

Tetrahedron 54 (1998) 6259-6292

TETRAHEDRON

Synthesis and Oxidative Fragmentation of Catharanthine Analogs. Comparison to the Fragmentation - Coupling of Catharanthine and Vindoline

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Received 25 September 1997; revised 5 February 1998; accepted 27 March 1998

Abstract: Two new analogs of catharanthine have been synthesized in racemic form. They differ from catharanthine in the fusion of the indole ring to the non-aromatic portion of the *iboga* skeleton, with the [2,3] fusion present in catharanthine being replaced by [2,1] and [3,2] fusions. The corresponding deethyl analogs were also prepared and methodological improvements were applied to an existing synthesis of deethylcatharanthine and a formal synthesis of racemic catharanthine. The reactivity of the catharanthine analogs toward coupling with vindoline was examined. Coupling was attempted by both the amine oxide fragmentation (Potier) and Fe³⁺ methods. Under Potier conditions the [2,1] fused analogs give low yields of coupling products in which vindoline is attached to the 3-position of the indole ring. The [3,2] isomers undergo fragmentation of the C16-C21 bond, as observed for catharanthine, but no coupling to vindoline occurs. The reactivity, oxidation potentials and conformation of the analogs are compared with catharanthine, deethylcatharanthine and N-methylcatharanthine. © 1998 Elsevier Science Ltd. All rights reserved.

The dimeric vinca alkaloids vincristine (VCR) and vinblastine (VLB), originally reported in 1960, are of major significance in cancer chemotherapy.¹ Early successes in treatment of leukemia and Hodgkin's disease have been followed by development of many combination therapies which include VCR or VLB as one of the agents.² More recently the semi-synthetic analog vinorelbine has been introduced into clinical use.³



 R^1 = Me, R^{20} = Et, X = (=C15), n = 2 Anhydrovinblastine (AVLB) R^1 = Me, R^{20} = Et, X = OH, n = 2 Vinblastine (VLB) R^1 = CHO, R^{20} = Et, X = OH, n = 2 Vincristine (VCR) R^1 = Me, R^{20} = Et, X = (=C15), n = 1 Vinorelbine

0040-4020/98/\$19.00 © 1998 Elsevier Science Ltd. All rights reserved. *PII:* S0040-4020(98)00289-0 The importance of the dimeric *vinca* alkaloids has resulted in numerous studies of the synthesis and biological evaluation of analogs. The methods explored for synthesis of analogs include modification of the naturally occurring dimer,⁴ construction of dimers by coupling modified versions of the monomeric precursors catharanthine or vindoline,⁵ and construction of the catharanthine-derived portion by total synthesis.⁶

I. Synthesis of Catharanthine Analogs

In previous work we reported the synthesis of 6-nor-^{7a} and 5,6-homo-20-deethylcatharanthine,^{7b} as well as related 15-oxo derivatives.^{7c,7d} In this paper we report the synthesis of four new catharanthine analogs having alternative fusions of the indole ring to the isoquinuclidine portion of the *iboga* skeleton in which the natural indole [2,3] fusion is replaced by [2,1] (8b) and [3,2] fusions (18b). The racemic C20-deethyl analogs were also synthesized (8a, 18a). Improvements in our earlier synthesis of (\pm)-20-deethylcatharanthine⁸ (10a) are also reported. Those improvements were applied to a formal synthesis of (\pm)-catharanthine (10b).



Compound 6a, a key intermediate in the synthesis of 20-deethylcatharanthine (10a), is also a potential precursor of the [2,1]-isomer 8a. In the synthesis of 10a, photocyclization of 6a results in closure of the C6-C7 bond to give 9a.⁸ It seemed likely that a base-catalyzed reaction would lead to C6-N1 cyclization by the indole anion and formation of the isomer 7a. The starting material 3a is available from the Diels-Alder reaction of methyl 2-(1-phenylsulfonylindol-2-yl)propenoate (1) and 1-benzyloxycarbonyl-1,2-dihydropyridine (2a). An improved N4-deprotection of 3a was developed using trimethylsilyl iodide for cleavage. Chloroacetylation to give 5a was then carried out using chloroacetic anhydride.

A selective removal of the phenylsulfonyl protecting group without affecting the chloroacetyl group was achieved by use of the photochemical desulfonylation conditions developed by Yonemitsu and coworkers.⁹ Photolysis through Vycor with 1,5-dimethoxynaphthalene and ascorbic acid as reductant resulted in a 66% yield of **6a** without competing photolysis of the chloroacetyl group. Intermediate **6a** was then cyclized to **7a** in 83% yield by treatment with NaH in THF. If, instead, **6a** was subjected to photolysis through Vycor in methanol, 5-

oxo-20-deethylcatharathine 9a, a precursor of 10a, was obtained in 45% yield.

The same approach was applied to the Diels-Alder adduct 3b obtained from 1 and 2b. Racemic 5-oxocatharanthine (9b) was obtained. The material had spectroscopic properties identical to those reported by Szantay who obtained 9b by an alternate route.¹⁰ This constitutes a formal synthesis of (\pm)-catharanthine (10b) since Szantay has previously converted 9b to (\pm)-10b.¹⁰ Treatment of 6b with NaH in THF led to the N1-C6 cyclization product 7b in 81% yield.

Scheme 1.



Conditions. a) 100°C, 60h. b) TMSI, CH₂Cl₂. c) chloroacetic anhydride, DMAP, CH₂Cl₂. d) hv, pyrex filter, 1,5-dimethoxynaphthalene, ascorbic acid. e) hv, vycor filter. f) NaH, THF, rt. g) NaBH₄, BF₃-OEt₂, THF.

Our previous method for reduction of 5-oxocatharanthine analogs involved reduction of the corresponding S-methyl thioimmonium salts.⁸ However, Szantay and coworkers used *in situ* generation of diborane for this reduction, despite the presence of the C15-C20 double bond.¹¹ Application of their method to **9a** gave excellent conversion to **10a**. These improvements reduce the number of steps from **3a** to 20-deethylcatharanthine from 7 to 4 and improve the yield to 24% compared to 18% for the published synthesis.

Use of the *in situ* diborane conditions for reduction of the [2,1]-analog 7a was much more problematic. The reaction was highly sensitive to reaction temperature and the solvent used for extractive work up. The best yield achieved was 40%. Among the key variables were reaction temperature (0°C) and extraction solvent (CH₂Cl₂, EtOAc >> CHCl₃). Although the competing processes have not been identified, boronation at the open indole ring position may be the origin of the problem.¹² The reduction completed the synthesis of 8a in overall 8% yield from 3a. A parallel series of reactions starting with 3b led to 8b in overall 14% yield from 3b.

An attempt to develop a parallel synthesis of the [3,2]-isomer 18a using methyl 2-(1-phenylsulfonylindol-3yl) propenoate failed because this substance was very prone to dimerization.¹³ No Diels-Alder adduct could be isolated after attempted reaction with the dihydropyridine 2a. However an adaptation of a route to catharanthine used by Szantay¹⁰ and Raucher¹⁴ provided access to 18a and 18b. Methyl 2-chloropropenoate is known to react with 2b to give a mixture of *exo-* and *endo* Diels-Alder adducts 11a and 12a.^{10,11,14} These adducts were separated and deprotected with TMSI. Coupling of 11a with indole-2-acetic acid using DCCI then gave 16a. As compared with Szantay's and Raucher's photocyclizations which occur at the 2-position of the indole ring, the photocyclization of 16a involves the 3-position of the indole ring. The direct photolysis of 16a proceeds in 20-25% yield which is comparable to Szantay's results (16% yield) for the indole-3-acetic acid amide.¹¹ A similar reaction sequence converted 11b to 16b. It was photocyclized to 17b in 23% yield. The bromo analog 16c was also prepared but it afforded no improvement in yield. The photocyclization products 17a and 17b were reduced using the *in situ* borane procedure in good yield to 18a and 18b, respectively.

Scheme 2.



Conditions. a) toluene, 80°C. b) TMSI, rt. c) 2-indolylacetic acid, DCC, CH_2CI_2 , rt, overnight. d) hv, Vycor filter, sodium carbonate, methanol-water, 5h. e) NaBH₄, BF₃*OEt₂, THF.

II. Reactivity Under Coupling Conditions

Several methods are now available for coupling of catharanthine with vindoline¹⁵⁻¹⁹ The most prominent are the fragmentation of catharanthine-N-oxide by trifluoroacetic anhydride $(TFAA)^{15}$ (Potier fragmentation) and Fe³⁺ oxidative fragmentation.¹⁶ The amine oxide fragmentation has proven to be quite limited in its structural scope, however. The following structural modifications all result in failure of coupling under these conditions: (1) saturation of the C15-C20 double bond;¹⁶ (2) deletion of C6 to give *nor*-catharanthine;²⁰ (3) addition of an extra carbon in the tryptamine bridge to give C5-C6-*homo*-catharanthine.²¹ No catharanthine analogs have been used in the Fe³⁺ coupling procedure. We have now studied the reactivity of the catharanthine analogs **8a**, **8b**, **18a** and **18b** with vindoline under both the Potier and Fe³⁺ coupling procedures.

A. [2,1]-Analogs 8a and 8b

Several groups have reported Fe^{3+} -mediated coupling of catharanthine and vindoline.¹⁶ We used the conditions of Szantay et al. which use 6-20 equivalents of FeCl₃ in aqueous 0.05 M glycine solution.^{16d} The conditions used for the amine oxide fragmentation were those developed by Potier *et. al.* and involve low temperature treatment of the amine oxide with TFAA.^{15a} In both procedures, the coupled product anhydrov-inblastine (AVLB) is isolated after NaBH₄ reduction of a dihydropyridinium intermediate.

When [2,1]-analogs 8a and 8b were subjected to Fe^{3+} conditions, both in the presence and absence of vindoline, the reactants were recovered unchanged. The Fe^{3+} -glycine system evidently is not a sufficiently strong oxidant to oxidize the [2,1]-isomers, which have a somewhat higher oxidation potential than either the [2,3] and [3,2] analogs, vide infra.

Conversion of 8a to its N-oxide (19a) and reaction with vindoline under the Potier conditions resulted in a product mixture containing mainly unreacted vindoline and a small amount (10-15% each) of two apparent coupling products, as indicated by appearance of two new sets of singlets at 6.2 and 6.6 ppm and at 6.1 and 6.9 ppm, suggestive of C10-substitution on vindoline. Essentially all of the 19a reacted. However, no peaks attributable to products derived from 19a, except those assigned to the coupled products, were evident in the crude product. Purification of the reaction mixture by flash chromatography followed by radial chromatography lead to the isolation (2-3% yield each) of two substances, each with apparent molecular ions at 763 amu which showed two singlets in the NMR at 6.2 and 6.6 and 6.1 and 6.9, respectively, confirming substitution at C10 of

vindoline. The observed M^{+} is two amu less than anticipated for a fragmentation-coupling-reduction analogous to the formation of AVLB from catharanthine and vindoline. Precedented coupling products of MW 763 might include elimination followed by addition of vindoline at C3 or C5²⁰ or C5-C6 fragmentation followed by addition of vindoline at C5 and N1-C6 cyclization.^{15a,22} The absence of an ¹H-NMR signal attributable to a indole-3H excluded such structures. The presence of peaks characteristic of an intact isoquinuclidine structure with C15-C20 unsaturation, including the C21 (4.13 ppm), C17 (1.80) protons, and consistent COSY couplings indicated the "dimers" were the diastereomers 20a and 21a. The formation of structures 20a and 21a can be rationalized by a C16-C21 fragmentation analogous to that observed for catharanthine but with capture of vindoline at C7 rather than C16. Reformation of the C16-C21 bond could occur through a Mannich-type addition. (Scheme 3)

Scheme-3



In an effort to determine the fate of the remainder of the 19a it was subjected to Potier conditions in the absence of vindoline, followed by the NaBH₄ reductive work-up. A very good recovery of 8a was obtained. A similar experiment was also done using NaBD₄. The recovered 8a contained no deuterium. These results indicate that no fragmentation of 19a occurred in the absence of vindoline and that it was simply reduced back to 8a by NaBH₄ or NaBD₄. The implication of these experiments is that vindoline <u>catalyzes or induces</u> the fragmentation reaction, a phenomena previously observed for catharanthine-N-oxide.²³

The results with 8b were very similar. All of the amine oxide 19a reacted to give a mixture containing mainly unreacted vindoline, but showing NMR peaks suggesting one or more coupling products. A dimeric material 20b was characterized having spectral features closely analogous to one of the 20a, 21a diastereomers. In particular, the signal for C21 proton appears at 4.2 ppm as a singlet, consistent with the assignment to the bridgehead position.

B. [3,2]-Analogs 18a and 18b

The principal product from reaction of 18b under Fe^{3+} coupling conditions with NaBH₄ work-up was the allylic alcohol *E*-24b. The structure assignment was based on ¹H-NMR, COSY and ¹³C-NMR spectra. The ¹H-NMR data are given in Table 1. Nearly all of the vindoline was recovered unchanged. When the NaBH₄ reduction step was eliminated, the crude product mixture contained the aldehyde *Z*-23b and its *E*-isomer in about 40% total yield. The same aldehydes were formed in 30% yield when vindoline was omitted from the reaction mixture. Under the Potier conditions, 18b gave a 1.2:1 mixture of aldehydes *E*-23b and *Z*-23b in about 45% yield. The fragmentation proceeded with or without vindoline in the reaction mixture.

Similarly, 18a underwent oxidative fragmentation by Fe^{3+} but no coupling product was observed. The aldehyde 23a was obtained as a 10:1 *E:Z* mixture in about 35% yield. When a reductive work-up with NaBH₄ was done, the allylic alcohol *E*-24a was isolated. The same aldehydes were observed, along with some unreacted 18a when vindoline was omitted from the reaction mixture. Reaction of 18a and vindoline under Potier conditions gave no indication of a coupling product. Most of the vindoline remained unchanged but no product due to the 18a could be isolated.²⁴

The results from attempted couplings of 18a and 18b are summarized in Table 2. While the details of the individual reactions varied, the common pattern is that C16-C21 fragmentation occurs, but there is no evidence that the fragmentation product is trapped by vindoline. Instead the dominant process is N4-C16 bond formation, followed by ring opening between N4 and C21 leading to aldehydes 23a and 23b (Scheme 4).

C. Catharanthine Derivatives.

For comparison with the [2,1] and [3,2] analogs we also examined the reaction of catharanthine with Fe^{3+} in the presence and absence of vindoline. Anhydrovinblastine (AVLB) was obtained in good yield, in accord with previous work.¹⁶ Surprisingly, we found that almost no reaction occurred when vindoline was absent from the reaction mixture. Under conditions where >90% fragmentation-coupling occurs in the presence of vindoline, < 10% conversion of catharanthine occurred. Only trace amounts of an unidentified possible oxidation product were visible in the NMR spectrum. The implication is that vindoline participates in the rate-determining step for Fe^{3+} oxidation of catharanthine. To the best of our knowledge, this is a new observation.²⁵

Scheme 4



20-Deethylcatharanthine (10a) has previously been coupled with vindoline under Potier conditions.²⁶ Two diastereometric adducts 25a and 26a were isolated in 15% yield each. 10a has not previously been coupled with vindoline under Fe^{3+} conditions. The reaction provided the same two adducts obtained from the Potier coupling in about 30% yield each. The adducts had the features expected for deethyl analogs of AVLB and the ¹H NMR chemical shifts are in agreement with those reported earlier.²⁶ As with catharanthine, the omission of vindoline significantly retarded oxidation. Only about 20% of conversion occurred under the standard conditions, with the remainder of the 10a being recovered (Scheme 5).

N-Methylcatharanthine (10c) has previously been shown to undergo oxidative fragmentation under Potier conditions, although coupling with vindoline has not been established.²³ In the Fe^{3+} system, 10c was nearly completely converted in the presence of vindoline but no evidence of coupling was found. Most of the vindoline was recovered, but no oxidation products derived from 10c could be identified. In the absence of vindoline, 10c was recovered unchanged. Thus each of the catharanthine analogs is considerably less reactive under the Fe^{3+} conditions when vindoline is not included.

<u>Table 1.</u>	IWN	R Data for 23a, 23b, 24a, 24b ^a					
E-23a	<u>1.96s</u>	<u>3CH₂, 5CH₂, 6CH₂, 14CH, 17CH₂ 3.6-3.7m; 3.35 dd (J = 14.0, 6.0); 3.1-3.2 m (2H); 3.07 ddd (J = 17.0), 11.5, 5.5); 2.8-2.95 m (2H); 2.68 m 2.57 bd</u>	<u>9CH, 10CH, 11CH, 12CH</u> 7.67 d (J = 7.8); 7.31 d (J = 7.8); 7.21 t (J = 7.8); 7.13 t (J = 7.8)	<u>15CH</u> 6.94 dd (J = 15.5, 8.0)	<u>R</u> 6.50 dd (J = 15.5, 7.5)	<u>8.5</u> d J = 7.5	<u>CO₂CH₃</u> 3.73s (3H)
E-24a	8.02s	3.8-4.0 m (2H); 3.40 m, 3.02-3.2 m (3H); 2.75 dd (J = 11.0, 7.0); 2.4-2.7 m (2H)	7.78 d (J = 7.8); 7.35 d (J = 7.8); 7.19 t (J = 7.8); 7.13 t (J = 7.8)	5.78 dd (J = 16.5, 8.0)	5.61 dt (J = 16.5, 6.5, 6.5)	4.08 m (2H)	3.75s (3H)
E-23b	7.98s	3.5-3.8 m (2H); 3.36 dd (J = 14.0, 5.15); 3.0-3.3 m (3H); 2.90 dd (J =	7.66 d (J = 7.8); 7.34 d (J = 7.8); 7.18 t (J = 7.8); 7.17 t (J = 7.8)	6.57 d (J = 10.3)	2.19 q (2H, J = 7.5); 0.98 t (3H, J = 7.5)	9.90s	3.72s (3H)
Z-23b	7.94s	12.5, 8.0); 2.58 m (.h) 3.6-3.8 m; 3.4 m; 3.0-3.3 m (4H); 2.8- 3.0 m; 2.6 m (2H)	7.67d (J = 7.8); 7.35 d (J = 7.8); 7.22 t (J = 7.8); 7.14 t (J = 7.8)	6.63 d (J = 8.8)	2.10 q (2H, J = 7.5); 1.00 (3H, J = 7.5)	9.93s	373s (3H)
E-24b	7.93s	3.55-3.65 m (2H); 3.31 dd (J = 14.0, 5.4); 2.98-3.12 m (3H); 2.90 m; 2.76 dd (J = 12.1, 7.2); 2.50 dd (J = 16.5, 4.3); 2.39 dd (J = 12.4, 8.1)	7.68 d (J = 7.8); 7.35 d (J = 7.8); 7.18 t (J = 7.8); 7.12 t (J = 7.8)	5.40 d	2.10 q (2H, J = 7.5); 1.02 t (3H, J = 7.5)	3.98 q (J _{gem} = 12)	3.70s (3H)

(a) 300 Mhz in CDCl₃. Unless indicated otherwise, all peaks represent a single (1H) proton. Coupling constants are \pm 0.3 Hz.

	Η	arric Chloride	e (Produ	ct Ratios	from NMR) ^a	Potier-	Polono	<u>vsky (Prod</u> i	uct Ratios from NMR) ^a
	Re	actants	Rec Rea	overed	Products	Reactant	Rec <u>s Rea</u>	overed	Products
Analog	NIN	Fe ³⁺ (eq)	NIN	Analog		NIN	NIN	Analog	
10a	- 0	10	30%	10% 80%	25a (30%) 26a (30%)	1.1	%0	%0	25a (16%) 26a (16%) ^{Ref. 26}
10b	1 0	∞ ∞	20%	20% 90%	AVLB (62%)	1.1	%0	%0	AVLB (50-60%) ^{Ref.15a}
10c	1 0	16 16	100%	%06 >00%	unidentified	experi	ment not	lone	
18a	- 0 0	7 11 25	100%	40% 90% 50%	Z-23a (3%) E-23a (32%) E-24a ^b E-23a (30%)	1.1 0	%06	%0	unidentified unidentified
18b	- 0 0	30 6 27	80%	0 90% 60%	E-24b (40%) ^b E-23b (5%) Z-23b (5%) E-23b (25%)	1.1 0	%06	%0 %0	unidentified Z-23b (30%)
Sa	1 0	6-22 6-30	100%	100% 100%	no reaction no reaction	1.1 0	80%	0% 75%	20a (10%) 21a (10%) unidentified
8b	1 0	0 9	100%	100% 100%	no reaction no reaction	1.1 experi	70% ment not	0% done	20b (12%) 21b (12%)
a Product viald	S WPTP PC	timated from the	mace hal		1. 2 is - a f t - a - b - a - a		-		

a. Product yields were estimated from the mass balance of the crude product and the ratio of characteristic peaks in the NMR spectrum. Relative accuracy is estimated to be ±20%. There was some variation in extent of conversion from run to run. b. Reductive workup. c. See footnote 23.

Table 2 Summary of Results

Scheme-5



In order to explore this observation further, an additional experiment was done in which racemic 10a was present in 2:1 excess to vindoline under Fe^{3+} coupling conditions. The reaction proceeded normally and all of the vindoline reacted. The NMR of the recovered 10a was examined with Eu(hfc), shift reagent (0.3-0.5 equiv.) under conditions where excellent separation of the NH resonances of the 10a enantiomers could be observed. Within the accuracy of the measurement (±10%), there was no selectivity with respect to the enantiomers. This experiment indicates that, whatever the nature of the interaction that promotes reaction in the presence of vindoline, it is not highly enantioselective for the 10a enantiomers. It should be noted that the Potier coupling of (±)-10a²⁶ and (±)-catharanthine²⁷ also are not very stereoselective

III. Electrochemistry

The electrochemistry of catharanthine has been previously reported 18 and the voltammograms of 10a, 10b, 10c, 8a, 8b, 18a, 18b, and vindoline were obtained in CH₃CN-0.1 M LiClO₄ solution at glassy carbon electrode. The voltammetry of 10a was studied in some detail. A close analogy with the results for catharanthine was noted.

The cyclic voltammogram of 10a presented in Figure 1a shows two anodic peaks at 0.52 and 1.2 V vs. SCE, respectively. The first wave corresponds to the oxidation of 10a and the second wave is due to $10aH^+$ from the protonation of the parent molecule by protons liberated in the electrode reaction along the first wave. The ratio of the peak currents between the first and second wave was increased by increasing of the sweep rate, v. The average value of the transfer coefficient obtained from the peak widths (Ep-Ep/2) for the first wave at eight sweep rates fits well for α =0.60 and n=1. When the cyclic voltammogram was run in the presence of 2,6-lutidine, the current of the first peak increased (Fig. 1b). The height of the peak current corresponds to a two electron oxidation, assuming that the value of the diffusion coefficient for 10a is the same as previously

determined for catharanthine (D = $1.4\times10-5$ cm²/s).¹⁸ The wave obtained in the presence of 2,6-lutidine is broader and the average value of the transfer coefficient obtained from the peak widths for 10a (α =0.52) is similar to catharanthine (α =0.51) under the same experimental conditions. This result indicates that 2,6-lutidine provides an additional driving force for C16-C21 fragmentation of the intermediate radical cation of 10a.²⁷

The coulometry at controlled potential of 10a was performed at a platinum gauze electrode in CH₃CN-0.1 M LiClO₄ solution containing 2,6-lutidine. Cyclic voltammograms under constant conditions were obtained as a function of the charge passed. The results show that under controlled potential electrolysis, the oxidation wave at 0.51 V vs. SCE decreases at a rate corresponding to the consumption of 2 F/mol. At the end of the electrolysis a new peak at 0.65 V vs. SCE appeared, presumably corresponding to the oxidation product formed. The n value calculated from the cyclic voltammograms of 10a (see Experimental) as a function of the sweep rates, v, was found to increase by increasing log v. In the presence of 2,6-lutidine, the voltammograms of 10a showed n=2 (Fig. 1). The increase of n values with the sweep rates for 10a without the presence of 2,6-lutidine can be explained by the equations (1-3).

$$10a - 2e \rightarrow [10a]^{+2} \tag{1}$$

$$[10a]^{+2} + H_2O \Rightarrow [10a-OH_2]^{+2}$$
 (2)

$$[10a-OH_2]^{+2} + 10a \rightarrow [10a-OH]^{+} + [10aH]^{+}$$
 (3)

A two electron oxidative fragmentation of 10a gives dication, $[10a]^{2+}$ (eq. 1) which is attacked by water, presumably at the position C21, giving $[10a-OH_2]^{2+}$ (eq. 2). A diffusing molecule of 10a acts as a base deprotonating $[10a-OH_2]^{2+}$ (eq. 3) and therefore lowering n value from n=2 to n=1. This reaction scheme can be considered as a self-protonation for which a detailed kinetic studies were reported.²⁹ The theoretical treatment of self-protonation reaction demonstrates that the apparent number of electrons n is particularly affected by the variation of the rate of the following chemical reaction and sweep rate. At slower anodic sweep rates there is enough time for the following chemical reactions (eq. 2 and 3) to occur which results with the lower n values. At higher sweep rates, n gradually increases because the chemical reactions do not compete with diffusion.

The cyclic voltammograms of the other compounds, listed in Table 1, are also characterized by the same basic patterns. The relative ease of removing an electron from the tertiary amine moiety varies substantially among these compounds. The range for the peak potentials, E_p , varies from 0.50 to 0.64 V vs. SCE for catharanthine, 10a and the [3,2]-derivatives. The [2,1]-analogs 18a and 18b are harder to oxidize and showed

 E_p values of 0.83 and 0.77 V vs. SCE, respectively. All experimentally determined values for the transfer coefficient, α , fell in the range 0.54 to 0.68. The irreversibility is the result of the slow electron transfer. Oxidation at N4 is accompanied with the changing of hybridization of the amine moiety from sp³ to sp² of the resulting radical cation. Strain energy distorts the planar configuration thereby increasing the free energy for electron transfer. The electrochemistry of vindoline was studied previously³⁰ and results in Table 3 are also given for the purpose of comparison.

The shape of the voltammograms for all compounds listed in Table 3 in the presence of vindoline were studied in acetonitrile solution. Three types of voltammograms, i.e. A, B, and C in Figure 2, were obtained. Type A voltammograms, exemplified by that for 10a with and without vindoline are shown in Figure 2a. The peak potential of 10a in the presence of vindoline was shifted about 30 mV cathodically indicating that the overall rate of the fragmentation/coupling reaction is enhanced by the presence of vindoline, which is consistent with previous chemical²³ and electrochemical¹⁸ studies on catharanthine. The cyclic voltammogram for 18a (Fig. 2b) represents type B in which there is no essential change of the oxidation wave with or without vindoline. Type C voltammograms (Fig. 2c), as found for 8b in the presence of vindoline, shows increase of the peak current which is result of the overlapping waves for 8b and vindoline due to their close oxidation potentials.

Compound	E _p 1 ^a	E _p 2	αn	Shape of Cv ^b
	(V vs. SCE)	(V vs. SCE)		
10 a	0.51	1.17	0.60	A
10b	0.50	1.15	0.59	Α
10c	0,64	1.18	0.68	В
18a	0.56	1.32	0.56	В
18b	0.52	1.24	0.62	В
8a	0.83	1.34	0.54	С
8b	0.77	1.26	0.54	С
Vindoline	0.76	0.92	0.68	-

Table 3. Oxidation peak potentials and shapes of cyclic voltammograms

a) Sweep rate: 50 mV/s; GCE (A = 0.05 cm^2); CH₃CN- 0.1 M LiClO₄.

b) In the presence of vindoline.



Left: Cyclic voltammograms in CH₃CN-0.1 M LiClO₄; GCE; v=75 mV/s; (a) 10a (1 mM); (b) plus 2,6-lutidine (4 mM); Right: Sweep rate dependance of the apparent number of electrons n for oxidation of 10a (1 mM) o: plus 2,6-lutidine (4 mM).

Figure 2



Cyclic voltammograms in CH₃CN-0.1 M LiClO₄; GCE; v=50 mV/s; A (a) DECATH (1 mM), (b) plus VIND (1 mM); B (a) *inv*-DECATH (3.8 mM), (b) plus VIND (1.3 mM); C (a) *iso*-CATH (1.78 mM), (b) plus VIND (1.4 mM).

Figure 1

IV. Conformational Analysis

Since fragmentation of the C16-C21 bond is expected to involve participation by the indole ring, the orientation of that bond with respect to the indole ring is presumably a crucial structural factor. Molecular models suggest two potential conformations with values of the dihedral angle θ between the plane of the indole ring and the C16-C21 bond of about 0° and 60°, corresponding to boat-like and chair-like conformations of the seven-membered C-ring. To pursue this point, minimizations using the MacroModel routine were done for 10a, 10c, 8a and 18a. The dihedral angles for the minima were 33.4°, 12.2°, 39.4° and 32.8°, respectively. Thus while it appears that interaction of the N-methyl group with the C16 ester substituent in 10c favors a boat-like conformation of the C-ring, the other analogs adopt conformations which are very similar to one another, suggesting that reactivity differences among the analogs must be primarily electronic in origin.

Discussion

Our results on coupling of catharanthine and its deethyl analog 10a are consistent with prior studies and extend the information available on Fe³⁺-mediated coupling. 10a is coupled by Fe³⁺ but there is no evidence of significant enantioselectivity, consistent with the prior results of Potier coupling.^{26,27} The most significant new result, which pertains to catharanthine, 10a, and 10c is that the reaction of the catharanthine analogs with Fe³⁺ is significantly catalyzed or induced by vindoline.

The [2,1]-analogs were unreactive toward Fe^{3+} , a result which is consistent with their higher oxidation potential. Reaction did occur under Potier conditions. However trapping by vindoline was inefficient and coupling occurs at C7 rather than C16 (*iboga* numbering). As has previously been observed with catharanthine,²³ the fragmentation reaction of **19a** appears to be catalyzed or induced by vindoline. When vindoline was omitted from Potier reactions, the only product found after reductive work up was **8a**. Furthermore, no deuterium was incorporated when NaBD₄ was used for reductive work-up. This excludes a C3-N4 or C5-N4 elimination followed by reduction as the source of the recovered **8a**. Instead it appears **19a** did not react in the absence of vindoline and was simply reduced to **8a** on workup.

The [3,2]-analogs underwent C16-C21 fragmentation under both Fe^{3+} and Potier conditions. The oxidation products 23a and 23b can be isolated in fair yield from the Fe^{3+} oxidations, but there was no evidence of coupling with vindoline. The formation of these products can be rationalized by C16-C21 oxidative fragmentation analogous to catharanthine, followed by trapping by water (Scheme 4). Under Potier conditions, 18a gave several unidentified products and small amounts of 23a. The aldehyde Z-23b was the main product from 18b under Potier conditions. Both the Fe^{3+} and Potier reactions occurred at comparable rates in the presence and absence of vindoline. Thus the [3,2]-series appears to undergo C16-C21 fragmentation in a manner analogous to catharanthine but there is no trapping by vindoline under either the aqueous Fe^{3+} conditions or the anhydrous Potier conditions. The failure to react with vindoline may reflect both decreased reactivity at the C16 carbon and greater steric shielding by the benzenoid portion of the indole ring.

In seeking to clarify the differences between the catharanthine series and the [2,1]- and [3,2]-analogs, we considered the following structural factors. (1) Catharanthine, 10a, 18a and 18b can undergo fragmentation assisted by N1-deprotonation. Because of N-alkylation, 10c, 8a and 8b cannot benefit from such assistance. (2) The indole ring is a better electron donor at C3 than C2 (indole numbering). As a result the fragmentation products from 18a and 18b should be more stable than those from catharanthine, 10a and 8a, 8b. (3) If factors (1) and (2) work together in a kinetic sense, the predicted ease of oxidative fragmentation is 18a, 18b >10a, 10b > 8a, 8b. This is in qualitative agreement with the observed reactivity, particularly when it is noted that vindoline acceleration appears to be required for the [2,3]- and [2,1]-series but not the [3,2]-series.

These chemical results are in qualitative agreement with the electrochemical studies. Most notably, vindoline was found to shift the oxidation potential of catharanthine and deethylcatharanthine by 30 mV. No such shift is seen with 18a or 18b and the reactivity of these analogs is unaffected by the presence of vindoline under chemical conditions. The [2,1]-series is the most difficult to oxidize, both under chemical and electrochemical conditions. The fact that vindoline is oxidized at a similar potential obscures any shift it might cause in the potential of 8a. However, vindoline does appear to accelerate Potier fragmentation of the 19a.

We refer to the accelerating effect of vindoline on both the Fe³⁺ and Potier oxidation on catharanthine, 10a and 10c and on Potier oxidation of 19a as the "vindoline effect." We have previously proposed that for Potier fragmentation of catharanthine, vindoline accelerates fragmentation simply by acting as a base to deprotonate the indole-N-H and facilitate C16-C21 fragmentation, but we noted that Potier fragmentation of 10c was also accelerated by vindoline.²³ Since 10c and the [2,1]-analogs are substituted at nitrogen, the deprotonation mechanism is not applicable. Furthermore, the observation of the "vindoline effect" in Fe³⁺ oxidation 0.05 M glycine solution, where solution acidity should be independent of vindoline, suggests some mechanism besides N-deprotonation must operate. The "vindoline effect" in the Fe³⁺ oxidations is the formal equivalent of a "concerted" fragmentation mechanism.^{16b}

We have considered two limiting mechanistic descriptions of the "vindoline effect". One is base catalysis. We have previously cited evidence that N-deprotonation triggers C16-C21 fragmentation under Potier conditions in the case of catharanthine.²³ The base-catalysis explanation is not applicable to 10c or the [2,1]-series, yet acceleration of Potier fragmentation is noted for 10c and 19a and Fe³⁺ oxidation is also accelerated for 10c. Acceleration might come from π - π electronic interaction through which vindoline increases the effective electron-density of the indole ring. The extreme for such a π - π interaction would be concerted electron transfer leading to coupling.

From a synthetic perspective, the salient outcome of the present work is further evidence of the extremely tight structural parameters which limit the catharanthine-vindoline coupling process. While deethylcatharanthine 10a retains the ability to couple, albeit with somewhat reduced efficiency, under both the Fe^{3+} and Potier conditions, both the [2,1] and [3,2]-series fall outside these parameters, despite their close structural relationship to catharanthine. The other noteworthy observation is the evidence for vindoline participation in the Fe^{3+} -mediated coupling, which parallels previously noted effects in Potier oxidation and electrochemistry.

Experimental Section

Methyl 2-(benzyloxycarbonyl)-7-exo-(1-phenylsulfonylindol-2-yl)-2-azabicyclo[2.2.2]oct-5-ene-7-endo carboxylate (3a) A mixture of indoleacrylate 1 (6.0 g, 0.018 mol) and dihydropyridine 2a (19.0 g, 0.088 mol) was heated in an oil bath at 100°C for 60 h under inert atmosphere. Flash chromatography (ether: CH₂Cl₂: hexane 1:1:2) gave an oil, which was crystallized from ether and hexane to give 8.29 g (84%) of 3a, m.p. 182-184°C. ¹H NMR (CDCl₃, δ ppm, 300MHz): 7.69 (t, 1H, J = 5 Hz), 7.52 (d, 2H, J = 7.5 Hz), 7.40 (m, 2H), 7.28 (m, 2H), 7.23 (s, 2H), 7.14 (m, 2H), 6.57 (t, 1H, J = 7.5 Hz), 6.37 (t, 1H, J = 7.5 Hz), 5.46 (d, 1H, J = 6 Hz), 5.19 (d, 1H, J = 12 Hz), 3.58 (s, 3H), 2.97 - 3.06 (dd, 2H, $J_1 = 13.5$ Hz, $J_2 = 10$ Hz), 2.94 (d, 2H, J = 13.5 Hz). MW calcd. for $C_{31}H_{28}N_2O_6S$ (556.56), found MS (CI) m/z 557 (M +1⁺). Methyl 7-exo-(1-phenylsulfonylindol-2-yl)-2-azabicyclo[2.2.2]oct-5-ene-7-endo-carboxylate (4a) TMSI was prepared in situ from iodine (9.38 g, 0.035 mol) and hexamethyldisilane (16.4 ml, 0.084 mol). The mixture was carefully heated to 100°C until the purple color faded, and then cooled to rt. A 500 ml flask was charged with 8.0 g (0.014 mol) of carbamate 3a in 400 ml of dry CH₂Cl₂ and cooled to 0°C. The TMSI solution was slowly added. The mixture was stirred at 0°C for one h then at rt for 30 min. When starting material was consumed, 120 ml of 0.3 M HCl in methanol was added and the mixture was stirred for one h. Then 100 ml of 10% HCl was added and stirred for 10 min. The organic solvents were removed using a rotary evaporator and the aqueous residue was extracted with ether (3 x 100 ml). The aqueous layer was then basified to pH 9 with a cold conc. NaOH solution. The mixture was extracted with EtOAc (4 x 75 ml), dried (Na₂CO₃), and concentrated to give 3.52 g (58%) of 4a as a white foam (> 90% endo-carboxylate by NMR). ¹H NMR (CDCl₃, δ ppm, 300MHz): 7.73 (t, 1H, J = 4.5 Hz), 7.56 (d, 2H, J = 7.5 Hz), 7.47 (t, 1H, J = 4.5 Hz), 7.41 (t, 1H, J = 7.5 Hz), 7.31 (d, 1H, J = 7.5 Hz), 7.27 (d, 1H, J = 4.5 Hz), 7.15 (t, 2H, $J_{I} = 4.5$ Hz), 7.15 (t, 2H, J_{I} = 4.5 Hz), 7.15 (t, 2H, J_{I} = 4.5 H

7.5 Hz, $J_2 = 4.5$ Hz), 7.05 (s, 1H), 6.54 (t, 1H, J = 7.5 Hz), 6.36 (t, 1H, J = 7.5 Hz), 4.26 (d, 1H, J = 7.5 Hz), 3.57 (s, 3H), 2.90 (d, 1H, J = 15 Hz), 2.72 (s, 1H), 2.67 (d, 1H, J = 10.5 Hz), 2.40 (d, 1H, J = 10.5 Hz), 2.05 (d, 1H, J = 15 Hz). MW calcd. for C₂₃H₂₂N₄O₄S (422.37), found MS (CI) m/z 423 (M +1⁺). Methyl 2-(chloroacetyl)-7-exo-(1-phenylsulfonylindql-2-yl)-2-azabicyclo[2.2.2]oct-5-ene-7-endo-carboxylate (5a) The amine 4a (2.20 g, 5.21 mmol) was dissolved in 100 ml of dry CH₂Cl₂. This solution was cooled to 0°C and chloroacetic anhydride (2.67 g, 15.6 mmol) in 10 ml of dry CH₂Cl₂ was added followed by DMAP (191 mg, 1.56 mmol). After 20 min at 0°C, Et₃N (0.87 ml, 6.25 mmol) was added and stirring was continued at 0°C for 5 h. The reaction mixture was washed with 5% NaHCO₃, dried with Na₂CO₃, concentrated and subjected to flash chromatography (EtOAc:hexane 2:5) to yield 1.81 g (70%) of 5a, m.p. 184-186°C. ¹H NMR (CDCl₃, δ ppm, 300MHz): 7.65 (t, 1H, J = 4.5 Hz), 7.50 (d, 2H, J = 7.5 Hz), 7.43 (dd, 2H, $J_I = 21$ Hz, $J_2 = 7.5$ Hz), 7.29 (t, 2H, $J_I = 7.5$ Hz, $J_2 = 9$ Hz), 7.15 (s, 1H), 7.12 (t, 2H, J = 7.5 Hz), 6.37 (t, 1H, J = 7.5 Hz), 5.84 (d, 1H, J = 6 Hz), 3.84 (s, 2H), 3.59 (s, 3H), 3.25 (d, 1H, J = 9 Hz), 3.10 (d, 1H, J = 15 Hz), 3.06 (d, 1H, J = 9 Hz), 3.02 (s, 1H), 2.18 (d, 1H, J = 15 Hz). MW calcd. for C₂₅H₂₃ClN₂O₅S (498.91), found MS (CI) m/z 499, 501 (3 : 1) (M +1⁺).

Methyl 2-(chloroacetyl)-7-exo-(indol-2-yl)-2-azabicyclo[2.2.2]oct-5-ene-7-endo-carboxylate (6a) Chloroacetamide 5a (500 mg, 1.0 mmol) and 1,5-dimethoxynaphthalene (93 mg, 0.5 mmol) were dissolved in 400 ml of EtOH. This solution was then transferred into a 500 ml photolysis apparatus. Ascorbic acid (530 mg, 3.0 mmol), 50 ml EtOH and 50 ml water were added in sequence. The mixture was stirred under nitrogen and subjected to photolysis using a 450W mercury-arc Hanovia lamp in a quartz immersion well equipped with a pyrex filter sleeve for 5 h. The solvent was removed by rotary evaporation. The residue was dissolved in CH₂Cl₂, dried with K₂CO₃ and concentrated. Flash chromatography (EtOAc:hexane 2:5) gave a foam, which was crystallized with EtOAc and hexane to give 6a, m.p. 210-211°C, lit¹¹ 210-211°C (273 mg, 65%). ¹H NMR (CDCl₃, δ ppm, 300MHz): 9.20 (s, 1H), 7.52 (d, 1H, J = 7.5 Hz), 7.38 (d, 1H, J = 7.5 Hz), 7.16 (t, 1H, J = 7.5 Hz), 7.06 (t, 1H, J = 7.5 Hz), 6.59 (t, 1H, J = 7.5 Hz), 6.45 (t + s, 2H, J = 7.5 Hz), 6.06 (d, 1H, J = 7.5 Hz), 3.83 (dd, 2H, $J_I = 16.5$ Hz, $J_2 = 10.5$ Hz), 3.60 (s, 3H), 3.33 (d, 1H, J = 9 Hz), 3.17 (d, 1H, J = 9 Hz), 3.04 (s, 1 H), 2.97 (d, 1H, J = 15 Hz), 2.19 (d, 1H, J = 15 Hz). MW calcd for C1₉H₁₉ClN₂O₃ (358.82), found MS (CI) m/z 359, 361 (3 : 1) (M +1⁺).

[2.1]-Deethyllactam (7a) Sodium hydride (22 mg, 0.76 mmol, 80%/wt suspension in mineral oil) was added to a dry 25 ml flask and was washed with hexane. The NaH was then suspended in 5 ml of dry THF. Chloroacetamide 6a (138 mg, 0.385 mmol) was dissolved in 8 ml of dry THF and added via syringe. The mixture was allowed to stir at rt for 3 h under Ar. The reaction was quenched with 1 ml of water and the THF was removed by rotary evaporation. The residue was partitioned between CH_2Cl_2 and water. The organic phase was dried over MgSO₄, and the solvent was removed solvent *in vacuo* to yield 121 mg (97.5%) of crude material which was purified by flash chromatography (EtOAc:hexane = 2:1) to yield 104 mg (83%) of pure 7a. An analytical sample was recrystallized from EtOAc and hexane, mp. 193-194°C. ¹H NMR (CDCl₃, δ ppm, 300 MHz): 7.50 (d, 1H, J = 7.5 Hz), 7.36 (d, 1H, J = 8.4 Hz), 7.24 (t, 1H, J = 8.4Hz), 7.11 (t, 1H, J = 7.5 Hz), 6.74 (t, 1H, J = 7.2Hz), 6.38 (t, 1H, J = 6.9 Hz), 6.38 (s, 1H), 5.39 (d, 1H, J = 6.3Hz), 5.24 (d, 1H, J = 14.1Hz), 5.01 (d, 1H, J = 14.1Hz), 3.72 (dd, 1H, $J_I = 10.6$ Hz, $J_2 = 2.5$ Hz), 3.62 (s, 3H), 2.99 (dd, 1H, $J_I = 15.4$ Hz, $J_2 = 2.3$ Hz), 2.95 (m, 2H), 1.78 (d, 1H, J = 12.3 Hz). ¹³C NMR (CDCl₃) 172.54, 169.84, 139.96, 139.88, 137.62, 128.28, 127.94, 122.77, 120.90, 120.80, 110.00, 103.64, 54.74, 53.37, 51.33, 51.03, 50.19, 34.70, 31.66. MW calcd. for C₁₉H₁₈N₂O₃ (322.2), found MS (CI) m/z 323 (M+1⁺). Anal. Calcd.: C, 70.79; H, 5.63; N, 8.69. Found: C, 70.53; H, 5.73; N, 8.59.

Deethyl[2,1]-Analog (8a) The procedure Szantay^{10,11} developed for catharanthine was used. In a 50 ml flask, lactam 7a (20 mg, 0.062 mmol) was dissolved in 8 ml of dry THF. NaBH₄ (102 mg, 2.69 mmol) was added to the flask in one portion. The flask was cooled to 0°C and BF₃-OEt₂ (0.48 ml, 3.90 mmol) was syringed in over one minute. The ice bath was removed and the reaction was allowed to stir at room temperature for 3 h. The solvent was removed *in vacuo*. The residue was dissolved in 20 ml of CH₂Cl₂. Water (20 ml) was slowly added. The organic phase was dried over Na₂SO₄ and concentrated to yield 17 mg of crude material, which was subjected to flash chromatography (acetone:CH₂Cl₂ = 4:1) to yield 7 mg (37%) of pure 8a. ¹H NMR (CDCl₃, δ ppm, 300MHz): 7.48 (d, 1H, *J* = 7.8 Hz), 7.27 (d, 1H, *J* = 6.9 Hz), 7.18 (t, 1H, *J* = 7.6 Hz), 7.06 (t, 1H, *J* = 7.5 Hz), 6.50 (t, 1H, *J* = 6.8 Hz), 6.38 (t, 1H, *J* = 6.8 Hz), 5.96 (s, 1H), 4.58 (m, 1H), 4.27 (dt, 1H, *J₁* = 15 Hz, *J₂* = 2.7 Hz), 3.77 (s, 3H), 3.55 - 3.35 (m, 2H), 3.01 (d, 1H, *J* = 9.6 Hz), 2.93 - 2.80 (m, 3H), 1.79 (d, 1H, *J* = 12.3 Hz). ¹³C NMR (CDCl₃) 174.15, 141.39, 138.39, 134.62, 134.06, 127.64, 122.05, 120.68, 120.18, 109.24, 102.38, 59.01, 57.52, 53.88, 52.79, 49.78, 41.59, 33.36, 30.74. HRMS (EI) m/z calcd. for C₁₉H₂₀N₂O₂ (308.1525), found 308.1534 (M⁺).

Methyl 7-exo-(1-phenylsulfonylindol-2-yl)-2-(benzyloxycarbonyl)-6-ethyl-2-azabicyclo[2.2.2]oct-5-ene-7endo-carboxylate (3b) A mixture of dihydropyridine 2b (6.18 g, 25.4 mmol) and the indolyl acrylate 1 (2.5 g, 7.32 mmol) was heated without any solvent at 100°C for 60 h (under argon). The product was purified by flash column chromatography on silica gel (10% EtOAc in CH₂Cl₂) to yield 2.01 g (47%) of pure 3b (exclusively the *exo*-indolyl adduct), as an oil. ¹H NMR (CDCl₃, δ ppm 300 MHz) 7.45 - 6.95 (m, 15 H), 6.08 (dd, 1H, $J_I = 6.8$ Hz, $J_2 = 1.5$ Hz), 5.44 - 5.01 (dd + dd', 2H, $J_I = 43.95$ Hz, $J_2 = 12.7$ Hz), 3.56 (s, 3H), 3.10 - 2.81 (m, 5H), 2.10 - 2.00 (m, 3H), 1.08 + 1.05 (t + t', 3H, J = 7.3 Hz). MW calcd. for C₃₃H₃₂N₂O₆S (584.61), found MS (CI) m/z 585 (M +1⁺).

<u>Methyl</u> 7-exo-(1-phenylsulfonylindol-2-yl)-6-ethyl-2-azabicyclo[2.2.2]oct-5-ene-7-endo-carboxylate (4b) Trimethylsilyl iodide (13.5 mmol) was prepared *in situ* as described for 4a. Compound 3b (2.81 g, 4.81 mmol) was dissolved in 80 ml of dry CH₂Cl₂ and cooled to 0°C. The freshly prepared TMSI was slowly syringed into the reaction flask and the solution was allowed to stir for 30 min at 0°C and for 60 min at rt. The progress of the reaction was monitored by TLC. When 3b was consumed, the reaction mixture was processed as for 4a to yield 1.32g (61%) of 4b that was >80% the *exo* stereoisomer. ¹H NMR (CDCl₃, δ ppm 300 MHz) 7.73 (m, 1H), 7.57 (d, 1H, J = 7.5Hz), 7.51 - 7.11 (m, 7H), 7.05 (s, 1H), 6.11 (d, 1H, J =5.4 Hz), 4.08 (s, 1H), 3.58 (s, 3H), 2.92 (dt, 1H, $J_I = 13.4$ Hz, $J_2 = 3.2$ Hz), 2.71 (m, 2H), 2.65 (d, 1H, J =9.7 Hz), 2.41 (dt, 1H, $J_I = 9.4$ Hz, $J_2 = 2.4$ Hz), 2.10 + 2.04 (m + m', 2H), 1.11 (t, 3H, J = 7.5 Hz). MW calcd. for C₂₅H₂₆N₂O₄S (450.56), found MS (CI) m/z 451 (M +1⁺).

(±) Methyl 7-exo-(1-phenylsulfonylindol-2-yl)-2-chloroacetyl-6-ethyl-2-azabicyclo[2.2.2]oct-5-ene-7-endocarboxylate (5b) The deprotected amine 4b (1.02 g, 2.27 mmol) was dissolved in 80 ml of CH₂Cl₂ and converted to 5b as described for 5a, except that diisopropylethylamine was used in place of Et₃N. The crude product was purified by flash column chromatography on silica gel (EtOAc: hexane 1:1) to give 0.97 g (81%) of 5b, mp. 74-76°C. ¹H NMR (CDCl₃, δ ppm 300 MHz) 7.64 (m, 1H), 7.51 - 7.29 (m, 4H), 7.29 (t, 2H, J = 7.8 Hz), 7.16 (s, 1H), 7.11 (m, 2H), 6.10 (dd, 1H, $J_I = 6.3$ Hz, $J_2 = 1.5$ Hz), 5.64 (d, 1H, J = 1.5Hz), 3.84 (d, 2H, J = 1.0 Hz), 3.59 (s, 3H), 3.20 (dd, 1H, $J_I = 8.8$ Hz, $J_2 = 1.5$ Hz), 3.17 - 2.91 (m, 4H), 2.19 - 2.05 (m, 3H), 1.07 (t, 3H, J = 7.3 Hz). MW calcd. for C₂₇H₂₇ClN₂O₄S (527.04), found MS (CI) m/z 527, 529 (3 : 1) (M +1⁺).

(±) Methyl 2-(chloroacetyl)-6-ethyl-7-exo-(indol-2-yl)-2-azabicyclo[2.2.2]oct-5-ene-7-endo-carboxylate (6b) Chloroacetamide 5b (0.500 g, 0.950 mmol) was desulfonylated as described for 6a. The crude material was purified by flash column chromatography on silica gel (7% EtOAc in CH₂Cl₂) to yield 0.243 g (66%) of 6b. ¹H NMR (CDCl₃, δ ppm 300 MHz) 9.21 (bs, 1H), 7.53 (d, 1H, J = 6.3 Hz), 7.39 (d, 1H, J = 7.8 Hz), 7.17 (t, 1H, J = 6.4 Hz), 7.05 (t, 1H, J = 7.3 Hz), 6.42 (s, 1H), 6.16 (d, 1H, J = 5.9 Hz), 5.90 (s, 1H), 3.82 (dd, 2H, $J_1 =$ 16.5Hz, $J_2 = 10.5$ Hz), 3.61 (s, 3H), 3.39 (s, 1H), 3.31 (d, 1H, J = 9 Hz), 3.15 (d, 1H, J = 9 Hz), 3.01 - 2.93 (m, 2H), 2.25 - 2.11 (m, 3H), 1.07 (t, 3H, J = 7.3 Hz). MW calcd. for C₂₁H₂₃ClN₂O₃ (386.88), found MS (CI) m/z 387, 389 (3 : 1) (M +1⁺).

[2,1]-Lactam (7b) Chloroacetamide 6b (0.071 g, 0.183 mmol) was cyclized using NaH in THF as described for 7a to give 0.052 g (81%) of 7b. An analytical sample was purified by flash chromatography (10% EtOAc in CH₂Cl₂), and recrystallized from EtOAc and hexane, mp. 242-244°C (decomp). ¹H NMR (CDCl₃, δ ppm 300 MHz) 7.50 (d, 1H, J = 7.8 Hz), 7.36 (d, 1H, J = 8.4 Hz), 7.23 (dt, 1H, $J_I = 8.1$ Hz, $J_2 = 1.0$ Hz), 7.10 (dt, 1H, $J_I = 7.8$ Hz, $J_2 = 1.0$ Hz), 6.36 (dd, 1H, $J_I = 6.6$ Hz, $J_2 = 1.2$ Hz), 5.28 (d, 1H, J = 14.1 Hz), 5.18 (s, 1H), 5.01 (d, 1H, J = 14.1 Hz), 3.66 (dd, 1H, $J_I = 10.5$ Hz, $J_2 = 2.7$ Hz), 3.61 (s, 3H), 3.07 - 2.85 (m, 3H), 2.22 (q, 2H, J = 7.5 Hz), 1.76 (dd, 1H, $J_I = 13.8$ Hz, $J_2 = 1.0$ Hz), 1.09 (t, 3H, J = 7.5 Hz). ¹³C NMR (CDCl₃) 172.26, 169.95, 142.77, 140.56, 137.60, 130.39, 127.98, 122.70, 120.87, 120.77, 110.01, 103.42, 55.76, 54.63, 53.10, 51.81, 50.27, 35.37, 31.44, 27.32, 11.81. MW calcd. for C₂₁H₂₂N₂O₃ (350.42), found MS (CI) m/z 351 (M +1⁺). Anal. Calcd.: C, 71.98; H, 6.33; N, 7.99. Found: C, 71.95; H, 6.40; N, 7.90. [2.1]-Analog (8b) Lactam 7b (0.100 g, 0.28 mmol) was reduced as described for 8a. The crude amine was obtained as a hard film (89 mg, 93% yield) and subjected to flash chromatography on silica gel (3:1, CHCl₃: acetone) to give 52 mg (54% yield) pure 8b. ¹H NMR (CDCl₃, δ ppm 300 MHz) 7.47 (d, 1H, J = 7.8 Hz), 7.27 (d, 1H, J = 7.3 Hz), 7.18 (t, 1H, J = 7.6 Hz), 7.06 (t, 1H, J = 7.3 Hz), 5.94 (~ 2H), 4.58 (ddd, 1H, $J_I =$ 16.6 Hz, $J_2 = 10.0$ Hz, $J_3 = 3.2$ Hz), 4.27 (dt, 1H, $J_I = 15.1$ Hz, $J_2 = <3$ Hz), 4.02 (s, 1H), 3.77 (s, 3H), 3.50-3.41 (m, 2H), 3.05-2.78 (m, 4H), 2.32-2.05 (m, 2H), 1.75 (d, 1H, J = 13.7 Hz), 1.06 (t, 3H, J = 7.3 Hz). NMR (CDCl₃) 174.74, 148.08, 137.43, 126.75, 123.88, 121.08, 119.75, 119.25, 108.46, 101.46, 62.89, 56.29, 52.99, 51.61, 49.62, 40.64, 33.04, 29.82, 25.82, 10.34. HRMS (EI) m/z calcd for C₂₁H₂₄N₂O₂ (336.1838), found 336.1828 (M⁺).

<u>±-20-Deethyl-5-oxocatharanthine 9a</u> Chloroacetamide 6a (500 mg, 1.4 mmol) was dissolved in 900 ml of MeOH and 820 mg of NaHCO₃ was added. The solution was photolyzed under nitrogen for 6 h using a 450 W mercury-arc Hanovia lamp in a quartz immersion well with a Vycor filter sleeve. The methanol was evaporated, yielding a solid which was partitioned between CH₂Cl₂ and water. The CH₂Cl₂ layer was dried over K₂CO₃ and subjected to flash chromatography (EtOAc: hexane = 1:1.5) to yield 202 mg (45%) of 9a as an off-white solid. m.p. 295-296°C; lit⁸ 292-294°C. ¹H NMR (CDCl₃, δ ppm, 300 MHz): 8.06 (bs, 1 H), 7.56 (d, 1 H, J = 7.5 Hz), 7.27 (d, 1 H, J = 7.5 Hz), 7.21 - 7.11 (m, 2 H, J = 7.5 Hz), 6.68 (t, 1 H, J = 7.5 Hz), 6.49 (t, 1 H, J = 7.5 Hz), 5.33 (d, 1 H, J = 6.3 Hz), 4.17 (d, 1 H, J = 15.6 Hz), 3.78 (d, 1 H, J = 15.6 Hz), 3.67 (s, 3 H), 3.59 (dd, 1 H, J₁ = 10.7, J₂ = 3 Hz), 3.00 (d, 1 H, J = 10.7 Hz), 2.95 (m, 1 H), 2.65 (dt, 1 H, J₁ = 13.4 Hz, J₂ = 4 Hz), 1.80 (d, 1 H, J = 13.4 Hz). MW calcd for C₁₉H₁₈ N₂O₃ (322.2) found MS (CI) m/z 323 (M+1⁺).

<u>±-20-Deethylcatharanthine 10a</u> Lactam 9a (460 mg, 1.43 mmol) was dissolved in 70 ml dry THF. NaBH₄ (2.35 g, 62.2 mmol) was added in one portion. The reaction mixture was cooled to 0°C and BF₃•OEt₂ (11.2 ml, 90 mmol) was syringed in dropwise slowly at 0°C and the mixture (yellow suspension) stirred at room temperature for 3 h under nitrogen. The THF was evaporated and methanol (50 ml), H₂O (10 ml) and 10% HCl (5 ml) were added. This acidic solution was stirred at room temperature for 4 h. The methanol was evaporated and the residue mixed with CH₂Cl₂ and 10% NaOH at pH 8-9. The water layer was extracted with CH₂Cl₂. The combined organic layers were dried with anhydrous K₂CO₃ and concentrated to a yellow foam (438 mg, 99.6%). The purity was checked by TLC (pure EtOAc). m.p. 155-160°C; lit⁸ 155-160°C. ¹H NMR (CDCl₃, δ ppm, 300 MHz): 7.72 (bs, 1 H), 7.48 (d, 1 H, J = 7.5 Hz), 7.25 (d, 1 H, J = 7.5 Hz), 7.15 (t, 1 H, J = 7.5 Hz), 7.10 (t, 1 H, J = 7.5 Hz), 6.60 (t, 1 H, J = 7.5 Hz), 6.39 (t, 1 H, J = 7.5 Hz), 4.38 (d, 1 H, J = 6.2 Hz), 3.75 (s, 3 H), 3.55 (m, 1 H), 3.26 - 3.88 (m, 2 H), 2.91 (m, 1 H), 2.90 (s, 2 H), 2.76 (m, 1 H), 2.69 (d, 1 H, J = 13 Hz), 1.83 (dd, 1

H, $J_1 = 13.5$ Hz, $J_2 = 2$ Hz). ¹³C NMR (CDCl₃, δ ppm): 174.7, 135.9, 135.1, 134.8, 133.1, 129.0, 121.9, 119.5, 118.2, 110.7, 110.5, 56.8, 55.5, 53.0, 52.6, 48.8, 38.2, 30.7, 21.3. MW calcd for C₁₉H₂₀N₂O₂ 308.15, found MS (CI) m/z; 309 (M+1⁺).

(\pm)-5-Oxocatharanthine 9b To a 190 ml photolysis cell was added 5b (90 mg, 0.170 mmol), 1,5-dimethoxynaphthalene (94 mg, 0.499 mmol), NaBH₃CN (10 mg, 0.159 mmol), and NaHCO₃ (100 mg, 1.19 mmol) in 190 ml of methanol. The solution was irridated under nitrogen with light from a 450 W Hanovia lamp for 5 h (Vycor filter)The methanol was removed by rotary evaporation and the residue partitioned between CH₂Cl₂ and water. The organic phase was dried (K₂CO₃) and concentrated. The residue was chromatographed on silica gel (10% EtOAc in CH₂Cl₂) to yield 15 mg (25%) of 9b. This material was then crystallized from acetone and hexane. This compound matched the expected ¹H NMR and MS data.^{10 1}H NMR (CDCl₃, δ ppm, 300 MHz): 7.97 (bs, 1H), 7.56 (d, 1H, J = 7.3 Hz), 7.27 - 7.10 (m, 3H), 6.26 (d, 1H, J = 6.4 Hz), 5.11 (s, 1H), 4.20 (d, 1H, J = 15.6 Hz), 3.78 (d, 1H, J = 15.2 Hz), 3.66 (s, 3H), 3.54 (dd, 1H, J₁ = 10.2 Hz, J₂ = 2.4 Hz), 2.95 (d, 1H, J = 10.3 Hz), 2.90 (m, 1H), 2.71 (m, 1H), 2.28 (q, 2H, J = 6.8 Hz), 1.77 (d, 1H, J = 13.2 Hz), 1.10 (t, 3H, J = 7.3 Hz). MW calcd. for C₂₁H₂₂N₂O₃ (350.42), found MS (Cl) m/z 351 (M+1⁺).

<u>Methyl 2-Chloropropenoate</u>³¹ To a 100 ml one-neck round-bottom flask charged with methyl 2,3-dichloropropionate³² (12.0 ml, 71.0 mmol), a solution (in 40 ml water) of NaHCO₃ (15.0 g, 141 mmol) and *tetra*butylammonium bromide (1.26 g, 3.90 mmol) was added. A small amount (about 20 mg) of phenothiazine, along with 30 ml of CH₂Cl₂, was added. This mixture was refluxed for 1.5 h. The mixture was cooled to room temperature and the layers were separated. The organic phase was dried over MgSO₄ and carefully concentrated by rotary evaporation. The residue was distilled at 55°C under aspirator pressure into a flask (cooled to 0°C) containing a small amount of hydroquinone to yield 5.68 g (66%) of 10a a colorless liquid. ¹H NMR (CDCl₃, δ ppm, 300MHz): 6.53 (d, 1H, J = 1.5 Hz), 6.01 (d, 1H, J = 1.5 Hz), 3.85 (s, 3H).

Methyl 2-benzyloxycarbonyl-7-chloro-2-azabicyclo[2.2.2]oct-5-ene-7-carboxylate (11a and 12a) Dihydropyridine 2a (8.9 g, 41.3 mmol), 10a and hydroquinone (50 mg) were mixed in 1 ml of dry toluene. This mixture was stirred at 80-85°C under argon for 36-48 h. After cooling, the solvents and unreacted dienophile was removed by vacuum pump. The residue was purified by flash column chromatography (silica gel, 25% EtOAc in hexane) and gave 4.46 g (34%) of the *endo*-chloro adduct 12a and 3.34 g (25%) of the *exo*chloro adduct 11a. The *endo*-chloro adduct elutes before the *exo*-chloro adduct. 12a: ¹H NMR (CDCl3, δ ppm 300 MHz) 7.43 - 7.29 (m, 5H), 6.59 - 6.41 (m, 2H), 5.27 + 5.23 (d + d', 1H, J = 6.2 Hz), 5.21 - 5.01 (m, 2H), 3.70 + 3.52 (s + s', 3H), 3.27 + 3.20 (dd + dd', 1H, $J_I = 9.9$ Hz, $J_2 = 1.5$ Hz), 3.07 - 2.81 (ca, 3H), 1.84 (dd, 1H, $J_I = 14.4$ Hz, $J_2 = 2.1$ Hz). 11a: mp. 91°C, lit¹¹ value mp. = 85-86°C. ¹H NMR (CDCl3, δ ppm 300 MHz) 7.45 - 7.29 (m, 5H), 6.47 + 6.45 (t + t', 1H, J = 6.7 Hz), 6.40 - 6.27 (m, 1H), 5.30 (d, 1H, $J_I = 6.0$ Hz), 5.25 - 5.11 (m, 2H), 3.77 + 3.76 (s + s', 3H), 3.51 (dt, 1H, $J_I = 10.2$ Hz, $J_2 = 2.4$ Hz), 3.06 (dt, 1H, $J_1 = 10.5$ Hz, $J_2 = 2.5$ Hz), 2.93 - 2.83 (ca, 1H), 2.73 (dt, 1H, $J_1 = 14.4$ Hz, $J_2 = 3.0$ Hz), 1.98 (dt, 1H, $J_1 = 14.4$ Hz, $J_2 = 3.0$ Hz). MW calcd. for C₁₇H₁₈ClNO₄ (335.78) (both epimers), found MS (CI) m/z 336, 338 (3 : 1) (M + 1⁺).

Methyl 7-exo-chloro-2-azabicyclo[2.2.2]oct-5-ene-7-endo-carboxylate hydrogen iodide salt (15a) Trimethylsilyl iodide (20 mmol) was prepared *in situ* as described for 4a. Exo adduct 11a (6.12 g, 0.0183 mol) was dissolved in 50 ml of CH₂Cl₂ and cooled in an ice bath. The freshly prepared TMSI solution was added via syringe. The mixture was stirred at 0°C for 45 min. The reaction was quenched by careful addition of methanol (a total of 5 ml) and the solvents were carefully removed by rotary evaporation and then under vacuum. The residue was treated with acetone/ether to induce crystallization. The solvent mixture was decanted and the remaining solids were dried under vacuum to yield 2.8 g (46%) of 15a. ¹H NMR (CDCl₃, δ ppm 300 MHz) 10.09 (bs, 1H), 9.05 (bs, 1H), 6.71 (t, 1H, J = 7.0 Hz), 6.41 (t, 1H, J = 7.0 Hz), 4.79 (d, 1H, J = 6.0 Hz), 3.80 (s, 3H), 3.57 (d, 1H, J = 10.2 Hz), 3.11 - 3.07 (m, 2H), 2.72 (dt, 1H, $J_I = 14.9$ Hz, $J_2 = 3.2$ Hz), 2.19 (dd, 1H, $J_I = 14.1$ Hz, $J_2 = 1.8$ Hz). MW calcd. for C₉H₁₈ClINO₂ (329.5) found MS (CI) m/z 202, 204 (3 : 1) (M + 1⁺) for cation.

2-{1-[2-(Indol-2-yl)-1-oxoethyl]}-7-exo-chloro-2-azabicyclo[2.2.2]oct-5-ene-7-endo-carboxylic acid methyl ester (16a) Indole-2-acetic acid³³ (1.49 g, 8.5 mmol) and 15a (2.8 g, 8.5 mmol) were dissolved in 50 ml of dry CH₂Cl₂ and stirred at rt for 5 min. DCCI (3.51 g, 17.0 mmol) dissolved in an additional 50 ml of CH₂Cl₂ was added. The reaction mixture was allowed to stir overnight at room temperature. The solution was filtered to remove the urea by-product. The filtrate was washed with dilute HCl (10 ml), NaHCO₃ (10 ml), and water (10 ml). The organic phase was dried over MgSO₄ and concentrated. The crude material was subjected to flash column chromatography (silica gel, 1:1 EtOAc: hexane) to yield 1.77 g (58 %) of 16a as a gummy material. ¹H NMR (CDCl₃, δ ppm 300 MHz) 9.13 + 9.05 (bs + bs', 1H), 7.53 (t, 1H, *J* = 6.6 Hz), 7.32 (d, 1H, *J* = 8.1 Hz), 7.13 (dt, 1H, *J*₁ = 7.7 Hz, *J*₂ = 1.1 Hz), 7.06 (t, 1H, *J* = 7.4 Hz), 6.49 + 6.47 (t + t', 1H, *J* = 7.5 Hz), 6.35 + 6.29 (s + s', 1H), 6.31 + 6.23 (t + t', 1H, *J* = 6.3 Hz), 5.78 + 5.04 (d + d', 1H, *J* = 6.0 Hz), 4.06 (dd, 2H, *J*₁ = 35.0 Hz, *J*₂ = 16.0 Hz), 3.66 + 3.57 (dd + dd', 1H, *J*₁ = 9.3 Hz, *J*₂ = 2.2 Hz), 3.20 = 3.12 (dt + dt', 1H, *J*₁ = 9.0 Hz, *J*₂ = 2.5 Hz), 2.94 (ca, 1H), 2.79 - 2.70 (m, 1H), 1.98 (m, 1H). MW calcd. for C₁₉H₁₉ClN₂O₃ (358.81), found MS (CI) m/z 359, 361 (3 : 1) (M + 1⁺).

<u>Deethyllactam (17a)</u> A 500 ml photolysis cell was charged with 16a (0.500 g, 1.39 mmol) and NaHCO₃ (0.228 g, 2.71 mmol) dissolved in 350 ml of methanol and 150 ml of water. The reaction cell was continuously purged with nitrogen and irradiated with ultraviolet light from a 400W Hanovia mercury pressure lamp with a Vycor filter for 5 h. The cell was drained and as much of the methanol was removed by rotary evaporation as possible. The remaining aqueous solution was extracted with EtOAc (4 x 70 ml) and the extract dried over MgSO₄. The crude material was crystallized with a minimal amount of hot EtOAc to yield 0.105 g (23%) of 17a mp. >300°C (decomp). ¹H NMR (CDCl₃, δ ppm 300 MHz) 9.10 (s, 1H), 7.45 (d, 1H, J = 8.1 Hz), 7.30 (d, 1H, J = 8.1 Hz), 7.13 (t, 1H, J = 7.4 Hz), 7.04 (t, 1H, J = 7.2 Hz), 6.77 (t, 1H, J = 7.0 Hz), 6.38 (t, 1H, J = 7.0 Hz), 5.33 (d, 1H, J = 7.2 Hz), 4.53 (d, 1H, J = 15.6 Hz), 3.61 (~ 1H), 3.57 (s, 3H), 3.51 (d, 1H, J = 15.6 Hz), 3.06 - 2.98 (~ 3H), 1.69 (d, 1H, J = 11.7 Hz). ¹³C NMR (CDCl₃) 174.01, 173.00, 139.69, 135.73, 128.39, 127.59, 126.27, 122.46, 120.42, 119.59, 114.50, 111.50, 53.60, 53.01, 52.66, 50.72, 37.08, 32.86. MW calcd. for C₁₉H₁₈N₂O₃ (322.36), found MS (CI) m/z 323 (M +1⁺). Anal. Calcd.: C, 70.79; H, 5.63; N, 8.69. Found: C, 70.84; H, 5.69; N, 8.60.

Deethyl[3,2]-analog (18a) Lactam 17a (0.100 g, 0.31 mmol) was suspended in 15 ml dry THF (under argon). NaBH₄ (0.51 g, 13.5 mmol) was added in one portion. The reaction vessel was cooled to 0°C in an ice bath. BF₃-OEt₂ (2.4 ml, 19.5 mmol) was added dropwise by syringe. The ice bath was removed and the mixture was stirred at room temperature for 2-3 h. The solvents were removed *in vacuo*. The residue was partitioned between CHCl₃ and water. The organic layer was dried (Na₂SO₄) and concentrated. This residue was dissolved in 5 ml of methanol and 100 mg of dry Na₂CO₃ was added. This mixture was refluxed for 1 h under an argon atmosphere. The methanol was removed *in vacuo* and the residue was partitioned between CHCl₃ and water. The organic phase was dried (Na₂SO₄) and concentrated to give nearly pure 18a (89 mg, 93% yield). The material was subjected to flash column chromatography on silica gel (acetone) to give 52 mg (54% yield) of pure 18a. ¹H NMR (CDCl₃, δ ppm, 300MHz): 7.82 (bs, 1H), 7.30 (d, 1H, J = 8.1 Hz), 7.26 (d, 1H, J = 8.1 Hz), 6.54 - 6.43 (m, 2H), 4.42 (d, 1H, J = 5.4 Hz), 3.63 (s, 3H), 3.35 - 3.20 (m, 3H), 3.11 - 2.99 (m, 4H), 2.77 (d, 1H, J = 7.8 Hz), 1.68 (dd, 1H, $J_I = 12.8$ Hz, $J_2 = 2.0$ Hz). ¹³C NMR (CDCl₃) 175.59, 135.30, 134.63, 134.27, 133.92, 127.84, 121.85, 120.12, 119.05, 115.61, 110.88, 58.19, 52.91, 52.75, 51.78, 36.92, 31.47, 26.65. HRMS (EI) m/z calcd. for C₁₉H₂₀N₂O₂ (308.1525), found 308.1519 (M⁺).

Methyl 2-benzyloxycarbonyl-7-exo-chloro-6-ethyl-2-azabicyclo[2.2.2]oct-5-ene-7-endo-carboxylate (11b) Dihydropyridine 2b (7.05 g, 0.0287 mol), methyl 2-chloroacrylate (3.46 g, 0.0287 mol), and a small amount of hydroquinone (25 mg) were dissolved in 2 ml of dry toluene and stirred at 80°C for 24-36 h. The toluene and unreacted acrylate was removed by vacuum pump. The material was purified by flash chromatography on silica gel (1:4 EtOAc:hexane) to yield 3.67g (35%) of the *exo*-chloro adduct (11b) (approximately the same amount of the *endo*-chloro adduct 12b was formed). The *endo*-chloro adduct has a slightly higher R_f and elutes before the *exo*-chloro adduct 11b: ¹H NMR (CDCl₃, δ ppm 300 MHz) 7.40 - 7.30 (m, 5H), 6.05 - 5.98 (m, 1H), 5.18 (d, 1H, J = 1.4Hz), 3.77 +3 .76 (s + s', 3H), 3.46 + 3.43 (dd + dd', 1H, $J_I = 10.2$ Hz, J_2 =2.1 Hz), 3.03 + 2.99 (q+q', 1H, J = 2.4Hz), 2.86 - 2.78 (m, 1H), 2.77 + 2.72 (q + q', 1H, J = 2.4Hz), 2.11 (dq, 2H, $J_I = 7.5$ Hz, $J_2 = 1.5$ Hz), 1.98 + 1.93 (d + d', 1H, $J_I = 2.1$, $J_2 = 1.8$ Hz), 1.00 + 0.96 (t+t', 3H, J =7.2Hz). MW calcd. for C₁₉H₂₂ClNO4 (363.84), found MS (CI) m/z 364, 366 (3:1) (M +1⁺). Methyl 7-exo-chloro-6-ethyl-2-azabicyclo[2.2.2]oct-5-ene-7-endo-carboxylate hydrogen iodide salt (15b) Exo-Chloro adduct 11b (3.50 g, 9.62 mmol) was dissolved in 20 ml of CH₂Cl₂ and cooled with an ice bath. It was deprotected using TMSI as described for 15a to yield 1.48 g (47.6%) of 15b. ¹H NMR (d₆-DMSO, δ ppm, 300MHz) 9.45 (bs, 1H), 8.86 (bs, 1H), 6.27 (d, 1H, J = 6.0 Hz), 4.58 (s, 1H), 3.72 (s, 3H), 3.20 (m, 1H), 3.01 (m, 1H), 2.69 (m, 1H), 2.12 (q, 2H, J = 7.2 Hz), 0.93 (t, 3H, J = 7.2 Hz). MW calcd for C₁₁H₁₇ClINO₂(357.62), found MS (CI) m/z 230 (cation).

Methyl 2-{1-(indol-2-yl)-1-oxoethyl)}-7-exo-chloro-6-ethyl-2-azabicyclo[2.2.2]oct-5-ene-7-endo-carboxylate (16b) The hydrogen iodide salt 15b (1.26 g, 4.05 mmol) was converted to 16b as described for 16a to give 2.99 g crude 16b. This material was purified by flash chromatography on silica gel (EtOAc: hexane = 1:1) to yield 1.20 g (76.7%) of pure 16a as a gummy material. ¹H NMR (CDCl₃, δ ppm 300 MHz) 9.12 + 9.04 (bs + bs', 1H), 7.53 + 7.52 (d + d', 1H, J = 7.5Hz), 7.32 (d, 1H, J = 8.1Hz), 7.13 (t, 1H, J = 7.5Hz), 7.06 (t, 1H, J = 7.8Hz), 6.37 + 6.29 (s + s', 1H), 6.12 + 6.03 (dd + dd', 1H, $J_1 = 6.6$ Hz, $J_2 = 1.5$,), 5.64 + 4.86 (d + d', 1H, J = 1.5Hz), 4.08 (dd, 1H, J = 15.3, 35.7Hz), 3.81 + 3.77 (s + s', 3H), 3.62 + 3.59 (d + d', 1H, J = 2.1Hz), 3.54 + 3.50 (d + d', 1H, J = 2.1Hz), 3.10 + 3.06 (dt + dt', 1H, $J_1 = 10.8$ Hz, $J_2 = 2.7$ Hz,), 2.89 (~ 1H), 2.82 - 2.72 (m, 1H), 2.13 (q, 2H, J = 6.3Hz), 2.08 - 1.90 (m, 2H), 0.97 + 0.87 (t + t', 3H, J = 7.5Hz). MW calcd. for C₂₁H₂₃ClN₂O₃ (386.8), found MS (C1) m/z 387, 389 (3 : 1) (M + 1⁺).

[3,2]-Lactam 17b A 500 ml photolysis cell was charged with 16b (0.500 g, 1.29 mmol) and NaHCO₃ (0.228 g, 2.71 mmol) that had been dissolved in 350 ml of methanol and 150 ml of water. The reaction cell was purged with nitrogen while irradiated with ultraviolet light from a Hanovia mercury pressure lamp with a Vycor filter for 5 h. The product was isolated as for 17a to yield 0.105 g (23%) of 17b, mp. 297°C (decomp). ¹H NMR (CDCl₃, δ ppm 300 MHz) 8.61 (bs, 1H), 7.41 (d, 1H), 7.30 (d, 1H, J = 8.1 Hz), 7.14 (dt, 1H, $J_I = 7.5$ Hz, $J_2 = 1.1$ Hz), 7.04 (dt, 1H, $J_I = 7.5$ Hz, $J_2 = 1.1$ Hz), 6.31 (dd, 1H, $J_I = 7.2$ Hz, $J_2 = 1.2$ Hz), 5.08 (s, 1H), 4.58 (d, 1H, J = 15.6Hz), 3.55 (s, 3H), 3.53 (~ 1H), 3.49 (d, 1H, J = 15.6Hz), 3.07 - 2.98 (m, 1H), 2.95 (dd, 2H, $J_I = 11.4$ Hz, $J_2 = 2.1$ Hz), 2.20 (dq, 2H, $J_I = 7.2$ Hz, $J_2 = 1.8$ Hz), 1.61 (d, 1H, J = 12.9Hz), 1.10 (t, 3H, J = 7.5Hz). ¹³C NMR (CDCl₃) 173.82, 173.05, 142.83, 135.69, 129.81, 127.55, 126.21, 122.43, 120.39, 119.44, 114.96, 111.46, 58.08, 52.73, 52.49, 51.55, 37.15, 33.41, 32.61, 27.27, 11.81. MW calcd. for C₂₁H₂₂N₂O₃ (350.42), found MS (Cl) m/z 351 (M +1⁺). Anal. Calcd.: C, 71.98; H, 6.33; N, 7.99. Found: C, 72.02; H, 6.36; N, 7.97.

[3,2]-Analog 18b Lactam 17b (0.100 g, 0.28 mmol) was suspended in 15 ml of dry THF. Reduction was carried out as for 18a to give 18b (89 mg, 93% yield). Pure material was obtained by flash chromatography on silica gel (1:1, CHCl₃ acetone) to give 52 mg (54% yield) of 18b. ¹H NMR (CDCl₃, δ ppm 300 MHz) 7.82 (s, 1H), 7.26 (d, 1H, J = 7.3 Hz), 7.24 (d, 1H, J = 6.3 Hz), 7.11 (t, 1H, J = 7.6 Hz), 7.00 (t, 1H, J = 7.1 Hz), 5.96 (d, 1H, J = 6.4 Hz), 4.14 (s, 1H), 3.66 (m, 1H), 3.61 (s, 3H), 3.35 - 3.20 (m, 2H), 3.05 (dt, 2H, J_I

= 13.2 Hz, J_2 = 2.9 Hz), 2.91 (m, 1H), 2.79 (d, 2H, J = 8.3 Hz), 2.29 + 2.02 (m + m², 2H), 1.63 (d, 1H, J = 13.0 Hz), 1.07 (t, 3H, J = 7.3 Hz). ¹³C NMR (CDCl₃) 174.56, 147.87, 134.55, 133.20, 127.12, 123.25, 120.97, 119.20, 118.16, 115.44, 109.98, 62.57, 52.01, 51.67, 51.58, 51.07, 37.07, 30.67, 25.94, 10.23. HRMS (EI) m/z calcd. for C₂₁H₂₄N₂O₂ (336.1838), found 336.1844 (M⁺).

Methyl 2-bromopropenoate (10b) In a 500 ml flask, Na₂CO₃ (34.26 g, 0.324 mol) and tetrabutylammonium bromide (2.38 g, 7.37 mmol) were dissolved in 200 ml of water. A small amount of phenothiazine and methyl 2,3-dibromopropionate³⁴ (36.26 g, 0.147 mol) were dissolved in 100 ml of CH₂Cl₂ and added to the flask. This mixture was stirred at reflux for 30 min, cooled and the layers separated. The organic layer was dried over MgSO₄ and carefully concentrated by rotary evaporation. The pure ester was obtained by low pressure distillation (hydroquinone was added to the collection flask as a stabilizer). 10b was obtained (18.2 g, 75%) as a clear liquid. ¹H NMR (CDCl₃, δ ppm, 300MHz): 6.96 (d, 1H, *J* = 1.5 Hz), 6.27 (d, 1H, *J* = 1.5 Hz), 3.84 (s, 3H).

Methyl 2-benzyloxycarbonyl-7-bromo-2-azabicyclo[2.2.2]oct-5-ene-7-carboxylate (13a,14a) Methvl 2bromopropenoate (4.0 ml, 0.039 mol) and 2a (9.49 g, 0.044 mol) were mixed with a small amount of hydroquinone and heated at 50°C overnight. Reaction progress was slow as determined by TLC (EtOAc: hexane = 4:1). The temperature was raised to 75°C and the reaction was allowed to stir for an additional 24 h. The reaction vessel was cooled and unreacted materials were removed by vacuum pump. After flash column chromatography (silica gel, 1:4 EtOAc:hexane) 2.26 g (15.2%) of exo-bromo adduct 13a was obtained as an oil. 13a: ¹H NMR (CDCl₃, δ ppm, 300MHz): 7.42 - 7.28 (m, 5H), 6.45 (dd (both rotamers), 1H, $J_1 = 12.3$ Hz, $J_2 = 5.4$ Hz), 6.39 - 6.30 (m (both rotamers), 1H), 5.37 (d, 1H, J = 5.7 Hz) + 5.25 (d', 1H, J = 6.0 Hz), 5.22 - 5.15 (m, 2H), 3.76 (s, 3H) + 3.75 (s', 3H), 3.54 - 3.48 (m, 1H), 3.08 (t, 1H, J = 2.6Hz) + 3.04 (t', 1H, J = 2.7 Hz), 2.90 - 2.81 (ca, 1H), 2.80 (d, 1H, J = 2.7 Hz) + 2.75 (δ' , 1H, J = 2.4 Hz), 2.20 (t, 1H, J = 1.8 Hz) + 2.15 (t', 1H, J = 1.8Hz). Endo-Bromo adduct 14a: ¹H NMR (CDCl₃, d ppm, 300MHz): 7.44 - 7.28 (~ 5H), 6.55 - 6.45 (m, 2H), 5.38 (m, 1H) + 5.31 (d', 1H, J = 6.0 Hz), 5.24 (d, 1H, J = 12.0 Hz) + 5.07 (d', 1H, J = 12.0 Hz), 3.68 (s, 3H) + 3.53 (s', 3H), 3.24 (dd, 1H, $J_I = 10.5$ Hz, $J_2 = 2.1$ Hz), 3.10 - 2.93 (ca, 2H), 2.90 - 2.79 (ca, 1H), 2.03 (d, 1H, J = 15 Hz). MW calcd. for C₁₇H₁₈BrNO₄ (380.23) (both isomers), found MS (CI) m/z 380, 382 (1 :1) (M +1⁺).

Methyl 7-exo-bromo-2-azabicyclo[2.2.2]oct-5-ene-7-endo-carboxylate hydrogen bromide salt (15c) 30% wt HBr in acetic acid (10 ml) was added dropwise by syringe to 13a (2.26 g, 5.94 mmol). The mixture was stirred at rt (under argon) until all the starting material was consumed, as determined by TLC. The AcOH/HBr and some benzyl bromide were removed under vacuum. The residue was treated with acetone to induce crystallization. The crystals were collected to yield 1.09 g (56%) of 15c. ¹H NMR (d₆-DMSO, 8 ppm, 300MHz) 6.71 (t, 1H, J = 7.5 Hz), 6.29 (t, 1H, J = 7.5 Hz), 4.76 (d, 1H, J = 6.0 Hz), 3.71 (s, 3H), 3.27 (d, 1H, J = 12.3 Hz), 3.01 (ca, 1H), 2.71 (dt, 1H, $J_I = 11.1$ Hz, $J_2 = 2.4$ Hz), 2.56 (dt, 1H, $J_I = 15.0$

Hz, $J_2 = 3.1$ Hz), 2.25 (dd, 1H, $J_1 = 15.0$ Hz, $J_2 = 2.1$ Hz).

Methyl 2-{1-[2-(indol-2-y])-1-oxoethyl]}-7-exo-bromo-2-azabicyclo[2.2.2] oct-5-ene-7-endo-carboxylate (16c) Indole-2-acetic acid³³ (0.270 g, 1.54 mmol) and DCC (0.320 g, 1.55 mmol) were suspended in 10 ml of dry CH₂Cl₂ and stirred at rt for 5 min. A mixture of Hunig's Base (0.30 ml, 1.72 mmol) and bromide salt 15c (0.500 g, 1.53 mmol) in 5 ml of CH₂Cl₂ was added to the reaction vessel and the mixture was allowed to stir overnight at rt (under argon). The mixture was then filtered and the filtrate was washed with 10 ml of 3% HCl, 3% NaHCO₃, and brine. The organic phase was dried (Na₂SO₄) and concentrated. Flash column chromatography (silica gel, 50% EtOAc in hexane) afforded 0.220 g (35%) of 16c as a brownish foam. ¹H NMR (CDCl₃, δ ppm, 300MHz): 9.13 + 9.07 (bs + bs', 1H), 7.53 (dd, 1H, J_I = 6.6 Hz, J_2 = 6.3 Hz), 7.32 (d, 1H, J = 8.1 Hz), 7.14 (dt, 1H, J_I = 7.5 Hz, J_2 = 1.2 Hz), 7.06 (dt, 1H, J_I = 7.2 Hz, J_2 = 0.9 Hz), 6.48 (q, 1H, J = 15.0 Hz), 6.35 + 6.30 (d + d', 1H, J = 1.2 Hz), 6.33 + 6.24 (dt + dt', 1H, J_I = 6.0 Hz, J_2 = 1.5 Hz), 5.85 + 5.12 (d + d', 1H, 6.2Hz), 4.17 - 4.01 (dd, 1H, J_I = 28.5 Hz, J_2 = 16.2 Hz) + 3.86 (s', 1H), 3.83 + 3.77 (s + s', 3H), 3.67 + 3.56 (dd + dd', 1H, J_I = 9.9 Hz, J_2 = 2.1Hz). MW calcd. for C₁₉H₁₉BrN₂O₃ (403.27), found MS (CI) m/z 403, 405 (1 : 1) (M + 1⁺).

<u>N-Methylcatharanthine (10c)</u>. Catharanthine (100 mg, 0.3 mmol) was added to a suspension of NaH (7 mg, 0.3 mmol, 80%/wt suspension in mineral oil) in 3 ml of THF at 0°C. The solution was heated for 5 min at reflux to complete evolution of H₂ and then recooled to 0°C. Methyl iodide (19 µl, 0.3 mmol) was added. The solution was allowed to stir at 0°C for 30 min and at rt for 30 min. The solvent was removed by rotary evaporation and the residue was partitioned between CH₂Cl₂ and water. The organic layer was dried (Na₂SO₄) and concentrated. The product was separated from unreacted 10b by radial chromatography (1:1 EtOAc:hexane) to give 21 mg (20 %) of 10c as a film. ¹H NMR (CDCl₃, δ ppm, 300 MHz): 7.54 (d, 1H, *J* = 7.8 Hz), 7.26 - 7.10 (m, 3H), 6.05 (d, 1H, *J* = 5.9 Hz), 4.55 (s, 1H), 3.71 (m, 1H), 3.59 (s, 3H), 3.50 (s, 3H), 3.30 (ddd, 1H, *J_I* = 18 Hz, *J₂* = 13 Hz, *J₃* = 5 Hz), 3.13 - 3.05 (m, 2H), 3.01 (dt, 1H, *J_I* = 13.9 Hz, *J₂* = 2.0 Hz), 2.95 - 2.85 (m, 2H), 2.75 (m, 1H), 2.50 (d, 1H, *J* = 8.3 Hz), 2.35 + 1.97 (dq + dq, 2H, *J_I* = 16.6 Hz, *J₂* = 7.3 Hz), 1.72 (d, 1H, *J* = 12.1 Hz), 1.09 (t, 3H, *J* = 7.1 Hz).

Amine oxide, 22b. 18b (56 mg, 0.166 mmol) was dissolved in 5 ml of dry CH₂Cl₂ and cooled to 0°C. *m*-Chloroperoxybenzoic acid (MCPBA) (34 mg, 0.166 mmol) was added in one portion. The mixture was allowed to stir at 0°C for 45 min. At this point TLC (2:1 CHCl₃:acetone) showed that all starting material had been consumed. The CH₂Cl₂was removed by rotary evaporation and the residue dried in vacuum to yield 85 mg (94%) of a greenish film. ¹H NMR (CDCl₃, δ ppm, 300 MHz): 9.48 (s, 1H), 8.06 (s, 1H), 7.96 (d, 1H, J = 8.0 Hz), 7.56 (d, 1H, J = 8.0 Hz), 7.40 (t, 1H, J = 7.8 Hz), 7.30 (d, 1H, J = 8.1 Hz), 7.23 (d, 1H, J = 8.1 Hz), 7.10

(t, 1H, J = 7.5 Hz), 7.04 (t, 1H, J = 7.5 Hz), 6.27 (t, 1H, J = 5.9 Hz), 5.12 (dd, 1H, $J_1 = 14.0$ Hz, $J_2 = 6.9$ Hz), 4.17 (dd, 1H, $J_1 = 17.7$ Hz, $J_2 = 10.2$ Hz), 3.96 (dd, 1H, $J_1 = 12.4$ Hz, $J_2 = 10.2$ Hz), 3.92 - 3.76 (m, 3H), 3.45 (s, 3H), 3.30 - 3.11 (m, 3H), 2.50 + 2.26 (dq + dq, 2H, $J_1 = 7.5$ Hz, $J_2 = 2.1$ Hz), 1.56 (d, 1H, J = 13.4 Hz), 1.11 (t, 3H, J = 7.5 Hz). MW calcd. for C₂₁H₂₄N₂O₃ (352.41), found MS (CI) m/z 353 (M+1⁺).

<u>Amine oxide 22a</u>. 18a (60 mg, 0.195 mmol) was dissolved in 5 ml of dry CH₂Cl₂ and cooled to 0°C. MCPBA (39 mg, 0.195 mmol) was added in one portion. The mixture was allowed to stir at 0°C for 45 min. The CH₂Cl₂ was removed by rotary evaporation and the product dried in vacuum to yield 96 mg (96%) of a greenish film. ¹H NMR (CDCl₃, δ ppm, 300 MHz): 9.77 (s, 1H), 9.40 - 8.60 (bs, 1H), 8.04 (s, 1H), 7.93 (d, 1H, J = 7.5 Hz), 7.47 (d, 1H, J = 7.5 Hz), 7.33 (t, 1H, J = 8.1 Hz), 7.30 - 7.21 (m, 2H), 7.09 (t, 1H, J = 7.5 Hz), 7.03 (t, 1H, J = 7.5 Hz), 6.66 (t, 1H, J = 7.3 Hz), 6.43 (t, 1H, J = 6.7 Hz), 5.38 (d, 1H, J = 5.4 Hz), 4.92 (m, 1H), 3.96 - 3.71 (m, 4H), 3.51 (s, 3H), 3.26 - 3.05 (m, 3H), 1.53 (d, 1H, 14.0 Hz). MW calcd. for C₁₉H₂₀N₂O₃ (324.38), found MS (CI) m/z 325 (M+1⁺).

<u>Amine oxide 19b.</u> **8b** (18 mg, 0.053 mmol) was dissolved in 2 ml of dry CH₂Cl₂ and cooled to 0°C. MCPBA (11 mg, 0.053 mmol) was added in one portion. The mixture was allowed to stir at 0°C for 25 min. The CH₂Cl₂ was removed by rotary evaporation and the product dried in vacuum to yield 29 mg (100%) of a greenish film. ¹H NMR (CDCl₃, δ ppm, 300 MHz): 8.05 (s, 1H), 7.93 (d, 1H, J = 8.0 Hz), 7.52 (d, 1H, J = 8.0 Hz), 7.48 (m, 1H), 7.35 (t, 1H, J = 8.1 Hz), 7.28 - 7.10 (m, 3H), 6.21 (d, 1H, J = 5.9 Hz), 6.11 (s, 1H), 4.95 - 4.73 (m, 2H), 4.50- 4.33 (m, 1H), 4.18 (d, 1H, J = 12.4 Hz), 3.84 - 3.80 (ca, 4H), 3.18 - 3.02 (m, 2H), 2.50 + 2.28 (dq + dq, 2H, $J_I = 7.5$ Hz, $J_2 = 2.1$ Hz), 1.71 (d, 1H, J = 14.0 Hz), 1.13 (t, 1H, J = 7.5 Hz). MW calcd. for C₂₁H₂₄N₂O₃ (352.41), found MS (CI) m/z 353 (M+1⁺).

<u>Amine oxide 19a</u>. **8a** (33 mg, 0.107 mmol) was dissolved in 5 ml of dry CH₂Cl₂. and cooled to 0°C. MCPBA (22 mg, 0.107 mmol) was added in one portion. The mixture was allowed to stir at 0°C for 30 min. The CH₂Cl₂ was removed by rotary evaporation and the product dried in vacuum to yield 55 mg (100%) of a greenish film. ¹H NMR (CDCl₃, δ ppm, 300 MHz): 8.05 (s, 1H), 7.94 (d, 1H, J = 7.0 Hz), 7.55 - 7.47 (m, 2H), 7.35 (t, 1H, J = 8.0 Hz), 7.28 - 7.09 (m, 3H), 6.70 - 6.35 (m, 2H), 6.13 (s, 1H), 5.05 (ca, 1H), 4.89 - 4.82 (m, 2H), 4.50 - 4.39 (m, 2H), 4.22 (d, 1H, J = 12.4 Hz), 3.90 (d, 1H, J = 12.4 Hz), 3.84 (s, 3H), 3.16 (m, 1H), 3.02 (d, 1H, J = 14.5 Hz), 1.77 (d, 1H, J = 14.0 Hz). MW calcd. for C₁₉H₂₀N₂O₃ (324.38), found MS (CI) m/z 325 (M+1⁺).

General Procedure for Ferric Chloride Reactions. In a 50 ml round-bottom three-neck flask 10 ml of 0.10 M glycine solution (7.5 g glycine and 5.85 g sodium chloride in 1000 ml water) was purged with argon for 10 min at room temperature. FeCl₃·6H₂O (6 - 30 eq) was added to the flask as a solid.³⁵ Approximately 15 mg of the appropriate amine was dissolved in 1 ml of methanol and transferred to the flask by pipet. When vindoline was used, 1.0 - 1.2 equivalents was added. The mixture was allowed to stir at rt for 3 h under a constant purge of

argon. Then 1 - 2 eq. of NaBH₄ was added in 1-2 ml of concentrated NH₄OH solution and the mixture was allowed to stir for 5 min (NOTE: in some reactions the NaBH₄ was deleted to provide a non-reductive workup). The pH was tested with pH paper to be certain that the solution was slightly basic (around pH 8-9). This basic solution was extracted with CH₂Cl₂ (4 x 15 ml), dried (Na₂SO₄) and concentrated under reduced pressure. At this point the ¹H NMR was examined to determine the crude product composition. The products and any unreacted starting material were separated by column chromatography. The isolation procedures, yield and spectroscopic information are listed for individual compounds.

General Procedure for Potier Reactions. Approximately 15 mg of amine-N-oxide as its MCBA salt was dissolved in 1 ml of dry CH₂Cl₂ and transferred by syringe to a 5 ml side-neck flask (under argon) cooled to 0°C. If vindoline was included, 1.0-1.2 eq was dissolved in 0.5 ml of CH₂Cl₂ and added by syringe. The flask was then cooled to -60°C and TFAA (3-5 eq, distilled from P₂O₅ immediately before use) was rapidly added by syringe. The solution was allowed to stir under argon for 3 h at -55°C. The reaction could then be worked up two ways: a) reductive or b) nonreductive. a) <u>Reductive Workup</u>: The CH₂Cl₂ solution was quickly transferred to a cold (-40°C) solution of ethanol and excess NaBH₄ or NaBD₄ and allowed to stir and warm to room temperature over 5 min. The excess BH₄ was then quenched with 1-2 ml of acetone. The solvents were removed by rotary evaporation. The residue was partitioned between CH₂Cl₂ and aqueous Na₂CO₃. The organic layer was dried (Na₂SO₄) and concentrated. b) <u>Nonreductive Workup</u>: The reaction was quenched with 0.5 ml of methanol and the solvents were removed by rotary evaporation and dried under vacuum. This residue was then partitioned between CH₂Cl₂ and aqueous Na₂CO₃. The organic layer was dried (Na₂SO₄) and concentrated. The ¹H NMR was examined to determine the crude product composition. The products were separated by column and radial chromatography using 1:5 methanol:EtOAc.

<u>20-Deethylanhydrovinblastine dimers 25a and 26a</u>. Glycine solution (10 ml) was purged with argon for 10 min at rt. FeCl₃-6H₂O (130 mg, 0.480 mmol) was added to the flask as a solid. Amine 10a (31 mg, 0.100 mmol) was dissolved in 1 ml of methanol and transferred to the round-bottom flask by pipet. Vindoline (23 mg, 0.50 mmol) was then added as a solid. The mixture was allowed to stir at room temperature for 3 h under argon. NaBH₄ (1-2 eq.) was added in 1-2 ml of concentrated ammonium hydroxide solution and the mixture was allowed to stir for 5 min. The pH was checked with pH paper and was 8-9. The basic solution was extracted with CH₂Cl₂ (4 x 15 ml), dried (Na₂SO₄) and concentrated under reduced pressure to give 48 mg (89%) of crude material. At this point the ¹H NMR was examined to determine the amount of coupling. The two "dimers" were present in a 1:1 ratio. All of the vindoline had been consumed and some 10a was still present. The "dimers" were separated by radial chromatography (silica gel, 1:6 MeOH:EtOAc) 25a: 12 mg, 25% yield; 26a:14 mg, 30% yield) and the unreacted 10a was recovered by preparative layer chromatography (silica gel, 1:6 MeOH:EtOAc). The recovered 10a (2 mg) was treated with Eu(hfc)₃ (4.2 mg, 0.4 equivalents). The ¹H NMR in CDCl₃ showed a 1:1.1 ratio of the indole N-H signals. Dimer 25a (key assignments): ¹H NMR (CDCl₃, δ ppm, 300MHz): 8.03 (bs, 1H), 7.52 (d, 1H, J = 7.8 Hz), 7.20 - 7.10 (m, 3H), 6.61 (s, 1H), 6.12 (s, 1H), 5.86 (dd, 1H, $J_1 = 10.4$ Hz, $J_2 = 3.8$ Hz), 5.74 (bs, 2H), 5.46 (s, 1H), 5.29 (d, 1H, J = 10.3 Hz), 3.82 (s, 3H), 3.80 (s, 3H), 3.62 (s, 3H), 2.72 (s, 3H), 2.11 (s, 3H), 0.79 (t, 3H, J = 7.3 Hz). Dimer 26a (key assignments): ¹H NMR (CDCl₃, δ ppm, 300MHz): 8.08 (s, 1H), 7.52 (d, 1H, J = 7.3 Hz), 7.28 - 7.10 (m, 3H), 6.74 (s, 1H), 6.16 (s, 1H), 5.78 (m, 3H), 5.46 (s, 1H), 5.24 (d, 1H, J = 10.0 Hz), 3.81 (s, 3H), 3.80 (s, 3H), 3.56 (s, 3H), 2.73 (s, 3H), 2.07 (s, 3H), 0.37 (t, 3H, J = 7.3 Hz).

<u>Methyl</u> 7H-2-(E-3-hydroxymethyl-1-butenyl)-1,2,5,6-tetrahydroindolizidino[7,8-b]indole-11c-carboxylate 24b. The general ferric chloride (25 eq) reductive procedure was used on 15 mg of 18b. After CH₂Cl₂ extraction from the basified aqueous solution from reductive workup, the crude material was chromatographed on a basic alumina column (CH₂Cl₂ \rightarrow 30% EtOAc in CH₂Cl₂) to yield 4 mg of recovered 18b and 4 mg (25%) of 24b. MW calcd. for C₂₁H₂₆N₂O₃ (354.45), found MS (CI) m/z 355 (M+1⁺).

Methyl 7H-2-(E-3-formyl-1-butenyl)-1,2,5,6-tetrahydroindolizidino[7,8-b]indole 11c-carboxylate (E-23b). FeCl₃·6H₂O (325 mg, 1.20 mmol) and 18b (14 mg, 0.0417 mmol) were allowed to react in glycine solution (10 ml) for 3 h under argon. The solution was made basic and processed in the standard way. The crude material was chromatographed on a silica gel column (1:4 MeOH:EtOAc) to give 3 mg of 18b and 4 mg of E-23b (27% yield) as a film. MW calcd. for $C_{21}H_{24}N_2O_3$ (354.45), found MS (C1) m/z 355 (M+1⁺).

Methyl7H-2-(Z-3-formyl-1-butenyl-1,2,5,6-tetrahydroindolizidino[7,8-b]indole-11c-carboxylate(Z-23b).Amine oxide 22b (18 mg, 0.0354 mmol) was dissolved in 1 ml of dry CH2Cl2 and transferred by syringe to a 5

ml side-neck flask (under argon) cooled to 0°C. Et₃N (6 μ l, 0.043 mmol) was added by syringe. The flask was then cooled to -60°C and TFAA (3 - 5 eq), was rapidly added by syringe. The solution was allowed to stir under argon for 3 h at -55°C. The reaction was quenched with 1 ml of methanol and the mixture was allowed to stir until the flask reached rt. The crude product (15 mg) was obtained by the usual workup and chromatographed on a silica gel column (1:5 MeOH:EtOAc) to give 1.6 mg (14% yield) of Z-23b as film. MW calcd. for C₂₁H₂₄N₂O₃ (352.44), found MS (CI) m/z 353 (M+1⁺).

<u>Methyl 7H-2-(E-3-oxo-1-propenyl)-1,2,5,6-tetrahydroindolizidino[7,8-b]indole-11c-carboxylate (E-23a)</u>. Ferric chloride hexahydrate (512 mg, 1.89 mmol) and 18a (24 mg, 0.0779 mmol) in 10 ml of glycine solution were allowed to stir at room temperature for 3 h under argon. The solution was made basic (around pH 8-9) and subjected to the standard workup. The crude material was chromatographed on a silica gel column (1:4 MeOH:EtOAc) to give aldehyde E-23a as a film (5 mg, 26% yield). MW calcd. for C₁₉H₂₀N₂O₃ (324.45), found MS (CI) m/z 325 (M+1⁺).

<u>Dimers (20a, 21a)</u>. Amine oxide 19a (15 mg, 0.0312 mmol) was dissolved in 1 ml of dry CH₂Cl₂ and transferred by syringe to a 5 ml side-neck flask (under argon) cooled to 0°C. Vindoline (19 mg, 0.0415 mmol) was dissolved in 0.5 ml of CH₂Cl₂ and added to the flask. The mixture was then cooled to -60°C and TFAA (25 μ l,

5.7 eq) was added. The solution was allowed to stir under argon for 3 h at -55°C. The reaction solution was quickly transferred to a cold (-40°C) solution of ethanol and excess NaBH4 and allowed to stir and warm to rt over 5 min. The excess borohydride was then quenched with 1 - 2 ml of acetone. The solvents were removed by rotary evaporation. The residue was partitioned between CH₂Cl₂ and aqueous Na₂CO₃. The CH₂Cl₂ layer was dried (Na₂SO₄) and concentrated to give 22 mg of crude material. The NMR of the crude material indicated approximately 10% of each "coupled" material based on comparison with the vindoline signals. This reaction was repeated twice with similar results. The combined materials were chromatographed by radial chromatography (1:5 MeOH: EtOAc) to give vindoline (24 mg) and the two "dimers" (3 mg, 4.3 %). Dimer 20a (kev assignments): ¹H NMR (d₆-acetone, δ ppm, 300MHz): 7.28 (d, 1H, J = 8.3 Hz), 6.99 (dt, 1H, J₁ = 7.6 Hz, $J_2 = 1.5$ Hz), 6.78 (d, 1H, J = 7.8 Hz), 6.71 (t, 1H, J = 8.3 Hz), 6.56 (s, 1H), 6.20 (s, 1H), 6.11 (m, 2H), 5.71 (dd, 1H, $J_1 = 10.3$ Hz, $J_2 = 3.9$ Hz), 5.43 (s, 1H), 5.29 (d, 1H, J = 10.6 Hz), 4.41 (ddd, 2H, $J_1 = 15.1$ Hz, $J_2 = 10.3$ Hz, $J_3 = 2.1$ Hz), 4.36 (ddd, 2H, $J_1 = 15.1$ Hz, $J_2 = 4.9$ Hz, $J_3 = 2.0$ Hz), 4.13 (d, 1H, J = 4.8 Hz), 3.60 (s, 3H), 3.56 (s, 3H), 3.51 (a, 1), 3.44 (m), 3.33 (dd, 1H J = 15.5, $J_2 = 5$, 3.1-3.3 (m), 3.01 (s, 3H), 2.60 (s, 3H), 3.61 (s, 3H), 3 3H), 2.65 (J = 7.8) 2.0-2.2 (m), 1.86 (s, 3H), 1.80 (d, J = 14.5 Hz), 1.58 (dg, J = 7.5, 1), 0.69 (t, 3H, J = 7.3Hz). MW calcd. for C44H51N4O8 (763.87), found MS (EI) m/z 763 (M^+); Dimer 21a (key assignments): ¹H NMR (d₆-acetone, 8 ppm, 300 Mhz): 7.30 (d, J = 8 Hz), 7.00 (t, J = 7 Hz), 6.94 (d, J = 7 Hz); 6.78, 6.76 (t and s); 6.15 (s); 5.90 dd (J = 10, 4 Hz); 5.38 (d, J = 11 Hz) 5.31 (s); 4.4-4.6 (m), 4.09 (m); 3.60 s; 3.56 s; 3.2-3.4 (m) 2.97 (s) 2.6-2.8 (m); 2.62 (s) 2.20 (m): 1.53, 1.65 (AB quartets) 0.69 (t, J = 7 Hz). MW calcd for $C_{44}H_{51}N_4O_8$ (763.87), found MS (EI m/z 763 M⁺).

<u>Dimer (20b)</u>. Amine oxide 19b (15 mg, 0.0295 mmol) was dissolved in 1 ml of dry CH₂Cl₂ and transferred by syringe to a 5 ml side-neck flask (under argon) cooled to 0°C. Vindoline (15 mg, 0.0330 mmol) was dissolved in 0.5 ml of CH₂Cl₂ and added by syringe. The flask was then cooled to -60°C and TFAA (19 µl, 4 eq.) was rapidly added. The solution was allowed to stir under argon for 3 h at -55°C. The reaction solution was treated as described for 20a/21a to give 21 mg of crude material. This reaction was repeated once more with similar results. The combined materials were purified by radial chromatography (silica, 1:5 MeOH:EtOAc) to give unreacted vindoline (17 mg) and 20b (1 mg, 2.1 %). Key assignments, ¹H NMR (d₆-acetone, δ ppm, 300MHz): 7.30 (d, 1H, J = 7.8 Hz), 7.00 (t, 1H, J = 7.8 Hz), 6.78 (t, 1H, J = 7.8 Hz), 6.71 (d, 1H, J = 7.8 Hz), 6.58 (s, 1H), 6.21 (s, 1H), 5.72 (m, 3H), 5.42 (s, 1H), 5.30 (dd, 1H, $J_I = 10.2$ Hz, $J_2 = 1.0$ Hz), 4.41 (m, 2H), 3.61 (s, 3H), 3.56 (s, 3H), 3.03 (s, 3H), 2.60 (s, 3H), 1.87 (s, 3H), 0.85 (t, 3H, J = 7.3 Hz), 0.69 (t, 3H, J = 7.3 Hz). MW calcd. for C4₆H₅₅N₄O₈ (791.94), found MS (EI) m/z 791 (M⁺).

<u>Electrochemistry</u>. The voltammetric measurements were performed in a 15 ml conical cell with a glassy carbon electrode (GCE; $A = 0.05 \text{ cm}^2$), with platinum as a counter and saturated calomel electrode (SCE) as a reference. Before each measurement GCE was sonicated for 1 min in acetone and polished on a filter paper. All electrochemical measurements were carried out using PAR M-175 electrochemical analysis system and the

voltammograms were recorded on Kipp & Zonen X-Y recorder. The cyclic voltammograms were recorded in the range of 0.015 to 1 V/s. The value of transfer coefficient was obtained from the peak widths for each voltammogram using the following equation.³⁶

$$E_{p}-E_{p}/2 = 1.857 (RT/\alpha nF)$$

For each voltammogram the peak current and transfer coefficient were measured and assuming the value of the diffusion constant for DECATH (D = $1.4 \times 10-5$ cm²/s) the apparent number of electrons n was calculated at different sweep rates using the following equation.³⁷

$$i_p = (2.99 \times 10^5) n (\alpha n)^{1/2} ACD^{1/2} v^{1/2}$$

Controlled potential coulometry of DECATH was performed in divided cell filled with CH₃CN-0.1 M LiClO₄ using Pt-gauze anode (3×5 cm), graphite cathode and SCE. Into 35 ml of analyte DECATH (22 mg, 0.071 mmol) and 2,6-lutidine (38 mg, 0.35 mmol) were added. The potential was increased gradually from 0.5 to 0.6 V vs. SCE. The cyclic voltammograms (GCE, 50 mV/s) were recorded during the electrolysis and the amount of charge was monitored by built-in digital coulometer.

Acknowledgment

We thank Dr. Dean Harman and Mr. Stanley Kolis for preliminary cyclic voltametric measurements and Mr. Kim Harich, VPI&SU, for the HRMS and high molecular weight mass spectra.

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