

DIRECT SCIENCE

Bioorganic & Medicinal Chemistry Letters 13 (2003) 4277-4279

BIOORGANIC & MEDICINAL CHEMISTRY LETTERS

A Simple Stereoselective Synthesis and Biological Evaluation of FR181157: Orally Active Prostacyclin Mimetic

Kouji Hattori,* Seiichiro Tabuchi, Osamu Okitsu and Kiyoshi Taniguchi

Medicinal Chemistry Research Laboratories, Fujisawa Pharmaceutical Co. Ltd, 1-6, Kashima 2-Chome, Yodogawa-Ku, Osaka, Japan

Received 18 August 2003; accepted 29 September 2003

Abstract—Synthetic method of novel prostaglandin (PG) mimetic: FR181175 without PG skeleton are described. The key to success is creation of a chiral epoxide using Sharpless AD reaction with high ee yield. FR181157 shows high potency and agonist efficacy at the IP receptor and has good bioavailability.

© 2003 Elsevier Ltd. All rights reserved.

Introduction

Prostacyclin (PGI₂): 1 is primarily derived from vascular endothelium and plays an extremely important inhibitory role in platelet aggregation and as a vasodilator in maintaining homeostatic circulation.¹ Despite fascinating pharmacological properties, the inherent instability of 1 limits its therapeutic applicability. An Edinburgh University group,² a Bristol-Myers Squibb group,³ and an Ono group⁴ has already disclosed novel PGI₂ analogues without a PG skeleton. These investigation led us to create of novel PGI₂ mimetic with improved on both chemical and metabolic stability. After extensive research, we found the non prostanoid structure FR181157: (S)-4 having potent PGI_2 activity with improved pharmacokinetic property. In order to generate multigram quantities of (S)-4 for continuing biological evaluation and its toxicity, an efficient synthetic route had to be developed. In this paper, we would like to describe a practical and simple stereoselective synthesis and biological activity of (S)-4 (Fig. 1).

Chemistry

Our synthetic strategy is illustrated in Scheme 1. We address to make this compound by especially controlling stereochemistry at a benzyl center and a double bond of a cyclohexene ring. The key substrate that was

0960-894X/\$ - see front matter \odot 2003 Elsevier Ltd. All rights reserved. doi:10.1016/j.bmcl.2003.09.054

the focus for construction of the desired stereochemistry of 4 was a chiral epoxide. A ring opening of the epoxide by a nucleophile creates a chiral center at the benzyl position and hydroxyl function favorable arranged for the control of an *exo*-olefin in the process of elimination.⁵

The required optically active epoxide 8 was obtained via the Sharpless AD reaction and epoxidation shown in Scheme 2. Commercially available 1-cyclohexenylcarboxylic acid 5 was easily transformed into 1-cyclohexenyloxazole 6 by condensation with benzoin followed by treatment with ammonium acetate in acetic acid. Asymmetric dihydroxylation of 6 by the standard procedure⁶ with 1 mol% of AD-mix- α provided the diol



^{*}Corresponding author. Tel.: +81-6-6390-1220; fax: +81-6-6304-5435; e-mail: kouji_hattori@po.fujisawa.co.jp

7 in 98% ee. Treatment by crystallization with hexaneether gave the optically pure product in 80% yield and >99% ee as determined by HPLC analysis with a chiral column (Chiralcel AD). The diols 7 was smoothly converted to the epoxide 8 using the method involving a cyclic acetoxonium intermediate.⁷ The key chiral epoxide 8 was effectively produced on 100–300 g scale without any purification by column chromatography.

The completion of the synthesis of (S)-4 from 8 was accomplished by the route outline in Scheme 3. Ring opening of 8 with methoxybenzylmagnesium chloride (5 equiv) in the presence of a catalytic amount of CuBr provided quantitatively the alcohol compound 9 after purification of short column chromatography. Elimination of water successfully proceeded with the predicted regioselectivity upon treatment with *p*-toluenesulfonic acid (0.2 equiv) under toluene azeotrope. The crude product was purified by crystallization from hexane-ether to obtain the *exo*-olefin 10 as a single isomer in 64% yield and >99% ee (as determined by HPLC). This reaction gave the olefin compounds with a ratio of exo/ $endo = 90:10.^{8}$ The regioselectivity of the obtained olefin compounds may be controlled under kinetic control. When the exo-olefin was treated with p-toluenesulfonic acid under toluene azeotrope, we could not observe isomerization to the endo-olefin. On the other hand, the

Table 1. In vitro effect of FR181157 and the derivatives

	Function assay (IC ₅₀ nM)	Binding assay (K _i nM)	
(S)-4: FR181157	60	54	
(<i>R</i>)-4	110	NT	
14	530	NT	
3 : BMY 42393	3400	NT	
2: Iloprost	2.5	6.5	

treatment of the *endo*-olefin isolated by column chromatography with the same condition could not also occur isomerization. These results suggest that olefin are not prone to rapid equilibration. Enantiomerically pure 10 was subsequently treated with BBr₃ followed by alkylation with ethylbromoacetate gave the ethyl ester 13. Finally, hydrolysis of 13 furnished (S)-4 in 81% yield and >99% ee from 10.⁹

Biological activity

FR181157 possesses potent PGI_2 agonist activity and especially good pharmacokinetic properties. Table 1 was shown the in vitro activity. PGI_2 receptor binding was examined by the conventional ligand binding assay



Scheme 1.



Scheme 2. (a) $SOCl_2$, CH_2Cl_2 ; (b) benzoin, Et_3N , CH_2Cl_2 ; (c) $ACONH_4$, AcOH; (d) AD-*mix*- α , 'BuOH-H₂O; (e) $Me(OMe)_3$, cat. *p*-TsOH, CH_2Cl_2 ; (f) AcBr, CH_2Cl_2 ; (g) K_2CO_3 , MeOH.



Scheme 3. (a) m-MeOBzMgBr, cat. CuBr, THF; (b) cat. p-TsOH, Tol; (c) BBr₃, CH₂Cl₂; (d) BrCH₂CO₂Et, K₂CO₃; DMF; (e) NaOH, EtOH.

Table 2. Pharmacokinetics profiles of FR181157

	(po, fasted)				(iv)		F (%)
	Dose (mg/kg)	C _{max} (ng/mL)	T _{max} (h)	AUC ₀₋₂₄ (ng h/mL)	$\frac{t_{1/2}\beta}{(h)}$	CL _{tot} (mL/min/kg)	
Rat Dog	0.32 0.032	16.4 38.7	0.3 2.3	147.0 391.7	6.6 26	17.7 1.09	50 72

based on the displacement of [3H]-Iloprost from the cloned human IP receptor.10 IC50 values of the functional assay was obtained by measuring inhibition of ADP-induced platelet aggregation using human platelet rich plasma. FR181157 exhibited high binding affinity for the IP receptor with a K_i of 54 nM and anti-aggregative potency with an IC₅₀ of 60 nM. The enatiomer isomer (R)-4 was 2-fold less potent, and also the *endo*olefin 14 was 9-fold less potent under the same condition. It is well known that this class of PGI₂ mimetic has species difference.¹ FR181157 was also observed the same result, inhibition of ADP-induced platelet aggregation using rat and dog platelet rich plasma was 20-fold less potent than human (rat: $IC_{50} = 1.2 \mu M$, dog: $IC_{50} = 1.3 \mu M$). Table 2 was shown the pharmacokinetic profiles of rat and dog. FR181157 without a PG skelton displayed a good oral bioavailability (rat: F = 50%, dog: F = 72%) and long duration time in the both species. The detailed pharmacological and pharmacokinetic properties of FR181157 will be published in future.

Conclusion

In this communication, we have reported an operationally simple method for the highly stereoselective synthesis and a biological evaluation of FR181157, a potent and orally active prostacylin mimetic. This is a 12step synthesis from commercially available starting materials, and involves only simple purification to give a 100-g scale. This route should be suitable for industrial order.

Acknowledgements

We express our thanks to Dr. Hirokazu Tanaka, Dr. Kazuo Sakane, and Dr. Hisashi Takasugi for their practical guidance and Dr. David Barrett for his critical reading of the manuscript.

References and Notes

1. (a) Wise, H.; Jones, R. L. *TiPS* **1996**, *17*, 17. (b) Meanwell, N. A.; Romine, J. L.; Seiler, S. M. *Drugs Future* **1994**, *19*, 361.

(c) Vane, J., O'Grady, J., Eds. Therapeutic Applications of Prostaglandins; Edward Arnold: London, 1993. (d) Dusting, G. J.; MacDonald, P. S. Pharmacol. Ther. 1990, 48, 323. 2. Muir, G.; Jones, R. L.; Will, S. G.; Winwick, T.; Peesapatil, V.; Wilson, N. H.; Griffiths, N.; Nicholson, W. V.; Taylor, P.; Sawyer, L.; Blake, A. J. Eur. J. Med. Chem. 1993, 28, 609. 3. (a) Meanwell, N. A.; Rosenfeld, M. J.; Wright, J. J. K.; Brassard, C. L.; Buchanan, J. O.; Federici, M. E.; Fleming, J. S.; Seiler, S. M. J. Med. Chem. 1992, 35, 389. (b) Meanwell, N. A.; Rosenfeld, M. J.; Trehan, A. K.; Wright, J. J. K.; Brassard, C. L.; Buchanan, J. O.; Federici, M. E.; Fleming, J. S.; Gamberdella, M.; Zavoico, G. B.; Seiler, S. M. J. Med. Chem. 1992, 35, 3483. (c) Meanwell, N. A.; Rosenfeld, M. J.; Trehan, A. K.; Romine, J. L.; Wright, J. J. K.; Brassard, C. L.; Buchanan, J. O.; Federici, M. E.; Fleming, J. S.; Gamberdella, M.; Zavoico, G. B.; Seiler, S. M. J. Med. Chem. 1992, 35, 3498. (d) Meanwell, N. A.; Rosenfeld, M. J.; Wright, J. J. K.; Brassard, C. L.; Buchanan, J. O.; Federici, M. E.; Fleming, J. S.; Gamberdella, M.; Hartl, K. S.; Zavoico, G. B.; Seiler, S. M. J. Med. Chem. 1993, 36, 3871. (e) Meanwell, N. A.; Romine, J. F.; Rosenfeld, M. J.; Martin, S. W.; Trehan, A. K.; Wright, J. J. K.; Malley, M. F.; Gougoutas, J. Z.; Brassard, C. L.; Buchanan, J. O.; Federici, M. E.; Fleming, J. S.; Gamberdella, M.; Hartl, K. S.; Zavoico, G. B.; Seiler, S. M. J. Med. Chem. 1993, 36, 3884.

4. (a) Hamanaka, N.; Takahashi, K.; Nagao, Y.; Toritsu, K.; Tokumoto, H.; Kondo, K. *Bioorg. Med. Chem. Lett.* **1995**, *5*, 1065. (b) Hamanaka, N.; Takahashi, K.; Nagao, Y.; Toritsu, K.; Takada, H.; Tokumoto, H.; Kondo, K. *Bioorg. Med. Chem. Lett.* **1995**, *5*, 1071. (c) Hamanaka, N.; Takahashi, K.; Nagao, Y.; Toritsu, K.; Tokumoto, H.; Kondo, K. *Bioorg. Med. Chem. Lett.* **1995**, *5*, 1077. (d) Hamanaka, N.; Takahashi, K.; Nagao, Y.; Toritsu, K.; Tokumoto, H.; Kondo, K. *Bioorg. Med. Chem. Lett.* **1995**, *5*, 1083. (e) Hamanaka, N.; Takahashi, K.; Nagao, Y.; Toritsu, K.; Shigeoka, S.; Hamada, S.; Kato, H.; Tokumoto, H.; Kondo, K. *Bioorg. Med. Chem. Lett.* **1995**, *5*, 1083.

5. (a) McNiven, N. L.; Read, J. J. Chem. Soc. **1952**, 153. (b) Hughes, E. D.; Wilby, J. J. Chem. Soc. **1960**, 4094.

Sharpless, K. B.; Amberg, W.; Bennani, Y. L.; Crispino,
G. A.; Hartung, J.; Jeong, K.-S.; Kwong, H.-L.; Morikawa,
K.; Wang, Z.-M.; Xu, D.; Zhang, X.-L. J. Org. Chem. 1992,
57, 2768.

7. Kolb, H. C.; Sharpless, K. B. *Tetrahedron* **1992**, *48*, 10515. 8. The ratio of the crude product was determined by ¹H NMR. *endo*-Olefin was completely removed by crystallization. ¹H NMR of benzyl proton in CDCl₃, **10**: δ 2.53 (1H, dd, *J*=10.2, 12.8 Hz), 2.31 (1H, dd, *J*=3.2, 12.8 Hz), **12**: δ 4.09 (2H, s).

9. The assignment of the absolute configuration of FR181157 was determined by X-ray structural analysis. (*S*)-4 $[\alpha]_{D}^{24}$ +93.0 (*c* 0.20, MeOH); HPLC (chiralcel AGP, 10 cm×0.4 cm I.D., flow rate 0.8 mL/min, 20% acetonitrile/0.02 M phosphate buffer) R_t =4.0 min (>99%). [(*R*)-4: R_t =6.0 min]. This compound should be handled with extreme caution because of PGI₂ agonist activity and high permeability of skin.

10. Katsuyama, M.; Sugimoto, Y.; Namba, T.; Irie, A.; Negishi, M.; Narumiya, S.; Ichikawa, A. *FEBS Lett.* **1994**, *344*, 74. Boie, Y.; Ruchmore, T. H.; Darmon-Goodwin, A.; Grygorczyk, R.; Slipetz, D. M.; Metters, K.; Abramovitz, M. *J. Biol. Chem.* **1994**, *269*, 12173.