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PAPER

Triazolophostins: A Library of Novel and Potent Agonists of IP₃ Receptors[†]

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IP₃ receptors are channels that mediate release of Ca^{2+} from the intracellular stores of cells stimulated by hormones or neurotransmitters. Adenophostin A (AdA) is the most potent agonist of IP₃ receptors, with the β -anomeric adenine contributing to the increased potency. The potency of AdA and its stability towards the enzymes that degrade IP₃ have aroused interest in AdA analogs for biological studies. The

- $_{10}$ complex structure of AdA poses problems that have necessitated optimization of synthetic conditions for each analog. Such lengthy one-at-a-time syntheses limit access to AdA analogs. We have addressed this problem by synthesizing a library of triazole-based AdA analogs, triazolophostins, by employing click chemistry. An advanced intermediate having all the necessary phosphates and a β -azide at the anomeric position was reacted with various alkynes under Cu(I) catalysis to yield triazoles, which upon
- ¹⁵ deprotection gave triazolophostins. All eleven triazolophostins synthesized are more potent than IP₃ and some are equipotent with AdA in functional analyses of IP₃ receptors. We show that a triazole ring can replace adenine without compromising the potency of AdA and provide facile routes to novel AdA analogs.

Introduction

- ²⁰ Many biological processes are regulated by changes in the intracellular concentration of Ca^{2+,1} A major pathway for these Ca²⁺ signals is the release of Ca²⁺ from intracellular stores sequestered within the endoplasmic reticulum (ER) via IP₃ receptors (IP₃R)² IP₃R are a family of intracellular Ca²⁺ channels
- ²⁵ that are expressed largely in ER membranes.³ IP₃ (Fig. 1A, 1) is produced when cell-surface receptors stimulate phospholipase C activity. IP₃ then binds to IP₃R causing its channel to open and release Ca²⁺ into the cytosol.⁴ The ability of IP₃ to interact with IP₃R is terminated by its dephosphorylation or phosphorylation
- ³⁰ by specific 5-phosphatase and 3-kinase enzymes.⁵ While terminating the Ca²⁺-mobilizing ability of IP₃, these steps also initiate recycling of inositol to the lipid from which IP₃ is generated and the synthesis of more complex bioactive phosphoinositols including the pyrophosphates,⁶ many of which
- ³⁵ have been recent synthetic targets.⁷ Adenophostin A (AdA), a fungal metabolite isolated from the culture broth of *Penicillium brevicompactum*, is a more potent agonist than IP₃ of IP₃R.⁸ Moreover, AdA is resistant to the enzymes that metabolize IP₃. Many analogs of AdA have been synthesized to address
- ⁴⁰ structure-activity relationships and the determinants of the increased potency of AdA.⁹ These studies revealed that the three phosphates and 2"-OH (color coded in Fig. 1A, **2**), which structurally mimic the essential pharmacophore of IP₃,¹⁰ are







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Adenophostin A (AdA, 2)

Fig. 1 (A) Molecular structures of IP₃ (1) and AdA (2); (B) Conventional synthesis of AdA-analogs. The question marks represent the complexity of conventional syntheses.

- Recent studies suggested that a cation- π interaction between ⁵ adenine and a cationic residue (Arg504) of the IP₃R might be responsible for the increased potency of AdA.^{9b,12} Additional base-modified analogs of AdA would be useful in both testing this proposal and as potent and metabolically stable agonists of IP₃R to probe Ca²⁺ signaling mechanisms.
- The major hurdle in developing such AdA analogs lies in 10 the synthesis of these structurally complex base-modified analogs (disaccharide nucleotides).^{9e,13} The traditional synthesis involves N-glycosylation of a nucleobase with an orthogonally protected disaccharide derivative followed by chemoselective demasking 15 and phophorylation of specific hydroxyl groups in the nucleoside (Fig. 1B). This early introduction of the nucleobase necessitates an almost individualy tailored synthesis (branching at an early stage of synthesis) for each base-modified analog. Further complications arise from the lack of a generally applicable 20 method for glycosylation and phosphorylation of such complex molecules. For instance glycosylation of nucleobases is challenging with respect to yield¹⁴ and regioselectivity,^{13b,15} and each glycosylation condition has to be optimized individually. Chemoselective O-phosphorylation in the presence of 25 nucleobases having nucleophilic groups/sites is also challenging.¹⁶ Thus syntheses of analogs by a one-at-a-time method is cumbersome, time-consuming and impractical. We herein report a general strategy for making a combinatorial library of structurally diverse AdA analogs that are potent 30 agonists of IP₃R.



Fig. 2 (A) Imidophostin and Triazolophostins. (B) Retrosynthetic analysis of triazolophostins.

Results and discussion

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Imidophostin (Fig. 2A), though weaker than AdA, is more potent than IP₃, suggesting that even a 5-membered aromatic heterocyclic ring can be a partial functional mimic of adenine. As triazole is isosteric with imidazole, we envisioned that triazolebased AdA analogs would also be potent agonists of IP₃R. This 40 and the earlier reports on use of click reaction in biologically important molecules¹⁷ prompted us to adopt azide-alkyne click chemistry to make a library of triazolophostins, AdA analogs wherein the nucleobase is replaced by substituted triazoles. These molecules can easily be prepared by reacting appropriately ⁴⁵ protected tris-phosphate derivative **II** having an azide at anomeric position with aryl or heteroaryl alkynes under Cu(I) catalysis (Fig. 2B). Such an approach would solve three major issues associated with conventional AdA analog synthesis: (i) Ensure the necessary β -anomeric stereochemistry of the base-surrogate 50 (ii) As the triazole-based base-surrogate is introduced only at the penultimate step (after phosphorylation), the interference of nucleobases during phosphorylation can be avoided (iii) Diversification at the penultimate step provides access to many diverse analogs from a common advanced structurally



Scheme 1 Synthesis of triazolophostins. a) H_2SO_4 -silica, Ac_2O , DCM, rt, 12 h, 90%; b) TMSN₃, AlCl₃, DCM, 0 °C, 10 min, 99%; c) NaOMe, MeOH, rt, 30 min, 96%; d) i. (iPr)₂NP(OBn)₂, ImOTf, DCM, rt, 1 h, ii. m-CPBA, -78 °C, 45 ⁶⁰ min, 90%; e) i. Trimethylsilylacetylene, Cu, CuSO₄, H₂O:¹BuOH, rt, 12 h; ii. TBAF, THF, rt, 2 h, (95% for two steps); f) Alkyne, Cu, CuSO₄, H₂O:¹BuOH, rt, 12 h, 48-97%; g) Cyclohexene, Pd(OH)₂, MeOH:H₂O, 80 °C, 4 h, Bn = benzyl, Ac = acetyl, Me = methyl.

We have synthesized orthogonally protected disaccharide **4** in ⁶⁵ thirteen steps from D-glucose and D-xylose in a convergent manner as reported previously.¹⁸ Acetolysis¹⁹ of the ketal **4** gave tetraacetate **5** as anomeric mixtures in very good yield. Lewis acid catalysed azidation of the acetate **5** gave anomerically pure

 β -azide 6.²⁰ Methanolysis of the acetate groups in 6 provided the triol 7 in excellent yield. Phosphitylation of the triol 7 using phosphoramidite followed by in situ oxidation using m-CPBA provided the fully protected trisphosphate 8. Compound 8 with 5 three phosphates in their appropriate relative position and β oriented azide is the common advanced intermediate for coupling with different alkynes. Copper (I) catalysed click reaction of azide 8 with various alkynes provided fully protected triazolophostins 9a-k. As the reaction is catalyzed by copper(I), 10 1,4-substituted triazoles²¹ were obtained exclusively in all the cases. Global removal of benzyl protecting groups by transfer hydrogenolysis provided triazolophostins 10a-k in very good yields. Though in a recent elegant communication, Potter et al reported a functionally active triazole based cADPR analog,²² 15 ours is the first report on the use of click chemistry to generate a library of IP₃R agonists.

Table 1. Ca2+ release via IP₃R evoked by triazolophostins

но—					
² O ₃ PO -2O ₃ PO	HO O OPO3-2				
Ligand	R =	pEC ₅₀	EC ₅₀	EC_{50}	max.
			(nM)	w.r.t.	response
				1	(%)
IP ₃ (1)		6.75 ± 0.08	178 ±	1	73 ± 3
			30		
AdA (2)		8.05 ± 0.04	9 ± 1	20	73 ± 2
10a	н	7.87 ± 0.00	13 ± 0	13	71 ± 1
10b	N CH ₃	7.75 ± 0.03	18 ± 2	10	72 ± 2
10c		7.81 ± 0.05	15 ± 1	11	65 ± 3
10d		7.77 ± 0.03	17 ± 1	10	64 ± 1
10e	\square	7.70 ± 0.07	20 ± 3	9	75 ± 4
10f	F	8.00 ± 0.03	10 ± 1	18	69 ± 1
10g	F	7.86 ± 0.04	14 ± 2	13	71 ± 2
10h	F	7.64 ± 0.01	23 ± 1	8	67 ± 2
10i	OMe	7.45 ± 0.03	35 ± 2	5	65 ± 1
10j		7.78 ± 0.03	17 ± 2	11	68 ± 2
10k	CHO	7.89 ± 0.02	13 ± 1	14	70 ± 1

Results show the maximal Ca^{2+} release (% of Ca^{2+} content of intracellular ²⁰ stores), EC₅₀ and pEC₅₀ values (means ± SEM from at least 3 experiments). Hill coefficients were not significantly different from 1.

The abilities of the newly synthesized ligands to evoke Ca^{2+} release via IP_3R were assessed using permeabilized cells that

express only type 1 IP₃R.²³ A low-affinity Ca²⁺ indicator trapped ²⁵ within the ER allowed the effects of IP₃ to be directly measured using a fluorescence plate-reader equipped to allow automated additions. All the triazolophostins fully release the IP₃-sensitive Ca²⁺ stores and all are more potent than IP₃ (Table 1, Fig. 3). It is interesting to note that the parent analog triazolophostin **10a** ³⁰ having only the five-membered ring is 13-fold more potent than IP₃. This remarkable AdA-like potency of **10a** suggests that a single triazole ring can effectively replace the adenine base in AdA without compromising potency. In contrast, imidophostin was previously reported to be only slightly (1.3-fold) more potent ³⁵ than IP₃, and it was therefore suggested that a fused bicyclic nucleobase/surrogate is essential for AdA-like potency.^{18a} Among all triazolophostins, the 3-fluoro derivative **10f** is the most potent, and equipotent to AdA: it is the most potent AdA analog without

⁴⁰ literature, very few match the potency of AdA.^{9c,12b,13b,14,18a,24,25} In this context, our late-stage diversification method is important because it allows many potent analogs to be made in an easy and combinatorial way from the same advanced intermediate.

a purine nucleobase. Of the several AdA analogs reported in the



Fig. 3 Concentration-dependent release of Ca²⁺ (%) by IP₃, AdA and triazolophostins from the intracellular stores of cells expressing IP₃R1. Results show means \pm SEM from at least 3 experiments.

The docking study suggests that triazolophostin 10a and AdA 50 exhibit similar interactions with the IP₃-binding core of IP₃R1 (IBC, residues 224-604).²⁴ The triazolyl ring of 10a forms a cation- π interaction with Arg504 similar to the proposed interaction between the adenine of AdA and Arg504. This supports previous reports that a cation- π interaction between the 55 base and IP₃R contributes to the increased potency of AdA and analogs.^{9c,24} All other triazolophostin analogs showed a similar interaction. The small differences in activity among these analogs seem to arise from steric, rather than electronic factors. For instance, attaching aromatic rings with different electrostatic 60 surface potentials to the triazole ring did not cause any systematic change in biological activity. The fact that all the analogs are both more potent than IP₃, and show a conserved cation- π interaction between the triazole ring and Arg504 suggests that this critical interaction is unaffected by aromatic substitution, irrespective of 65 the electronic nature of the aromatic substituent. The single bond connection between the triazole ring and the substituent (aromatic ring) allow torsional freedom (freely rotatable) and hence the sterically favorable non-coplanar conformation of the two aromatic rings. As a result of such non-coplanarity (lack of 70 conjugation), the electron density on the triazole ring cannot be influenced by the aromatic unit. This could be the reason why

there is no clear trend in activity when the aromatic ring is modified with electron-deficient or electron-rich substituents.

- One reason for the increased potency of AdA is believed to be its conformational flexibility allowing it to adopt an optimal ⁵ conformation for binding to IP₃R.²⁶ The additional conformational freedom due to the single bond linkage of this hetero-biaryl unit (rather than fused rings in AdA) allows the second ring to orient to an electronically favorable position, such that it can have additional electrostatic interactions. Docking ¹⁰ results suggest that the most potent analog, **10f**, has additional
- interactions between F and IP_3R side-chains (D566, Q565 and Q507) that may enhance its activity (Fig. 4).



Is Fig. 4 Docking of 3-fluorophenyl triazolophostin with IBC showing preferred binding mode and the important cation-π and fluorine-oxygen interactions with residues Arg504 and Asp566, respectively. The cation-π distance r = 2,97 Å. The fluorophenyl moiety orients perpendicular to the triazole ring. Color code for ligand: carbon, gray; oxygen, red; nitrogen, blue; 20 phosphorous, yellow; fluorine, cyan.

Conclusions

- In conclusion, addressing the problems associated with conventional one-at-a-time synthesis of AdA analogs, we have synthesized a library of new analogs using azide-alkyne click ²⁵ chemistry. The simple reaction conditions, high yield and atomeconomy of the click reaction to install base-surrogates are
- advantageous over the conventional Vorbruggen method of installation of nucleobase. By this easy and rapid method, we generated a variety of ligands with diverse structures from a 30 common advanced intermediate. All the analogs synthesized by
- this method are more potent than IP_3 and some (10a, 10f, 10g, 10k) are as potent as AdA. Though replacement of purine with pyrimidines or other aromatic groups are reported to give agonists that are more potent than IP_3 , none of the previously
- ³⁵ reported non-purinated AdA analogs was as potent as AdA. We have shown that AdA-like potency can be achieved without a nucleobase. Our study reveals that a five-membered aromatic ring, triazole, is sufficient as a base surrogate to exhibit AdA-like potency. Easy access to potent, non-metabolizable ligands of
- ⁴⁰ IP₃R and the ease of conjugation with reporter groups such as fluorescent probes, photoaffinity probes, etc offers avenues for advanced biological exploration. This report that triazoles can be functional mimics of nucleobases has broader implication and might trigger such attempts in other nucleobase-derived signaling

45 molecules.

Experimental section

1. General Methods

Chromatograms were visualized under UV light and by dipping plates into either phosphomolybdic acid in MeOH or ⁵⁰ anisaldehyde in ethanol, followed by heating. ¹H NMR, COSY, NOESY and HMOC spectra were recorded on a 500 MHz NMR spectrometer. Proton chemical shifts are reported in ppm (\delta) relative to the internal standard tetramethylsilane (TMS, δ 0.0 ppm) or with the solvent reference relative to TMS employed as ⁵⁵ the internal standard (CDCl₃, δ 7.26 ppm; D₂O, δ 4.79 ppm). Data are reported as follows: chemical shift (multiplicity [singlet (s), doublet (d), doublet of doublet (dd), triplet (t), quartet (q), and multiplet (m)], coupling constants [Hz], integration and peak identification). All NMR signals were assigned on the basis of ¹H NMR, ¹³C NMR, COSY and HMQC experiments. ¹³C spectra were recorded with complete proton decoupling. Carbon chemical shifts are reported in ppm (δ) relative to TMS with the respective solvent resonance as the internal standard. All NMR data were collected at 25 °C. The concentration of the compounds 65 for ¹H NMR was 5 mg per 0.5 mL and for ¹³C NMR it was 20 mg per 0.5 mL for protected compounds and 5-7 mg per 0.5 mL for final compounds in case of ¹H and ¹³C NMR. Each triazolophostin (10a-k) was quantified using Briggs's/Ame's phosphate assay. Melting point was determined using melting 70 point apparatus and is uncorrected. Flash column chromatography was performed using silica gel 230-400 mesh. Where ever needed, the reactions were carried out under argon or nitrogen atmosphere employing oven dried glassware.

X-ray intensity data measurements of freshly grown crystals 75 of **4** was carried out at 293-296 K on a Bruker-KAPPA APEX II CCD diffractometer with graphite-monochromatized (MoK = 0.71073Å) radiation. The X-ray generator was operated at 50 kV and 30 mA. Data were collected with scan width of 0.3° at different settings of φ (0°, 90° and 180°) keeping the sample to 80 detector distance fixed at 40 mm and the detector position (2θ) fixed at 24°. The X-ray data collection was monitored by SMART program. All the data were corrected for Lorentzian, polarization and absorption effects using SAINT and SADABS programs. SHELX-97 was used for structure solution and full 85 matrix least-squares refinement on *F*2. Molecular and packing diagrams were generated using Mercury-3.1. Geometrical calculations were performed using SHELXTL and PLATON.

2. Synthesis of 1,2,3',4'-tetra-*O*-acetyl-2',5,6'-tri-*O*-benzyl-3-*O*-α-D-glucopyranosyl-D-ribofuranose (5)

⁹⁰ To a solution of disaccharide 4 (1.0 g, 1.41 mmol) and Ac₂O (0.4 mL, 4.24 mmol) in dry DCM (20 mL), 0.05 g of freshly prepared H₂SO₄-silica¹⁹ was added. The reaction mixture was stirred at room temperature for 12 h under nitrogen atmosphere and the reaction was monitored using TLC. When the starting material ⁹⁵ disappeared, the reaction mixture was quenched by the addition of solid NaHCO₃. Solid material was filtered off and the organic layer was washed with a saturated aq. NaHCO₃ solution. The organic layer was dried over Na₂SO₄ and evaporated under reduced pressure. The crude product thus obtained was purified ¹⁰⁰ by flash column chromatography using 40% ethyl acetate in petroleum ether as eluent to get the known^{18a} tetraacetate 5 (0.96 g, 90%) as a sticky mass.

3. Synthesis of 2,3',4'-tri-*O*-acetyl-2',5,6'-tri-*O*-benzyl-3-*O*-α-D-glucopyranosyl-β-D-ribofuranosyl azide (6)

To a solution of tetraacetate 5 (0.51 g, 0.68 mmol) and TMSN₃ (0.44 mL, 3.4 mmol) in anhydrous dichloromethane (10 mL) was 5 added AlCl₃ (0.09 g, 0.68 mmol) at 0°C. The reaction mixture was stirred at 0°C under nitrogen atmosphere for 10 min. When the starting material disappeared (TLC), the reaction mixture was quenched by the addition of aqueous NaHCO₃. The reaction mixture was partitioned between dichloromethane and aqueous 10 layers in a separating funnel. The dichloromethane layer was washed with brine, dried over Na2SO4 and evaporated under reduced pressure. The crude product thus obtained was purified by flash column chromatography using 25% ethyl acetate in petroleum ether as eluent to get triacetate azide 6 (0.495 g, 99%)¹⁵ as a colourless gum. ¹H NMR (500 MHz, CDCl₃) δ : 1.79 (s, 6H, 2 x COCH₃), 1.86 (s, 3H, COCH₃), 3.22 (dd, J = 10.0, 4.0 Hz, 1H, H-6A'), 3.26 (dd, J = 10.0, 2.0 Hz, 1H, H-6B'), 3.46 (dd, J = 10.0, 3.4 Hz, 1H, H-2'), 3.52 (dd, J = 11.0, 4.0 Hz, 1H, H-5A), 3.61 (dd, J = 11.0, 3.0 Hz, 1H, H-5B), 3.77 (d, J = 9.8, Hz, 1H, H-²⁰ 5'), 4.22 (d, J = 12.0, Hz, 1H, 0.5 x PhCH₂), 4.29-4.30 (m, 1H, H-4), 4.38-4.43 (m, 3H, 1.5 x PhCH2), 4.46-4.54 (m, 3H, H-3 and 1 x PhCH₂), 4.88 (d, J = 3.3 Hz, 1H, H-1'), 4.95 (t, J = 9.8 Hz, 1H, H-4'), 5.02 (d, J = 2.6 Hz, 1H, H-2), 5.27-5.29 (m, 2H, H-1 and H-3'), 7.16-7.22 (m, 15H, 3 x Ph); ¹³C NMR (125 MHz, CDCl₃) 25 S: 20.5, 20.7, 20.9 (COCH₃), 67.6 (C-6'), 69.0 (C-4' and C-5'), 69.5 (C-5), 71.9 (C-3'), 73.3, 73.5, 73.5 (3 x CH₂Ph), 73.9 (C-2), 74.5 (C-3), 76.6 (C-2'), 81.4 (C-4), 92.77 (C-1), 96.7 (C-1'), 127.5, 127.7, 127.7, 127.8, 128.0, 128.4, 128.4, 128.5 (Ar-C), 137.5, 137.7, 137.9 (ipso Ar-C), 169.8, 170.5, 170.5 (3 x ³⁰ COCH₃); Anal. calcd for C₃₈H₄₃N₃O₁₂: C, 62.20; H, 5.91; N, 5.73. found: C, 62.12; H, 6.09; N, 5.71.

4. Synthesis of 2',5,6'-tri-*O*-benzyl-3-*O*-α-D-glucopyranosyl-β-D-ribofuranosyl azide (7)

- To the suspension of triacetate **6** (0.42 g, 0.57 mmol) in MeOH $_{35}$ (10 mL) was added sodium methoxide (0.015 g, 0.28 mmol) and stirred at room temperature. Reaction mixture became clear after 30 min. When TLC showed complete disappearance of starting material, the reaction was quenched by adding Dowex 50-ion exchange resin. The reaction mixture was then filtered through a $_{40}$ filter paper and washed successively with methanol (2 times).
- The filtrate was concentrated under reduced pressure to get a thick mass. The thick mass was then purified by flash column chromatography using 50% ethyl acetate in petroleum ether as eluent to get triol azide 7 (0.34 g, 96%) as a white solid. mp: 69-
- ⁴⁵ 71 °C; ¹H NMR (500 MHz, CDCl₃) δ : 3.33 (dd, J = 9.5, 3.5 Hz, 1H, H-2'), 3.43–3.54 (m, 5H, H-4', H-5_A, H-5_B, H-6_A', H-6_B'), 3.66 (ddd, J = 9.8, 8.7, 3.9, 1H, H-5'), 3.82-3.85 (m, 2H, H-3' and H-2), 4.16-4.18 (m, 2H, 3-H and H-4), 4.38-4.48 (m, 4H, 2 x CH_2 Ph), 4.61-4.70 (m, 3H, H-1' and 1 x CH_2 Ph), 5.20 (d, J = 1.4
- ⁵⁰ Hz, 1H, H-1), 7.18–7.29 (m, 15H, 3 x Ar-*H*); ¹³C NMR (125 MHz, CDCl₃) δ: 69.2 (C-5 or C-6'), 70.2 (C-5 or C-6'), 70.6 (C-5'), 71.1 (C-4'), 73.3 (C-3'), 73.5 and 73.6 (2 x CH₂Ph), 74.0 (C-2), 74.3 (1 x CH₂Ph), 78.3 (C-3 or C-4), 78.4 (C-2'), 81.4 (C-3 or C-4), 95.1 (C-1), 98.0 (C-1'), 127.6, 127.7, 127.8, 127.9, 128.4,
- $_{55}$ 128.5, 128.6, 128.8 (Ar-C), 137.1, 137.6, 137.9 (Ipso Ar-C); Anal. calcd for $C_{32}H_{37}N_3O_9$: C, 63.25; H, 6.14; N, 6.92; found: C, 63.23; H, 6.16; N, 6.90.

5. Synthesis of 2',5,6'-tri-O-benzyl-2,3',4'-tris(dibenzyloxyphosphoryl)-3-O- α -D-glucopyranosyl- β -D-

60 ribofuranosyl azide (8)

A solution of the triol 7 (0.1 g, 0.16 mmol), bis(benzyloxy) N,Ndiisopropylaminophosphine (0.27 mL, 0.82 mmol), and imidazolium triflate (0.20 g, 0.90 mmol) in anhydrous DCM (10 mL) was stirred at room temperature under N₂ atmosphere for 30 65 min. The reaction mixture was then cooled to -78 °C, and then m-CPBA (0.093 g, 0.54 mmol) was added. The reaction mixture was then stirred for 45 min at -78 °C and then at room temperature for an additional 30 min. The reaction was quenched by the addition of aqueous NaHCO₃. The reaction mixture was 70 partitioned between DCM and aqueous layer in a separating funnel. The DCM layer was washed with brine, dried over Na₂SO₄ and evaporated under reduced pressure. The crude product thus obtained was purified by flash column chromatography using 25% acetone in petroleum ether as eluent $_{75}$ to get pure **8** (0.206 g, 90%) as a colourless gum: ¹H NMR (500 MHz, CDCl₃) δ: 3.43-3.58 (m, 5H, H-6_A', H-6_B', H-5_B, H-5_A, H-2'), 3.70-3.72 (m, 1H, H-5'), 4.17-4.22 (m, 2H, H-4, and 0.5 x PhCH₂), 4.27-4.34 (m, 3H, H-3 and 1 x PhCH₂), 4.37-4.42 (m, 2H, 1 x PhCH₂), 4.47 (dd, 19.05, 9.75 Hz, 1H, H-4'), 4.55-4.59 80 (m, 2H, H-2, 0.5 x PhCH2), 4.65 (dd, 11.95, 8.5 Hz, 1H, 0.5 x PhCH₂), 4.78-4.94 (m, 12H, H-3' and 5.5 x PhCH₂), 5.01 (d, 3.55

- Hz, 1H, H-1'), 5.30 (d, 2.6 Hz, 1H, H-1), 6.96-7.25 (m, 45H, 9 x Ph), 13 C NMR (125 MHz, CDCl₃) δ : 68.1 (C-6'), 69.1 (C-5), 69.3, 69.3, 69.4, 69.5, 69.8, 69.83, 69.9, (31 P coupled, PhCH₂OP), 70.0 ss (C-5'), 72.0 (1 x PhCH₂OC), 73.2 (C-3), 73.3, 73.5 (2 x
- PhCH₂OC), 74.4 (³¹P coupled, C-4'), 76.9 (C-2'), 77.6 (C-3', ³¹P coupled), 78.1 (C-2, ³¹P coupled), 81.3 (C-4), 92.4 (C-1), 95.2 (C-1'), 127.6, 127.65, 127.70, 127.8, 128.06, 128.08, 128.26, 128.28, 128.31, 128.34, 128.39, 128.44, 128.49, 128.52, 128.62,
- ⁹⁰ 128.64, 128.68, 128.69 (aromatic carbons), 135.40, 135.45, 135.70, 135.77, 135.85, 135.90, 136.14, 136.16, 136.20, 136.23, 137.6, 137.9, 138.1, (³¹P coupled, ipso carbons of POCH₂Ph); ³¹P NMR (202.4 MHz, CDCl₃) δ : -1.20, -1.90, -2.15; Anal. calcd for C₇₄H₇₆N₃O₁₈P₃: C, 64.02; H, 5.52; N, 3.03. Found: C, 64.33; H, ⁹⁵ 5.45; N, 3.19.

6. Synthesis of fully protected triazolophostins: a) Synthesis of 1-{5-*O*-benzyl-2-*O*-bisbenzyloxyphosphoryl-3-*O*-[2,6-di-*O*-benzyl-3,4-di-*O*-(bisbenzyloxyphosphoryl)-*α*-D-glucopyranosyl]-*β*-D-ribofuranosyl]-1,2,3-triazole (9a)

¹⁰⁰ A mixture of tris-phosphate 8 (0.1 g, 0.07 mmol), trimethylsilylacetylene (0.011 mL, 0.071 mmol), Cu (0.018 g, 0.28 mmol) and CuSO₄ (0.004 g, 0.014 mmol) in H₂O/⁴BuOH (1/1, v/v, 1 mL) was stirred at room temperature for 12 h. After complete consumption of the starting material (TLC, 12 h), TBAF (1.0 M in THF, 0.5 mL) was added to the reaction and the mixture was stirred for additional 2 h at room temperature. The reaction mixture was then filtered and washed with ethyl acetate. The filtrate was extracted with ethyl acetate (2 x 10 mL) and washed with water and brine. The organic layer was dried over 110 MgSO₄ and concentrated under reduced pressure. The crude product thus obtained was purified by flash column chromatography using a mixture of acetone and petroleum ether (3:7 v/v) as eluent to get pure 9a (0.098 g, 97%) as a colourless gum. ¹H NMR (500 MHz, CDCl₃) δ: 3.43-3.45 (m, 1H, H-5_A'),

3.47-3.57 (m, 3H, H-2", H-5_B', H-6_A"), 3.59-3.61 (m, 1H, H-6_B") 3.75 (ddd, J = 9.2, 4.65, 4.2 Hz, 1H, H-5"), 4.21-4.23 (m, 1H, 0.5 x PhCH₂), 4.29-4.32 (m, 4H, H-4', 1.5 x PhCH₂), 4.39-4.46 (m, 3H, H-3', H-4" and 0.5 x PhCH₂), 4.58-4.66 (m, 3H, 1.5 x ⁵ PhCH₂), 4.68-4.72 (m, 3H, 1.5 x PhCH₂), 4.80-4.96 (m, 8H, H-3", 3.5 x PhC H_2), 5.13 (d, J = 3.0 Hz, 1H, H-1"), 5.27 (ddd, J =8.75, 8.2, 4.6 Hz, 1H, H-2'), 6.31 (d, 4.9 Hz, 1H, H-1'), 6.99-7.27 (m, 45H, 9 x C_6H_5), 7.48 (s, 1H, H-5), 7.61 (s, 1H, H-4); ¹³C NMR (125 MHz, CDCl₃) δ: 68.5 (C-6"), 69.1 (C-5'), 69.2, 69.36, ¹⁰ 69.40, 69.5, 69.6, 69.8, 69.83, 69.9, 70.0, 70.02, (³¹P coupled, PhCH2OP), 70.3 (C-5"), 71.9, 73.4, 73.6 (3 x PhCH2OC), 73.9 (C-3'), 74.5 (³¹P coupled, C-4"), 76.9 (C-2'), 78.0 (C-3", ³¹P coupled), 78.6 (C-2', ³¹P coupled), 82.81 (C-4'), 90.0 (C-1'), 95.6 (C-1"), 122.4 (C-5), 127.6, 127.7, 127.74, 127.77, 127.8, 127.9, 15 128.0, 128.07, 128.1, 128.2, 128.3, 128.34, 128.4, 128.5, 128.52, 128.55, 128.59, 128.7, (aromatic carbons), 135.2, 135.28, 135.7, 135.74, 135.8, 135.87, 136.1, 136.2, 137.4, 137.6, 138.0, (³¹P coupled, ipso carbons of POCH₂Ph); ³¹P NMR (202.4 MHz, CDCl₃) δ: -1.427, -1.908, -2.128; Anal. calcd for C₇₆H₇₈N₃O₁₈P₃: 20 C, 64.54; H, 5.56; N, 2.97. Found: C, 64.31; H, 5.42; N, 3.01.

b) Synthesis of 1-{5-O-benzyl-2-O-bisbenzyloxyphosphoryl-3-O-[2,6-di-O-benzyl-3,4-di-O-(bisbenzyloxyphosphoryl)- α -D-glucopyranosyl]- β -D-ribofuranosyl}-4-(1-methylimidazol-5-yl)-1,2,3-triazole (9b)

- ²⁵ A mixture of tris-phosphate 8 (0.2 g, 0.14 mmol), 5-ethynyl-1-methyl-1*H*-imidazole (0.016 mL, 0.15 mmol), Cu (0.036 g, 0.57 mmol) and CuSO₄ (0.008 g, 0.028 mmol) in H₂O/BuOH (1/1, v/v, 2 mL) was stirred at room temperature for 12 h. After completion of the reaction, the mixture was filtered and washed ³⁰ successively with ethyl acetate. The filtrate was extracted with ethyl acetate and washed with water and brine. The organic layer was dried over MgSO₄ and concentrated under reduced pressure. The resulting crude product was purified by flash column chromatography using a mixture of acetone, diethyl ether and ³⁵ petroleum ether (4:1:15 v/v/v) as eluent to get pure **9b** (0.205 g, 95%) as a colourless gum. ¹H NMR (500 MHz, CDCl₃) δ: 3.55-3.76 (m, 8H, 1 x N-CH₃, 6_A", H-6_B", H-5_B', H-2" and H-5_A'), 3.88
- (bs, 1H, H-5"), 4.31-4.45 (m, 5H, H-4' and 2 x PhC*H*₂), 4.51-4.54 (m, 3H, H-4" and 1 x PhC*H*₂), 4.70-4.82 (m, 6H, H-3', and 2.5 x ⁴⁰ PhC*H*₂), 4.94-5.03 (m, 8H, H-3" and 3.5 x PhC*H*₂), 5.26 (bs, 1H,
- H-1"), 5.41 (bs, 1H, H-2'), 6.42 (s, 1H, H-1'), 6.97-7.34 (m, 47H, Ar-*H*), 7.75 (s, 1H, H-5); ¹³C NMR (125 MHz, CDCl₃) δ : 33.4 (N-CH₃), 68.6 (C-6"), 69.2 (C-5'), 69.4, 69.42, 69.5, 69.59, 69.8, 69.9, 70.03, (³¹P coupled PhCH₂OP), 70.3 (C-5"), 71.92, 73.4,
- ⁴⁵ 73.7 (3 x PhCH₂OC), 74.3 (C-3'), 74.5 (C-4", ³¹P coupled), 77.0 (C-2', ³¹P coupled), 78.0 (C-2"), 78.8 (C-3", ³¹P coupled), 83.1 (C-4'), 90.4 (C-1', ³¹P coupled), 95.77 (C-1"), 119.41 (C-5), 127.64, 127.67, 127.71, 127.80, 127.8, 127.96, 128.0, 128.1, 128.14, 128.3, 128.5, 128.52, 128.6, 128.67, (aromatic carbons),
- $_{50}$ 135.1, 135.2, 135.6, 135.76, 135.80, 135.9, 136.1, 136.18, 137.2, 137.6, ($^{31}\mathrm{P}$ coupled, ipso carbons of POCH_2*Ph*), 138.0, 138.8 (ipso carbons of COCH_2*Ph*); $^{31}\mathrm{P}$ NMR (202.4 MHz, CDCl_3) δ : 1.453, -1.894, -2.080; Anal. calcd for C_{80}H_{82}N_5O_{18}P_3: C, 64.29; H, 5.53; N, 4.69. Found: C, 63.99; H, 5.23; N, 4.81.

55 c) Synthesis of 1-{5-O-benzyl-2-O-bisbenzyloxyphosphoryl-3-O-[2,6-di-O-benzyl-3,4-di-O-(bisbenzyloxyphosphoryl)-α-D-

glucopyranosyl]-β-D-ribofuranosyl}-4-phenyl-1,2,3-triazole (9c)

- A mixture of tris-phosphate **8** (0.1 g, 0.07 mmol), phenylacetylene (0.016 mL, 0.158 mmol), Cu (0.018 g, 0.28 mmol) and CuSO₄ (0.004 g, 0.014 mmol) in H₂O/⁴BuOH (1/1, v/v, 1 mL) was stirred at rt for 12 h. After completion of the reaction (TLC) the mixture was filtered. The filtrate was extracted with ethyl acetate and washed successively with water ⁶⁵ and brine. The organic layer was dried over MgSO₄ and concentrated under reduced pressure. The crude product thus obtained was purified by flash column chromatography using a
- mixture of acetone and petroleum ether (3:7 v/v) as eluent to get pure **9c** (0.101 g, 94%) as a colourless gum. ¹H NMR (500 MHz, 70 CDCl₃) δ : 3.46-3.52 (m, 2H, H-2" and H-5_A'), 3.55-3.58 (m, 2H,
- H-6_B" and H-5_B'), 3.64-3.66 (m, 1H, H-6_A"), 3.77-3.80 (m, 1H, H-5"), 4.21-4.35 (m, 5H, H-4' and 2 x PhC H_2), 4.41-4.46 (m, 3H, H-3', H-4" and 0.5 x PhC H_2), 4.58-4.64 (m, 3H, 1.5 x PhC H_2), 4.68-4.72 (m, 3H, 1.5 x PhC H_2), 4.81-4.94 (m, 8H, H-3", and 3.5 x PhC H_2), 5.16 (d, J = 3.5 Hz, 1H, H-1"), 5.33 (ddd, J = 9.4, 8.55, 5.0, Hz, 1H, H-2'), 6.33 (d, 5.4 Hz, 1H, H-1'), 6.99-7.47 (m, 50H, 10 x C₆ H_5), 7.83 (s, 1H, H-5); ¹³C NMR (125 MHz, CDCl₃)
- δ: 68.7 (C-6"), 69.2 (C-5'), 69.2, 69.4, 69.45, 69.5, 69.5, 69.6, 69.85, 69.88, 70.0, 70.02, 70.1 (31 P coupled, Ph*C*H₂OP), 70.4 (C-5"), 72.0, 72.5, 72.7, (32 P, CH, OC), 74.5, (32 P, OC), 74.5,
- ⁸⁰ 5"), 72.0, 73.5, 73.7 (3 x PhCH₂OC), 74.5 (C-3'), 74.6 (³¹P coupled, C-4"), 77.0 (³¹P coupled, C-2"), 78.1 (C-3", ³¹P coupled), 78.8 (C-2'), 83.0 (C-4'), 90.1 (C-1'), 95.9 (C-1"), 118.2 (C-5), 125.8 (ArC), 127.6, 127.7, 127.8, 127.86, 128.0, 128.1, 128.2, 128.3, 128.4, 128.45, 128.50, 128.53, 128.6, 128.7
 ⁸⁵ (aromatic carbons), 130.3, 135.2, 135.23, 135.3, 135.7, 135.8,
- ⁸⁵ (aromatic carbons), 130.3, 135.2, 135.2, 135.3, 135.7, 135.8, 135.87, 135.93, 136.2, 136.25, 137.4, 137.6, 138.1, 148.1; (³¹P coupled, ipso carbons of POCH₂Ph); ³¹P NMR (202.4 MHz, CDCl₃) δ : -1.551, -1.932, -2.105; Anal. calcd for C₈₂H₈₂N₃O₁₈P₃: C, 66.08; H, 5.55; N, 2.82. Found: C, 65.95; H, 5.72; N, 2.97.

⁹⁰ d) Synthesis of 1-{5-*O*-benzyl-2-*O*-bisbenzyloxyphosphoryl-3-*O*-[2,6-di-*O*-benzyl-3,4-di-*O*-(bisbenzyloxyphosphoryl)- α -Dglucopyranosyl]- β -D-ribofuranosyl}-4-(2-pyridyl)-1,2,3triazole (9d)

A mixture of tris-phosphate 8 (0.2 g, 0.14 mmol), 2-95 ethynylpyridine (0.016 mL, 0.15 mmol), Cu (0.036 g, 0.57 mmol) and CuSO₄ (0.008 g, 0.028 mmol) in H₂O/^tBuOH (1/1, v/v, 2 mL) was stirred at rt for 12 h. After completion of the reaction (TLC) the mixture was filtered. The filtrate was extracted with ethyl acetate and washed successively with water and brine. The 100 organic layer was dried over MgSO4 and concentrated under reduced pressure. The crude product thus obtained was purified by flash column chromatography using a mixture of acetone and petroleum ether (3:7 v/v) as eluent to get pure 9d (0.104 g, 48%) as a colourless gum. ¹H NMR (500 MHz, CDCl₃) δ: 3.43-3.45 (m, 105 1H, H-5_A'), 3.48-3.59 (m, 4H, H-2", H-5_B', H-6_B"and H-6_A"), 3.64-3.71-3.76 (m, 1H, H-5"), 4.20-4.23 (m, 1H, 0.5 x PhCH₂), 4.27-4.37 (m, 4H, H-4' and 1.5 x PhCH₂), 4.41-4.45 (m, 3H, H-3', H-4" and 0.5 x PhCH₂), 4.60-4.62 (m, 1H, 0.5 x PhCH₂), 4.66-4.72 (m, 5H, 2.5 x PhCH₂), 4.80-4.93 (m, 8H, H-3", and 3.5 x

¹¹⁰ PhC H_2), 5.15 (d, J = 2.9 Hz, 1H, H-1"), 5.34-5.35 (m, 1H, H-2'), 6.30 (d, J = 5.0 Hz, 1H, H-1'), 6.99-7.19 (m, 45H, 9 x C6H5), 7.28 (d, J = 7.0 Hz, 1H, pyr-H), 7.72 (t, J = 7.0 Hz, 1H, pyr-H), 8.03 (d, J = 7.0 Hz, 1H, pyr-H), 8.44 (s, 1H, H-5), 8.52 (d, J = 4.0

Hz, 1H, pyr-H); ¹³C NMR (125 MHz, CDCl₃) δ: 68.4 (C-6'), 69.1 (C-5'), 69.13, 69.18, 69.4, 69.42, 69.5, 69.55, 69.8, 69.85, 69.87, 69.9, 70.0, 70.04 (³¹P coupled, PhCH₂OP), 70.3 (C-5''), 72.0, 73.38, 73.6 (3 x PhCH₂OC), 74.2 (C-3'), 74.5 (³¹P coupled, ⁵ C-4''), 77.1 (³¹P coupled, C-2''), 78.1 (C-3'', ³¹P coupled), 78.7 (C-2'), 83.1 (C-4'), 90.4 (C-1'), 95.7 (C-1''), 120.6 (C-5), 123.0 (ArC), 127.6, 127.7, 127.8, 127.9, 128.0, 128.1, 128.2, 128.3, 128.4, 128.43, 128.49, 128.5, 128.55 (aromatic carbons), 135.2, 135.25, 135.28, 135.30, 136.19, 136.25, 137.37, 137.60, 138.11; ¹⁰ (³¹P coupled, ipso carbons of POCH₂Ph); ³¹P NMR (202.4 MHz,

CDCl₃) δ : -1.60, -1.95, -2.13; Anal. calcd for C₈₁H₈₁N₄O₁₈P₃: C, 65.23; H, 5.47; N, 3.76. Found: C, 65.01; H, 5.21; N, 3.92.

e) Synthesis of 1-{5-O-benzyl-2-O-bisbenzyloxyphosphoryl-3-O-[2,6-di-O-benzyl-3,4-di-O-(bisbenzyloxyphosphoryl)-α-D Is glucopyranosyl]-β-D-ribofuranosyl}-4-(2-fluorophenyl)-1,2,3-

triazole (9e) A mixture of tris-phosphate 8 (0.2 g, 0.14 mmol), 2-fluorophenyl acetylene (0.021 mL, 0.15 mmol), Cu (0.036 g, 0.57 mmol) and CuSO₄ (0.008 g, 0.028 mmol) in H₂O/^tBuOH (1/1, v/v, 2 mL) 20 was stirred at rt for 12 h. After completion of the reaction (TLC) the mixture was filtered. The filtrate was extracted with ethyl acetate and washed successively with water and brine. The organic layer was dried over MgSO4 and concentrated under reduced pressure. The crude product thus obtained was purified 25 by flash column chromatography using a mixture of acetone and petroleum ether (1:3 v/v) as eluent to get 9e (0.203 g, 93%) as a colourless gum. ¹H NMR (500 MHz, CDCl₃) δ: 3.53-3.55 (m, 1H, H-5_A'), 3.61-3.73 (m, 4H, H-2", H-5_B', H-6_A" and H-6_B"), 3.88 (bs, 1H, H-5"), 4.33-4.35 (m, 1H, 0.5 x PhCH₂), 4.42-4.48 (m, 30 4H, H-4' and 1.5 x PhCH₂), 4.53-4.58 (m, 3H, H-3', H-4" and 0.5 x PhCH₂), 4.72-4.81 (m, 6H, 3 x PhCH₂), 4.96-5.06 (m, 8H, H-3", 3.5 x PhCH₂), 5.27 (bs, 1H, H-1"), 5.46 (bs, 1H, H-2'), 6.42 (bs, 1H, H-1'), 7.13-7.40 (m, 48H, Ar-H), 8.21 (s, 1H, H-5), 8.30 (bs, 1H, Ar-H); ¹³C NMR (125 MHz, CDCl₃) δ: 68.4 (C-6"), 69.0 35 (C-5'), 69.1, 69.2, 69.4, 69.42, 69.5, 69.6, 69.8, 69.85, 69.89, 70.0, 70.05 (³¹P coupled, PhCH₂OP), 70.2 (C-5"), 71.9, 73.4, 73.6 (3 x PhCH₂OC), 74.2 (C-3'), 74.4 (³¹P coupled, C-4"), 77.1 (³¹P coupled, C-2"), 78.0 (C-3", ³¹P coupled), 78.6 (C-2'), 83.0 (C-4'), 90.2 (C-1'), 95.7 (C-1"), 115.7 (C-5), 121.6, 121.7, 124.5, 40 124.6, 127.6, 127.9, 128.0, 128.1, 128.5, 128.6 (aromatic carbons), 129.4, 129.5, 135.2, 135.22, 135.3, 135.7, 135.8, 135.84, 135.89, 136.2, 136.22, 137.3, 137.6, 138.1, 141.48, 141.5, 158.3, 160.3 (³¹P coupled, ipso carbons of POCH₂Ph); ³¹P NMR (202.4 MHz, CDCl₃) δ: -1.632, -1.930, -2.134; ¹⁹F NMR 45 (470.68 MHz, CDCl₃) δ: -114.22; Anal. calcd for C₈₂H₈₁FN₃O₁₈P₃: C, 65.29; H, 5.41; N, 2.79. Found: C, 65.43; H, 5.59; N, 2.67.

f) Synthesis of 1-{5-*O*-benzyl-2-*O*-bisbenzyloxyphosphoryl-3-*O*-[2,6-di-*O*-benzyl-3,4-di-*O*-(bisbenzyloxyphosphoryl)-α-D-

- ⁵⁰ glucopyranosyl]-β-D-ribofuranosyl}-4-(3-fluorophenyl)-1,2,3triazole (9f)
- A mixture of azide **8** (0.2 g, 0.14 mmol), 3-fluorophenyl acetylene (0.021 mL, 0.15 mmol), Cu (0.036 g, 0.57 mmol) and CuSO₄ (0.008 g, 0.028 mmol) in H₂O/BuOH (1/1, v/v, 2 mL) ⁵⁵ was stirred at rt for 12 h. After completion of the reaction (TLC),
- the mixture was filtered, the filtrate was extracted with ethyl

acetate and washed successively with water and brine. The organic layer was dried over MgSO4 and concentrated under reduced pressure. The crude product thus obtained was purified 60 by flash column chromatography using a mixture of acetone and petroleum ether (1:3 v/v) as eluent to get 9f (0.188 g, 86%) as a colourless gum; ¹H NMR (500 MHz, CDCl₃) δ : 3.58 (dd, J = 10.85, 2.45 Hz, 1H, H- 5_{A}), 3.62 (dd, J = 9.65, 3.65 Hz, 1H, H-2"), 3.66-3.69 (m, 2H, H- 6_A ", H- 5_B), 3.78 (dd, J = 10.8, 1.5 Hz, $_{65}$ 1H, H- $_{B}''$) 3.90 (ddd, J = 10.8, 9.2, 4.2 Hz, 1H, H-5''), 4.33-4.38 (m, 2H, 1 x PhCH₂), 4.41-4.44 (m, 2H, 1 x PhCH₂), 4.46-4.47 (m, 1H, H-4'), 4.52-4.56 (m, 3H, H-3', H-4" and 0.5 x PhCH₂), 4.70-4.75 (m, 3H, 1.5 x PhCH₂), 4.80-4.84 (m, 3H, 1.5 x PhCH₂), 4.92-5.06 (m, 8H, H-3", 3.5 x PhC H_2), 5.28 (d, J = 3.6 Hz, 1H, 70 H-1"), 5.43 (ddd, J = 8.85, 5.2, 4.65 Hz, 1H, H-2'), 6.44 (d, 5.6 Hz, 1H, H-1'), 7.09-7.39 (m, 49H, Ar-H), 7.90 (s, 1H, H-5); ¹³C NMR (125 MHz, CDCl₃) δ: 68.7 (C-6"), 69.1 (C-5'), 69.4, 69.5, 69.8, 69.6, 69.7, 69.87, 69.9, 69.94, 70.07, 70.1 (³¹P coupled, PhCH2OP), 72.0 (C-5"), 72.1, 73.5, 73.7 (3 x PhCH2OC), 74.5 75 (C-3'), 74.53 (³¹P coupled, C-4"), 77.1 (³¹P coupled, C-2"), 78.1 (C-3", ³¹P coupled), 78.9 (C-2'), 83.1 (C-4'), 90.3 (C-1'), 95.9 (C-1"), 112.7, 114.8 (Ar-C), 118.5 (C-5), 121.4 (Ar-C), 127.6, 127.7, 127.8, 127.83, 127.9, 128.0, 128.07, 128.14, 128.31, 128.34, 128.43, 128.48, 128.5, 128.51, 128.69, (aromatic carbons), 130.2, 80 130.27, 136.2, 136.23, 137.31, 137.61, 138.1, 148.1, 162.3, 164.2 (³¹P coupled, ipso carbons of POCH₂Ph); ³¹P NMR (202.4 MHz, CDCl₃) δ: -1.515, -1.920, -2.064; ¹⁹F NMR (470.68 MHz, CDCl₃) δ: -112.766; Anal. calcd for C₈₂H₈₁FN₃O₁₈P₃: C, 65.29; H, 5.41;

g) Synthesis of 1-{5-*O*-benzyl-2-*O*-bisbenzyloxyphosphoryl-3-*O*-[2,6-di-*O*-benzyl-3,4-di-*O*-(bisbenzyloxyphosphoryl)- α -Dglucopyranosyl]- β -D-ribofuranosyl}-4-(4-fluorophenyl)-1,2,3triazole (9g)

N, 2.79. Found: C, 65.50; H, 5.21; N, 2.66.

A mixture of azide 8 (0.2 g, 0.14 mmol), 4-fluorophenyl 90 acetylene (0.018 mL, 0.15 mmol), Cu (0.036 g, 0.57 mmol) and CuSO₄ (0.008 g, 0.028 mmol) in H₂O/^tBuOH (1/1, v/v, 2 mL) was stirred at rt for 12 h. After completion of the reaction (TLC), the mixture was filtered, the filtrate was extracted with ethyl acetate and washed successively with water and brine. The 95 organic layer was dried over MgSO4 and concentrated under reduced pressure. The resulting crude product thus obtained was purified by flash column chromatography using a mixture of 25% acetone in petroleum ether as eluent to get pure 9g (0.207 g, 95%) as a colourless gum; ¹H NMR (500 MHz, CDCl₃) δ: 3.47-3.52 100 (m, 2H, H-2" and H-5_A'), 3.55-3.59 (m, 2H, H-6_A", H-5_B'), 3.65-3.67 (m, 1H, H-6_B"), 3.77-3.80 (m, 1H, H-5"), 4.21-4.28 (m, 2H, 1 x PhCH₂), 4.30-4.35 (m, 3H, H-4'), 4.39-4.45 (m, 3H, H-4" and 1 x PhCH₂), 4.61-4.65 (m, 3H, H-3', and 1 x PhCH₂), 4.67-4.73 (m, 3H, 1.5 x PhCH₂), 4.81-4.97 (m, 8H, H-3", 3.5 x PhCH₂), 105 5.16 (d, J = 3.6 Hz, 1H, H-1"), 5.32 (ddd, J = 9.2, 8.5, 5.0 Hz, 1H, H-2'), 6.34 (d, 5.6 Hz, 1H, H-1'), 6.94-7.37 (m, 49H, Ar-H), 7.79 (s, 1H, H-5); ¹³C NMR (125 MHz, CDCl₃) δ: 68.6 (C-6"), 69.2 (C-5'), 69.22, 69.4, 69.45, 69.5, 69.56, 69.6, 69.8, 69.9, 69.91, 70.03, 70.07 (³¹P coupled, PhCH₂OP), 70.3 (C-5"), 72.0, 110 73.5, 73.7 (3 x PhCH₂OC), 74.4 (C-3'), 74.5 (³¹P coupled, C-4"), 77.3 (³¹P coupled, C-2"), 78.0 (C-3", ³¹P coupled), 78.9 (C-2'), 83.0 (C-4'), 90.2 (C-1'), 95.9 (C-1"), 115.6 (C-5), 117.9, 127.5, 127.51, 127.62, 127.67, 127.7, 127.8, 127.87, 128.0, 128.01,

128.07, 128.1, 128.16, 128.20, 128.35, 128.39, 128.47, 128.5, 128.55, 128.65, 128.67 (aromatic carbons), 136.14, 136.20, 137.4, 137.6, 138.0, 147.5, 162.1, 164.4 (31 P coupled, ipso carbons of POCH₂Ph); 31 P NMR (202.4 MHz, CDCl₃) δ : -1.49, - 5 1.920, -2.092; 19 F NMR (470.68 MHz, CDCl₃) δ : -113.585; Anal. calcd for C₈₂H₈₁FN₃O₁₈P₃: C, 65.29; H, 5.41; N, 2.79. Found: C, 64.98; H, 5.11; N, 2.93.

h) Synthesis of 1-{5-*O*-benzyl-2-*O*-bisbenzyloxyphosphoryl-3-*O*-[2,6-di-*O*-benzyl-3,4-di-*O*-(bisbenzyloxyphosphoryl)-α-D ¹⁰ glucopyranosyl]-β-D-ribofuranosyl}-4-(3,5-difluorophenyl)-

1,2,3-triazole (9h) A mixture of azide **8** (0.2 g, 0.14 mmol), 3,5-difluorophenyl

- acetylene (0.018 mL, 0.15 mmol), Cu (0.036 g, 0.57 mmol) and CuSO₄ (0.008 g, 0.028 mmol) in H₂O/^tBuOH (1/1, v/v, 2 mL) 15 was stirred at rt for 12 h. After completion of the reaction (TLC), the mixture was filtered. The filtrate was extracted with ethyl acetate and washed successively with water and brine. The organic layer was dried over MgSO4 and concentrated under reduced pressure. The crude product thus obtained was purified 20 by flash column chromatography using a mixture of 25% acetone in petroleum ether as eluent to get pure 9h (0.2 g, 91%) as a colourless gum;. ¹H NMR (500 MHz, CDCl₃) δ : 3.46 (dd, J = 10.8, 2.1 Hz, 1H, $H-5_{A'}$), 3.51 (dd, J = 9.65, 3.6 Hz, 1H, H-2''), 3.54-3.58 (m, 2H, H- $6_A''$ and H- $5_B'$), 3.68 (dd, J = 10.7, 1.3 Hz, $_{25}$ 1H, H-6_B"), 3.79 (ddd, J = 5.95, 4.85, 3.95 Hz, 1H, H-5"), 4.24 $(dd, J = 11.5, 7.5 Hz, 2H, 1 x PhCH_2), 4.29-4.34 (m, 2H, 1 x$ PhC H_2), 4.36 (d, J = 2.3 Hz, 1H, H-4'), 4.38-4.45 (m, 3H, H-3', H-4" and 0.5 x PhCH2), 4.59-4.62 (m, 3H, 1.5 x PhCH2), 4.68-4.72 (m, 3H, 1.5 x PhCH₂), 4.81-4.94 (m, 8H, H-3", 3.5 x ³⁰ PhC H_2), 5.18 (d, J = 3.6 Hz, 1H, H-1"), 5.31 (ddd, J = 8.85, 5.2, 4.65 Hz, 1H, H-2'), 6.33 (d, J = 5.75 Hz, 1H, H-1'), 6.97-7.29 (m,
- 48H, Ar-*H*), 7.77 (s, 1H, H-5); ¹³C NMR (125 MHz, CDCl₃) δ: 68.7 (C-6"), 69.2 (C-5'), 69.23, 69.4, 69.45, 69.5, 69.57, 69.6, 69.8, 69.85, 69.9, 70.02, 70.07 (³¹P coupled, Ph*C*H₂OP), 70.4 (C-³⁵ 5"), 72.0, 73.48, 73.7 (3 x Ph*C*H₂OC), 74.5 (C-3"), 74.54 (³¹P coupled, C-4"), 77.1 (³¹P coupled, C-2"), 78.0 (C-3", ³¹P
- coupled), 78.9 (C-2'), 83.3 (C-4'), 90.4 (C-1'), 95.8 (C-1"), 108.7 (C-5), 118.9 127.6, 127.7, 127.73, 127.85, 127.92, 128.0, 128.1, 128.14, 128.3, 128.35, 128.39, 128.5, 128.51, 128.54, 128.58, 40 128.66, 128.79 (aromatic carbons), 135.2, 135.23, 135.25, 135.7,
- ⁴³ 126.00, 126.77 (aromatic carbons), 155.2, 155.22, 155.23, 155.7, 135.71, 135.8, 135.89, 136.1, 136.2, 137.2, 137.23, 137.6, 146.45, 162.5, 163.3 (³¹P coupled, ipso carbons of POCH₂Ph); ³¹P NMR (202.4 MHz, CDCl₃) δ : -1.502, -1.902, -2.068; ¹⁹F NMR (470.68 MHz, CDCl₃) δ : -109.331; Anal. calcd for ⁴⁵ C₈₂H₈₀F₂N₃O₁₈P₃: C, 64.52; H, 5.28; N, 2.75. Found: C, 64.37; H,
- 5.58; N, 2.91.

i) Synthesis of 1-{5-*O*-benzyl-2-*O*-bisbenzyloxyphosphoryl-3-*O*-[2,6-di-*O*-benzyl-3,4-di-*O*-(bisbenzyloxyphosphoryl)-α-Dglucopyranosyl]-β-D-ribofuranosyl}-4-(4-methoxyphenyl)-⁵⁰ 1,2,3-triazole (9i)

A mixture of azide 8 (0.2 g, 0.14 mmol), 4-methoxyphenyl acetylene (0.02 mL, 0.15 mmol), Cu (0.036 g, 0.57 mmol) and CuSO₄ (0.008 g, 0.028 mmol) in H₂O/^tBuOH (1/1, v/v, 2 mL) was stirred at rt for 12 h. After completion of the reaction (TLC)

55 the mixture was filtered. The filtrate was extracted with ethyl acetate and washed successively with water and brine. The

organic layer was dried over MgSO4 and concentrated under reduced pressure. The crude product thus obtained was purified by flash column chromatography using a mixture of acetone and 60 petroleum ether (1:3 v/v) as eluent to get 9i (0.193 g, 88%) as a colourless gum; ¹H NMR (500 MHz, CDCl₃) δ: 3.49-3.56 (m, 5H, $6_A''$, $H-6_B''$, $H-5_B'$, H-2'' and $H-5_A'$), 3.77 (bs, 4H, 1 x OCH₃ and H-5"), 4.21-4.35 (m, 5H, H-4' and 2 x PhCH₂), 4.40-4.45 (m, 3H, H-3', H-4" and 0.5 x PhCH₂), 4.58-4.72 (m, 6H, and 3 x 65 PhCH₂), 4.84-4.93 (m, 8H, H-3" and 3.5 x PhCH₂), 5.16 (bs, 1H, H-1"), 5.32 (bs, 1H, H-2'), 6.33 (d, J = 3.2 Hz, 1H, H-1'), 6.80 (d, J = 7.5 Hz, 2H, Ar-H), 7.00-7.28 (m, 45H, 9 x C₆H₅), 6.38 (d, J =7.5 Hz, 2H, Ar-H), 7.75 (s, 1H, H-5); ¹³C NMR (125 MHz, CDCl₃) δ: 55.4 (OCH₃), 68.6 (C-6"), 69.16 (C-5'), 69.2, 69.4, 70 69.44, 69.5, 69.58, 69.85, 69.9, 70.07, (³¹P coupled PhCH₂OP), 70.3 (C-5"), 72.0, 73.4, 73.6 (3 x PhCH₂OC), 74.4 (C-3' and C-4", ³¹P coupled), 77.0 (C-2', ³¹P coupled), 78.0 (C-2"), 78.6 (C-3", ³¹P coupled), 83.0 (C-4'), 90.3 (C-1', ³¹P coupled), 95.9 (C-1"), 114.2, 117.5 (C-5), 127.1, 127.2, 127.6, 127.7, 127.8, 127.82, 75 127.85, 128.0, 128.03, 128.07, 128.1, 128.2, 128.3, 128.4, 128.45, 128.5, 128.52, 128.6, 128.64, (aromatic carbons), 135.1, 135.15, 135.2, 135.65, 135.70, 135.8, 135.84, 136.1, 136.16, 137.4, 137.6, (³¹P coupled, ipso carbons of POCH₂Ph), 138.0 (ipso carbon of COCH₂Ph); ³¹P NMR (202.4 MHz, CDCl3) δ: -

 80 1.576, -1.941, -2.118; Anal. calcd for $C_{83}H_{84}N_3O_{19}P_3$: C, 65.56; H, 5.57; N, 2.76. Found: C, 65.66; H, 5.77; N, 2.95.

j) Synthesis of 1-{5-*O*-benzyl-2-*O*-bisbenzyloxyphosphoryl-3-*O*-[2,6-di-*O*-benzyl-3,4-di-*O*-(bisbenzyloxyphosphoryl)-α-Dglucopyranosyl]-β-D-ribofuranosyl}-4-(2-formylphenyl)-1,2,3ss triazole (9j)

A mixture of azide 8 (0.2 g, 0.14 mmol), 2-ethynylbenzaldehyde (0.02 g, 0.15 mmol), Cu (0.036 g, 0.57 mmol) and CuSO₄ (0.008 g, 0.028 mmol) in H₂O/^tBuOH (1/1, v/v, 2 mL) was stirred at rt for 12 h. After completion of the reaction (TLC) the mixture was 90 filtered. The filtrate was extracted with ethyl acetate and washed successively with water and brine. The organic layer was dried over MgSO₄ and concentrated under reduced pressure. The crude product thus obtained was purified by flash column chromatography using a mixture of acetone and petroleum ether 95 (1:3 v/v) as eluent to obtain pure 9j (0.19 g, 87%) as a colourless gum;. ¹H NMR (500 MHz, CDCl₃) δ: 3.57-3.64 (m, 2H, H-2" and H-5_A'), 3.68-3.70 (m, 2H, H-5_B' and H-6_A"), 3.76-3.78 (m, 1H, H-6B"), 3.90 (bs, 1H, H-5"), 4.33-4.44 (m, 4H, 2 x PhCH2), 4.49-4.58 (m, 4H, H-4', H-4" and 1 x PhCH₂), 4.72-4.86 (m, 6H, H-3', 100 and 2.5 x PhCH₂), 4.93-5.07 (m, 8H, H-3", 3.5 x PhCH₂), 5.29 (bs, 1H, H-1"), 5.47 (bs, 1H, H-2'), 6.48 (d, 5.0 Hz, 1H, H-1'), 7.13-7.41 (m, 46H, Ar-H), 7.51-7.59 (m, 2H, Ar-H), 8.00 (s, 1H, H-5), 8.05 (d, J = 7.4 Hz, 1H, Ar-H), 10.35 (s, 1H, CHO); ¹³C NMR (125 MHz, CDCl₃) δ: 68.6 (C-6"), 68.66 (C-5'), 69.2, 105 69.22, 69.3, 69.4, 69.45, 69.6, 69.61, 69.87, 69.9, 69.94, 70.07, 70.1 (³¹P coupled, PhCH₂OP), 70.4 (C-5"), 72.0, 73.5, 73.7 (3 x PhCH₂OC), 74.3 (C-3'), 74.5 (³¹P coupled, C-4"), 77.3 (³¹P coupled, C-2"), 78.0 (C-3", ³¹P coupled), 78.9 (C-2'), 83.2 (C-4'), 90.5 (C-1'), 95.8 (C-1"), 121.7 (C-5), 127.7, 127.71, 127.73, 110 127.8, 127.83, 127.86, 128.0, 128.1, 128.14, 128.19, 128.35, 128.38, 128.5, 128.54, 128.59, 128.69 (aromatic carbons), 129.9, 132.8, 134.1, 136.2, 137.2, 137.6, 138.1, 145.6 (³¹P coupled, ipso carbons of POCH₂Ph), 192.21 (CHO); ³¹P NMR (202.4 MHz,

CDCl₃) δ : -1.32, -1.89, -2.07; Anal. calcd for C₈₃H₈₂N₃O₁₉P₃: C, 65.65; H, 5.44; N, 2.77. Found: C, 65.41; H, 5.73; N, 2.90.

k) Synthesis of 1-{5-*O*-benzyl-2-*O*-bisbenzyloxyphosphoryl-3-*O*-[2,6-di-*O*-benzyl-3,4-di-*O*-(bisbenzyloxyphosphoryl)- α -Ds glucopyranosyl]- β -D-ribofuranosyl}-4-(6-methoxynaphth-2yl)-1,2,3-triazole (9k)

The mixture of azide 8 (0.1 g, 0.07 mmol),2-ethynyl-6methoxynaphthalene (0.014 g, 0.077 mmol), Cu (0.018 g, 0.28 mmol) and CuSO₄ (0.004 g, 0.014 mmol) in H₂O/^tBuOH (1/1, 10 v/v, 1 mL) was stirred at rt for 12 h. After completion of the reaction (TLC) the mixture was filtered. The filtrate was extracted with ethyl acetate and washed successively with water and brine. The organic layer was dried over MgSO4 and concentrated under reduced pressure. The crude product thus 15 obtained was purified by flash column chromatography using 25% acetone in petroleum ether as eluent to get pure, protected trizolophostin 9k (93 mg, 82%) as a colourless gum; ¹H NMR (500 MHz, CDCl₃) δ: 3.47-3.52 (m, 2H, H-2" and H-5₄'), 3.55-3.59 (m, 2H, H-5_B' and H-6_A"), 3.64-3.66 (m, 1H, H-6_B"), 3.77-20 3.80 (m, 1H, H-5"), 3.85 (s, 3H, OCH₃), 4.21-4.24 (m, 1H, 0.5 x PhCH₂), 4.28-4.36 (m, 4H, H-4' and 1.5 x PhCH₂), 4.41-4.48 (m, 3H, H-3', H-4" and 1 x PhCH₂), 4.59-4.65 (m, 3H, 1.5 x PhCH₂), 4.69-4.72 (m, 3H, 1.5 x PhCH₂), 4.80-4.97 (m, 8H, H-3", 3.5 x PhC H_2), 5.18 (d, J = 3.3 Hz, 1H, H-1"), 5.37 (ddd, J = 9.1, 8.5, 25 4.9 Hz, 1H, H-2'), 6.35 (d, 5.3 Hz, 1H, H-1'), 7.00-7.19 (m, 45H, 9 x C₆ H_5), 7.29 (d, J = 7.5 Hz, 2H, Ar-H), 7.48 (d, J = 7.5 Hz, 1H, Ar-H), 7.63 (t, J = 7.5 Hz, 2H, Ar-H), 7.89 (s, 1H, Ar-H), 8.05 (s, 1H, H-5); ¹³C NMR (125 MHz, CDCl₃) δ: 55.4 (OCH₃), 68.7 (C-6"), 69.2 (C-5'), 69.4, 69.6, 69.8, 69.9, 70.1 (³¹P coupled, 30 PhCH₂OP), 70.3 (C-5"), 72.0, 73.5, 73.6 (3 x PhCH₂OC), 74.5 (C-3'), 74.54 (³¹P coupled, C-4"), 77.0 (³¹P coupled, C-2"), 78.1 (C-3", ³¹P coupled), 78.8 (C-2'), 83.1 (C-4'), 90.1 (C-1'), 95.9 (C-1"), 105.9, 119.20 (C-5), 124.5, 127.2, 127.6, 127.63, 127.7, 127.8, 127.86, 127.89, 127.97, 128.0, 128.1, 128.13, 128.2, 35 128.3, 128.36, 128.38, 128.45, 128.5, 128.52, 128.57, 128.6, 128.7, 129.0, (aromatic carbons), 129.8, 134.4, 136.2, 137.2, 137.5, 137.6, 138.1 (³¹P coupled, ipso carbons of POCH₂Ph); ³¹P NMR (202.4 MHz, CDCl₃) δ: -1.524, -1.920, -2.082; Anal. calcd for C₈₇H₈₆N₃O₁₉P₃: C, 66.53; H, 5.52; N, 2.68. Found: C, 66.74; 40 H, 5.39; N, 2.88.

7. General Procedure for the Synthesis of Triazolophostins:

To a solution of protected triazolophostins **9a-k** (80-200 mg, 0.06-0.13 mmol) and cyclohexene (2 mL) in a mixture of methanol and water (9:1 v/v, 5 mL), Pd(OH)₂ (20% on carbon, 50 ⁴⁵ mg) was added and the mixture was stirred at 80 °C for 4 h. The reaction mixture was then cooled, filtered through a membrane filter washing successively with methanol and water. The combined filtrate was evaporated under reduced pressure. The crude product thus obtained was purified by ion-exchange

so column chromatography on Q-Sepharose matrix using $0 \rightarrow 1.0$ M triethylammonium bicarbonate (TEAB) as eluent to get pure triazolophostins **10a-k**.

a) 3'-O- α -D-glucopyranosyl-1- β -D-ribofuranosyl-1,2,3-triazole 2',3'',4''-trisphosphate [triazolophostin] (10a)

55 By using 80 mg (0.056 mmol) of **9a**, 33 mg (97%) of

triazolophostin 10a was obtained as a white hygroscopic solid: ¹H NMR (500 MHz, D_2O) δ : 3.74-3.80 (m, 7H, H-4", H-5", H-5_A', H-5_B', H-6_A", H-6_B" and H-2"), 4.03 (bs, 1H, H-4'), 4.41 (bs, 2H, H-3' and H-3"), 5.14 (bs, 1H, H-2'), 5.23 (bs, 1H, H-1"), 6.38 (bs, ⁶⁰ 1H, H-1'), 7.80 (s, 1H, H-5), 8.18 (s, 1H, H-4); ¹³C NMR (125 MHz, D_2O) δ : 60.3 (C-5'), 60.8 (C-6"), 70.9 (C-2", ³¹P coupled), 71.8 (C-5", ³¹P coupled), 72.6 (C-4", ³¹P coupled), 73.6 (C-3', ³¹P coupled), 73.8 (C-4'), 90.8 (C-1', ³¹P coupled), 98.0 (C-1"), 125.1 (C-4, C-5); ³¹P solution (C-2), δ : 3.496, 3.570, 4.382; HRMS (ESI)

mass calcd for $C_{13}H_{24}N_3O_{18}P_3$ [M]⁺, 603.0268, found: 603.0271.

b) 4-(1-methyl-imidazol-5-yl)-triazolophostin (10b)

By using 105 mg (0.07 mmol) of 9b, 45 mg (94%) of triazolophostin 10b was obtained as a white hygroscopic solid: $_{70}$ ¹H NMR (500 MHz, D₂O) δ : 3.74-3.81 (m, 5H, H-2", H-5_A', H- $5_{B'}$, H- $6_{A''}$ and H- $6_{B''}$), 3.87 (dd, 1H, J = 12.8, 3.05 Hz, H-4"), 3.94 (s, 3H, N-CH₃), 4.08-4.10 (m, 1H, H-5"), 4.47-4.50 (m, 2H, H-4' and H-3"), 4.66-4.70 (m, 1H, obscured by HDO peak, H-3'), 5.24 (dd, 1H, J = 8.8, 4.45 Hz,, H-2'), 5.27 (d, J = 3. 3 Hz, 1H, H- $_{75}$ 1"), 6.46 (d, J = 3.9 Hz, 1H, H-1'), 7.76 (s, 1H, ImH-4), 8.63 (s, 1H, H-5), 8.80 (s, 1H, ImH-2); ¹³C NMR (125 MHz, D₂O) δ: 34.8 (N-CH₃), 60.3 (C-5'), 60.7 (C-6"), 70.6 (C-2", ³¹P coupled), 71.7 (C-5", ³¹P coupled), 72.8 (C-4", ³¹P coupled), 73.6 (C-3', ³¹P coupled), 76.5 (C-2', ³¹P coupled), 77.7 (C-3", ³¹P coupled), 84.2 ⁸⁰ (C-4'), 91.2 (C-1', ³¹P coupled), 98.0 (C-1"), 118.8 (C-5), 124.8 (ImC-4), 125.0 (C-4), 134.1 (ImC-5), 136.4 (ImC-2); ³¹P NMR (202.4 MHz, CDCl₃) δ: 3.62, 3.69, 4.41; HRMS (ESI) mass calcd for C₁₇H₂₈N₅O₁₈P₃ [M+H]⁺, 684.072, found 684.0699.

c) 4-phenyl triazolophostin (10c)

⁸⁵ By using 100 mg (0.067 mmol) of **9c**, 41 mg (91%) of triazolophostin **10c** was obtained as a white hygroscopic solid: ¹H NMR (500 MHz, D₂O) δ: 3.73-3.85 (m, 6H, H-2", H-5_A', H-4", H-5_B', H-6_A", H-6_B"), 4.07-4.09 (m, 1H, H-5"), 4.43 (bs, 1H, H-4'), 4.47-4.49 (m, 1H, H-3"), 4.64-4.75 (m, 1H, H-3'), 5.19-5.20
⁹⁰ (m, 1H, H-2'), 5.24 (s, 1H, H-1"), 6.39 (d, 1H, 3.65 Hz, H-1'), 7.38-7.41 (m, 1H, Ar-H), 7.46 (t, *J* = 7.5 Hz, 2H, Ar-H), 7.78 (d, *J* = 7.5 Hz, 2H, Ar-H), 8.48 (s, 1H, H-5); ¹³C NMR (125 MHz, D₂O) δ: 60.2 (C-5'), 60.8 (C-6''), 70.4 (C-2", ³¹P coupled), 71.6 (C-5", ³¹P coupled), 73.0 (C-4", ³¹P coupled), 73.8 (C-3', ³¹P
⁹⁵ coupled), 76.6 (C-2', ³¹P coupled), 78.1 (C-3", ³¹P coupled), 83.9 (C-4'), 91.0 (C-1', ³¹P coupled), 98.1 (C-1"), 121.3 (C-5), 125.9, 126.0, 129.1, 129.2, 129.3 (Ar-H); ³¹P NMR (202.4 MHz, D₂O) δ: 3.551, 3.597, 4.343; HRMS (ESI) mass calcd for C₁₉H₂₈N₃O₁₈P₃[M-H]⁺, 678.0503, found 678.0482.

100 d) 4-(2-pyridyl)-triazolophostin (10d)

By using 100 mg (0.067 mmol) of **9d**, 21.2 mg (47%) of triazolophostin **10d** was obtained as a sticky mass: ¹H NMR (500 MHz, D₂O) δ: 3.79-3.90 (m, 5H, H-6_A", H-6_B", H-5_A', H-5_B', H-2"), 4.11-4.12 (m, 1H, H-5"), 4.49-4.51 (m, 2H, H-4' and H-4"), ¹⁰⁵ 4.72 (m, 2H, obscured by HDO peak, H-3' and H-3"), 5.26-5.29 (m, 2H, H-2' and H-1"), 6.51 (s, 1H, H-1'), 7.98 (s, 1H, Pyr-*H*), 8.40 (d, *J* = 6.5 Hz, 1H, Pyr-*H*), 8.62 (s, 1H, Pyr-*H*), 8.74 (s, 1H, Pyr-*H*), 9.04 (s, 1H, H-5); ¹³C NMR (125 MHz, D₂O) δ: 60.3 (C-5'), 60.7 (C-6"), 70.6 (C-2", ³¹P coupled), 71.8 (C-5", ³¹P 110 coupled), 73.0 (C-4", ³¹P coupled), 73.8 (C-3', ³¹P coupled), 76.7

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 $\begin{array}{l} (C\mbox{-}2',\ ^{31}\mbox{P}\ coupled),\ 78.0\ (C\mbox{-}3'',\ ^{31}\mbox{P}\ coupled),\ 84.5\ (C\mbox{-}4'),\ 91.4 \\ (C\mbox{-}1',\ ^{31}\mbox{P}\ coupled),\ 98.1\ (C\mbox{-}1''),\ 124.9\ (C\mbox{-}5),\ 126.2,\ 139.4,\ 141.3, \\ 142.4,\ 147.6\ (A\mbox{r}\mbox{-}C);\ ^{31}\mbox{P}\ NMR\ (202.4\ MHz,\ D_2O)\ \delta:\ 3.637, \\ 3.720,\ 4.448;\ HRMS\ (ESI)\ mass\ calcd\ for\ C_{18}H_{27}N_4O_{18}P_3\ [M\mbox{s}\ 5\ H]^+,\ 679.0455,\ found\ 679.0431. \end{array}$

e) 4-(2-Fluorophenyl)-triazolophostin (10e)

By using 190 mg (0.12 mmol) of 9e, 79 mg (91%) of triazolophostin **10e** was obtained as a white hygroscopic solid: ¹H NMR (500 MHz, MeOD) 3.58-3.81 (m, 6H, H-5", H-6_A", H-6_B", ¹⁰ H-5_A', H-5_B', H-2"), 4.09-4.11 (m, 1H, H-4"), 4.30 (bs, 1H, H-4'), 4.51-4.53 (m, 1H, H-3"), 4.68 (t, J = 5.0 Hz, 1H, H-3"), 5.15 (bs, 1H, H-1"), 5.22 (bs, 1H, H-2'), 6.31 (bs, 1H, H-1'), 7.10-7.14 (m, 1H, Ar-H), 7.18 (t, J = 7.5 Hz, 1H, Ar-H), 7.28 (dd, J = 13.0, 6.5 Hz, 1H, Ar-H), 8.02 (t, J = 7.5 Hz, 1H, Ar-H), 8.51 (s, 1H, H-5); 15 ¹³C NMR (125 MHz, MeOD) δ: 61.9 (C-5'), 62.1 (C-6"), 72.7 (C-2", ³¹P coupled), 73.6 (C-5", ³¹P coupled), 74.6 (C-4", ³¹P coupled), 76.8 (C-3', ³¹P coupled), 79.1 (C-2', ³¹P coupled), 79.5 (C-3", ³¹P coupled), 85.3 (C-4'), 93.0 (C-1', ³¹P coupled), 100.6 (C-1"), 116.9 (C-5), 124.0, 125.8, 128.8, 131.1, 142.4, 159.8, 20 161.7 (Ar-C); ³¹P NMR (202.4 MHz, MeOD) δ: 0.601 (2P), -0.550; ¹⁹F NMR (470.68 MHz, MeOD) δ: -116.02; HRMS (ESI) mass calcd for $C_{19}H_{27}FN_3O_{18}P_3$ [M]⁺, 697.0486, found 697.0490.

f) 4-(3-Fluorophenyl)-triazolophostin (10f)

By using 180 mg (0.11 mmol) of 9f, 78 mg (93%) of ²⁵ triazolophostin **10f** was obtained as a white hygroscopic solid: ¹H NMR (500 MHz, MeOD) 3.61-3.62 (m, 1H, H-2"), 3.69-3.84 (m, 5H, H-5", H-6_A", H-6_B", H-5_A', H-5_B'), 4.13-4.18 (m, 1H, H-4"), 4.32-4.33 (m, 1H, H-4'), 4.53-4.55 (m, 1H, H-3"), 4.69 (t, J = 5.0 Hz, 1H, H-3'), 5.18 (d, J = 3.0 Hz, 1H, H-1"), 5.27 (t, J = 3.8 Hz, $_{30}$ 1H, H-2'), 6.32 (d, J = 3.0 Hz, 1H, H-1'), 7.01 (dt, J = 8.5, 2.0 Hz, 1H, Ar-H), 7.38 (dd, J = 8.0, 14.0 Hz, 1H, Ar-H), 7.51 (d, J = 9.7 Hz, 1H, Ar-*H*), 7.57 (d, *J* = 7.5 Hz, 1H, Ar-*H*), 8.58 (s, 1H, H-5); ¹³C NMR (125 MHz, MeOD) δ: 61.9 (C-5'), 62.0 (C-6"), 72.5 (C-2", ³¹P coupled), 73.4 (C-5", ³¹P coupled), 75.2 (C-4", ³¹P ³⁵ coupled), 76.6 (C-3', ³¹P coupled), 79.3 (C-2', ³¹P coupled), 80.0 (C-3", ³¹P coupled), 85.4 (C-4'), 92.7 (C-1', ³¹P coupled), 100.5 (C-1"), 113.4 (C-5), 116.2, 121.8, 122.6, 132.0, 133.9, 148.0 (Ar-C); ³¹P NMR (202.4 MHz, MeOD) δ: -0.755, -0.131, 0.279; ¹⁹F NMR (470.68 MHz, MeOD) δ: -114.722; HRMS (ESI) mass ⁴⁰ calcd for C₁₉H₂₇FN₃O₁₈P₃ [M]⁺, 697.0486, found 697.0472.

g) 4-(4-Fluorophenyl)-triazolophostin (10g)

By using 200 mg (0.13 mmol) of **9g**, 88 mg (95%) of triazolophostin **10g** was obtained as a white hygroscopic solid: 1 H NMR (500 MHz, MeOD) 3.60-3.83 (m, 6H, H-2", H-5", H-6_A",

- ⁴⁵ H-6_B", H-5_A', H-5_B'), 4.13 (bs, 1H, H-4"), 4.31 (bs, 1H, H-4'), 4.54 (bs, 1H, H-3"), 4.68 (bs, 1H, H-3'), 5.17 (bs, 1H, H-1"), 5.25 (bs, 1H, H-2'), 6.30 (bs, 1H, H-1'), 7.09 (t, J = 8.5 Hz, 2H, Ar-H), 7.76 (d, J = 5.5 Hz, 2H, Ar-H), 8.50 (s, 1H, H-5); ¹³C NMR (125 MHz, MeOD) δ: 61.9 (C-5'), 62.1 (C-6"), 72.6 (C-2", ³¹P
- ⁵⁰ coupled), 73.5 (C-5", ³¹P coupled), 74.9 (C-4", ³¹P coupled), 76.7 (C-3', ³¹P coupled), 79.1 (C-2', ³¹P coupled), 79.7 (C-3", ³¹P coupled), 85.4 (C-4'), 92.8 (C-1', ³¹P coupled), 100.5 (C-1"), 116.8 (C-5), 116.9, 121.2, 128.0, 128.8, 128.9, 148.2, 163.3, 165.2 (Ar-C); ³¹P NMR (202.4 MHz, MeOD) δ: -0.602, -0.269, ⁵⁵ 0.473; ¹⁹F NMR (470.68 MHz, MeOD) δ: -115.406; HRMS (ESI)

mass calcd for $C_{19}H_{27}FN_3O_{18}P_3\ \mbox{[M]}^+,\,697.0486,\,found\,\,697.0481.$

h) 4-(3,5-diflurorophenyl)-triazolophostin (10h)

By using 200 mg (0.13 mmol) of **9h**, 79 mg (84%) of triazolophostin **10h** was obtained as a white hygroscopic solid: ⁶⁰ ¹H NMR (500 MHz, MeOD) 3.59-3.79 (m, 6H, H-2", H-5", H- 6_{A} ", H- 6_{B} ", H- 5_{A} ', H- 5_{B} '), 4.10 (bs, 1H, H-4"), 4.30 (bs, 1H, H-4'), 4.52 (bs, 1H, H-3"), 4.66 (bs, 1H, H-3'), 5.15 (bs, 1H, H-1"), 5.22 (bs, 1H, H-2'), 6.28 (s, 1H, H-1'), 6.83 (t, J = 8.9 Hz, 1H,

- Ar-H), 7.36 (d, J = 6.2 Hz, 2H, Ar-H), 8.60 (s, 1H, H-5); ¹³C
- ⁶⁵ NMR (125 MHz, MeOD) δ: 61.9 (C-5'), 62.1 (C-6"), 72.7 (C-2", ³¹P coupled), 73.6 (C-5", ³¹P coupled), 74.7 (C-4", ³¹P coupled), 76.7 (C-3', ³¹P coupled), 79.0 (C-2', ³¹P coupled), 79.5 (C-3", ³¹P coupled), 85.4 (C-4'), 92.9 (C-1', ³¹P coupled), 100.6 (C-1"), 104.2 (C-5), 109.3, 122.4, 135.2, 146.9, 163.9, 165.9 (Ar-C); ³¹P ⁷⁰ NMR (202.4 MHz, MeOD) δ: -0.650, 0.47, 0.81; ¹⁹F NMR (470.68 MHz, MeOD) δ: -111.10; HRMS (ESI) mass calcd for C₁₉H₂₆F₂N₃O₁₈P₃ [M]⁺, 715.0392, found 715.0402.

i) 4-(4-methoxyphenyl)-triazolophostin (10i)

By using 195 mg (0.12 mmol) of **9i**, 87 mg (96%) of ⁷⁵ triazolophostin **10i** was obtained as a white hygroscopic solid: ¹H NMR (500 MHz, D₂O) δ : 3.65-3.76 (m, 9H, H-2", H-5", H-6_A", H-6_B", H-5_A', H-5_B' and 1 x OCH₃), 4.00-4.02 (m, 1H, H-4"), 4.33 (bs, 1H, H-4'), 4.40-4.41 (m, 1H, H-3"), 4.54 (t, *J* = 5.1 Hz, 1H, H-3'), 5.08-5.15 (m, 1H, H-2'), 5.16 (bs, 1H, H-1"), 6.27 (d, *J* so = 3.2 Hz, 1H, H-1'), 6.89 (d, *J* = 8.6 Hz, 1H, Ar-H), 7.54 (d, *J* = 8.6 Hz, 1H, Ar-H), 8.25 (s, 1H, H-5); ¹³C NMR (125 MHz, D₂O) δ : 55.6 (OCH₃), 60.3 (C-5'), 61.0 (C-6"), 70.7 (C-2", ³¹P coupled), 71.7 (C-5", ³¹P coupled), 73.0 (C-4", ³¹P coupled), 73.9 (C-3', ³¹P coupled), 76.5 (C-2', ³¹P coupled), 78.0 (C-3", ³¹P so coupled), 83.8 (C-4'), 91.2 (C-1', ³¹P coupled), 98.2 (C-1"), 114.8 (C-5), 120.6, 122.1, 127.5, 147.4, 159.5 (Ar-C); ³¹P NMR (202.4 MHz, MeOD) δ : -0.55, 0.66 (2P); HRMS (ESI) mass calcd for C₂₀H₃₀N₃O₁₉P₃ [M]⁺, 709.0686, found 709.0691.

j) 4-(2-formylphenyl)-triazolophostin (10j)

- ⁹⁰ By using 190 mg (0.12 mmol) of **9j**, 78 mg (88%) of triazolophostin **10j** was obtained as a white hygroscopic solid: ¹H NMR (500 MHz, D₂O) δ: 3.75-3.87 (m, 6H, H-2", H-5", H-6_A", H-6_B", H-5_A' and H-5_B'), 4.09-4.11 (m, 1H, H-4"), 4.44 (bs, 1H, H-4'), 4.50 (bs, 1H, H-3"), 4.65-4.68 (m, 1H, H-3'), 5.21 (bs, 1H,
- ⁹⁵ H-2'), 5.25 (bs, 1H, H-1"), 6.40-6.42 (m, 1H, H-1'), 7.34 (bs, 1H, Ar-H), 7.46-7.47 (m, 1H, Ar-H), 7.60-7.61 (m, 1H, Ar-H), 7.77 (s, 1H, H-5 (triazole)), 8.36-8.48 (m, 1H, CH(OH)₂); ¹³C NMR (125 MHz, D₂O) δ: 60.3 (C-5'), 60.9 (C-6"), 71.5 (C-2", ³¹P coupled), 72.0 (C-5", ³¹P coupled), 72.2 (C-4", ³¹P coupled), 73.4
- ¹⁰⁰ (C-3', ³¹P coupled), 76.1 (C-2', ³¹P coupled), 77.1 (C-3'', ³¹P coupled), 83.8 (C-4'), 91.2 (C-1', ³¹P coupled), 97.8 (C-1''), 121.2 (C-5), 125.8, 126.3, 129.0, 129.3, 130.9, 136.5, 146.7, 147.4, (Ar-*C*); ³¹P NMR (202.4 MHz, D₂O) δ : 3.51, 3.63, 4.32; HRMS (ESI) mass calcd for C₂₀H₂₈N₃O₁₉P₃ [M]⁺, 707.0530, found 707.0549.

105 k) 4-(6-methoxynaphth-2-yl)-triazolophostin (10k)

By using 90 mg (0.057 mmol) of **9k**, 42 mg (97%) of triazolophostin **10k** was obtained as a white hygroscopic solid: ¹H NMR (500 MHz, MeOD) δ : 3.06 (d, J = 7.7 Hz, 1H, H-2"), 3.69-3.85 (m, 8H, H-5", H-5_A', H-5_B', H-6_A", H-6_B" and OCH₃),

4.13-4.15 (m, 1H, H-4"), 4.32-4.33 (m, 1H, H-4'), 4.53-4.55 (m, 1H, H-3"), 4.70 (t, J = 5.0 Hz, 1H, H-3'), 5.17 (bs, 1H, H-1"), 5.28 (bs, 1H, H-2'), 6.32 (d, J = 2.7 Hz, 1H, H-1'), 7.05 (dd, J = 9.0, 2.4 Hz, 1H, Ar-*H*), 7.15 (d, J = 2.2 Hz, 1H, Ar-*H*), 7.69 (d, J = 9.0 Hz, 1H, Ar-*H*), 7.73 (d, J = 9.0 Hz, 1H, Ar-*H*), 7.79 (s, 1H, H-5), 8.13 (s, 1H, Ar-*H*), 8.57 (s, 1H, Ar-*H*); ¹³C NMR (125 MHz, MeOD) δ : 55.9 (OCH₃), 61.9 (C-5'), 62.1 (C-6"), 72.6 (C-2", ³¹P coupled), 73.5 (C-5", ³¹P coupled), 75.0 (C-4", ³¹P coupled), 75.0 (C-4", ³¹P coupled), 77.0 (C-3', ³¹P coupled), 79.3 (C-2', ³¹P coupled), 79.5 ¹⁰ (C-3", ³¹P coupled), 85.4 (C-4'), 92.9 (C-1', ³¹P coupled), 100.7 (C-1"), 106.9 (C-5), 120.4, 121.2, 125.2, 125.5, 126.6, 128.7, 130.4, 130.7, 136.1, 149.4, 159.7 (Ar-C); ³¹P NMR (202.4 MHz, MeOD) δ : 3.605 (2P), 4.393; HRMS (ESI) mass calcd for C₂₄H₃₂N₃O₁₉P₃ [M-H]⁺, 758.0765, found 758.0785.

15 8. Biological Assay:

 Ca^{2+} release from the intracellular stores of saponinpermeabilized DT40 cells expressing only type 1 IP₃R was measured using a low-affinity Ca^{2+} indicator (Mag-fluo-4) as described previously.²¹ Briefly, the ER was loaded with Ca^{2+} to $_{20}$ steady state for ~120 s by addition of 1.5 mM MgATP in medium

containing *p*-trifluoromethoxyphenylhydrazone (FCCP) to inhibit mitochondria. IP₃, AdA or triazolophostins were added with cyclopiazonic acid (10 μ M) to inhibit further Ca²⁺ uptake. Ca²⁺ release was assessed 10–20 s after addition of the analog, and ²⁵ expressed as a fraction of the ATP-dependent Ca²⁺ uptake.

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35 Notes and rerences

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† Electronic Supplementary Information (ESI) available: Synthetic procedures and spectral data for all new compounds, crystal data (CCDC

45 1022279) for disaccharide 4 and details of the docking study. See DOI: 10.1039/b000000x/

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