl, TBS = tert-butyldimethylsilyl, Teoc = 2-(trimethylsilyl)ethoxycarbonyl.

carbon dioxide (from 6), and the rupture of the isoxazole ring

The construction of building blocks 5, 6, and 8 is summarized in Scheme 1. The synthetic route to allylic bromide 5, the precursor for the EF spirocyclic ring system, commenced with a modification of a known, four-step sequence to convert geranic acid (9) into methyl ester allylic alcohol 10 (Scheme 1 a).[12] Silylation (TBSCl, imidazole) of 10 followed by ester reduction (DIBAL-H) produced allylic alcohol 11 in 91 % overall yield. Low-temperature mesylation of the latter (MsCl, Et₃N) followed by displacement of the resulting mesylate moiety with LiBr produced the desired fragment 5 in quantitative yield (Scheme 1a). Synthesis of the cyclic anhydride 6 commenced from intermediate 13 [available in two steps from 4-chlororesorcinol (12) by a literature procedure, Scheme 1b].[13] Selective mono-demethylation of 13 (directed by the nearby ester moiety on the aromatic ring) was achieved through the use of BBr₃. Reprotection of the liberated phenolic group to give intermediate 14 was accomplished using Ag₂O/BnBr (66% yield for 2 steps). Saponification (aq NaOH) and anhydride formation (Ac₂O) produced the desired cyclic anhydride 6 (90% yield over 2 steps). The synthesis of isoxazole fragment 8 commenced from known Stork-Hagedorn isoxazole 16[3i,14] (Scheme 1c), which was prepared by a modified literature process involving sequential acylation of dimethylmalonate (AcCl, Et₃N, MgCl₂, 96% yield), [15] methylation of the resulting enol (Me₂SO₄, K₂CO₃, 54% yield), cyclization (H₂NOH·HCl, NaOMe, 48% yield), and subsequent benzylation of the free hydroxy group (BnBr, Ag₂O, 67% yield). Saponification of the so-produced methyl ester then furnished carboxylic acid 17 (aq NaOH, 99% yield). Conventional esterification methods to prepare the phenyl ester 18, such as through the intermediacy of the corresponding chloride or the mixed anhydride, failed, as did the use of peptide coupling reagents. The desired phenyl ester formation, however, could be achieved under Mitsunobu conditions (PPh3, PhOH, DIAD, 78% yield).[16] The need to install an electron-withdrawing—and ideally easily removable—group at the isoxazole methyl group was predicated on

Scheme 1. a) Synthesis of allylic bromide 5; b) synthesis of cyclic anhydride 6; and c) synthesis of isoxazole 8. Reagents and conditions: a) a) H₃PO₄ (0.2 equiv), toluene, reflux, 90 min; b) MeI (3.9 equiv), K_2CO_3 (2.0 equiv), acetone, 25 °C, 15 h; c) mCPBA (1.2 equiv), CH_2Cl_2 , 0→25 °C, 3 h; d) NaOMe (1.5 equiv), MeOH, reflux, 17 h, 70% yield over 4 steps; e) TBSCl (1.6 equiv), imidazole (2.0 equiv), CH₂Cl₂, 25 °C, 12 h; f) DIBAL-H (2.7 equiv), CH_2Cl_2 , $-78\rightarrow0$ °C, 70 min, 91 % over 2 steps; g) Et₃N (2.0 equiv), MsCl (1.7 equiv), CH₂Cl₂, -50 °C, 1 h; then LiBr (3.5 equiv), THF, $-50 \rightarrow -20$ °C, 1 h, quant.; b) a) MeI (4.0 equiv), K₂CO₃ (8.0 equiv), acetone, reflux, 15 h, 91%; b) NaH (6.0 equiv), DEM (4.0 equiv), THF, 0°C, 2.5 h; then LDA (1.0 equiv), THF, 0°C, 3.5 h, 65%; c) BBr₃ (1.35 equiv), CH_2Cl_2 , $-78\rightarrow25$ °C, 30 min; d) BnBr (1.1 equiv), Ag₂O (1.9 equiv), DMF, 25 °C, 15 h, 66% over 2 steps; e) NaOH (27 equiv), $H_2O/EtOH$ 5:7, reflux, 15 h; f) Ac_2O (1.1 equiv), toluene, reflux, 1 h, 90% over 2 steps; c) a) MgCl₂ (1.0 equiv), Et₃N (2.0 equiv), AcCl (1.0 equiv), MeCN, $0\rightarrow25$ °C, 23 h, 96%; b) Me₂SO₄ (1.3 equiv), K₂CO₃ (1.3 equiv), DMF, $0\rightarrow 25$ °C, 17 h, 54%; c) $H_2NOH\cdot HCl$ (1.4 equiv), NaOMe (3.1 equiv), MeOH, $0\rightarrow$ 25 °C, 24 h, 48 %; d) BnBr (1.2 equiv), Ag₂O (1.5 equiv), DMF, 25 °C, 18 h, 67%; e) NaOH (1.9 equiv), H₂O/EtOH 3:10, 25°C, 3 h, 99%; f) PPh₃ (1.05 equiv), PhOH (1.05 equiv), DIAD (1.05 equiv), THF, reflux, 3 h, 78%; g) LiHMDS (2.2 equiv), THF, -78°C, 30 min; then TeocCl (2.2 equiv), -78 °C, 2 h, 86 %. mCPBA = meta-chloroperoxybenzoic acid, DIBAL-H = diisobutylaluminum hydride, Ms = methanesulfonyl, DEM = diethylmalonate, LDA = lithium diisopropylamide, DMF = dimethylformamide, DIAD = diisopropylazodicarboxylate, PhOH = phenol, THF = tetrahydrofuran, LiHMDS = lithium hexamethyldisilazide.

AB ring model systems; such a group was found to be required to facilitate a later conjugate addition. To this end, deprotonation of the methyl group of phenyl ester 18 (LiHMDS) and quenching of the resulting anionic species with TeocCl produced the desired fragment 8 (86% yield). Quinone monoketal 7 was readily available in one step by a literature procedure.^[11]

With all four building blocks (5-8) in hand, the stage was now set for their union, as shown in Scheme 2a. Annulation of cyclic anhydride 6 with quinone monoketal 7 to afford tricyclic compound 19 was achieved by using a one-step Michael-Dieckmann/decarboxylation protocol (DBU, 65°C,



Scheme 2. a) Synthesis of heptacycle 27 and b) ORTEP representation of spirocycle 23 (thermal ellipsoids at 30% probability; gray = C, red = O, green = H). Reagents and conditions: a) 7 (3.0 equiv), NaH (3.0 equiv), THF, 0°C, 45 min; then 25°C, 1 h; b) DBU (5.0 equiv), toluene, 65°C, 4.5 h, 54% over 2 steps; c) CSA (0.02 equiv), CH₂Cl₂, 25°C, 30 min, 99%; d) 5 (1.1 equiv), Na₂CO₃ (10 equiv), DMF, 25°C, 1 h, 77%, d.r. ca. 1:1; e) BF₃·OEt₂ (0.15 equiv), CH₂Cl₂, 0°C, 20 min, 73%; f) PIDA (1.2 equiv), MeOH/CH₂Cl₂ 1:1, 0→25°C, 1 h; g) CSA (0.07 equiv), CH₂Cl₂, 0°C, 5 min, 85% over 2 steps; h) PIDA (1.2 equiv), MeOH/CH₂Cl₂ 10:1, 25°C, 1.5 h, 90%; i) 8 (1.1 equiv), tBuOK (1.2 equiv), toluene, 25°C, 15 min, 91%, d.r. ca. 2:1; j) TBAF (10 equiv), NH₄F (20 equiv), degassed THF, 25°C, 5 min, 86%. DBU = 1,8-diazabicyclo[5.4.0]undec-7-ene, CSA = (±)-camphor-10-sulfonic acid, PIDA = iodobenzene diacetate, TBAF = tetra-*n*-butylammonium fluoride.

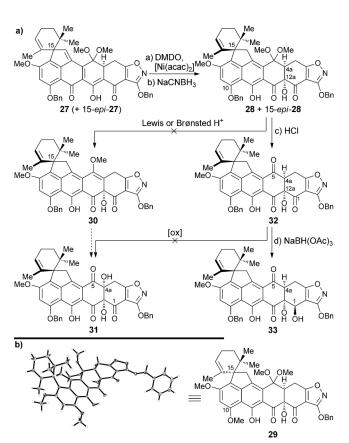
41 % yield). However, a two-step protocol involving exposure of **6** and **7** to NaH to induce anion generation and cycloaddition followed by work-up and subsequent treatment with DBU to promote decarboxylation was preferred, as it proved more efficient in producing **19** (54% yield overall). Reaction of ketal **19** with catalytic amounts of CSA caused extrusion of methanol, thereby furnishing the corresponding anthrone (99% yield), whose alkylation with allylic bromide **5**

proceeded under mild basic conditions (i.e. Na_2CO_3) to give intermediate **20** (77% yield, d.r. ca. 1:1). Much to our delight, ionization of the allylic TBS ether was readily achieved with catalytic quantities of $BF_3 \cdot OEt_2$, presumably forming highly stabilized but fleeting allylic cation **21**, whose intramolecular trapping by the nearby electron-rich arene led to spirocycle **22** as a single diastereomer (73% yield). The connectivity and relative stereochemical relationships within pentacycle **22** were assigned by comparing its NMR spectroscopic data with those of the related compound **23** (10-methoxy as opposed to 10-benzyloxy, Scheme 2b), whose structure was secured through X-ray crystallographic analysis $[m.p. = 114-116 \, ^{\circ}C \, (CHCl_3/EtOAc)$, see ORTEP representation, Scheme 2b]. [18]

To ready the newly acquired pentacycle 22 for cyclization with isoxazole 8, and thus forge ring A of the growing molecule, the former was treated with PhI(OAc)2 (PIDA, phenolic oxidation) to produce B-ring enone 24, which, however, did not undergo the desired Michael-Dieckmann reaction with isoxazole 8. We hypothesized that this reluctance was due to the adjacent electron-rich naphthalene ring, which decreases the electrophilicity of the enone moiety. Treatment of 24 with catalytic amounts of CSA (elimination of methanol, 85% yield over 2 steps) and subsequent exposure of the resulting C-ring quinomethide/B-ring pmethoxyphenol to PhI(OAc)₂ (PIDA) gave quinomethide 25 (90% yield), whose superior electrophilicity now allowed its facile fusion with its intended nucleophilic partner. Thus, the anticipated Michael-Dieckmann reaction between highly reactive enone 25 and isoxazole 8 was achieved using a slight excess of potassium tert-butoxide in toluene, which afforded heptacycle 26 and its isomer 15-epi-26 [91% yield, d.r. ca. 2:1 in favor of the natural C15 epimer (26), see below]. The relative stereochemical assignment of C4 and C4a in 26 was based on the observed ${}^{3}J_{4,4a} = 10.0 \text{ Hz}$ coupling constant, which is indicative of an H4,H4a anti relationship. From this point on, the two C15 diastereomers were carried forward as a mixture through several additional steps until separation became convenient. The use of a phenyl ester for the Michael-Dieckmann reaction was critical for success in the union of enone 25 and isoxazole 8, in agreement with previous observations by White et al.^[19] and Myers and co-workers.^[4d] Indeed, the methyl ester counterpart of isoxazole fragment 8 failed to enter this cyclization, leading instead to the initially formed conjugate addition adduct whose subsequently attempted ring closure under a variety of basic conditions failed.

The next task, that of removing the Teoc group with concomitant decarboxylation, proved problematic, initially leading to decomposition $[^{20a,b}]$ or low yields of the desired product (i.e. 27) $[^{20c]}$ under various conditions. After considerable experimentation it was found that using a freshly prepared solution of TBAF buffered with NH₄F^[21] in degassed THF achieved the desired goal of removing the Teoc group in 86% yield. We hypothesized that these buffered conditions served to keep the C1,C12 β -diketone moiety of 26 in its fully protonated state, thereby allowing for smooth anion formation during the decarboxylation step. This transformation marked the successful completion of the entire carbon framework of viridicatumtoxin B (2).

Soon after arriving at compound 27, we realized that its conversion into the more advanced intermediates of obligatory higher oxygenation (at C4a and C12a) would present challenges. Among the most serious issues were the multitude of functional groups present in the substrate, a problem exacerbated by the scarcity of protons around the AB ring system, which made spectroscopic analysis rather tedious, as well as the practical problems arising from the insoluble nature of the intermediates. After extensive experimentation, and as shown in Scheme 3 a, the hydroxylation of 27 at C12a



Scheme 3. a) Unsuccessful attempts to install the C4a hydroxy group and synthesis of C1 hydroxy compound **33** and **b**) ORTEP representation of heptacycle **29** (thermal ellipsoids at 30% probability; gray = C, red = O, blue = N, green = H). Reagents and conditions: a) [Ni(acac)₂] (0.2 equiv), DMDO (5.1 equiv), CH_2Cl_2 , $-78 \rightarrow -60^{\circ}C$, 6.5 h, 36%, 60% brsm, 50% after one recycle; b) NaCNBH₃ (10 equiv), THF, $-78 \rightarrow -60^{\circ}C$, 90 min, 39% for **28**, 19% for 15-*epi-***28**, chromatographically separated; c) 2 N aq HCl/THF 1:10, 25 °C, 5 h, quant.; d) NaBH-(OAc)₃ (1.2 equiv), EtOAc/acetone 1:1, 40°C, 105 min, 47%; acac = acetylacetonate, DMDO = dimethyldioxirane.

was finally achieved with dimethyldioxirane (DMDO) in the presence of catalytic amounts of [Ni(acac)₂]^[22] at -78 °C in dichloromethane,^[23] thereby leading to the desired hydroxylated product in 50% yield after one recycle of the 40% recovered starting material. Reduction of the quinomethide moiety of the latter with NaCNBH₃, as a soft hydride source, was then carried out to afford naphthalene derivative **28** (+15-*epi*-**28**) in 58% combined yield for the two diastereomers (**28**/15-*epi*-**28** ca. 2:1). At this stage, the two isomers

were separated chromatographically, with the natural diastereomer taken through the remaining steps. The assignment of the structural configurations of **28** and 15-*epi-28* were based on comparisons of the NMR spectroscopic data of the two isomers with those of the 10-methoxy counterpart **29** (Scheme 3b), whose structure had been unambiguously proven by X-ray crystallographic analysis [m.p. = 213–215 °C-(decomp) (EtOAc), see ORTEP representation, Scheme 3b]. [18]

From intermediate 28, a number of strategies and tactics were envisioned for the installment of the C4a hydroxy group (see Scheme 3a). Our initial efforts were directed at the generation of enol ether 30 and its hydroxy-directed α epoxidation, followed by hydrolysis of the resulting epoxide to obtain compound 31. However, all attempts to remove a molecule of methanol from ketal 28 under a variety of both Lewis and Brønsted acidic conditions did not lead to any detectable enol ether product (30).[24] Faced with this predicament, we proceeded to hydrolyze the dimethyl ketal moiety of 28 under acidic conditions (aq HCl, quant.) with the intention of converting the resulting triketone (32) into the desired product (31) through oxygenation of its enolate (at C4a). All attempts, however, to achieve this conversion were met with failure (see 32 to 31, Scheme 3a). The reluctance of these substrates to react as projected may be attributed to unwanted rearrangement pathways, including β-elimination/ aromatization or ring-expansion/lactone formation involving the C12a hydroxy group. [4a,25]

Speculating that a C4a,C5 diol could be introduced through dihydroxylation, we next attempted to reduce the C5 ketone within 32 (see Scheme 3a) to its corresponding alcohol by employing 1,3-directed reduction reagents^[26] such as NaBH(OAc)₃, expecting that we might eventually be able to dehydrate the latter to the desired C4a,C5 olefinic bond within ring B. Surprisingly, however, treatment of triketone 32 with a slight excess of NaBH(OAc)₃ in EtOAc/acetone (1:1) at 40 °C produced the 1,2-directed reduction product 33 (47 % yield) instead, as evident from diagnostic HMBC correlations. This result seemingly stands in contrast to the recently reported 1,3-reduction of a similar system by Tatsuta et al., [27] but is most likely a result of the electron-withdrawing effect exerted by the adjacent isoxazole system which renders the C1 ketone more electrophilic. Interestingly, the use of solvents other than EtOAc/acetone (1:1) resulted in intractable mixtures of reduction products. Recognizing the potential implications of this unexpected reduction, and building on the intelligence we had accumulated thus far on the reactivity of our system, we reasoned that a substrate such as 33 could be less prone to the suspected undesired reactions such as β elimination followed by a thermodynamically downhill aromatization of ring A.

Following this reasoning, and to exploit the fortuitous formation of **33**, the C1 hydroxy group of the latter was silylated to afford TBS ether **34** (TBSOTf, 2,6-lutidine, 61 % yield, Scheme 4). Much to our delight, the desired C4a hydroxylation was successfully performed by generating the trianion (**35**) of **34** with excess KHMDS at $-78\,^{\circ}$ C and quenching it with the Davis oxaziridine reagent, thereby leading to C4a-hydroxylated compound **36** (20 % yield



Scheme 4. C4a oxidation and completion of the synthesis. Reagents and conditions: a) TBSOTf (40 equiv), 2,6-lutidine (60 equiv), CH_2Cl_2 , $0\rightarrow25\,^{\circ}C$, 1 h, 61%; b) KHMDS (3.4 equiv), THF, $-78\,^{\circ}C$, 1 h; then Davis ox. (3.9 equiv), $-78\,^{\circ}C$, 1.7 h, 20% + 45% recovered **34**; c) HF-py (excess), MeCN, $0\rightarrow50\,^{\circ}C$, 25 h, 61%; d) DMP (3.0 equiv), DCE, $0\rightarrow50\,^{\circ}C$, 7.5 h, 66%; e) H_2 , Pd black (4.9 equiv), 1,4-dioxane/MeOH 1:1, 25 $\,^{\circ}C$, 8 min, 98%; TBSOTf = tert-butyldimethylsilyl trifluoromethanesulfonate, KHMDS = potassium hexamethyldisilazide, Davis ox. = (\pm) -trans-2-(phenylsulfonyl)-3-phenyloxaziridine, py = pyridine, DMP = Dess-Martin periodinane, DCE = 1,2-dichloroethane.

+45% recovered starting material, 36% brsm). Key to the success of this reaction were the free phenol at C11, an unprotected alcohol at C12a (which allows alkoxide formation, thereby preventing β-elimination), and protection of the secondary alcohol at C1. Structural deviations from this substrate resulted in no conversion, rearrangement pathways, or β-elimination of the C12a oxygen atom. The stereochemical outcome of this hydroxylation was revealed by NOESY correlations of product 36 (see the Supporting Information), and was in agreement with our expectations based on steric considerations (see structure 35, Scheme 4).

Removal of the TBS group from compound **36** with HF·py then led to the expected hydroxy compound **37** (61% yield). From NMR studies, we surmised that compound **37** exists in equilibrium with its cyclic hemiacetal isomer **37**′, with the two isomers interconverting slowly as evidenced from their unusually broad 1H NMR signals at ambient temperature. In support of this notion was the fact that the 1H NMR spectrum of this mixture acquired at $-40\,^{\circ}\text{C}$ displayed two sets of signals. Finally, oxidation of **37**/37′ with DMP^[29] furnished the corresponding C1 ketone (66% yield), which was subjected to hydrogenolysis^[4d] (two benzyl ethers and an N–O bond; H_2 , Pd black, 98% yield) to afford compound (\pm)-**38** as shown in Scheme 4. The physical properties of (\pm)-**38** matched those reported for viridicatumtoxin B^[7] except for

the presence of the C5 signal at $\delta = 194.1$ ppm in the ¹³C NMR spectrum of (\pm)-38 and the absence of the reported^[7] C5 signal at $\delta = 116.4$ ppm.^[30] No spectroscopic evidence for the existence of an epoxy hemiacetal structural motif was observed in the ¹³C NMR spectrum of (\pm)-38, thus compelling us to revise the structure of natural viridicatumtoxin B to that of 38. Pleasantly, synthetic viridicatumtoxin B (38) crystallized in crystals suitable for X-ray analysis from CH₂Cl₂/EtOH [m.p. = 245–247 °C(decomp), see ORTEP representa-

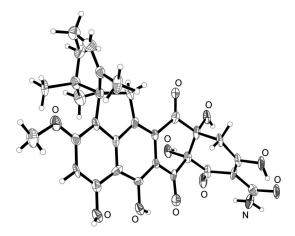


Figure 3. ORTEP representation of synthetic viridicatumtoxin B (38; thermal ellipsoids at 30% probability; gray = C, red = O, blue = N, green = H).

tion, Figure 3], which provided unambiguous proof of its structure. [18]

In conclusion, the total synthesis of racemic viridicatum-toxin B has been achieved through a highly convergent synthetic strategy from four easily accessible building blocks. This accomplishment led to the revision of the originally assigned epoxy hemiacetal structure of viridicatumtoxin B (2, Figure 1) to its hydroxy ketone form (38, Scheme 4). Efforts currently in progress aim to achieve an enantioselective total synthesis of viridicatumtoxin B (38) and confirm its absolute configuration. The developed chemistry sets the stage for further advances to occur in the field, including the design, synthesis, and biological evaluation of analogues of the viridicatumtoxins as potential leads for drug discovery to combat bacterial infections.

Received: May 30, 2013
Published online:

Keywords: antibiotics \cdot natural products \cdot structural revision \cdot tetracyclines \cdot total synthesis

a) F. von Nussbaum, M. Brands, B. Hinzen, S. Weigand, D. Häbich, Angew. Chem. 2006, 118, 5194-5254; Angew. Chem. Int. Ed. 2006, 45, 5072-5129; b) K. C. Nicolaou, J. S. Chen, D. J. Edmonds, A. A. Estrada, Angew. Chem. 2009, 121, 670-732; Angew. Chem. Int. Ed. 2009, 48, 660-719.

- [2] a) C. Walsh, Antibiotics, Actions, Origins, Resistance, 1st ed., ASM, Washington, DC, 2003; b) S. Rachakonda, L. Cartee, Curr. Med. Chem. 2004, 11, 775-793; c) S. B. Levy, B. Marshall, Nat. Med. 2004, 10, S122-S129; d) C. Nathan, Nature 2004, 431, 899-902.
- [3] a) L. H. Conover, K. Butler, J. D. Johnston, J. J. Korst, R. B. Woodward, J. Am. Chem. Soc. 1962, 84, 3222-3224; b) H. Muxfeldt, W. Rogalski, J. Am. Chem. Soc. 1965, 87, 933-934; c) A. I. Gurevich, M. G. Karapetyan, M. N. Kolosov, V. G. Korobko, V. V. Onoprienko, S. A. Popravko, M. M. Shemyakin, Tetrahedron Lett. 1967, 8, 131-134; d) J. J. Korst, J. D. Johnston, K. Butler, E. J. Bianco, L. H. Conover, R. B. Woodward, J. Am. Chem. Soc. 1968, 90, 439-457; e) H. Muxfeldt, G. Hardtmann, F. Kathawala, E. Vedejs, J. B. Mooberry, J. Am. Chem. Soc. 1968, 90, 6534-6536; f) D. H. R. Barton, P. D. Magnus, T. Hase, J. Chem. Soc. C 1971, 2215-2225; g) H. Muxfeldt, G. Haas, G. Hardtmann, F. Kathawala, J. B. Mooberry, E. Vedejs, J. Am. Chem. Soc. 1979, 101, 689-701; h) H. H. Wasserman, T.-J. Lu, A. I. Scott, J. Am. Chem. Soc. 1986, 108, 4237-4238; i) G. Stork, J. J. La Clair, P. Spargo, R. P. Nargund, N. Totah, J. Am. Chem. Soc. 1996, 118, 5304-5305; j) K. Tatsuta, T. Yoshimoto, H. Gunji, Y. Okado, M. Takahashi, Chem. Lett. 2000, 646-647; k) J. S. Wzorek, T. F. Knöpfel, I. Sapountzis, D. A. Evans, Org. Lett. 2012, 14, 5840-5843.
- [4] a) M. G. Charest, D. R. Siegel, A. G. Myers, J. Am. Chem. Soc. 2005, 127, 8292-8293; b) M. G. Charest, C. D. Lerner, J. D. Brubaker, D. R. Siegel, A. G. Myers, Science 2005, 308, 395-398; c) J. D. Brubaker, A. G. Myers, Org. Lett. 2007, 9, 3523-3525; d) C. Sun, Q. Wang, J. D. Brubaker, P. M. Wright, C. D. Lerner, K. Noson, M. Charest, D. R. Siegel, Y.-M. Wang, A. G. Myers, J. Am. Chem. Soc. 2008, 130, 17913-17927; e) P. M. Wright, A. G. Myers, Tetrahedron 2011, 67, 9853-9869; f) D. A. Kummer, D. Li, A. Dion, A. G. Myers, Chem. Sci. 2011, 2, 1710-1718
- [5] a) R. D. Hutchison, P. S. Steyn, S. J. Van Rensburg, *Toxicol. Appl. Pharmacol.* 1973, 24, 507-509. We propose that viridicatumtoxin be renamed "viridicatumtoxin A." It has been suggested that viridicatumtoxin A was isolated from *Penicillium aethiopicum* and not the originally reported *Penicillium viridicatum*; see b) J. C. Frisvad, O. Filtenborg, *Mycologia* 1989, 81, 837-861.
- [6] a) C. Kabuto, J. V. Silverton, T. Akiyama, U. Sankawa, R. D. Hutchison, P. S. Steyn, R. Vleggaar, J. Chem. Soc. Chem. Commun. 1976, 728–729; b) J. V. Silverton, C. Kabuto, T. Akiyama, Acta Crystallogr. Sect. B 1982, 38, 3032–3037.
- [7] C.-J. Zheng, H.-E. Yu, E.-H. Kim, W.-G. Kim, J. Antibiot. 2008, 61, 633–637.
- [8] J. Inokoshi, Y. Nakamura, Z. Hongbin, R. Uchida, K. Nonaka, R. Masuma, H. Tomoda, J. Antibiot. 2013, 66, 37–41.
- [9] N. Koyama, J. Inokoshi, H. Tomoda, *Molecules* 2013, 18, 204 224.
- [10] a) A. E. de Jesus, W. E. Hull, P. S. Steyn, F. R. van Heerden, R. Vleggaar, J. Chem. Soc. Chem. Commun. 1982, 902–904;
 b) R. M. Horak, V. J. Maharaj, S. F. Marais, F. R. van Heerden, R. Vleggaar, J. Chem. Soc. Chem. Commun. 1988, 1562–1564;
 c) H. Zhou, Y. Li, Y. Tang, Nat. Prod. Rep. 2010, 27, 839–868;
 d) Y.-H. Chooi, R. Cacho, Y. Tang, Chem. Biol. 2010, 17, 483–494;
 e) Y. Li, Y.-H. Chooi, Y. Sheng, J. S. Valentine, Y. Tang, J. Am. Chem. Soc. 2011, 133, 15773–15785;
 f) Y.-H. Chooi, P. Wang, J. Fang, Y. Li, K. Wu, P. Wang, Y. Tang, J. Am. Chem. Soc. 2012, 134, 9428–9437.
- [11] Prepared in one step from 4-methoxyphenol: A. Pelter, S. Elgendy, *Tetrahedron Lett.* 1988, 29, 677-680.

- [12] I. Kadás, G. Árvai, G. Horváth, Org. Prep. Proced. Int. 1998, 30, 79–85.
- [13] W. E. Bauta, D. P. Lovett, W. R. Cantrell, Jr., B. D. Burke, J. Org. Chem. 2003, 68, 5967 – 5973.
- [14] G. Stork, A. A. Hagedorn III, J. Am. Chem. Soc. 1978, 100, 3609–3611.
- [15] M. W. Rathke, P. J. Cowan, J. Org. Chem. 1985, 50, 2622-2624.
- [16] V. P. Fitzjarrald, R. Pongdee, Tetrahedron Lett. 2007, 48, 3553– 3557.
- [17] Mechanistic studies of the one-pot procedure involving the use of DBU on related systems suggested a stepwise pathway. The use of a strong base (i.e. NaH or LDA) with homophthalic anhydrides and quinones (Tamura Diels-Alder reaction) is typically depicted as a concerted [4+2] cycloaddition, but the synchronicity of the reaction has been questioned: a) C. D. Cox, T. Siu, S. J. Danishefsky, Angew. Chem. 2003, 115, 5783-5787; Angew. Chem. Int. Ed. 2003, 42, 5625-5629; b) M. González-López, J. T. Shaw, Chem. Rev. 2009, 109, 164-189.
- [18] CCDC 941202, 941203, and 945867 contain the supplementary crystallographic data for compounds 23, 29, and 38, respectively for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc. cam.ac.uk/data_request/cif. Compounds 23 and 29 were prepared by similar routes to those described herein and will be reported in the full account of this work.
- [19] a) J. D. White, E. G. Nolen, Jr., C. H. Miller, J. Org. Chem. 1986, 51, 1150–1152; b) J. D. White, F. W. J. Demnitz, Q. Xu, W. H. C. Martin, Org. Lett. 2008, 10, 2833–2836.
- [20] a) C. Gioeli, N. Balgobin, S. Josephson, J. B. Chattopadhyaya, Tetrahedron Lett. 1981, 22, 969–972; b) K. C. Nicolaou, K. R. Reddy, G. Skokotas, F. Sato, X. Y. Xiao, C. K. Hwang, J. Am. Chem. Soc. 1993, 115, 3558–3575; c) S. E. Denmark, T. Kobayashi, C. S. Regens, Tetrahedron 2010, 66, 4745–4759.
- [21] A. Fürstner, H. Weintritt, J. Am. Chem. Soc. 1998, 120, 2817– 2825
- [22] W. Adam, A. K. Smerz, Tetrahedron 1996, 52, 5799-5804.
- [23] M. Gibert, M. Ferrer, F. Sánchez-Baeza, A. Messeguer, *Tetrahedron* 1997, 53, 8643–8650.
- [24] Electron-rich α-methoxystyrenes are known to be very labile, even at near-neutral pH values; see, for example: J. P. Richard, K. B. Williams, J. Am. Chem. Soc. 2007, 129, 6952–6961.
- [25] A. I. Scott, E. Yamaguchi, S.-K. Chung, Tetrahedron Lett. 1975, 16, 1369–1372.
- [26] D. A. Evans, K. T. Chapman, E. M. Carreira, J. Am. Chem. Soc. 1988, 110, 3560 – 3578.
- [27] K. Tatsuta, T. Fukuda, T. Ishimori, R. Yachi, S. Yoshida, H. Hashimoto, S. Hosokawa, *Tetrahedron Lett.* 2012, 53, 422–425.
- [28] F. A. Davis, O. D. Stringer, J. Org. Chem. 1982, 47, 1774–1775.
- [29] D. B. Dess, J. C. Martin, J. Org. Chem. 1983, 48, 4155-4156.
- [30] We thank Professor W.-G. Kim for kindly providing us with the 1 H and 13 C NMR spectra of natural viridicatumtoxin B obtained at 300 MHz and 226 MHz, respectively, from a small sample of natural material. These spectroscopic data were in accord with those obtained by us at 600 MHz and 151 MHz for synthetic material, thus confirming the identity of the two samples (except for the racemic nature of ours). Furthermore, we noted certain discrepancies in the reported chemical shift values in Ref. [7] compared to those actually found in the provided spectra of the natural product. In particular, the originally unreported carbonyl resonance near $\delta = 194$ ppm which we assigned as C5 was indeed present in the scanned authentic spectrum of the natural product. For the spectra of the natural and synthetic materials and further details, see the Supporting Information.