## Stereoselective Synthesis of a Potent Human NK<sub>1</sub> Receptor Antagonist via Acyl-Claisen Rearrangement

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This work is dedicated to Professor Jerzy Wicha on the occasion of his 70th birthday.

**Abstract:** Stereoselective synthesis of the tetrahydropyran derivative **1** is reported. Diastereoselective acyl-Claisen rearrangement was employed for formation of C3 and C4 chiral centres on the tetrahydropyran ring.

**Key words:** acyl-Claisen rearrangement, asymmetric synthesis, total synthesis, stereoselectivity, NK<sub>1</sub> receptors

Neurokinin-1 (NK<sub>1</sub>) receptor antagonists are of continuing interest since the natural ligand for the NK1 receptor -Substance P - has been implicated in the pathophysiology of a wide range of disease conditions including neurogenic inflammation, transmission of pain, emesis and depression.<sup>1,2</sup> Recently, the Merck NK<sub>1</sub> antagonist, aprepitant (Emend®), has been approved for the prevention of acute and delayed chemotherapy-induced nausea and vomiting (Figure 1). Since 1991, when the first non-peptidic antagonist was reported,<sup>3</sup> numerous selective and structurally diverse NK1 ligands have been identified and subsequently developed.<sup>1,4</sup> Recently, syntheses of *trans,trans*-substituted tetrahydropyran derivatives have been described.4g,h In the search for selective hNK<sub>1</sub> receptor antagonists, we were interested in examining 3,4,5-trans,trans-trisubstituted tetrahydropyran derivatives. Compound 1 was identified for synthesis and biological evaluation.





In this communication, we report our approaches to the stereoselective synthesis of a 3,4,5-trisubstituted tetrahydropyran **1**, a potent NK<sub>1</sub> receptor antagonist.

Conceptually, 1 can be disconnected into two fragments, a *trans* lactone 2 and the chiral spiropiperidine 3 (Scheme 1).

We envisaged that the optically pure *trans* lactone 2 would be accessible from the chiral allyl ester 4 through the diastereoselective Ireland–Clasien rearrangement<sup>5</sup> and subsequent lactonisation of the resulting hydroxy acid. In turn, 4 could be assembled from commercially



**Scheme 1** Ar = 3,5-bis(trifluoromethyl)phenyl

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Scheme 2 Reagents and conditions: a) tert-butyl bromoacetate, NaH, THF, 90%; b) TFA,  $CH_2Cl_2$ , 56%; c) (COCl)<sub>2</sub>,  $CH_2Cl_2$ , cat. DMF, quant.

available compounds: bromoacetate **5**, (R)-1-[3,5-bis(tri-fluoromethyl)phenyl]ethanol (**6**), methyl acrylate (**7**) and 4-fluorobenzaldehyde (**8**).

Chiral alcohol **6** was chosen as a starting nonracemic building block. We assumed that the chiral benzylic centre might be a stereocontrolling element in the [3+3] sigmatropic rearrangement and would facilitate definition of stereochemistry of new chiral centres on the tetrahydropyran ring.

The synthesis started with alkylation of the commercially available R alcohol **6** with *tert*-butyl bromoacetate followed by removal of the *tert*-butyl group providing the acid **9** in 50% overall yield (Scheme 2).



Scheme 3 Reagents and conditions: a) methyl acrylate, DABCO, 60 °C, 66–70%; b) PBr<sub>3</sub>, Et<sub>2</sub>O, 0 °C to r.t.; c) Et<sub>3</sub>N, HCO<sub>2</sub>H, MeCN, reflux; d) concd HCl, MeOH, 62–70% (3 steps); e) TBSCl, DMF, imidazole, quant.; f) DIBAL-H, CH<sub>2</sub>Cl<sub>2</sub>, 86%; g) **13a**, **9**, EDC, Et<sub>3</sub>N, DMAP, CH<sub>2</sub>Cl<sub>2</sub>, 86%. Ar = 3,5-bis(trifluoromethyl)phenyl.

Having prepared the acid 9, our next problem was selective preparation of Z-allyl alcohols 13 and 13a (Scheme 3). This was accomplished by a six-step sequence beginning with the Baylis–Hillman reaction of methyl acrylate and corresponding benzaldehyde (8, 8a) followed by treatment with PBr<sub>3</sub> in diethyl ether to afford allyl bromides 11 and 11a.<sup>6</sup> Displacement of bromine in 11 and 11a with formate and hydrolysis of the resulting formic acid ester gave allylic alcohols 12 and 12a in 40–

50% overall yields.<sup>6</sup> The hydroxy group in **12** and **12a** was protected as a *tert*-butyldimethylsilyl ether and the ester reduced using DIBAL-H to give Z-allylic alcohols 13 and 13a in 86% yield. Finally, the standard coupling of the alcohol 13a with the acid 9 provided the allylic ester 14 in 86% yield. The Ireland-Claisen rearrangement of the lithium enolate of 14 in the presence of trimethylsilyl chloride yielded a mixture of isomeric acids that were converted to methyl esters on treatment with trimethylsilyldiazomethane (Scheme 4). <sup>1</sup>H NMR analysis of the crude mixture of esters indicated the diastereoisomeric ratio of mixture **15a**:**15b**:**15c**:**15d** = 2:10:1:2. To establish relative stereochemistry of products of the rearrangement, the mixture of esters 15a-d was converted into a mixture of tetrahydropyrans **17a–d** as outlined in Scheme 4. Isomers were separated and the relative stereochemistry of diastereoisomers was assigned by <sup>1</sup>H NMR/NOE experiments. Disappointingly, we found that the undesired trans isomer, 17b, was a major product of this sequence.



Scheme 4 Reagents and conditions: a) LiHMDS, TMSCl, THF, then TMS-diazomethane, Et<sub>2</sub>O, MeOH, 65%; b) DIBAL-H, CH<sub>2</sub>Cl<sub>2</sub>, 77%; c) TsCl, Et<sub>3</sub>N, DMAP, CH<sub>2</sub>Cl<sub>2</sub>, 90%; d) TBAF, THF; e) BuLi, THF, 64% (two steps). Ar = 3,5-bis(trifluoromethyl)phenyl.

To overcome this obstacle, we turned our attention towards the acyl-Claisen rearrangement.<sup>7</sup> Recently, Mac-Millan and co-workers reported an enantioselective Lewis acid catalysed acyl-Claisen rearrangement of allylic amines and ketenes (Scheme 5).<sup>8</sup> Ketenes were generated in situ from corresponding acyl chlorides.





We have hypothesised that having a chiral centre on the allylic amine might invert stereoselectivity in the sigmatropic rearrangement. With the allylic alcohol **13a** in hand, we were in a position to test this hypothesis and briefly investigate this substrate control in the acyl-Claisen reaction.

We began with the synthesis of allyl amines 18a and 18b (Scheme 6). For our studies, we selected (R)-O-methylprolinol as a chiral amine and morpholine as an achiral analogue. Thus, treatment of the alcohol 13a with methanesulphonyl chloride followed by the corresponding amine afforded 18a<sup>9</sup> and 18b in 61-67% yield. Reaction of the allyl amine 18a and benzyloxyacetyl chloride in the presence of *i*-Pr<sub>2</sub>EtN and catalytic TiCl<sub>4</sub>-THF<sub>2</sub> yielded a mixture of two major syn diastereoisomers in the ratio **19a:19b** = 1:1. Pleasingly, reaction of the chiral acid chloride 10 and 18a proceeded smoothly to give a mixture diastereoisomers **20a–d** in of four the ratio **20a:20b:20c:20d** = 18:3:5:2 with the major *syn* diastereoisomer 20a<sup>10</sup> having the correct stereochemistry at C2 and C3 centres.<sup>11</sup> Reaction of the morpholine 18b with 10 furnished with a mixture of four morpholine amides 21a-d in the ratio **21a:21b:21c:21d** = 4.2:3:1:1 showing that both chiral centres are required for selectivity.



Scheme 6 Reagents and conditions: a) MsCl, Et<sub>3</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, 0 °C, then R<sup>1</sup>R<sup>2</sup>NH, 61–67%; b) BnOCH<sub>2</sub>COCl or **10**, cat. TiCl<sub>4</sub>–THF<sub>2</sub>, *i*-Pr<sub>2</sub>EtN, CH<sub>2</sub>Cl<sub>2</sub>, –10 °C to 10 °C (58% for **20a**). Ar = 3,5-bis(trifluoromethyl)phenyl.

The morpholine amide 22 was prepared from the alcohol 13 in a similar way as described for 21a (Scheme 7). Acid-catalysed TBS-group deprotection–lactonisation on the amide 22 followed by reduction to the corresponding lactol using DIBAL-H gave a mixture of lactols which were reduced to the tetrahydropyran 23 using triethyl-silane in the presence of boron trifluoride–diethyl ether complex. Hydroboration–oxidation of the double bond in 23 furnished a 2:1 mixture of hydroxymethyl epimers in favour of the undesired 5R epimer. This mixture was oxidised to the aldehyde under Swern conditions and the mixture of aldehydes was allowed to equilibrate in the presence of catalytic 1,8-diazabicyclo[5.4.0]undec-1-ene (DBU) in dichloromethane providing the required 5S aldehyde 24 (de 94% by <sup>1</sup>H NMR) in 81% yield.

Spirocyclic piperidine **27** was prepared starting from ethyl isonipecotate **25** (Scheme 8). Aldol condensation of the lithium enolate of **25** with (*tert*-butyldimethylsilyl-oxy)acetaldehyde followed by reduction of the diester

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Scheme 7 Reagents and conditions: a) MsCl, Et<sub>3</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, 0 °C, then morpholine, 64%; b) **10**, cat. TiCl<sub>4</sub>–THF<sub>2</sub>, *i*-Pr<sub>2</sub>EtN, CH<sub>2</sub>Cl<sub>2</sub>, -70 °C to r.t., 27%; c) *p*-TSA, MeOH, PhMe, 71%; d) DIBAL-H, CH<sub>2</sub>Cl<sub>2</sub>, -78 °C, quant.; e) Et<sub>3</sub>SiH, BF<sub>3</sub>·OEt<sub>2</sub>, CH<sub>2</sub>Cl<sub>2</sub>, 93%; f) BH<sub>3</sub>·THF, -78 °C to r.t.; then H<sub>2</sub>O<sub>2</sub>, NaOH, 81%; g) (COCl)<sub>2</sub>, DMSO, Et<sub>3</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, -60 °C to 0 °C; h) DBU (cat.), CH<sub>2</sub>Cl<sub>2</sub>, 99% (two steps). Ar = 3,5-bis(trifluoromethyl)phenyl.

using lithium borohydride, removal of the TBS group with tetrabutylammonium fluoride and subsequent cycloetherification of the resulting triol under Mitsunobu conditions afforded the racemic alcohol **26** in 44% overall yield.



Scheme 8 Reagents and conditions: a) TBSOCH<sub>2</sub>CHO, LiHMDS, THF, 76%; b) LiBH<sub>4</sub>, THF; then TBAF, THF, 76%; c) DEAD, Ph<sub>3</sub>P, THF, 76%; d) RCl, Et<sub>3</sub>N, DMAP, CH<sub>2</sub>Cl<sub>2</sub>, 38%, de 98%; e) Pd/C, H<sub>2</sub>, TFA, EtOH, quant.

Acylation of **26** with (–)-camphanic acid chloride and subsequent crystallisation of the mixture of diasteroisomeric esters from methanol–water gave the ester **27** as a white solid (38% yield, de 98%).<sup>12</sup> Removal of the benzyl carbamate provided the spirocyclic piperidine **28** in quantitative yield.

Finally, reductive amination of the aldehyde 24 with piperidine 28 and sodium triacetoxyborohydride followed by saponification of the camphanate ester furnished  $1^{13}$  in



Scheme 9 *Reagents and conditions*: a) NaHB(OAc)<sub>3</sub>, DCE; b) NaOH, MeOH–H<sub>2</sub>O, 50%. Ar = 3,5-bis(trifluoromethyl)phenyl.

50% yield (Scheme 9). The tetrahydropyran **1** was tested in the NK<sub>1</sub> receptor binding affinity assay in vitro<sup>14</sup> and it was found to antagonise the human NK<sub>1</sub> receptor with  $IC_{50} = 0.15$  nM.

In conclusion, the stereoselective synthesis of novel NK<sub>1</sub> antagonist based upon the tetrahydropyran framework was developed. Diastereoselectivity in the acyl-Claisen sigmatropic rearrangement of **18a,b** and **10** was investigated allowing inversion of the distereoselective outcome of Ireland–Claisen rearrangement of **14**. The synthesis of tetrahydropyran **1** was accomplished and the binding affinity of **1** at the human NK<sub>1</sub> receptor was determined. The tetrahydropyran derivative **1** exhibited excellent binding affinity at the human NK<sub>1</sub> (IC<sub>50</sub> 0.15 nM).

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## **References and Notes**

- (1) Seward, E. M.; Swain, C. J. *Expert Opin. Ther. Pat.* **1999**, *9*, 571.
- (2) Kramer, M. S.; Cutler, N.; Feighner, J.; Shrivastava, R.; Carman, J.; Sramek, J. J.; Reines, S. A.; Liu, G.; Snavely, D.; Wyatt-Knowles, E.; Hale, J. J.; Mills, S. G.; MacCoss, M.; Swain, C. J.; Harrison, T.; Hill, R. G.; Hefti, F.; Scolnick, E. M.; Cascieri, M. A.; Chicchi, G. G.; Sadowski, S.; Williams, A. R.; Hewson, L.; Smith, D.; Carlson, E. J.; Hargreaves, R. J.; Rupniak, N. M. J. Science **1998**, 281, 1640.
- (3) Snider, R. M.; Constantine, J. W.; Lowe, J. A. III; Longo, K. P.; Lebel, W. S.; Woody, H. A.; Drozda, S. E.; Desai, M. C.; Vinick, F. J.; Spencer, R. W.; Hess, H.-J. *Science* 1991, 251, 435.
- (4) For recent advances, see: (a) Shaw, D.; Chicchi, G. G.; Elliott, J. M.; Kurtz, M. M.; Morrison, D.; Ridgill, M. P.; Szeto, N.; Watt, A. P.; Williams, A. R.; Swain, C. J. *Bioorg. Med. Chem. Lett.* 2001, *11*, 3031. (b) Elliott, J. M.; Castro, J. L.; Chicchi, G. G.; Cooper, L. C.; Dinnell, K.; Hollingworth, G. J.; Ridgill, M. P.; Rycroft, W.; Kurtz, M. M.; Shaw, D. E.; Swain, C. J.; Tsao, K.-L.; Yang, L. *Bioorg. Med. Chem. Lett.* 2002, *12*, 1755. (c) Cooper, L. C.; Carlson, E. J.; Castro, J. L.; Chicchi, G. G.; Dinnell, K.; Di Salvo, J.; Elliott, J. M.; Hollingworth, G. J.; Kurtz, M. M.;

Ridgill, M. P.; Rycroft, W.; Tsao, K.-L.; Swain, C. J. Bioorg. Med. Chem. Lett. 2002, 12, 1759. (d) Seward, E. M.; Carlson, E.; Harrison, T.; Haworth, K. E.; Herbert, R.; Kelleher, F. J.; Kurtz, M. M.; Moseley, J.; Owen, S. N.; Owens, A. P.; Sadowski, S. J.; Swain, C. J.; Williams, B. J. Bioorg. Med. Chem. Lett. 2002, 12, 2515. (e) Williams, B. J.; Cascieri, M. A.; Chicchi, G. G.; Harrison, T.; Owens, A. P.; Owen, S. N.; Rupniak, N. M. J.; Tattersall, F. D.; Williams, A.; Swain, C. J. Bioorg. Med. Chem. Lett. 2002, 12, 2719. (f) Gale, J. D.; O'Neill, B. T.; Humphrey, J. M. Expert Opin. Ther. Pat. 2001, 11, 1837. (g) Huffman, M. A.; Smitrovich, J. H.; Rosen, J. D.; Boice, G. N.; Qu, C.; Nelson, T. D.; McNamara, J. M. J. Org. Chem. 2005, 70, 4409. (h) Nelson, T. D.; Rosen, J. D.; Smitrovich, J. H.; Payack, J.; Craig, B.; Matty, L.; Huffman, M. A.; McNamara, J. Org. Lett. 2005, 7, 55.

- (5) (a) Pereira, S.; Srebnik, M. Aldrichimica Acta 1993, 26, 17.
  (b) Enders, D.; Knopp, M.; Schiffers, R. Tetrahedron: Asymmetry 1996, 7, 1847.
- (6) Beltaief, I.; Hbaieb, S.; Besbes, R.; Amri, H.; Villieras, M.; Villieras, J. Synthesis 1998, 1765.
- (7) Gonda, J. Angew. Chem. Int. Ed. 2004, 43, 3516.
- (8) (a) Yoon, T. P.; Dong, V. M.; MacMillan, D. W. C. J. Am. Chem. Soc. 1999, 121, 9726. (b) Yoon, T. P.; MacMillan, D. W. C. J. Am. Chem. Soc. 2001, 123, 2911.
- (9) Experimental Procedure for the Preparation of 18a. Methanesulphonyl chloride (0.12 mL, 1.5 mmol) was added dropwise to an ice-bath-cooled, stirring mixture of 13a (280 mg, 1 mmol), Et<sub>3</sub>N (0.4 mL, 2.8 mmol) and CH<sub>2</sub>Cl<sub>2</sub> (2 mL). The mixture was stirred for 90 min and (R)-2-(methoxymethyl)pyrrolidine (0.2 mL, 1.6 mmol) was added. The mixture was stirred for 2 h and poured onto a sat. aq solution of NaHCO<sub>3</sub>. The mixture was extracted into CH<sub>2</sub>Cl<sub>2</sub>. The organic extract was dried (Na2SO4) and concentrated. The residue was purified on silica gel (CH<sub>2</sub>Cl<sub>2</sub>-2 M NH<sub>3</sub> in MeOH, 0–10%) to give the amine 18a (230 mg, 61%). <sup>1</sup>H NMR (360 MHz, CDCl<sub>3</sub>):  $\delta = 0.04$  (6 H, s), 0.90 (9 H, s), 1.61–1.77 (3 H, m), 1.86–2.01 (1 H, m), 2.21 (1 H, q, J = 8.5 Hz), 2.67 (1 H, m), 2.95 (1 H, d, J = 13.2 Hz), 3.04 (1 H, m), 3.25 (1 H, dd, J = 7.4, 9.1 Hz), 3.35 (3 H, s), 3.46 (1 H, dd, J = 4.5, 9.3 Hz), 3.74 (1 H, d, J = 13.1 Hz), 4.26 (2 H, s), 6.58 (1 H, s), 7.29-7.31 (5 H, m).
- (10) Experimental Procedure for the Preparation of 20a. TiCl<sub>4</sub>-THF<sub>2</sub> (33 mg, 0.1 mmol) was added to a stirred mixture of 18a (375 mg, 1 mmol), *i*-Pr<sub>2</sub>EtN (0.3 mL, 1.7 mmol) at -10 °C followed by 10 (510 mg, 1.5 mmol). The mixture was stirred at -10 °C for 30 min and warmed to 10 °C over 1 h. The mixture was treated with 1 M aq NaOH and the mixture was extracted into CH<sub>2</sub>Cl<sub>2</sub>. The organic extract was dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated. The residue (a mixture of 20a:20b:20c:20d in the ratio 18:3:5:2) was purified on silica gel (i-hexane-EtOAc, 5-35%) to give the amide 20a (390 mg, 58%, 5:1 mixture of rotamers). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, a major rotamer):  $\delta = -0.07$  (3 H, s), -0.06 (3 H, s), 0.83 (9 H, s), 1.31 (3 H, d, J = 6.4 Hz), 1.64– 2.00 (4 H, m), 3.26 (1 H, m), 3.29 (3 H, s), 3.33 (1 H, dd, *J* = 6.5, 9.3 Hz), 3.52 (1 H, m), 3.83–3.97 (3 H, m), 4.20 (1 H, m), 4.36 (1 H, d, J = 9.3 Hz), 4.55 (1 H, q, J = 6.6 Hz), 4.97 (1 H, s), 5.15 (1 H, s), 7.20-7.26 (5 H, m), 7.44 (2 H, s), 7.74 (1 H, s).
- (11) Diastereomeric products of the acyl-Claisen reaction were separable by flash chromatography and converted into the corresponding lactones **17a–d** as outlined in Scheme 4.
- (12) Diasteromeric excess was determined by HPLC analysis. The absolute stereochemistry of C3 centre of tetrahydrofuran ring was determined by X-ray analysis of close analogue of 1.

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- (13) Experimental Procedure for the Preparation of 1. Sodium triacetoxyborohydride (91 mg, 0.43 mmol) was added to a stirred mixture of 24 (100 mg, 0.215 mmol) and 28 (73 mg, 0.216 mmol) and DCE (0.5 mL). The mixture was stirred for 60 min and treated with a sat. aq solution of NaHCO<sub>3</sub> and extracted with CH<sub>2</sub>Cl<sub>2</sub>. The organic extract was dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated. The mixture was treated with MeOH (2 mL) and 4 M aq NaOH (0.3 mL) was added dropwise. The mixture was stirred for 15 min, diluted with H<sub>2</sub>O and extracted into CH<sub>2</sub>Cl<sub>2</sub>. The organic extract was dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated. The residue was purified on silica gel (CH<sub>2</sub>Cl<sub>2</sub>-MeOH, 0–10%) to give 1 (66 mg, 50%). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta = 1.30$  (3 H, d, J = 6.4
- Hz), 1.38-1.50 (3 H, m), 1.77 (1 H, dd, J = 3.0, 12.5 Hz), 1.81 (1 H, m), 1.97 (1 H, t, J = 10.7 Hz), 2.01-2.11 (2 H, m), 2.12-2.20 (1 H, m), 2.30 (1 H, t, J = 10.6 Hz), 2.40 (1 H, m), 3.12 (1 H, t, J = 11.5 Hz), 3.25 (1 H, t, J = 10.8 Hz), 3.42 (1 H, dt, J = 4.7, 9.8 Hz), 3.52 (1 H, AB, J = 8.6 Hz), 3.57 (1 H, AB, J = 8.6 Hz), 3.64 (1 H, dd, J = 2.2, 10.1 Hz), 3.88 (1 H, m), 4.01 (1 H, dd, J = 4.6, 10.1 Hz), 4.21 (1 H, dd, J = 4.5, 11.8 Hz), 4.25 (1 H, dd, J = 4.6, 11.0 Hz), 4.27 (1 H, q, J = 6.5 Hz), 6.83 (2 H, t, J = 8.9 Hz), 6.93 (2 H, m), 7.20 (2 H, s), 7.66 (1 H, s).
- (14) Cascieri, M. A.; Ber, E.; Fong, T. M.; Sadowski, S.; Bansal, A.; Swain, C. J.; Seward, E. M.; Frances, B.; Burns, D.; Strader, C. D. *Mol. Pharmacol.* **1992**, *47*, 458.