

# SYNTHESIS OF DIACETYLENIC ANALOGUES OF LEUKOTRIENE A<sub>4</sub> (LTA<sub>4</sub>) METHYL ESTER

J.P. LELLOUCHE, J. DESCHAMPS, C. BOULLAIS, J.P. BEAUCOURT \*

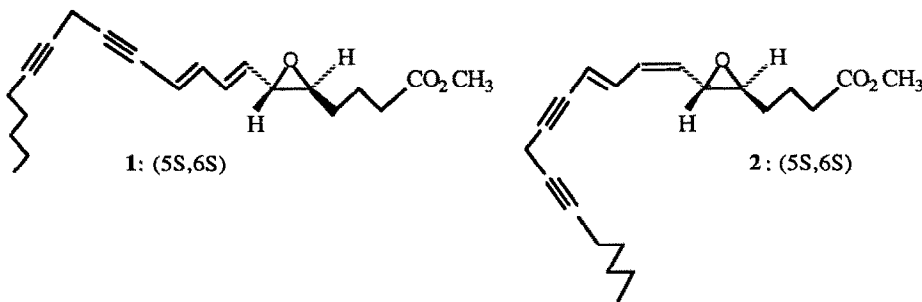
Service des Molécules Marquées, CEN Saclay, 91191 GIF-SUR-YVETTE, FRANCE.

## Summary

Methyl (5S,6S)-epoxy-11,14-eicosadiyne-7E,9E-dienoate **1** and its 7Z isomer **2** were prepared using Wittig methodology. **1** did not inhibit human neutrophil 5-lipoxygenase but rather stimulated LTB<sub>4</sub> formation.

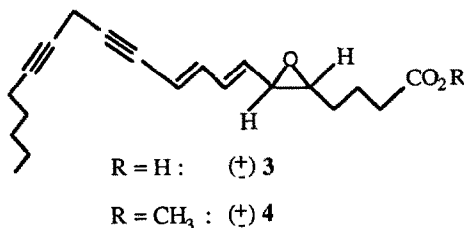
Arachidonic acid [AA or 20:4 (n-6)] is a polyunsaturated 20 carbon acid. It is an important component of membrane phospholipids in animal cells. Various phospholipases can release AA, which is metabolized by 5-lipoxygenase to give peptidoleukotrienes <sup>(1)</sup> through LTA<sub>4</sub>.

As a part of our studies on pharmacologically active lipids, we report here the syntheses of two diacetylenic analogues of LTA<sub>4</sub> methyl ester : methyl (5S,6S)-epoxy-11,14-eicosadiyne-7E,9E-dienoate **1** and its 7Z isomer **2** :



The synthesis of racemic epoxydiyne **4** via sulfonium ylid chemistry <sup>(2,3)</sup> has two major drawbacks :

- the time of reaction must not exceed 1 min at -25°C
- as **4** is a racemic, its ring opening by thiopeptides (glutathione, cysteinylglycine, cysteine) leads to diastereoisomeric diacetylenic peptidoleukotrienes which ought to be separable by HPLC. However, only the chromatographic resolution of the diastereomeric diacetylenic LTE<sub>4</sub> has been described <sup>(2)</sup>.



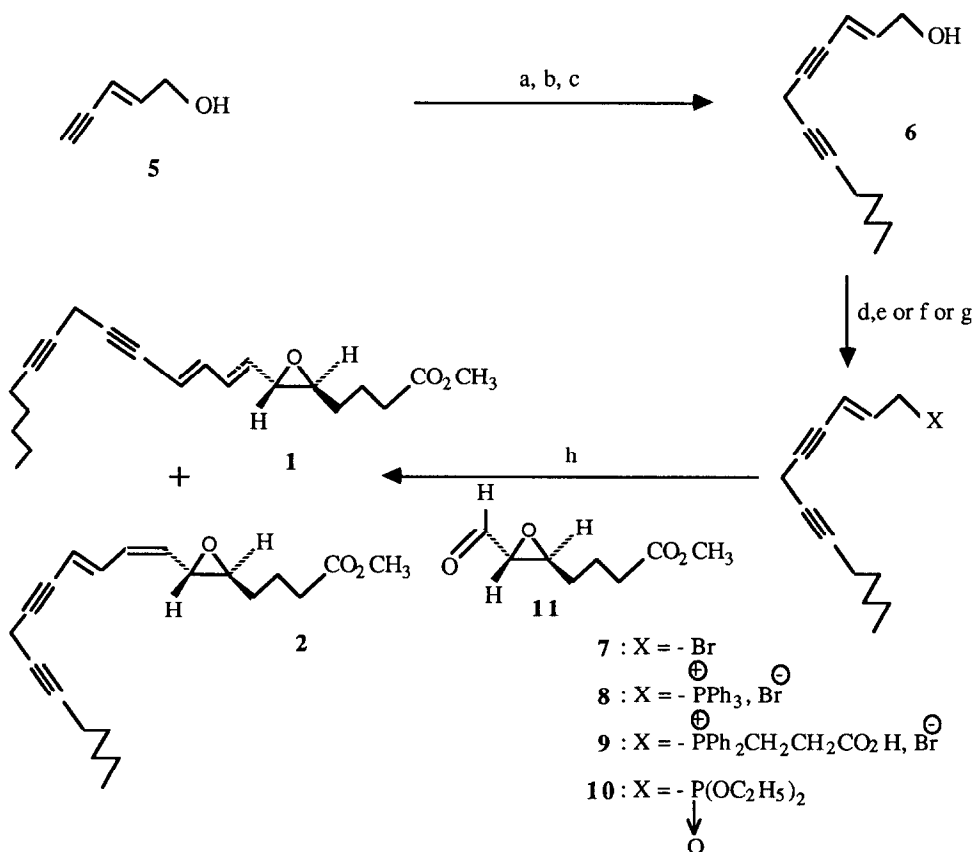
Both analogues **1** and **2**, and especially **1**, are particularly interesting because :

a) controlled Lindlar semi-hydrogenation of **1** with deuterium or tritium gas leads to 11,12,14,15-tetradeuterated or -tritiated LTA<sub>4</sub> methyl ester <sup>(11)</sup>.

b) **1** and **2** are precursors in the syntheses of acetylenic analogues of peptidoleukotrienes.

On the other hand, racemic epoxydiyne **3** has been shown to inhibit 50 % of SRS-A activity at a concentration of 10 μM. The (5*S*,6*R*)-LTE<sub>4</sub> analogue shows 50 % SRS-A activity at 11 μM <sup>(2)</sup>.

Thus, it seemed interesting to synthesize the chiral diacetylenic LTA<sub>4</sub>-methyl ester analogues **1** and **2** according to the following scheme, in order to test their biological properties as well as those of the corresponding peptidoleukotrienes.



a : CH<sub>2</sub> = CH-OC<sub>2</sub>H<sub>5</sub>, toluenesulfonic acid, 0°C, 1 h ; b : C<sub>2</sub>H<sub>5</sub>MgBr, THF, 20°C ; CuBr, 20°C ; n-C<sub>5</sub>H<sub>11</sub>-C≡C-CH<sub>2</sub>Br, 60°C, 45 min ; c : 0.5 N H<sub>2</sub>SO<sub>4</sub>, CH<sub>3</sub>COCH<sub>3</sub>, 3 h, 20°C ; d : PBr<sub>3</sub>, 5 % pyridine in ether, 18 h, 20°C ; e : PPh<sub>3</sub>, ether, 20°C, 48 h ; f : PPh<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>CO<sub>2</sub>H, ether, 20°C, 48 h ; g : P(OC<sub>2</sub>H<sub>5</sub>)<sub>3</sub>, CH<sub>3</sub>CN, 80°C, 7 h ; h : nBuLi : 1 eq for **8** and **10**, 2 eq for **9** ; anhydrous THF ; -80°C ; 30 min.

Diyne enol **6**, prepared from pent-4-yn-(2E)-en-1-ol **5** <sup>(4)</sup> according to ROSENBERGER et al.<sup>(3)</sup>, was brominated to **7** <sup>(5)</sup> (63 % yield). When **7** was treated with triphenylphosphine, 3-diphenylphosphinoyl-propanoic acid <sup>(6)</sup> or triethyl phosphite, phosphonium salts **8** (85 %) and **9** (50 %) or phosphonate **10** (72 %) were formed <sup>(5,7)</sup>. A solution of one equivalent of chiral epoxyaldehyde **11** <sup>(8)</sup> in anhydrous THF was added dropwise to the ylid of **8**, **9** or **10** over the period of one hour at 0°C. The crude mixture of **1** and **2** obtained was purified by column chromatography on silica gel (hexane/ethyl acetate/triethylamine : 70/30/02). Final purification allowed the isolation of 7E-**1** and 7Z-**2** <sup>(9)</sup> (Yield from ylid of **8** : 40 % of **1/2** (21/79), from ylid of **9** : 44 % of **1/2** (54/46), from ylid of **10** : 25 % of **1/2** (81/19)).

It must be noticed that the partially stabilized ylids react according to previously described results <sup>(10)</sup>.

The lithio phosphonate formed from **10** is probably too nucleophilic towards the fragile epoxy bridge of **11**, which can explain the lower yield of **1/2** obtained (25 %), 7E-**1** being predominant. Better yields were obtained with the less nucleophilic phosphonium salts of **8** and **9**. The best results were obtained with the ylid of **9** (44 % of **1/2** : 54/46 with a slightly predominant 7E isomer formation) due to the following factors :

- acidic phosphine oxide formed during the coupling reaction is eliminated in the aqueous phase, which makes easier further HPLC purification of **1/2**.

- in our experimental conditions the lithio carboxylic group of the ylid of **9** is compatible with the epoxy bridge.

#### *Biological evaluation of 1 .*

Saponified **1** <sup>(12)</sup> did not inhibit soybean 15-lipoxygenase confirming the structural specificity of the enzyme for a -(1Z,3Z)-pentadiene unit.

The formation and assay of 5-HETE at 234 mμ, LTB<sub>4</sub>, ω-hydroxy LTB<sub>4</sub> and ω-carboxy LTB<sub>4</sub> during the incubation of **1** with human neutrophils (10-30 mn incubation with 5 x 10<sup>6</sup> cells) showed that **1** did not inhibit human neutrophil 5-lipoxygenase <sup>(13)</sup>. It rather increased of about 30 % the amount of LTB<sub>4</sub> formed at concentrations of 1-10 μM <sup>(13)</sup>. The better bioavailability of endogeneous LTA<sub>4</sub> for LTA<sub>4</sub>-hydrolase could account for this result. Since diacetylenic peptidoleukotrienes (DAPTS) are devoid of any contractile activity on guinea pig ileum <sup>(14)</sup>, inhibition of glutathione-transferase by endogeneously biosynthesized DAPTS might be involved and explain our results as well as those reported by Rosenberger <sup>(2)</sup> on racemic (±) **3**.

On the other hand, **1** inhibited ω-hydrolase by 40 % at a concentration of 1μM and by 80 % at 10 μM <sup>(13)</sup>.

## REFERENCES AND NOTES

1 - S. HAMMARSTRÖM, R.C. MURPHY, B. SAMUELSSON, D.A. CLARK, C. MIOSKOWSKI, E.J. COREY, *Biochim. Biophys. Res. Commun.*, 1979, **91**, 1266 ; R.C. MURPHY, S. HAMMARSTRÖM, B. SAMUELSSON, *Proc. Natl. Acad. Sci. USA*, 1979, **76**, 4275.

2 - M. ROSENBERGER, *Eur. Pat. Appl. EP 36,663/30 Sept. 1981* (CA : **96**, 181124 u).

3 - M. ROSENBERGER, C. NEUKOM, *J. Amer. Chem. Soc.*, 1980, **102**, 5426.

4 - Manufactured by FARCHAN, Division Chempsampco, Inc.

5 - All intermediates had satisfactory <sup>1</sup>H-NMR, MS and UV data.

6 - H. DANIEL, M. LE CORRE, *Tetrahedron Lett.*, 1987, 28, 1165. We thank M. LE CORRE for providing us with acidic phosphine.

7 - Phosphonium salts **8** and **9** are stable when stored at 0°C. Phosphonate **10** has to be stored in anhydrous THF solution at -80°C (2mg of **10** in 1 ml THF).

8 - J. ROKACH, R. ZAMBONI, C.K.LAU, Y. GUINDON, *Tetrahedron Lett.*, 1981, 22, 2759.

9 - **1** UV (C<sub>2</sub>H<sub>5</sub>OH) :  $\lambda_{\max 1} = 271 \text{ m}\mu$  (44500) ;  $\lambda_{\max 2} = 283 \text{ m}\mu$  (35800).  $[\alpha]^{20} = -43^\circ$  (CHCl<sub>3</sub> ; 1.14 g/100 ml) ; SM : m/e = 328.5 (M<sup>+</sup> ; 14.4 %) ; RMN-<sup>1</sup>H (250 MHz ; CDCl<sub>3</sub> ;  $\delta$  en ppm ; TMS standard) ; 0.90 [t, 3H (H<sub>20</sub>), J<sub>19,20</sub> = 7.0 Hz] ; 1.32 to 1.80 [broad signal, 10H (H<sub>3</sub> + H<sub>4</sub> + H<sub>17</sub> + H<sub>18</sub> + H<sub>19</sub>)] ; 2.15 [t, 2H (H<sub>16</sub>), J<sub>16,17</sub> = 7.0 Hz] ; 2.38 [t, 2H (H<sub>2</sub>), J<sub>2,3</sub> = 7.2 Hz] ; 2.87 [t, 1H (H<sub>5</sub>), J<sub>4,5</sub> = 5.0 Hz] ; 3.13 [dd, 1H (H<sub>6</sub>), J<sub>5,6</sub> = 2.0 Hz, J<sub>6,7</sub> = 8.0 Hz] ; 3.32 [s, 2H (H<sub>13</sub>)] ; 3.68 [s, 3H (-OCH<sub>3</sub>)] ; 5.47 [dd, 1H (H<sub>7</sub>), J<sub>7,8</sub> = 15.0 Hz] ; 5.63 [d, 1H (H<sub>10</sub>), J<sub>9,10</sub> = 15.0 Hz] ; 6.40 [dd, 1H (H<sub>8</sub>), J<sub>8,9</sub> = 11.0 Hz] ; 6.53 [dd, 1H (H<sub>9</sub>)] ;

2 UV (C<sub>2</sub>H<sub>5</sub>OH) :  $\lambda_{\max 1} = 271.5 \text{ m}\mu$  (32800) ;  $\lambda_{\max 2} = 283.5 \text{ m}\mu$  (39900).  $[\alpha]^{20} = -8.5^\circ$  (CHCl<sub>3</sub> ; 6.1 g/100 ml) ; SM : m/e = 328.5 (M<sup>+</sup> ; 19 %) ; RMN-<sup>1</sup>H (250 MHz ; CDCl<sub>3</sub> ;  $\delta$  en ppm ; TMS standard) ; 0.92 [t, 3H (H<sub>20</sub>), J<sub>19,20</sub> = 7.0 Hz] ; 1.32 to 1.83 [broad signal, 10H (H<sub>3</sub> + H<sub>4</sub> + H<sub>17</sub> + H<sub>18</sub> + H<sub>19</sub>)] ; 2.17 [tt, 2H (H<sub>16</sub>), J<sub>16,17</sub> = 7.0 Hz, J<sub>13,16</sub> = 3.0 Hz] ; 2.40 [t, 2H (H<sub>2</sub>), J<sub>2,3</sub> = 7.2 Hz] ; 2.88 [multiplet, 1H (H<sub>5</sub>)] ; 3.33 [multiplet, 2H (H<sub>13</sub>)] ; 3.48 [dd, 1H (H<sub>6</sub>), J<sub>5,6</sub> = 2.0 Hz, J<sub>6,7</sub> = 9.0 Hz] ; 3.70 [s, 3H (-OCH<sub>3</sub>)] ; 5.10 [t, 1H (H<sub>7</sub>), J<sub>7,8</sub> = 10.0 Hz] ; 5.67 [d, 1H (H<sub>10</sub>), J<sub>9,10</sub> = 16.0 Hz] ; 6.23 [t, 1H (H<sub>8</sub>), J<sub>8,9</sub> = 10.0 Hz] ; 6.95 [dd, 1H (H<sub>9</sub>)] .

10 - *Organophosphorus Reagents in Organic Synthesis*, ed. J.L.G. CADOGAN, 1979, Academic Press, New-York.

11 - Partial reduction of **1** and **2** by deuterium gas will be described elsewhere (*Tetrahedron Letters*, following paper).

12 - F. FITZPATRICK, D. MORTON, M. WYNALDA, *J. Biol. Chem.*, 1982, 257, 4680.

13 - We thank Pr. DELAFORGE (UA 400 - CNRS, Université R. DESCARTES, 45 rue des Saints-Pères, 75270 PARIS CEDEX 06) for these data.

14 - These results will be soon reported : J.P. LELLOUCHE, F. AUBERT, J.P. BEAUCOURT, E. RECHENCQ, G. NIEL, J.P. GIRARD, J.C. ROSSI, M. BOUCARD, *Prostaglandins*, in press.

(Received in France 20 April 1988)