

Available online at www.sciencedirect.com



Bioorganic & Medicinal Chemistry Letters

Bioorganic & Medicinal Chemistry Letters 17 (2007) 1189-1192

## 5-((1*H*-Pyrazol-4-yl)methylene)-2-thioxothiazolidin-4-one inhibitors of ADAMTS-5

Adam M. Gilbert,<sup>a,\*</sup> Matthew G. Bursavich,<sup>a</sup> Sabrina Lombardi,<sup>a</sup> Katy E. Georgiadis,<sup>b</sup> Erica Reifenberg,<sup>b</sup> Carl R. Flannery<sup>b</sup> and Elisabeth A. Morris<sup>b</sup>

<sup>a</sup>Exploratory Medicinal Chemistry, Chemical and Screening Sciences, Wyeth Research, 401 North Middletown Road, Pearl River, NY 10965, USA

<sup>b</sup>Women's Health and Musculoskeletal Biology, Wyeth Research, 200 Cambridge Park Drive, Cambridge, MA 02140, USA

Received 25 October 2006; revised 7 December 2006; accepted 8 December 2006 Available online 12 December 2006

**Abstract**—A series of 5-((1*H*-pyrazol-4-yl)methylene)-2-thioxothiazolidin-4-one inhibitors of ADAMTS-5 (aggrecanase-2) is described. These compounds show  $\mu$ M functional inhibition of ADAMTS-5, and represent a new class of agents with the potential of inhibiting degradation of aggrecan, a major component of cartilage which is lost in osteoporosis. Compound **12** is noteworthy in that it has an ADAMTS-5 IC<sub>50</sub>: 1.1  $\mu$ M and shows >40-fold functional selectivity over ADAMTS-4 (aggrecanase-1). © 2006 Elsevier Ltd. All rights reserved.

Osteoarthritis (OA), a disease where aggrecan and collagen in the articular cartilage matrix are slowly degraded, often leads to debilitating chronic joint pain and inflammation. While collagen is the major structural component of aggrecan and provides strength to the tissue, aggrecan is responsible for the tissue's flexible and elastic nature. Aggrecan is lost in the initial phases of the disease, while collagen is lost in the disease's later stages. Thus an agent that would inhibit aggrecan breakdown could be a disease-modifying osteoarthritis treatment as opposed to treatments that only provide symptomatic relief (e.g., acetaminophen, COX-2 inhibitors, NSAIDs or intraarticular steroids).<sup>1</sup>

Aggrecan is catabolized by the proteolytic enzymes ADAMTS-4 (aggrecanase-1) and ADAMTS-5 (aggrecanase-2), two members of the "A Disintegrin And Metalloproteinase with ThromboSpondinmotifs" (ADAMTS) family of proteins. These enzymes cleave the aggrecan interglobular domain at the Glu373–Ala374 peptide bond.<sup>2</sup> Both enzymes are 1000-fold more efficient at cleaving aggrecan than any other enzymes although it is not known which enzyme is the primary source of aggrecanase activity in cartilage. Studies sug-

gest that ADAMTS-5 could be the major player as ADAMTS-5 knockout mice show reduced OA severity after surgical induction of joint instability, whereas the corresponding ADAMTS-4 knockouts show a normal OA progression.<sup>3</sup> ADAMTS-5 is also known to be the major ADAMTS in mouse cartilage and in a mouse model of inflammatory arthritis.<sup>4</sup> In this letter, we report our initial studies on a series of 5-((1*H*-pyrazol-4yl)methylene)-2-thioxothiazolidin-4-one ADAMTS-5 inhibitors 1 found from high throughput screening and show that they are useful as potentially selective ADAMTS-5 therapeutics to treat OA.<sup>5</sup> To our knowledge, these compounds represent the first disclosure of inhibitors of ADAMTS-5. Examples of general aggrecanase inhibitors have been previously disclosed.<sup>6–9</sup>



Thioxothiazolidin-4-one ADAMTS-5 inhibitors **1** are prepared according to Scheme 1.<sup>10</sup> 5-Hydroxypyrazoles **3** are synthesized by combining acetylacetic esters **2** with hydrazines.<sup>11</sup>

Keywords: ADAMTS-5; Aggrecanase-2; Thioxothiazolidin-4-ones; Osteoporosis.

<sup>\*</sup> Corresponding author. Tel.: +1 845 602 4865; fax: +1 845 602 5561; e-mail: gilbera@wyeth.com

<sup>0960-894</sup>X/\$ - see front matter @ 2006 Elsevier Ltd. All rights reserved. doi:10.1016/j.bmcl.2006.12.020



Scheme 1. Reagents and conditions: (a)  $R^1$ NHNH<sub>2</sub>, H<sub>2</sub>SO<sub>4</sub>, EtOH, reflux, 17 h; (b) POCl<sub>3</sub>, DMF, 80 °C, 12 h; (c)  $R^2$ SH, Et<sub>3</sub>N, DMSO, 40 °C, 12 h; (d)  $R^2$ NH, Et<sub>3</sub>N, DMF, 100 °C, 12 h; (e) *t*-BuOK, DMF, 23 °C, 12 h; (f) rhodanine, β-alanine, AcOH, 110 °C, 12 h.

The corresponding 4-formyl-5-chloropyrazoles **3** are prepared using POCl<sub>3</sub> in hot DMF.<sup>11</sup> The 5-chloro moiety can be displaced with the following nucleophiles: thiols to produce **5** (Et<sub>3</sub>N, DMSO, 40 °C), amines to produce **6** (Et<sub>3</sub>N, DMF, 100 °C), and alcohols to produce **7** (*t*-BuOK, DMF, 23 °C). The corresponding thioxothiazolidin-4-ones **1** are produced using rhodanine and  $\beta$ -alanine in hot HOAc.<sup>12</sup> <sup>1</sup>H NMR spectroscopy shows that thioxothiazolidin-4-ones **1** exist in a range

from nearly pure Z isomers (as drawn in Scheme 1) to 80:20 Z:E isomeric mixtures.

ADAMTS-5 activity was determined using a fluorescence resonance energy transfer (FRET) assay using a QF-peptide containing the aggrecanase cleavage site. Binding to target studies were conducted using fluorescence polarization experiments using a xanthene-ADAMTS-5 hydroxamic acid inhibitor. Several compounds were analyzed for ADAMTS-4 activity using the same QF-peptide used for the ADAMTS-5 activity studies.

Biological data of 3-CF<sub>3</sub>-1*H*-pyrazole thioxothiazolidin-4-ones are shown in Table 1. When  $R^1 = Me$  and  $R^5 = SPh (9)$  or SCH<sub>2</sub>Ph (10), weak ADAMTS-5 activity is seen. Replacing the Ph in 10 with the isosteric 2-thiophene in 11 produces similarly weak ADAMTS-5 potency. However the addition of a p-Cl moiety to 11 produces 12, a compound which displays micromolar ADAMTS-5 inhibitory activity (IC<sub>50</sub>: 1.1 µM). Movement of the *p*-Cl moiety in **12** to the *o*-position affords a compound with  $\sim 20$ -fold less activity (13: IC<sub>50</sub>, 23.2  $\mu$ M). The *p*-Cl moiety in **12** may be replaced by a *p-t*-Bu in 14 ( $\hat{IC}_{50}$ , 2.2  $\mu$ M), but not a *p*-OMe as in 15. Removing the benzyl  $CH_2$  methylene in 12 gives the *p*-Cl phenyl analog **16** which has reduced ADAMTS-5 inhibition. In addition to  $R^1 = Me$ ,  $R^1$  can be Ph (17:  $IC_{50}$ , 2.5  $\mu$ M) and cyclohexyl (18:  $IC_{50}$ , 4.2  $\mu$ M). The SCH<sub>2</sub> moiety in 12 and 17 can be replaced by a piperazine to give compounds with similar ADAMTS-5 activity (19: IC<sub>50</sub>, 9.0 µM and 20: IC<sub>50</sub>, 2.1 µM).

The 3-CF<sub>3</sub> moiety in **9–20** can be modified to produce active compounds if other pyrazole substituents are modified as well (Table 2). The 1,3-diphenyl-1*H*-pyrazole analog **21** shows ADAMTS-5 inhibition. This contrasts with the  $R^1 = Me$ ,  $R^3 = 3$ -CF<sub>3</sub> analog **11**. The 2-thiophene moiety in **21** may be changed to a furan

Table 1. In vitro ADAMTS-5 inhibition data for 5-((3-(trifluoromethyl)-1H-pyrazol-4-yl)methylene)-2-thioxothiazolidin-4-ones



Compound	$\mathbb{R}^1$	R <sup>5</sup>	$\%$ Inhib. @ $20^{\rm a}~(\mu M)$	$I{C_{50}}^a \ (\mu M)$
9	Me	SPh	28	
10	Me	SCH <sub>2</sub> Ph	44	
11	Me	SCH <sub>2</sub> (2-thiophene)	40	
12	Me	SCH <sub>2</sub> Ph(4-Cl)		1.1
13	Me	SCH <sub>2</sub> Ph(2-Cl)		23.2
14	Me	$SCH_2Ph(4-t-Bu)$		2.2
15	Me	SCH <sub>2</sub> Ph(4-OMe)	53	
16	Me	SPh(4-Cl)	47	
17	Ph	SCH <sub>2</sub> Ph(4-Cl)		2.5
18	СуН	SCH <sub>2</sub> Ph(4-Cl)		4.2
19	Me	PiperazinePh(4-Cl)		9.0
20	Ph	PiperazinePh(4-Cl)		2.1

<sup>a</sup> Values are means of two experiments, standard deviations are  $\pm 10\%$ .

Table 2. In vitro aggrecanase-2 inhibition data for 5-((1H-pyrazol-4-yl)methylene)-2-thioxothiazolidin-4-ones



Compound	$\mathbb{R}^1$	$\mathbb{R}^3$	R <sup>5</sup>	% Inhib. @ 20 ^ ( $\mu M)$	$IC_{50}{}^a$ ( $\mu M$ )
21	Ph	Ph	SCH <sub>2</sub> (2-thiophene)		3.7
22	Ph	Ph	SCH <sub>2</sub> (2-furan)		6.7
23	СуН	Ph	SCH <sub>2</sub> (2-furan)		4.6
24	Ph	$CF_3$	SCH <sub>2</sub> (2-furan)	42	
25	Ph	$CF_3$	SCH <sub>2</sub> Ph		16.9
26	Me	Ph	SCH <sub>2</sub> (2-furan)		14.1
27	Me	Ph	SPh(4-Me)		8.3
28	СуН	Ph	Piperazine(2-pyrimidine)		8.0
29	СуН	$CF_3$	Piperazine(2-pyrimidine)		11.7
30	Ph	Ph	Piperazine(Me)		4.6
31	Ph	$CF_3$	Piperazine(Me)	45	

<sup>a</sup> Values are means of two experiments, standard deviations are  $\pm 10\%$ .

22 with little change in activity, while the corresponding compound where  $R^1 = CyH 23$  has similar ADAMTS-5 potency. Interestingly, 24 where  $R^3 = CF_3$  does not show good ADAMTS-5 activity (compare with 22), but 25 shows moderate activity where  $R^5 = SCH_2Ph$ (compare with 10). The  $R^1 = Me$  and  $R^3 = Ph$  substitution pattern gives moderately active compounds (26 and 27) and piperazinyl moieties can be incorporated at  $R^5$ (28–31) as previously shown above (Table 1).

The sulfur at the  $R^5$  position of the pyrazole can be substituted with an oxygen substituent provided that an extra CH<sub>2</sub> group is added to the linker (Table 3). Thus, **32** possesses ADAMTS-5 activity (IC<sub>50</sub>, 4.2  $\mu$ M; compare with **17**). The corresponding *p*-F phenyl compound **33** also shows ADAMTS-5 inhibition (IC<sub>50</sub>, 7.1  $\mu$ M) as does the 2-thiophene analog **34** (IC<sub>50</sub>, 9.1  $\mu$ M). Similarly to **15**, the *p*-OMe-phenyl analog **35** shows reduced ADAMTS-5 activity again showing the importance of the *p*-substituent on phenyl group of the R<sup>5</sup> side chain (Table 4).

 Table 3. In vitro ADAMTS-5 inhibition data for 5-((5-alkoxy-1H-pyrazol-4-yl)methylene)-2-thioxothiazolidin-4-ones



Compound	<b>R</b> <sup>5</sup>	% Inhib. @ 20 <sup>a</sup> (µM)	$I{C_{50}}^a \ (\mu M)$
32	O(CH <sub>2</sub> ) <sub>2</sub> Ph(4-Cl)		4.2
33	$O(CH_2)_2Ph(4-F)$		7.1
34	O(CH <sub>2</sub> ) <sub>2</sub> (2-thiophene)		9.1
35	$O(CH_2)_2Ph(4-OMe)$	31	

<sup>a</sup> Values are means of two experiments, standard deviations are  $\pm 10\%$ .



Reduction of the double bond attached to the thioxothiazolidin-4-one headpiece using the Hantzsch ester<sup>13</sup> produces compounds with reduced ADAMTS-5 activity. Thus while **17** only loses 2-fold ADAMTS-5 potency on double bond reduction (**36**: IC<sub>50</sub>, 5.8  $\mu$ M), the case is worse for **24** and **31**; both compounds lose substantial potency when converted to their single bond analogs **37** and **38**.

Given the previously cited suggestive mouse ADAMTS-5 knockout data,<sup>3</sup> we have begun to look at ADAMTS-5/ ADAMTS-4 selectivity for a few of the thioxothiazolidin-4-ones presented in this manuscript using an ADAMTS-4 FRET assay. Two of the more potent ADAMTS-5 inhibitors show selectivity over ADAM-TS-4. The *p*-Cl-Ph analog **12** shows good functional selectivity over ADAMTS-4 (ADAMTS-5 IC<sub>50</sub>, 1.1  $\mu$ M; ADAMTS-4 IC<sub>50</sub>, 44  $\mu$ M), something that is not surprising since ADAMTS-5 has approximately 40% sequence homology to ADAMTS-4 overall and

 Table 4. In vitro ADAMTS-5 and ADAMTS-4 inhibition data for selected 5-((1*H*-pyrazol-4-yl)methylene)-2-thioxothiazolidin-4-ones

Compound	ADAMTS-5 IC <sub>50</sub> <sup>a</sup> (µM)	ADAMTS-4 IC <sub>50</sub> <sup>a</sup> (µM)
12	1.1	44
14	2.2	7.5

<sup>a</sup> Values are means of two experiments, standard deviations are  $\pm 10\%$ .

50% active site sequence similarity.<sup>14</sup> The closely related *t*-Bu-Ph analog **14** is less functionally selective (ADAMTS-5 IC<sub>50</sub>, 2.2  $\mu$ M; ADAMTS-4 IC<sub>50</sub>, 7.5  $\mu$ M). In addition, both **12** and **14** successfully displaced a xanthene-ADAMTS-5 hydroxamic acid inhibitor from the ADAMTS-5 protein showing that their inhibitory activity involved binding to ADAMTS-5.<sup>15</sup>

Thus, we have presented a series of 5-((1*H*-pyrazol-4yl)methylene)-2-thioxothiazolidin-4-one ADAMTS-5 inhibitors. This series of compounds has tractable SAR and several analogs show modest functional selectivity for ADAMTS-5 over ADAMTS-4. The continued development of selective ADAMTS-5 inhibitors is currently ongoing and will be reported in due course.

## Acknowledgments

We thank Dr. John Ellingboe for support of this work. We also thank Dr. Kristina Cunningham for performing fluorescence polarization binding to target studies.

## **References and notes**

- Wieland, H. A.; Michaelis, M.; Kirschbaum, B. J.; Rudolphi, K. A. Nat. Rev. Drug Disc. 2005, 4, 331.
- Liu, R. Q.; Trzaskos, J. M. Curr. Med. Chem. Anti-Inflam. Anti-Allergy Agents 2005, 4, 251.
- Glasson, S. S.; Askew, R.; Sheppard, B.; Carito, B. A.; Blanchet, T.; Ma, H. L.; Flannery, C. R.; Kanki, K.; Wang, E.; Peluso, D.; Yang, Z.; Majumdar, M. K.; Morris, E. A. Arthritis Rheum. 2004, 50, 2547.
- Stanton, H.; Rogerson, F. M.; East, C. J.; Golub, S. B.; Lawlor, K. E.; Meeker, C. T.; Little, C. B.; Last, K.; Farmer, P. J.; Campbell, I. K.; Fourie, A. M.; Fosang, A. J. *Nature* 2005, 434, 648.

- 5. For an accompanying letter describing a series of related aryl thioxothiazolidin-4-one ADAMTS-5 inhibitors, see: Bursavich, M. G.; Gilbert, A.M.; Lombardi, S.; Georgiadis, K.E.; Reifenberg, E.; Flannery, C.R.; Morris, E.A. *Bioorg. Med. Chem. Lett.* **2007**, *17*, in press.
- Xiang, J. S.; Hu, Y.; Rush, T. S.; Thomason, J. R.; Ipek, M.; Sum, P.-E.; Abrous, L.; Sabatini, J. J.; Georgiadis, K.; Reifenberg, E.; Majumdar, M.; Morris, E. A.; Tam, S. *Bioorg. Med. Chem. Lett.* 2006, 16, 311.
- Cherney, R. J.; Mo, R.; Meyer, D. T.; Wang, L.; Yao, W.; Wasserman, Z. R.; Liu, R.-Q.; Covington, M. B.; Tortorella, M. D.; Arner, E. C.; Qian, M.; Christ, D. D.; Trzaskos, J. M.; Newton, R. C.; Magolda, R. L.; Decicco, C. P. *Bioorg. Med. Chem. Lett.* **2003**, *13*, 1297.
- Yao, W.; Chao, M.; Wasserman, Z. R.; Liu, R.-Q.; Covington, M. B.; Newton, R.; Christ, D.; Wexler, R. R.; Decicco, C. P. *Bioorg. Med. Chem. Lett.* 2001, *12*, 101.
- Yao, W.; Wasserman, Z. R.; Chao, M.; Reddy, G.; Shi, E.; Liu, R.-Q.; Covington, M. B.; Arner, E. C.; Pratta, M. A.; Tortorella, M.; Magolda, R. L.; Newton, R.; Qian, M.; Ribadeneira, M. D.; Christ, D.; Wexler, R. R.; Decicco, C. P. J. Med. Chem. 2001, 44, 3347.
- 10. All newly prepared compounds were characterized by reversed phases-HPLC/MS spectroscopy. A selected number of compounds were also characterized by <sup>1</sup>H NMR.
- 11. Lee, L. F.; Schleppnik, F. M.; Schneider, R. W.; Campbell, D. H. J. Heterocycl. Chem. **1990**, 27, 243.
- Unangst, P. C.; Connor, D. T.; Cetenko, W. A.; Sorenson, R. J.; Kostlan, C. R.; Sircar, J. C.; Wright, C. D.; Schrier, D. J.; Dyer, R. D. J. Med. Chem. 1994, 37, 322.
- Hansen, M. M.; Harkness, A. R.; Khau, V. V.; Martinelli, M. J.; Deeter, J. B. *Tetrahedron: Asymmetry* 1996, 7, 2515.
- Abbaszade, I.; Liu, R.-Q.; Yang, F.; Rosenfeld, S. A.; Ross, O. H.; Link, J. R.; Ellis, D. M.; Tortorella, M. D.; Pratta, M. A.; Hollis, J. M.; Wynn, R.; Duke, J. L.; George, H. J.; Hillman, M. C., Jr.; Murphy, K.; Wiswall, B. H.; Copeland, R. A.; Decicco, C. P.; Bruckner, R.; Nagase, H.; Itoh, Y.; Newton, R. C.; Magolda, R. L.; Trzaskos, J. M.; Hollis, G. F.; Arner, E. C.; Burn, T. C. J. Biol. Chem. 1999, 274, 23443.
- 15. Studies to be published in the near future.