



A Journal of the Gesellschaft Deutscher Chemiker

# Angewandte Chemie

GDCh

International Edition

[www.angewandte.org](http://www.angewandte.org)

## Accepted Article

**Title:** Divergent Synthesis of Monosubstituted and Unsymmetrical 3,6-Disubstituted Tetrazines from Carboxylic Ester Precursors

**Authors:** Joseph M. Fox, Yixin Xie, Yinzhi Fang, Zhen Huang, Amanda Tallon, and Christopher am Ende

This manuscript has been accepted after peer review and appears as an Accepted Article online prior to editing, proofing, and formal publication of the final Version of Record (VoR). This work is currently citable by using the Digital Object Identifier (DOI) given below. The VoR will be published online in Early View as soon as possible and may be different to this Accepted Article as a result of editing. Readers should obtain the VoR from the journal website shown below when it is published to ensure accuracy of information. The authors are responsible for the content of this Accepted Article.

**To be cited as:** *Angew. Chem. Int. Ed.* 10.1002/anie.202005569

**Link to VoR:** <https://doi.org/10.1002/anie.202005569>

## RESEARCH ARTICLE

## Divergent Synthesis of Monosubstituted and Unsymmetrical 3,6-Disubstituted Tetrazines from Carboxylic Ester Precursors

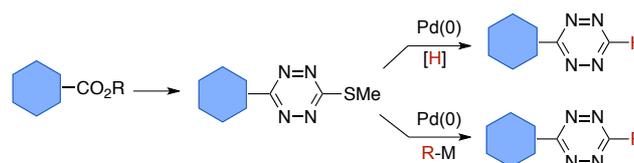
Yixin Xie,<sup>[a]</sup> Yinzhi Fang,<sup>[a]</sup> Zhen Huang,<sup>[c]</sup> Amanda M. Tallon,<sup>[a]</sup> Christopher W. am Ende,<sup>\*[b]</sup> and Joseph M. Fox<sup>\*[a]</sup>

**Abstract:** As tetrazines are important tools to the field of bioorthogonal chemistry, there is a need for new approaches to synthesize unsymmetrical and 3-monosubstituted tetrazines. Described here is a general, one-pot method for converting (3-methyloxetan-3-yl)methyl carboxylic esters into 3-thiomethyltetrazines. These versatile intermediates were applied as a platform for the synthesis of unsymmetrical tetrazines via Pd-catalyzed cross-coupling and in the first example of catalytic thioether reduction to access monosubstituted tetrazines. The method enables the development of new tetrazines possessing a favorable combination of kinetics, small size and hydrophilicity. The chemistry was applied to a broad range of aliphatic and aromatic ester precursors and to the synthesis of heterocycles including BODIPY fluorophores and biotin. In addition, a series of tetrazine probes for monoacylglycerol lipase (MAGL) were synthesized and the most reactive one was applied in labeling of endogenous MAGL in live cells.

## Introduction

Bioorthogonal reactions have become increasingly important to chemistry and biology since the term was initially coined early in this century.<sup>[1]</sup> The bioorthogonal reactions of tetrazines have become widely used in various fields,<sup>[2]</sup> including natural product synthesis,<sup>[3]</sup> cargo delivery,<sup>[4]</sup> genetic code expansion,<sup>[5]</sup> fluorogenic labeling,<sup>[6]</sup> radiochemistry,<sup>[7]</sup> coordination chemistry,<sup>[8]</sup> and material science.<sup>[9]</sup> Coupled to the growing utility of tetrazine-based bioorthogonal chemistry is the need for mild, safe and general methods of introducing tetrazine groups to complex molecules. Carboxylic esters are ubiquitous functional groups that have been used as handles for the introduction of preformed tetrazine groups via amide bond forming reactions.<sup>[10]</sup> For this approach, linkers are required which can add bulk and hydrophilicity to the conjugate.<sup>[11]</sup> The direct preparation of tetrazines from carboxylic precursors could greatly expand access to functionalized derivatives. Described here is a new strategy for the one-pot synthesis of 3-thiomethyltetrazines

## Installation of minimal tetrazines from carboxylic esters



**Figure 1.** Direct approach for the introduction of tetrazine functionality from carboxylic ester precursors.

from carboxylic esters of commercially available 3-methyl-3-oxetanemethanol. The compounds serve as versatile intermediates for the divergent synthesis of a broad range of functionalized tetrazines via Pd-catalyzed cross-coupling and in the first example of catalytic thioether reduction to form monosubstituted tetrazines (Figure 1). This 'carboxylate-to-tetrazine' strategy complements known approaches of tetrazine construction,<sup>[12]</sup> tolerates a range of heterocycles and functional groups, and generates classes of tetrazines that are most useful to bioorthogonal chemistry.

The most widely used method of preparing tetrazines involves condensation of nitriles and hydrazine followed by oxidation (Figure 2A).<sup>[13]</sup> Devaraj's method for Zn(OTf)<sub>2</sub> or Ni(OTf)<sub>2</sub> catalyzed condensation of nitriles in neat anhydrous hydrazine has greatly improved access to unsymmetrical tetrazines.<sup>[13d]</sup> More recently, thiols have been used as organocatalysts that function with hydrazine hydrate which is safer and more broadly available than anhydrous hydrazine.<sup>[13e]</sup> Mono-functional tetrazines have been prepared from nitriles, hydrazine and formamidine acetate under Lewis acid catalyzed conditions<sup>[13d, 13f, 13g]</sup> or with dichloromethane under sulfur-catalyzed conditions (Figure 2B).<sup>[14]</sup> Safety considerations for these procedures include the formation of volatile tetrazine byproducts with high nitrogen content and the direct addition of an oxidant to reaction mixtures containing hydrazine. Tetrazines have been used in metal catalyzed Heck-type reactions,<sup>[6c]</sup> CH activations<sup>[15]</sup> and cross-couplings.<sup>[6d, 6e, 16]</sup> 3,6-Dichlorotetrazine,<sup>[2, 17]</sup> 3,6-bis(3,5-dimethylpyrazol-1-yl) tetrazine<sup>[18]</sup>, and 3-bromotetrazine<sup>[19]</sup> have been used to create thiol, alcohol and amine functionalized tetrazines by nucleophilic aromatic substitution. While these methods for tetrazine synthesis have been enabling, there is a continuing need to broaden the types of precursors that can serve for the direct installation of tetrazines with improved functional group tolerance.

Carboxylic esters have played only a limited role as precursors to tetrazines.<sup>[12]</sup> Unsymmetrical 3-thiomethyltetrazines were synthesized in low yield through the condensation of S-methylisothiocarbonohydrazidium iodide (**2**) with either trimethylorthoacetate or trimethylorthoformate.<sup>[20]</sup> Recently, our group described a procedure for the conversion of trimethyl-

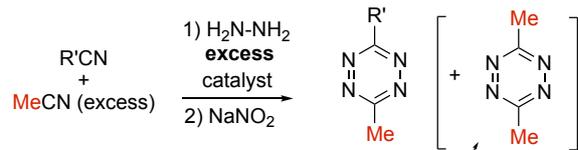
[a] Y. Xie, Y. Fang, A. M. Tallon and Prof. Dr. J. M. Fox  
Department of Chemistry and Biochemistry  
University of Delaware  
Newark, DE 19716 (USA)  
E-mail: [jmfox@udel.edu](mailto:jmfox@udel.edu)

[b] Dr. C. W. am Ende  
Pfizer Worldwide Research and Development  
Eastern Point Road, Groton, CT 06340 (USA)  
E-mail: [Christopher.amende@pfizer.com](mailto:Christopher.amende@pfizer.com)

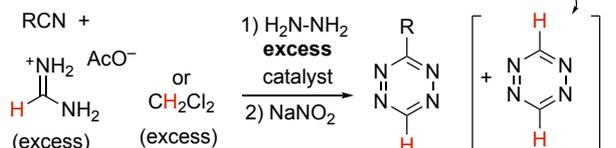
[c] Dr. Z. Huang  
Pfizer Worldwide Research and Development  
1 Portland Street, Cambridge, MA 02139 (USA)

## RESEARCH ARTICLE

## A Unsymmetrical Tetrazines from Nitriles with Hydrazine



## B Mono-substituted Tetrazines from Nitriles



## C This work: Tetrazines from Ester Precursors

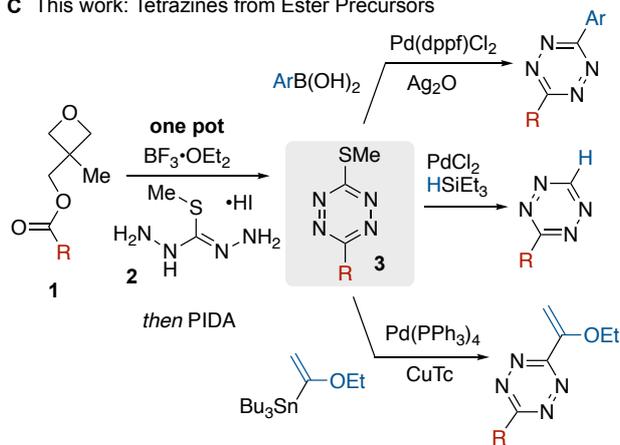


Figure 2. Selected methods of tetrazine synthesis

orthoacetate to a 3-methyl-6-thioalkyltetrazine that can in turn be used for the installation of minimal tetrazines through the silver-mediated Liebeskind-Srogl coupling with arylboronic acids.<sup>[21]</sup> This method was limited to trimethylorthoacetate and triethylorthoacetate, and we hoped to develop a general approach for directly converting ester precursors into tetrazines. However, trimethyl- and triethylorthoesters cannot be prepared directly from esters but instead require nitrile precursors and harshly acidic conditions.<sup>[22]</sup> Thus, our previous method did not enable the use of carboxylic acids or esters as precursors to tetrazines. Here, we describe a one-pot procedure for the conversion of (3-methyloxetan-3-yl)methyl carboxylic esters (**1**) to unsymmetrical 3-thiomethyltetrazines (**3**) via condensation of oxabicyclo[2.2.2] octyl (OBO) orthoester<sup>[23]</sup> intermediates with **2** and subsequent oxidation (Figure 2C). The activated esters **3** can be prepared simply and in high yield from the corresponding carboxylic acids and inexpensive 3-methyl-3-oxetanemethanol. These compounds provide a divergent platform for 3,6-disubstituted tetrazine synthesis and a new approach to monosubstituted tetrazines (Figure 2C).

## Results and Discussion

Table 1. Optimization of condensation between **1a'** and **2**.

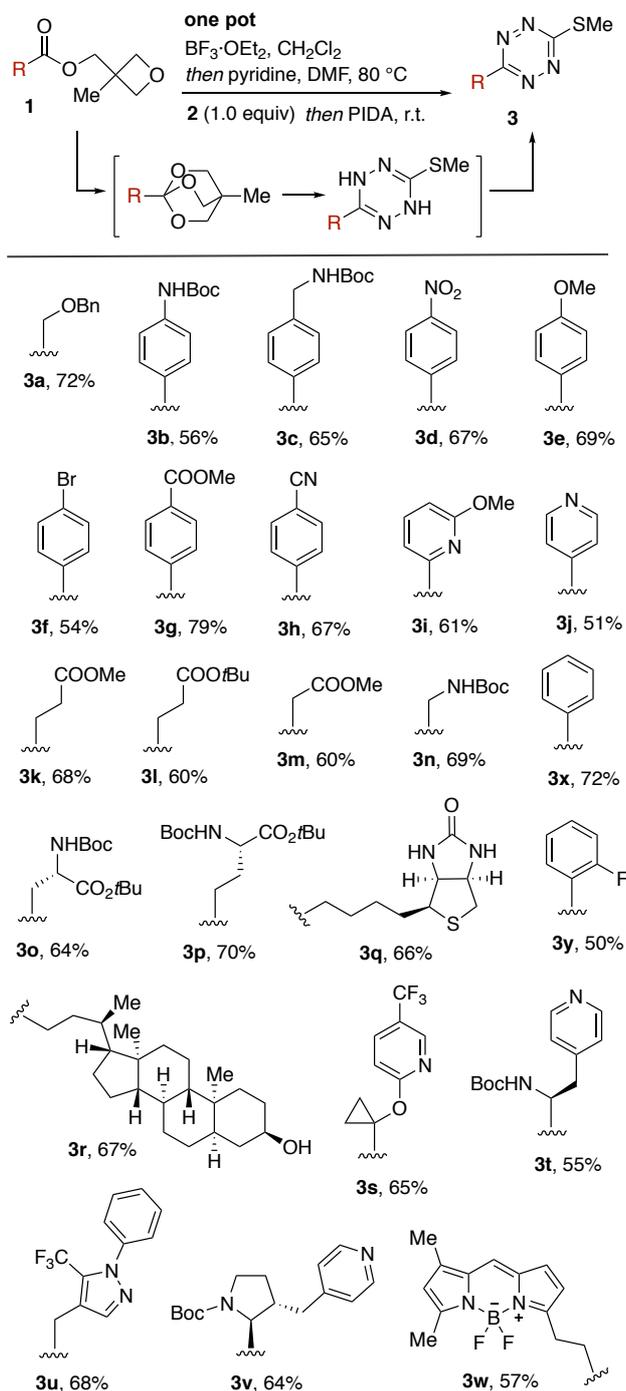
entry	additive	equiv of <b>1a'</b>	temp.	time	conc.	yield of <b>3a</b>
1	none	1.0	r.t.	6 h	0.2 M	18%
2	NEt <sub>3</sub> (1 equiv)	1.0	r.t.	6 h	0.2 M	0
3	DMAP (1 equiv)	1.0	r.t.	6 h	0.2 M	0
4	Cs <sub>2</sub> CO <sub>3</sub> (1 equiv)	1.0	r.t.	7 h	0.2 M	0
5	KOtBu (1 equiv)	1.0	r.t.	6 h	0.2 M	0
6	pyridine (1 equiv)	1.0	r.t.	7 h	0.2 M	24%
7	pyridine (3 equiv)	1.0	80 °C	1 h	0.2 M	23%
8	pyridine (3 equiv)	1.0	80 °C	20 min	1.0 M	54%
9	pyridine (3 equiv)	1.25	80 °C	20 min	1.0 M	75%
10	pyridine (3 equiv)	1.4	80 °C	20 min	1.0 M	87%

Compound **2** is available commercially or can be prepared in one step from iodomethane and thiocarbohydrazide.<sup>[24]</sup> The differential scanning calorimetry (DSC) profile of **2** has an onset temperature of 135 °C and a transition enthalpy of 875 J/g (Figure S3). The OBO orthoester **1a'** was prepared by treating its (3-methyloxetan-3-yl) methyl ester with BF<sub>3</sub>·OEt<sub>2</sub>. Table 1 shows conditions for optimizing the condensation of **1a'** with **2**, with subsequent oxidation by phenyliodosodiacetate (PIDA)<sup>[25]</sup> to provide thiomethyltetrazine **3a**. DMF was the only effective solvent likely due to the limited solubility of salt **2** in other solvents. Simple combination of **1a'** and **2** at r.t. gave **3a** in only 18% yield (Table 1, entry 1). Given the acid-lability of orthoesters, a number of basic additives were explored (entries 2-6). Pyridine was found to be uniquely effective, and further optimization showed that the temperature could be raised to 80 °C and reaction time shortened without sacrificing yield (entry 7). Using 3 equiv. of pyridine and increasing the concentration to 1 M had a significant impact and raised the yield to 54% (entry 8). Using 1.4 equiv. of **1a'** further raised the yield to 87% (entry 10).

We next developed a one-pot method for the synthesis of 3-thiomethyltetrazines **3** from esters **1**, which can be easily prepared in high yield (>90%) and inexpensive 3-methyl-3-oxetanemethanol from the corresponding acids by Steglich esterification. After initial treatment of **1** in CH<sub>2</sub>Cl<sub>2</sub> with BF<sub>3</sub>·OEt<sub>2</sub> to form the OBO orthoester, pyridine and **2** were added. The solvent was exchanged with DMF, and after heating at 80 °C, the dihydrotetrazine solution was cooled and directly treated with PIDA to provide tetrazines **3** in 50–79% yield (Scheme 1). Aromatic and alkyl substituents with a variety of functional groups were tolerated including Boc-amino (**3b-c**, **3n**), nitro (**3d**) and methoxy (**3e**) groups. The method also tolerates nitrile (**3h**) and ester (**3g**, **3k-m**) groups which are generally not compatible with tetrazine synthesis. Protected amino acids (**3o-p**), biotin (**3q**), BODIPY-FL (**3w**) and a number of heterocycles (**3i-j**, **3s-v**) were also successful and highlighted the ability to directly conjugate biologically relevant compounds. While the ester of 2-fluoro

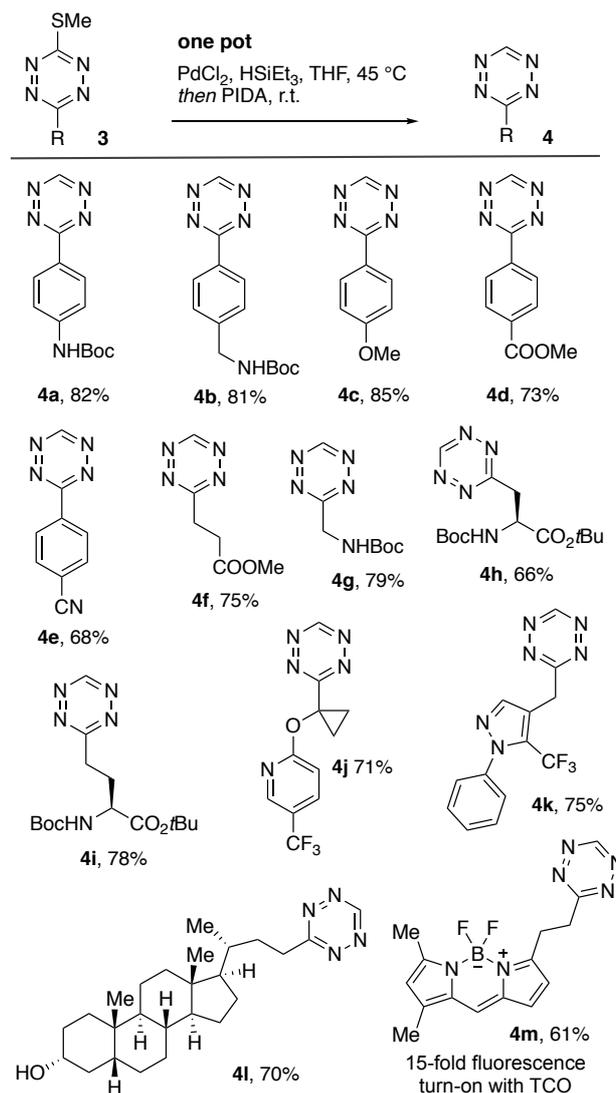
## RESEARCH ARTICLE

benzoic acid provides **3y** in 50% yield, esters with bulkier *ortho*-substituents (2-nitro or *N*-Boc-2-amino) were unsuccessful.



**Scheme 1.** One-pot synthesis of thiomethyltetrazines from oxetane esters.

Monosubstituted tetrazines are valued in bioorthogonal chemistry for their rapid kinetics and minimal size.<sup>[26]</sup> We sought to prepare monosubstituted tetrazines through thiomethyltetrazine reduction (Scheme 2). After optimization (see



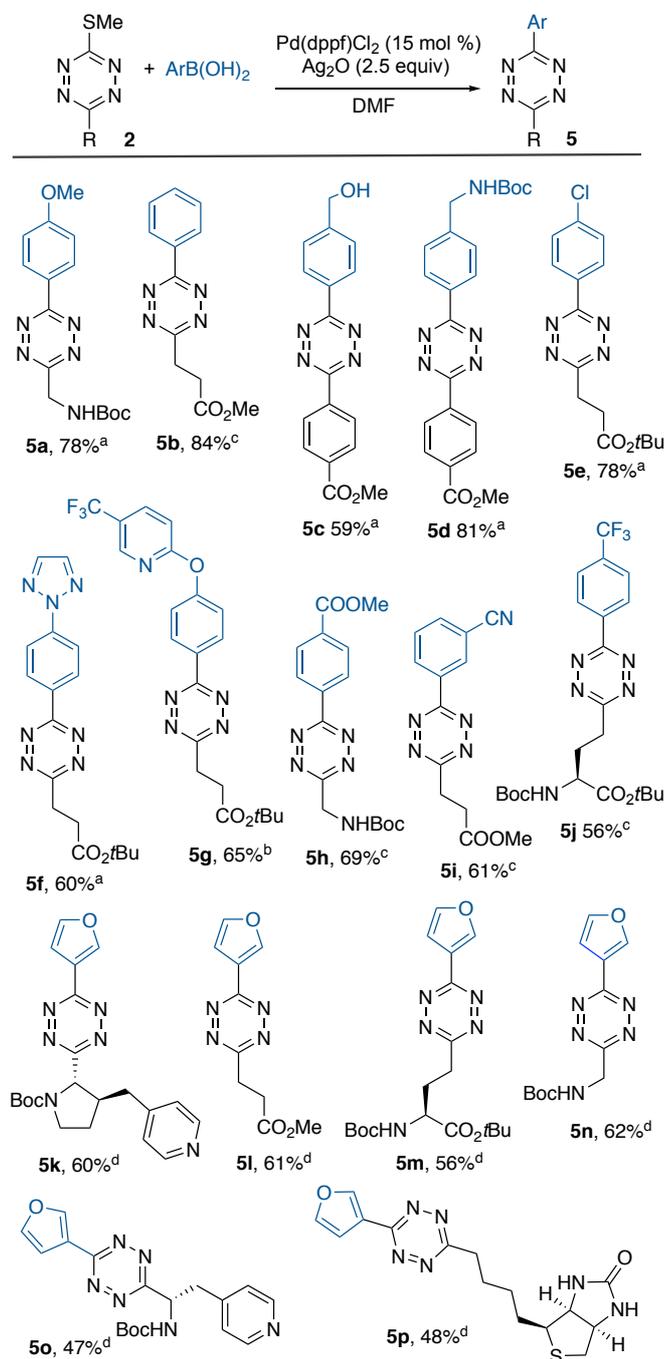
**Scheme 2.** Synthesis of 3-monosubstituted tetrazines. Conditions: **3** (1 equiv), PdCl<sub>2</sub> (10 mol %), HSiEt<sub>3</sub> (3 equiv.), THF, 45 °C, 24 h; PIDA (1.2 equiv.), r.t., 1h.

Table S1), it was found that efficient reduction could be realized with catalytic PdCl<sub>2</sub> (10 mol %) and triethylsilane<sup>[27]</sup> (3 equiv) in THF at 45 °C followed by treatment with PIDA to oxidize the initially formed dihydrotetrazine. The method provides monosubstituted tetrazines derived from aliphatic and aromatic esters with a variety of functional groups, including Boc-amino (**4a–b**, **4g**), methoxy (**4c**), ester (**4d**, **4f**) nitrile (**4e**), protected amino acids (**4h–i**), and heterocycles (**4j–k**). Reduction of **3w** gave BODIPY-FL derivative (**4m**), which shows a 15-fold turn-on of fluorescence upon reaction with *eq*-5-hydroxy-*trans*-cyclooctene (5-hydroxyTCO).<sup>[6a–e]</sup> Pyridine derivatives (**3i–j**, **3t**, **3v**) gave low conversions and attempted reduction of **3q** was unsuccessful, possibly due to overreduction of the biotin core.

Thiomethyltetrazines **3** also served as electrophiles for Ag-mediated Liebeskind-Srogl coupling reactions<sup>[21]</sup> with thioethers bearing either aliphatic or aromatic groups (Scheme 3). Tolerated functional groups include ether, chloride, ester, nitrile,

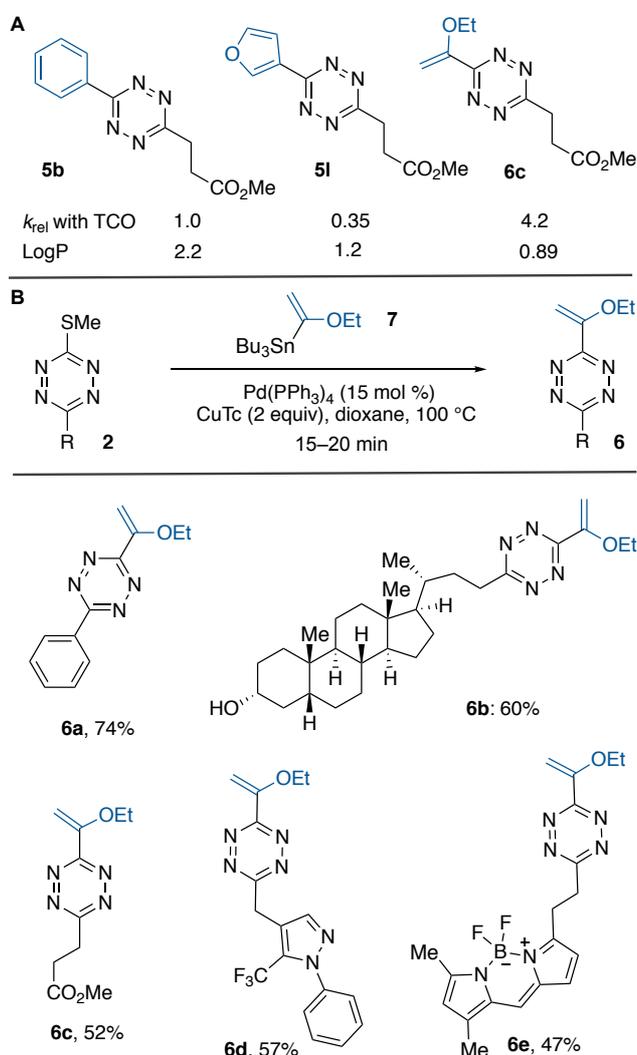
## RESEARCH ARTICLE

trifluoromethyl, alcohol, Boc-amino, and protected amino acids as well as pyridyl and triazole groups. Additionally, the 3-furyl group was readily introduced to a range of thiomethyltetrazines via Ag-mediated Liebeskind-Srogl coupling. In line with our previous observations, electron rich arylboronic acids were the most efficient nucleophiles.



**Scheme 3.** Synthesis of unsymmetrical tetrazines via Ag-mediated Pd-catalyzed Liebeskind-Srogl coupling. [a] ArB(OH)<sub>2</sub> (1.9 equiv.), 60 °C, 20 h. [b] ArB(OH)<sub>2</sub> (3 equiv.), 60 °C, 20 h. [c] ArB(OH)<sub>2</sub> (3 equiv.), microwave 100 °C, 3 h. [d] Pd(dppf)Cl<sub>2</sub> (30 mol %), Ag<sub>2</sub>O (5 equiv.), ArB(OH)<sub>2</sub> (6 equiv.), microwave 100 °C, 3 h.

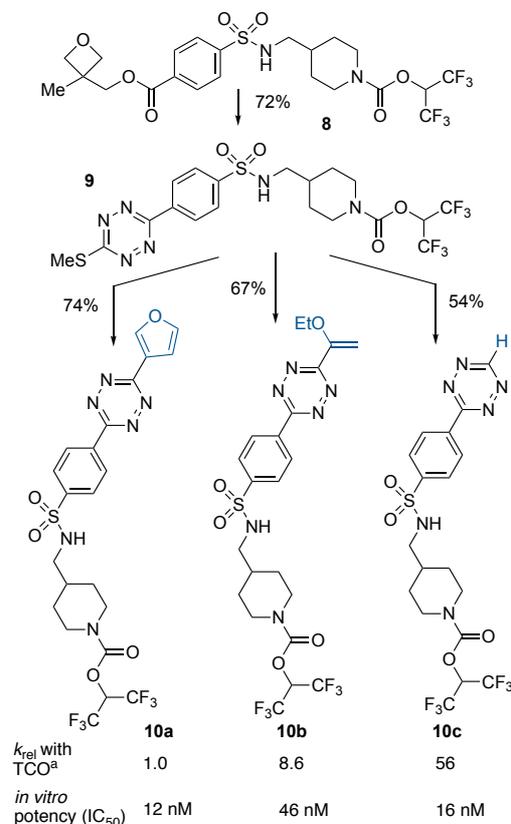
3,6-Disubstituted aromatic tetrazines have been used broadly for bioorthogonal chemistry due to their combination of stability and rapid kinetics but can be limited by their hydrophobicity. We anticipated that furyl or vinyl ether groups introduced through cross-coupling could serve as small and solubilizing alternatives to phenyl groups. Shown in Scheme 4A are the experimental logP values for three tetrazine analogs and their rate constants in reactions with 5-hydroxyTCO (Figure S2A-C). Furyl analog **5l** (logP 1.2) is considerably more hydrophilic albeit 2.9 times less reactive than phenyl derivative **5b** (logP 2.2). Vinyl ether **6c** is the most reactive and hydrophilic member of the series with logP 0.89 and reactivity that is 4.2 times faster than **5b**. Vinyl ether-substituted tetrazines represent a new class of dienophile with a favorable combination of kinetics, small size and hydrophilicity. Tetrazines **5b**, **5l** and **6c** all display >96% stability after 24 h incubation in PBS at 25 °C (Figure S1A). To introduce the 2-ethoxyvinyl group, we employed commercially available stannane



**Scheme 4.** Furyl- and vinyl ether-substituted tetrazines with favorable kinetics, small size and hydrophilicity. (A) Relative reactivity and logP value. (B) Synthesis via CuTc-mediated Liebeskind-Srogl coupling.

## RESEARCH ARTICLE

7. Here, coupling reactions were most efficient under conventional CuTc-promoted conditions with a brief reaction time

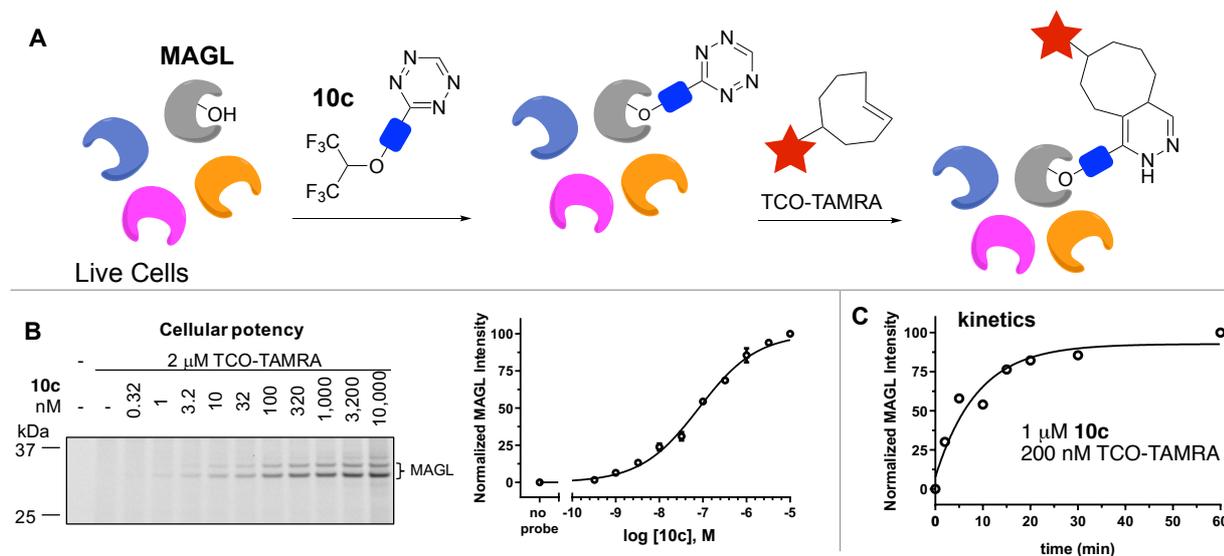


**Scheme 5.** Divergent synthesis of tetrazine-coupled MAGL-probes **10a-c** from common ester starting material and thiomethyltetrazine intermediate.

(see Table S2). As shown in Scheme 4B, these cross-coupling procedures can tolerate a broad range of functional groups and heterocycles of biological interest.

This divergent approach to synthesizing tetrazine-coupled probes was applied to the study of MAGL, a serine hydrolase involved in endocannabinoid signaling.<sup>[28]</sup> Our previous approach to studying this drug target was limited by the need for a scaffold with a relatively uncommon boronic acid group, thus restricting the types of drug candidates that could be studied.<sup>[21]</sup> We hypothesized that a divergent approach could be used to tune reaction kinetics and/or inhibitory affinity of a new class of MAGL-tetrazine probes. As shown in Scheme 5, probes **10a-c** were designed based on a 4-(arylsulfonamidomethyl)piperidine scaffold bearing an electrophilic hexafluoroisopropyl (HFIP) carbamate warhead.<sup>[29]</sup> Thus, reaction of **2** with the (3-methyloxetane-3-yl) methyl ester **8** gave 72% yield of thiomethyltetrazine **9**, which served as common intermediate for the synthesis of furyl-substituted **10a**, vinyloxy-substituted **10b**, and monosubstituted tetrazine **10c**. Stopped flow kinetics with 5-hydroxyTCO showed **10c** was 6.5-times more reactive than **10b**, which in turn was 8.6-fold more reactive than **10a**. All of these probes were 3–162 times more reactive than a previously described MAGL probe based on a different drug scaffold (**12**<sup>[21]</sup> Figure S4).

In an *in vitro* assay,<sup>[30]</sup> probes **10a-c** inhibited MAGL activity with IC<sub>50</sub>'s of 12 nM (**10a**), 46 nM (**10b**) and 16 nM (**10c**). The most reactive probe **10c** was further investigated for labeling of endogenous MAGL in live cells. Human brain vascular pericytes were treated with probe **10c** for 1 h, followed by labeling with 2 μM TCO-TAMRA for 30 min. After cell lysis, gel-based activity-based protein profiling<sup>[31]</sup> (ABPP, Figure 3A) showed strong labeling of MAGL. The labeling was dose responsive (Figure 3B), and treating cells with **10c** (1 μM, 1 h) followed by incubation with 200 nM TCO-TAMRA gave complete labeling with  $t_{1/2}$  of 6.1 min



**Figure 3.** (A) Live cells were treated with probe **10c** for 1 h, followed by TCO-TAMRA treatment for 30 min, cell lysis, and analysis by gel-based ABPP. (B) In-gel fluorescence and dose response fitting of probe **10c**. (C) Cellular labeling kinetics of **10c**

## RESEARCH ARTICLE

(Figure 3C). Similar kinetics were observed when the concentration of TCO-TAMRA was dropped to 50 nM (Figure S7), and with a higher concentration of TCO-TAMRA (2  $\mu$ M), labeling was complete when the first data point was collected after 2 min. By contrast, our previous 6-methyltetrazin-3-yl probe for MAGL displayed slower labeling kinetics ( $t_{1/2}$  of 13 min) with 2  $\mu$ M TCO-TAMRA (Figure S8).<sup>[21]</sup> Together, these experiments illustrate how modifying the structure of tetrazine-coupled probes can be used to tune the IC<sub>50</sub> and labeling kinetics.

## Conclusion

In conclusion, a one-pot method is described for the conversion of (3-methyloxetan-3-yl)methyl carboxylic esters into 3 thiomethyltetrazines that can subsequently serve as a platform for divergent tetrazine synthesis via Pd-catalyzed cross-coupling and the first example of monosubstituted tetrazine synthesis via catalytic thioether reduction. The utility of the method was demonstrated through the synthesis of aliphatic, aromatic and heterocycle substituted tetrazines and to the development and biological evaluation of a new series of tetrazine-coupled probes for MAGL. We anticipate that the greater access to tetrazine-small molecule conjugates available by this method will serve as an enabling tool in chemical biology.

## Acknowledgements

We would like to thank Jeffery Sperry for helpful discussions, Remzi Duzguner for DSC analysis, Edelweiss Evrard for outsourcing support and Lucy Stevens for the *in vitro* potency data on probes **10a-c**. This work was supported by NIH GM132460 and Pfizer. Instrumentation was supported by NIH awards P20GM104316, P30GM110758, S10RR026962, and S10OD016267 and NSF awards CHE-0840401, CHE-1229234, and CHE-1048367.

**Keywords:** tetrazine • bioorthogonal chemistry • minimal probe • divergent synthesis • orthoester

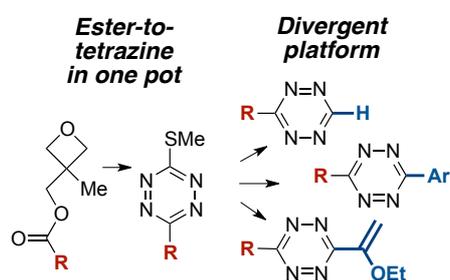
- [1] a) H. C. Hang, C. Yu, D. L. Kato, C. R. Bertozzi, *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 14846-14851; b) D. M. Patterson, L. A. Nazarova, J. A. Prescher, *ACS Chem. Biol.* **2014**, *9*, 592-605.
- [2] a) G. Clavier, P. Audebert, *Chem. Rev.* **2010**, *110*, 3299-3314. b) N. Saracoglu, N. *Tetrahedron* **2007**, *63*, 4199-4236.
- [3] J. Zhang, V. Shukla, D. L. Boger, *J. Org. Chem.* **2019**, *84*, 9397-9445.
- [4] a) R. Rossin, M. S. Robillard, *Curr. Opin. Chem. Biol.* **2014**, *21*, 161-169; b) J. Li, S. Jia, P. R. Chen, *Nat. Chem. Biol.* **2014**, *10*, 1003-1005; c) J. C. T. Carlson, H. Mikula, R. Weissleder, *J. Am. Chem. Soc.* **2018**, *140*, 3603-3612; d) J. L. Tu, M. H. Xu, R. M. Franzini, *ChemBioChem* **2019**, *20*, 1615-1627
- [5] a) K. Wang, A. Sachdeva, D. J. Cox, N. M. Wilf, K. Lang, S. Wallace, R. A. Mehl, J. W. Chin, *Nat. Chem.* **2014**, *6*, 393-403; b) K. Lang, J. W. Chin, *Chem. Rev.* **2014**, *114*, 4764-4806; c) I. Nikić, E. A. Lemke, *Curr. Opin. Chem. Biol.* **2015**, *28*, 164-173.
- [6] a) N. K. Devaraj, S. Hilderbrand, R. Upadhyay, R. Mazitschek, R. Weissleder, *Angew. Chem. Int. Ed.* **2010**, *49*, 2869-2872; b) J. C. T. Carlson, L. G. Meimetis, S. A. Hilderbrand, R. Weissleder, *Angew. Chem. Int. Ed.* **2013**, *52*, 6917-6920; c) H. Wu, J. Yang, J. Šečková, N. K. Devaraj, *Angew. Chem. Int. Ed.* **2014**, *53*, 5805-5809; d) A. Wiczorek, P. Werther, J. Euchner, R. Wombacher, *Chem. Sci.* **2017**, *8*, 1506-1510; e) A. Wiczorek, T. Buckup, R. Wombacher, *Org. Biomol. Chem.* **2014**, *12*, 4177-4185; f) L. G. Meimetis, J. C. T. Carlson, R. J. Giedt, R. H. Kohler, R. Weissleder, *Angew. Chem. Int. Ed.* **2014**, *53*, 7531-7534; g) Y. Lee, W. Cho, J. Sung, E. Kim, S. B. Park, *J. Am. Chem. Soc.* **2018**, *140*, 974-983.
- [7] a) F. C. J. van de Watering, M. Rijpkema, M. Robillard, W. J. G. Oyen, O. C. Boerman, *Front. Med.* **2014**, *1*, 1-11; b) J.-P. Meyer, P. Adumeau, J. S. Lewis, B. M. Zeglis, *Bioconjugate Chem.* **2016**, *27*, 2791-2807; c) M. Wang, R. Vannam, W. D. Lambert, Y. Xie, H. Wang, B. Giglio, X. Ma, Z. Wu, J. Fox, Z. Li, *Chem. Commun.* **2019**, *55*, 2485-2488.
- [8] a) M. Glöckle, W. Kaim, *Angew. Chem. Int. Ed.* **1999**, *38*, 3072-3074; b) W. Kaim, *Coord. Chem. Rev.* **2002**, *230*, 127-139.
- [9] a) H. Zhang, K. T. Dicker, X. Xu, X. Jia, J. M. Fox, *ACS Macro Lett.* **2014**, *3*, 727-731; b) S. Liu, H. Zhang, R. A. Remy, F. Deng, M. E. Mackay, J. M. Fox, X. Jia, *Adv. Mater.* **2015**, *27*, 2783-2790; c) D. L. Alge, M. A. Azagarsamy, D. F. Donohue, K. S. Anseth, *Biomacromolecules* **2013**, *14*, 949-953; d) H. Zhou, J. A. Johnson, *Angew. Chem. Int. Ed.* **2013**, *52*, 2235-2238.
- [10] H. Wu, N. K. Devaraj, *Acc. Chem. Res.* **2018**, *51*, 1249-1259.
- [11] a) T. Cañeque, S. Müller, R. Rodriguez, *Nat. Rev. Chem.* **2018**, *2*, 202-215; b) M. Baalman, M. J. Ziegler, P. Werther, J. Wilhelm, R. Wombacher, *Bioconjugate Chem.* **2019**, *30*, 1405-1414.
- [12] R. Stollé, *J. Prakt. Chem.* **1906**, *73*, 277-287.
- [13] a) A. Pinner, *Ber. Dtsch. Chem. Ges.* **1893**, *26*, 2126-2135; b) K. A. Hofmann, O. Ehrhart, *Ber. Dtsch. Chem. Ges.* **1912**, *45*, 2731-2740; c) T. Curtius, A. Hess, *J. Prakt. Chem.* **1930**, *125*, 40-53; d) J. Yang, M. R. Karver, W. Li, S. Sahu, N. K. Devaraj, *Angew. Chem. Int. Ed.* **2012**, *124*, 5312-5315; e) W. Mao, W. Shi, J. Li, D. Su, X. Wang, L. Zhang, L. Pan, X. Wu, H. Wu, *Angew. Chem. Int. Ed.* **2019**, *58*, 1106-1109; f) S. A. L. Jr, B. D. Johnson, E. Cohen, *J. Heterocycl. Chem.* **1975**, *12*, 1143-1153; g) M. R. Karver, R. Weissleder, S. A. Hilderbrand, *Bioconjugate Chem.* **2011**, *22*, 2263-2270.
- [14] Y. Qu, F. X. Sauvage, G. Clavier, F. Miomandre, P. Audebert, *Angew. Chem. Int. Ed.* **2018**, *57*, 12057-12061.
- [15] a) C. D. Mboyi, C. Testa, S. Reeb, S. Genc, H. Cattey, P. Fleurat-Lessard, J. Roger, J.-C. Hierso, *ACS Catal.* **2017**, *7*, 8493-8501; b) H. Xiong, Y. Gu, S. Zhang, F. Lu, Q. Ji, L. Liu, P. Ma, G. Yang, W. Hou, H. Xu, *Chem. Commun.* **2020**; c) C. Testa, É. Gigot, S. Genc, R. Decréau, J. Roger, J.-C. Hierso, *Angew. Chem. Int. Ed.* **2016**, *55*, 5555-5559; d) C. D. Mboyi, D. Vivier, A. Daher, P. Fleurat-Lessard, H. Cattey, C. H. Devillers, C. Bernhard, F. Denat, J. Roger, J.-C. Hierso, *Angew. Chem. Int. Ed.* **2020**, *59*, 1149-1154.
- [16] a) Z. Novák, A. Kotschy, *Org. Lett.* **2003**, *5*, 3495-3497; b) A. K.-W. Nicolas Leconte, Franck Suzenet, Gérald Guillaumet, *Synlett* **2007**, *2*, 204-210; c) Z. Li, J. Ding, N. Song, J. Lu, Y. Tao, *J. Am. Chem. Soc.* **2010**, *132*, 13160-13161; d) L. Pellegatti, E. Vedrenne, J.-M. Leger, C. Jarry, S. Routier, *Tetrahedron* **2010**, *66*, 4383-4389; e) C. Quinton, V. Alain-Rizzo, C. Dumas-Verdes, G. Clavier, L. Vignau, P. Audebert, *New J. Chem.* **2015**, *39*, 9700-9713; f) A. M. Bender, T. C. Chopko, T. M. Bridges, C. W. Lindsley, *Org. Lett.* **2017**, *19*, 5693-5696.
- [17] a) O. Stetsiuk, A. El-Ghayoury, F. Lloret, M. Julve, N. Avarvari, *Eur. J. Inorg. Chem.* **2018**, *2018*, 449-457; b) D. E. Chavez, D. A. Parrish, L. Mitchell, *Angew. Chem. Int. Ed.* **2016**, *55*, 8666-8669; c) P. Audebert, F. Miomandre, G. Clavier, M.-C. Vernières, S. Badré, R. Méallet-Renault, *Chem. Eur. J.* **2005**, *11*, 5667-5673.
- [18] a) D. E. Chavez, M. A. Hiskey, R. D. Gilardi, *Angew. Chem. Int. Ed.* **2000**, *39*, 1791-1793; b) M. H. V. Huynh, M. A. Hiskey, D. E. Chavez, D. L. Naud, R. D. Gilardi, *J. Am. Chem. Soc.* **2005**, *127*, 12537-12543.
- [19] a) S. D. Schnell, L. V. Hoff, A. Panchagnula, M. H. H. Wurzenberger, T. M. Klapötke, S. Sieber, A. Linden, K. Gademann, *Chem. Sci.* **2020**, *11*, 3042-3047; b) E. Ros, M. Bellido, X. Verdaguer, L. Ribas de Pouplana, A. Riera, *Bioconjugate Chem.* **2020**, *31*, 933-938.
- [20] S. C. Fields, M. H. Parker, W. R. Erickson, *J. Org. Chem.* **1994**, *59*, 8284-8287.

## RESEARCH ARTICLE

- [21] W. D. Lambert, Y. Fang, S. Mahapatra, Z. Huang, C. W. am Ende, J. M. Fox, *J. Am. Chem. Soc.* **2019**, *141*, 17068-17074.
- [22] Z. Wang. Pinner Reaction. In *Comprehensive Organic Name Reactions and Reagents*, John Wiley & Sons, Inc., **2010**, pp. 2237-2240.
- [23] E. J. Corey, N. Raju, *Tetrahedron Lett.* **1983**, *24*, 5571-5574.
- [24] E. S. Scott, L. F. Audrieth, *J. Org. Chem.* **1954**, *19*, 1231-1237.
- [25] R. Selvaraj, J. M. Fox, *Tetrahedron Lett.* **2014**, *55*, 4795-4797.
- [26] a) J. Sauer, *Chem. Heterocycl. Compd.* **1995**, *31*, 1140-1154; b) N. K. Devaraj, R. Weissleder, *Acc. Chem. Res.* **2011**, *44*, 816-827.
- [27] a) T. H. Graham, W. Liu, D.-M. Shen, *Org. Lett.* **2011**, *13*, 6232-6235; b) A. Modak, D. Maiti, *Org. Biomol. Chem.* **2016**, *14*, 21-35; c) T. Matsumura, T. Niwa, M. Nakada, *Tetrahedron Lett.* **2012**, *53*, 4313-4316.
- [28] J. L. Blankman, B. F. Cravatt, *Pharmacol. Rev.* **2013**, *65*, 849-871.
- [29] Jae W. Chang, Micah J. Niphakis, Kenneth M. Lum, Armand B. Cognetta, C. Wang, Megan L. Matthews, S. Niessen, Matthew W. Buczynski, Loren H. Parsons, Benjamin F. Cravatt, *Chem. Biol.* **2012**, *19*, 579-588.
- [30] C. R. Butler, E. M. Beck, A. Harris, Z. Huang, L. A. McAllister, C. W. am Ende, K. Fennell, T. L. Foley, K. Fonseca, S. J. Hawrylik, D. S. Johnson, J. D. Knafels, S. Mente, G. S. Noell, J. Pandit, T. B. Phillips, J. R. Piro, B. N. Rogers, T. A. Samad, J. Wang, S. Wan, M. A. Brodney, *J. Med. Chem.* **2017**, *60*, 9860-9873.
- [31] J. W. Chang, A. B. Cognetta, M. J. Niphakis, B. F. Cravatt, *ACS Chem. Biol.* **2013**, *8*, 1590-1599.

## RESEARCH ARTICLE

## Entry for the Table of Contents



**Divergent Tetrazine Synthesis:** Described is a new strategy for the one-pot synthesis of 3-thiomethyltetrazines from carboxylic ester precursors, providing a platform for the synthesis of unsymmetrical tetrazines via Pd-catalyzed cross-coupling and the first catalytic thioether reduction to access monosubstituted tetrazines.