

Contents lists available at ScienceDirect

Bioorganic & Medicinal Chemistry Letters

journal homepage: www.elsevier.com/locate/bmcl

Synthesis of 3,6-diazabicyclo[3.1.1]heptanes as novel ligands for neuronal nicotinic acetylcholine receptors

Gabriele Murineddu ^{a,*}, Caterina Murruzzu ^a, Maria M. Curzu ^a, Giorgio Chelucci ^b, Cecilia Gotti ^c, Annalisa Gaimarri ^c, Laura Legnani ^d, Lucio Toma ^d, Gerard A. Pinna ^a

^a Dipartimento Farmaco Chimico Tossicologico, Università di Sassari, via F. Muroni 23/A, 07100 Sassari, Italy

^b Dipartimento di Chimica, Università di Sassari, via Vienna 2, 07100 Sassari, Italy

^c CNR, Istituto di Neuroscienze, Farmacologia Cellulare e Molecolare, Dipartimento di Farmacologia Medica, Università di Milano, via Vanvitelli 32, 20129 Milano, Italy

^d Dipartimento di Chimica Organica, Università di Pavia, via Taramelli 10, 27100 Pavia, Italy

ARTICLE INFO

Article history: Received 10 September 2008 Revised 30 September 2008 Accepted 2 October 2008 Available online 5 October 2008

Keywords: 3,6-Diazabicyclo[3.1.1]heptanes Neuronal nicotinic acetylcholine receptors Binding affinity Modeling

ABSTRACT

A series of novel 3,6-diazabicyclo[3.1.1]heptane derivatives **4a–f** was synthesized and their affinity and selectivity towards $\alpha 4\beta 2$ and $\alpha 7$ nAChR subtypes were evaluated. The results of the current study revealed a number of compounds (**4a**, **4b** and **4c**) having a very high affinity for $\alpha 4\beta 2$ (K_i at $\alpha 4\beta 2$ ranging from 0.023 to 0.056 nM) versus $\alpha 7$ nAChR subtypes; among these compounds, the 3-(6-bromopyridin-3-yl)-3,6-diazabicyclo[3.1.1]heptane **4c** was found to be the most $\alpha 7\alpha 4\beta 2$ selective term in receptor binding assays ($\alpha 7\alpha 4\beta 2 = 1295$). Moreover, compound **4d** also had high affinity for the $\alpha 4\beta 2$ nAChR subtype ($K_i = 1.2$ nM) with considerably high selectivity ($\alpha 7/\alpha 4\beta 2 = 23300$).

© 2008 Elsevier Ltd. All rights reserved.

Inflammatory disease and neuropathic insults are often accompanied by severe pain states (physiological, inflammatory, and neuropathic) that can be treated with commonly prescribed analgesic agents, such as non-steroidal anti-inflammatory drugs (NSA-IDs) and opioids.¹ Unfortunately, the therapeutic potential of these powerful drugs has been limited by safety concerns, as well as by unpleasant side effects with consequent poor patient compliance.^{2–5}

Thus, the treatment of various form of pain is a hugely unmet medical need, and the search for novel and efficacious analgesics with improved therapeutic index receives considerable attention.^{6,7}

Among a number of novel approaches to pain relief currently under investigation, nicotinic acetylcholine receptors (nAChRs) hold considerable potential as therapeutic targets for the development of analgesic drugs.^{8,9} The neuronal nicotinic acethylcholine receptors are prototypic ligand-gated ion channel receptors that are widely expressed through the central and peripheral nervous system and mediate fast synaptic transmission.⁸ Neuronal nAChRs are pentameric proteins comprising either combinations of two different types of subunit (α and β) or five copies of the same α subunit symmetrically arranged around a central ion pore.¹⁰ Nine different types of α -subunit ($\alpha 2-\alpha 10$) and three kinds of β -subunits ($\beta 2-\beta 4$) have been cloned and characterized. The multiple

* Corresponding author. E-mail address: muri@uniss.it (G. Murineddu).

0960-894X/\$ - see front matter © 2008 Elsevier Ltd. All rights reserved. doi:10.1016/j.bmcl.2008.10.002

nAChR subunits form a plethora of different receptor subtypes among which the most abundant in the CNS are the $\alpha 4\beta 2^*$ and the $\alpha 7^*$ receptors (the asterisks indicate the potential presence of other subunits), whereas the $\alpha 3\beta 4$ receptor is the predominant subtype at ganglionic synapses.^{11,12}

The neuronal nAChRs are involved in a wide range of physiological and pathophysiological processes, and they have been proposed as potential therapeutic targets in a number of neurodegenerative and psychiatric disorders, in various form of pain, and in nicotinic addiction.^{8,13–15}

The $\alpha 4\beta 2$ receptor subtype has generated significant interest among academic and industrial researchers as a result of its role in pain reflexes.^{16,17} Thus, the generation of potent $\alpha 4\beta 2$ nAChR subtype selective ligands has been a goal for the advancement of novel nAChR-based analgesics with a minimum of side effects.⁹

The alkaloid epibatidine (1), the *exo*-2-(6-chloropyridin-3-yl)-7azabicyclo[2.2.1]heptane, isolated from the skin of the ecuadorian poison frog *Epidobates tricolour*,¹⁸ was identified as a potent ligand of α 4 β 2 receptors having powerful analgesic activity.

Because of the identification of various adverse effects of **1** on CNS responses and respiratory, gastrointestinal and cardiovascular function, synthetic modifications of its structure have been performed in order to improve potency and selectivity while reducing the toxicity.

These endeavors, based upon examination of the two key structural elements of **1**, that is, the (i) azabicyclo amine moiety and the (ii) pyridine heterocycle, led to a number of promising substances, many of them closely related but also not related to epibatidine. For example, tebanicline (2) described as an azetidine bioisoster of **1** with (*R*)-absolute configuration, retained a potent nAChR agonism with moderate enhancement of $\alpha 4\beta 2$ nAChR subtype selectivity as compared with 1. Tebanicline showed analgesic action in a spectrum of acute and chronic nociceptive models as well as in neuropathic pain but with only a modest therapeutic index.^{19–22}

Attempts to circumvent these problems induced W.H. Brunelle and coworkers to synthesize a series of N-(3-pyridinyl)-bridged bicyclic diamine derivatives as novel bridged nicotinoid, of which 2-(6-chloropyridin-3-yl)-2,5-diazabicyclo[2.2.1]heptane (**3**) is a representative term exhibiting exceptionally potent affinity for the $\alpha 4\beta 2$ receptors but, unfortunately, inadequate pharmacokinetic properties (poor CNS penetration).²³ As a consequence compound **3** was not selected as a candidate for clinical evaluation. Nevertheless, the diazabcycle **3** could be assumed as a lead compound for the design of novel high-affinity $\alpha 4\beta 2$ subtype-selective ligands.

In this context, the aim of this study was to investigate the effect of the variation of the 2,5-diazabicyclo[2.2.1]heptane scaffold of **3** with the 3,6-diazabicyclo[3.1.1]heptane skeleton synthesizing a range of isomers and analogues based on this diazabicyclic framework. This paper describes the synthesis of compounds 4 and the evaluation of their binding affinity at $\alpha 4\beta 2$ and $\alpha 7$ nAChR subtypes (Fig. 1).

The synthetic route to the desired final compounds **4** is outlined in scheme 1. The sequence commenced with the addition of the monoprotected 3,6-diazabicyclo[3.1.1]heptane **5**²⁴ and haloazines **6** to a toluenic mixture of Pd-BINAP catalyst in the presence of a base (Cs₂CO₃ or NaOtBu), resulting in coupling products **7a-d**. Synthesis of compound 7e was accomplished by refluxing 5 with 3,6-dichorop-



Figure 1. Epibatidine and related analogues.

iridazine in toluene and in the presence of TEA. Treatment of 7a-e with HCOOH underwent smooth deprotection of the Boc group leading to 4a-e in good-to-excellent yields (63-95%). Reaction of 4a with HCOOH/HCHO produced the N_6 -methylderivative **4f** in 72% yield.

The goal of this study was to prepare a series of 3,6-diazabicyclo[3.1.1]heptane analogues of the $\alpha 4\beta 2$ nAChR subtype ligand, 2-(6-chloropyridin-3-yl)-2,5-diazabicycloheptane,²³ and to measure their affinity at $\alpha 4\beta 2$ and $\alpha 7$ nAChR subtypes. The strategy of incorporating a novel diazabicycloheptane unit into the structure of **3** was expected to afford compounds with high affinity for $\alpha 4\beta 2$ nAChR subtype. Modifications were principally made on the pyridine ring to investigate the effect of different variations on $\alpha 4\beta 2$ and $\alpha 7$ nAChR subtype potency and selectivity.

Binding assays were performed on rat cortex tissues for $\alpha 4\beta 2$ and α 7 nAChR subtypes with [³H]-epibatidine and [¹²⁵I]- α -bungarotoxin as radioligands, respectively.^{25,26} The binding affinities (K_i values) for the new ligands,²⁷ as well as the reference compounds **3** and radioligands at $\alpha 4\beta 2$ and $\alpha 7$ nAChR subtypes are shown in Table 1.

In general, results showed a spectrum of $\alpha 4\beta 2$ nAChR subtype affinities ranging from 0.023 nM for compound 4a to 1.2 nM for compound **4d**, with α 7 nAChR subtype affinities ranging from 1.36 nM for compound 4b to 28,000 for compound 4d. Thus, the α 7 nAChR affinities of all of the investigated 3.6-diazabicyclo[3.1.1]heptanes are lower than their $\alpha 4\beta 2$ nAChR subtype affinities

Compound 4a, having a 3,6-diazabicyclo[3.1.1]heptane ring replacing the 2,5-diazabicyclo[2.1.1]heptane ring of $\mathbf{3}$, had K_i values of 0.023 nM and 4.1 nM for $\alpha 4\beta 2$ and $\alpha 7$ nAChR subtype, respectively, with a α 7: α 4 β 2 selectivity ratio of 178. Compound 4a serves as a convenient benchmark for all of the others in term of presenting structure-binding affinity relationships.

Table 1				
Binding affinity (K _i , nM)	^a of 4a–f to	$\alpha4\beta2$ and	α7 nAChR	subtypes.

Compound	α4β2: [³ H-Epibatidine] (<i>K</i> _i , nM)	α7: [¹²⁵ α-BgTx] (K _i , nM)	Selectivity K _i ratio α7/α4β2
4a	0.023 (34%)	4.1 (35%)	178
4b	0.056 (28%)	1.36 (42%)	24.3
4c	0.039 (36%)	50.5 (46%)	1295
4d	1.2 (45%)	28000 (52%)	23300
4e	0.168 (33%)	25 (48%)	149
4f	0.43 (34%)	20 (42%)	46.5
3	0.018 ^b	-	_
³ H-Epibatidine	0.083	0.8	9.6
¹²⁵ I-α-Bungarotoxin	-	0.9	-

^a The K_i values shown were the mean (% coefficient of variation) of three independent measurements.

^b Binding affinity of compound **3**²³ was determined by the inhibition of [³H] cytosine binding in homogenates of rat striatum.



Scheme 1. Reagents and conditions: (i) Pd₂(dba)₃, rac-BINAP, Cs₂CO₃ or NaOtBu, toluene, 100–110 °C, 22 h (for compounds 7a–d); TEA, toluene, 120 °C, 24 h (for compound 7e); (ii) HCOOH 99%, rt, 3.5 h; (iii) HCOOH 99%/HCHO 40%, 110 °C, 2.5 h.

De-chloro compound **4b** displayed 2.4-fold lower $\alpha 4\beta 2$ nAChR subtype affinity than **4a** whereas the $\alpha 7$ nAChR subtype affinity was increased displaying 3-fold greater affinity.

The bromo-substituted compound **4c** had 1.7-fold and 12.3-fold lower affinity than **4a** for α 4 β 2 and α 7, respectively, and possessed a selectivity ratio of 1295.

Substitution with a nitro group, compound **4d**, resulted in a reduction, about 52-fold, of $\alpha 4\beta 2$ receptor affinity and a much higher reduction of $\alpha 7$ receptor affinity with the highest selectivity ratio (23300).

These results indicate that factors such as electronics are likely involved in the binding of **4a** analogues.

Without doubts, the incorporation of a chlorine atom at C6' position of the pyridine moiety as in **4a** is favourable for $\alpha 4\beta 2$ affinity.

Compound **4e** has two basic nitrogen in the heteroaryl aromatic ring and is more hydrophilic than compound **4a**. This pyridazine derivative **4e** had 7.3-fold and 6.1-fold lower affinity than **4a** for $\alpha 4\beta 2$ and $\alpha 7$ nACh receptor subtypes, respectively.

 N_6 -Methyl substitution of **4a** as in compound **4f** lowered affinities for both receptors with K_i values of 0.43 nM for $\alpha 4\beta 2$ receptor and of 20 nM for $\alpha 7$ receptor.

The best compounds in the **4a–f** series show affinity for $\alpha 4\beta 2$ nAChR in the same range as their diazabicyclo[2.2.1]heptane analogue 3 suggesting similar geometrical and conformational properties. Thus, we performed a modeling study on the protonated form of these compounds through optimizations within the density functional approach at the B3LYP/6-311+G(d,p) level.²⁸ The energy of the optimized structures were recalculated using a continuum solvent model (PCM)²⁹ as implemented in Gaussian 03³⁰ to take into account the effect of water on these highly polar molecules. The main degree of conformational freedom of compounds 3 and 4 is the rotation of the aryl group around the single bond that connects it to the diazabicyclic system. This rotation cannot be expected to be completely free as the orientation of the aromatic ring can be influenced by the resonance with the lone pair of the nitrogen atom to which it is connected, so that conformations that make possible this conjugation should be preferred. This nitrogen atom usually does not assume a completely planar geometry but shows a certain degree of pyramidalization that originates two possible 'configurations'. Finally, in 4 the piperazine ring can, in principle, assume either a chair or a boat conformation contrarily to **3** where the same ring is forced into a twisted boat geometry.

Several minimum energy conformations were located for each compound and in Table 2 are reported the gas-phase and the water-solvated energies of **3** and **4a** together with selected geometrical features including the distance between the protonated

nitrogen and the pyridine nitrogen (A–B distance) or the center of the pyridine ring (A–C distance). The data in Table 2 and the three-dimensional plots, represented in Figure 2, show the very close similarity between **3** and **4a** that can justify the almost identical binding affinity to $\alpha 4\beta 2$ nAChR. In fact, each conformation of **4a** shows energy and geometry data in strict analogy with the corresponding conformation of **3**. Moreover, in all the conformations the orientation of the chloropyridinyl group allows a good conjugation with the adjacent nitrogen atom (τ_2 always about 90 or -90°). It is worthy pointing out that both compounds show A–B distances of about 6 Å and A–C distances of 5.0–5.5 Å confirming that these long distances are compatible with affinity for $\alpha 4\beta 2$ receptors in the subnanomolar range.³¹

The conformational behavior of **4b–f**, not reported in Table 2, are very similar to those of 4a. Thus, modulation of activity exerted by the substituent at C6' (**4b–d**), the second nitrogen in the aromatic ring (**4e**), or the N6-methyl substitution (**4f**) seems to derive from specific interactions of these groups rather than from their influence on the conformation of the molecules.



Figure 2. 3D plot of the most representative conformations of compounds 3 and 4a.

Table 2

Deleting an array and calcoted	manus atminal data of the DOLVE	1/C 211 C(d m) as low late d minimum	an anone conformations of some	maximala 7 and 4ad
Relative energy and selected	geometrical data of the BSLYP	2/10-311+G(010) Calculated munimum	energy conformations of com	DOUDOS 5 ADO 4a
neiunie energy und sereeted	geometrical data of the bobit	(d,p) carcalated minimum	energy connormations of com	poundo o una ma

Conformation	E _{rel} (gas-phase) (kcal/mol)	E _{rel} (water) (kcal/mol)	$A-B^{b}(Å)$	$A-C^{c}(Å)$	$\tau_1^{\mathbf{d}}(\circ)$	$\tau_2^{e}(\circ)$	α ^f (°)
3A	0.34	0.00	5.78	5.00	-21	-94	8.8
3B	0.00	0.05	5.67	4.98	154	81	8.9
3C	1.28	1.09	6.31	5.48	8	-100	-5.9
3D	1.18	1.13	6.24	5.46	-177	75	-4.5
4aA	0.02	0.00	6.29	5.49	-9	-88	3.1
4aB ^g	0.02	0.00	6.29	5.49	169	88	3.1
4aC	0.00	1.42	6.14	5.32	11	-91	-4.6
4aD ^h	0.00	1.42	6.14	5.32	-167	91	-4.6

^a Several other minimum energy conformation were located for **4a**, but they are not reported as their water-solvated energy is 5–7 kcal/mol higher than that of **4aA**.

^b Distance between the protonated nitrogen and the pyridine nitrogen.

^c Distance between the protonated nitrogen and the center of the pyridine ring.

 d τ_1 is C3–N2–C3'–C2' for 3 and C2–N3–C3'–C2' for 4a.

^e τ_2 is the inproper torsional angle N⁺-N2-C3'-C2' for **3** and N⁺-N3-C3'-C2' for **4a**.

^f Pyramidalization degree of the nitrogen atom bearing the aryl substituent defined as the difference between 360 and the sum of the three bond angles of the same atom, considered positive when it is pyramidalized in the direction of the methylene bridge and negative in the opposite case.

^g Conformation enantiomeric to 4aA.

^h Conformation enantiomeric to **4aC**.

In summary, we have prepared ligands for $\alpha 4\beta 2$ and $\alpha 7$ nAChR subtypes that combines a 3,6-diazabicyclo[3.1.1]heptane core with various pyridines and a pyridazine. Most of these ligands have very high affinity for $\alpha 4\beta 2$ nAChR subtypes. The best, compound **4a**,³² has a chloropyridine motif and binds 3.6 times better than epibatidine to $\alpha 4\beta 2$ nAChR subtypes in the meantime maintaining similar $\alpha 4\beta 2$ affinity respect to the lead **3**.

References and notes

- Willette, R. E. Analgesic agents. In Wilson and Gisvold's Textbook of organic medicinal and pharmaceutical chemistry; Block, J. H., Beale, J. M., Jr., Eds., eleventh ed.; Lippincot Williams and Wilkins: Philadelphia, 2004; pp 731–766.
 Jain K. Emerging drugs 2000 5, 241
- Jain, K. K. Emerging drugs 2000, 5, 241.
 Raith, K.; Hochlaus, G. Int. J. Clin. Pharmacol. Ther. 2004, 42, 191.
- Lewis, H. Nonsteroidal anti-inflammatory drugs: pathology and clinical presentation of hepatotoxicity. In *Drug-induced Liver Disease*; Kaplowitz, N., DeLeve, L. D., Eds.; Marcel Dekker, Inc.: New York, 2003; pp 377–404.
- 5. Mitchell, J. A.; Warner, T. D. Nat. Rev. Drug Discov. 2006, 5, 75.
- Dworkin, R. H.; O'Connor, A. B.; Backonja, M.; Farrar, J. T.; Finnerup, N. B.; Jensen, T. S.; Kalso, E. A.; Loeser, J. D.; Miaskowski, C.; Nurmikko, T. J.; Portenoy, R. K.; Rice, A. S. C.; Stacey, B. R.; Treede, R. D.; Turk, D. C.; Wallace, M. S. *Pain* **2007**, *132*, 237.
- 7. Kennedy, J. D. J. Med. Chem. 2007, 50, 2547.
- Jensen, A. A.; Frolund, B.; Lilijefors, T.; Krogsgaard-Larsen, P. J. Med. Chem. 2005, 48, 4705.
- 9. Bunnelle, W. H.; Dart, M. J.; Schrimpf, M. R. *Curr. Top. Med. Chem.* **2004**, *4*, 299. 10. Unwin, N. *Nature* **1995**, 373, 37.
- 11. Unwin, N. J. Mol. Biol. 1996, 257, 586.
- 12. Gotti, C.; Zoli, M.; Clementi, F. Trends Pharmacol. Sci. 2006, 27, 482.
- Levin, E. D. Nicotinic Receptors in the Nervous System; CRC press: Boca Raton, FL, 2002
- 14. Paterson, D.; Nordberg, A. Prog. Neurobiol. 2000, 61, 75.
- Cassel, B. K.; Bermudez, I.; Dajas, F.; Abin-Carriquiry, J. A.; Wannacot, S. Drug Discov. Today 2005, 10, 1657.
- 16. Lawand, N. B.; Lu, Y.; Westlund, K. N. Pain 1999, 80, 291.
- Marubio, L. M.; del Mar Arroyo-Jimenez, M.; Corsero-Erausquin, M.; Lena, C.; Le Novere, N.; de Kerchove d'Exaerde, A.; Huchet, M.; Damaj, M. I.; Changeux, J. P. *Nature* 1999, 398, 805.
- Spande, T. F.; Garaffo, M.; Edwards, M. W.; Yeh, J. C.; Pannel, L.; Daly, J. W. J. Am. Chem. Soc. 1992, 114, 3475.
- Lynch, J. J.; Wade, C. L.; Mikusa, J. P.; Decker, M. W.; Honore, P. Eur. J. Pharmacol. 2005, 509, 43.
- 20. Sorbera, L. A.; Revel, L.; Leeson, P. A.; Castagner, J. Drugs Future 2001, 26, 927.
- Holladay, M. W.; Wasicak, J. T.; Lin, N. H.; He, Y.; Ryther, K. B.; Bannon, A. W.; Buckley, M. J.; Kim, D. J. B.; Decker, M. W.; Anderson, D. J.; Campbell, J. E.; Kuntzweiler, T. A.; Donnelly-Roberts, D. L.; Piattoni-Kaplan, M.; Briggs, C. A.; Williams, M.; Arneric, S. P. J. Med. Chem. **1998**, *41*, 407.
- Decher, M. W.; Curzon, P.; Holladay, M. W.; Nikkel, A. L.; Bitner, R. S.; Bannon, A. W.; Donnelly-Roberts, D. L.; Puttfarcken, P. S.; Kuntzweiler, T. A.; Brigs, C. A.; Williams, M.; Arneric, S. P. J. Physiol. Paris 1998, 92, 221.
- Bunnelle, W. H.; Daanen, J. F.; Ryther, K. B.; Schrimpf, M. R.; Dart, M. J.; Gelain, A.; Meyer, M. D.; Frost, J. M.; Anderson, D. J.; Buckley, M.; Curzon, P.; Cao, Y. J.;

Puttfarcken, P.; Searle, X.; Ji, J.; Putman, C. B.; Surowy, C.; Toma, L.; Barlocco, D. J. Med. Chem. **2007**, *50*, 3627.

- Loriga, G.; Manca, I.; Murineddu, G.; Chelucci, G.; Villa, S.; Gessi, S.; Toma, L.; Cignarella, G.; Pinna, G. A. Bioorg. Med. Chem. 2006, 14, 676.
- Dellanoce, C.; Bazza, C.; Grazioso, G.; De Amici, M.; Gotti, C.; Riganti, L.; Clementi, F.; De Micheli, C. Eur. J. Org. Chem. 2006, 3746.
- Gotti, C.; Balestra, B.; Moretti, M.; Rovati, G. E.; Maggi, L.; Rossoni, G.; Berti, F.; Villa, L.; Pallavicini, M.; Clementi, F. Br. J. Pharmacol. 1984, 124, 1197.
- 27. Munson, P. J.; Rodbard, D. Anal. Biochem. 1980, 107, 220.
- Becke, A. D. J. Chem. Phys. 1993, 98, 5648; Lee, C.; Yang, W.; Parr, R. G. Phys. Rev. B 1988, 37, 785.
- 29. Barone, V.; Cossi, M.; Tomasi, J. J. Comput. Chem. 1998, 19, 404.
- 30. Gaussian 03, Revision B.02, Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Montgomery, Jr., J. A.; Vreven, T.; Kudin, K. N.; Burant, J. C.; Millam, J. M.; Iyengar, S. S.; Tomasi, J.; Barone, V.; Mennucci, B.; Cossi, M.; Scalmani, G.; Rega, N.; Petersson, G. A.; Nakatsuji, H.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Klene, M.; Li, X.; Knox, J. E.; Hratchian, H. P.; Cross, J. B.; Adamo, C.; Jaramillo, J.; Gomperts, R.; Stratmann, R. E.; Yazyev, O.; Austin, A. J.; Cammi, R.; Pomelli, C.; Ochterski, J. W.; Ayala, P. Y.; Morokuma, K.; Voth, G. A.; Salvador, P.; Dannenberg, J. J.; Zakrzewski, V. G.; Dapprich, S.; Daniels, A. D.; Strain, M. C.; Farkas, O.; Malick, D. K.; Rabuck, A. D.; Raghavachari, K.; Foresman, J. B.; Ortiz, J. V.; Cui, Q.; Baboul, A. G.; Clifford, S.; Cioslowski, J.; Stefanov, B. B.; Liu, G.; Liashenko, A.; Piskorz, P.; Komaromi, I.; Martin, R. L.; Fox, D. J.; Keith, T.; Al-Laham, M. A.; Peng, C. Y.; Nanayakkara, A.; Challacombe, M.; Gill, P. M. W.; Johnson, B.; Chen, W.; Wong, M. W.; Gonzalez, C.; Pople, J. A., Gaussian, Inc., Pittsburgh PA, 2003.
- Toma, L.; Quadrelli, P.; Bunnelle, W. H.; Anderson, D. J.; Meyer, M. D.; Cignarella, G.; Gelain, A.; Barlocco, D. J. Med. Chem. 2002, 45, 4011.
- 32. Experimental procedure for the synthesis of compound 4a. A round-bottom flask was charged with Pd2(dba)3 (0.92 g, 1.00 mmol) and BINAP (0.61 g, 1.00 mmol) in dry toluene (15 ml). The mixture was flushed with nitrogen for 10 min under magnetic stirring. In another round-bottom flask aryl iodide 6a (0.61 g, 2.52 mmol), amine 5 (0.50 g, 2.52 mmol) and Cs₂CO₃ (3.29 g, 10.1 mmol) were mixed with an additional amount of toluene (10 mL). Then, the Pd(OAc)₂/BINAP solution was added and the mixture was refluxed for 22 h. After cooling, the residue was partitioned between diethyl ether and water. The organic phase was washed with water, dried (Na₂SO₄) and evaporated. The resulting crude product was purified by flash column chromatography (petroleum ether/Ethyl acetate 7:3) to give 7a as a yellow solid (66%). Rf: 0.29; mp = 129–130 °C; ¹H NMR (200 MHz, CDCl₃, δ/ppm): 1.35 (s, 9H), 1.51 (d, 1H, *J* = 8.8 Hz); 2.58–2.78 (m, 1H); 3.26 (d, 2H, *J* = 10.6 Hz), 3.82–3.95 (m, 2H); 4.30 (d, 2H, J = 10.4 Hz) 6.97 (dd, 1H, J = 3.0 and 8.8 Hz); 7.17 (d, 1H, J = 8.8 Hz), 7.86 (d, 1H, J = 3.0 Hz). Anal. Calcd for C₁₅H₂₀ClN₃O₂: C, 58.16; H, 6.51; N, 18.56. Found: C, 58.20; H 6.40; N 18.60.A solution of **7a** (0.28 g, 0.90 mmol) in HCOOH 99% (5 ml) was stirred at room temperature for 3.5 h. The mixture was diluted with water and K₂CO₃ was added until neutral pH. The solution was extracted with CH₂Cl₂ dried (Na₂SO₄) and evaporated. The residue was triturated with diethyl ether to give the final compound **4a** as a white solid (81%). $R_{\rm f}$: 0.21 $(CH_2Cl_2/methanol 7:3); mp = 130-131 °C; ^1H NMR (200 MHz, CDCl_3, <math>\delta/ppm):$ 1.53 (br s, 1H), 1.62 (d, 1H, J = 8.8 Hz); 2.70–2.85 (m, 1H); 3.48–3.55 (m, 4H), 3.88–3.95 (m, 2H); 6.97 (dd, 1H, J = 2.8 and 8.8 Hz); 7.18 (d, 1H, J = 8.2 Hz), 7.87 (d, 1H, J = 3.2 Hz). Anal. Calcd for C₁₀H₁₂ClN₃: C, 57.28; H, 5.77; N, 20.04. Found: C, 57.16; H 5.74; N 19.90.