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Synthesis and Antiviral Evaluation of 3-Hydroxy-2-methylpyridin-4-one Dideoxynucleoside Derivatives

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Abstract—We describe the synthesis and the antiviral evaluation of novel α and β dideoxynucleoside derivatives in which the base has been replaced by a 3-hydroxy-2-methylpyridin-4-one. The syntheses were successfully achieved by the use of the standard Vorbrüggen coupling conditions. Moderate activity of these compounds were found on herpes simplex virus (HSV) type 1 and type 2.

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3-Hydroxypyridin-4-ones (HPOs) are currently one of the main candidates for the development of orally active iron chelators.¹ Indeed, the 1,2-dimethyl derivative (Deferiprone, Fig. 1), with an associated pFe³⁺ value of 19.4 is the only active iron chelator currently available for clinical use (marketed by Apotex Inc., Toronto, Canada as FerriproxTM).² It has been shown that iron chelation, which would make iron unavailable for redox reactions, could influence HIV replication in two possible ways:

- by inactivation of the iron-dependent cellular enzyme ribonucleotide reductase which is responsible for generating the building blocks for viral DNA.³
- via reduction of nuclear factor- κB (NF- κB) activation.⁴

Recently, a clear synergism in HIV-1 inhibition was observed by combining iron chelators with the anti-HIV nucleoside analogue ddI and it was suggested that in combination with existing antivirals, iron chelation could have a beneficial effect on HIV disease.⁵ These findings prompted us to synthesise several 2',3'-dideoxynucleosides

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Deferiprone, FerriproxTM $R_1 = R_2 = CH_3$

Figure 1.

in which the base has been changed to a 3-hydroxypyridin-4-one. Moreover, the base modification of these new nucleosides could confer activity against other viruses than HIV. For a start, the synthesised nucleosides contain sugar moieties similar to that of two currently FDA approved anti-HIV 2',3'-dideoxynucleosides: ddC and 3TC (Fig. 2).

Results and Discussion

3-Hydroxybenzyl-2-methylpyridin-4-one is a requisite for the Vorbrüggen conditions in the sugar-base coupling. It was obtained starting from maltol which was benzylated with benzyl bromide in a basic media (Scheme 1). Reaction of adduct 1 with ammonia was performed by reflux in 50% aqueous ethanol. The benzylated pyridinone 2 was isolated in a crystalline form

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Figure 2.



Scheme 1. (a) BnBr, NaOH, MeOH, reflux, 16 h; (b) NH₃, EtOH, reflux, 24 h.

as a free base in 58% yield.⁶ The primary alcohol function of the commercially available precursor (S)-(+)dihydro-5-(hydroxymethyl)-2-(3H)-furanone was protected with TBDPSCl in THF to give the lactone 3 (Scheme 2). This latter compound was then reduced with diisobutylaluminium (DIBAL-H), followed by trapping of the resulting lactol with acetic anhydride to give 4 in 78% yield. 3-Benzyloxy-2-methylpyridin-4-one 2 was silvlated under reflux for 2 h with hexamethyldisilazane (HMDS) in the presence of a catalytic amount of ammonium sulfate. After the removal of the excess of HMDS, the base was solubilised in 1,2-dichloroethane and a mixture of 4 and trimethylsilyltriflate (TMSOTf)⁷ in 1,2-dichloroethane was slowly added at room temperature to afford compounds 5 and 6 in 47% yield. The β/α ratio generally was around 1, and separation of both isomers was successfully accomplished by flash silica gel chromatography. The relative stereochemistry of these compounds was assigned by 1D and 2D ¹H NMR studies (Fig. 3). NOE interactions were observed between H-1' and H-4' suggesting the cis orientation of the less polar compound 5. The *trans* configuration was then assigned to the more polar isomer 6. Compounds 5



Scheme 2. (a) TBDPSCl, imidazole, dry DMF, rt, 18 h; (b) DIBAL, dry DCM, -90 °C then Ac₂O, pyridine, DMAP, rt, 72 h; (c) silylated 2, dry DCE, TMSOTf, rt, 20 h; (d) TBAF, dry THF, rt, 72 h; (e) H₂, Pd/C, MeOH, rt, 16 h.



Figure 3.

and 6 were converted to compounds 7 and 8 by treatment with TBAF in THF. Catalytic hydrogenation in MeOH over 10% palladium on carbon gave the *cis* nucleoside 9 and the *trans* nucleoside 10 in quantitative yields.⁸ In order to prepare the oxathiolane derivatives 17 and 18 (Scheme 3) the racemate thialactone 11 previously described by Choi et al.⁹ was reduced with DIBAL-H and acetylated by Ac_2O in presence of pyridine and 4-(dimethylamino)pyridine to give the corresponding acetate 12 in 64% yield. The heterocycle base 2 was silvlated and reacted with 12 in 1,2-dichloroethane and TMSOTf as catalyst. The relative stereochemistry of these compounds was assigned by NOESY and COSY experiments (Fig. 3). NOE interactions were observed between H-1' and H-4' suggesting the *cis* configuration of the less polar 13 and the trans configuration of the more polar 14. The racemic cis nucleoside 13 and the racemic trans nucleoside 14 were then deprotected by treatment with TBAF in THF, to give 15 and 16 in quantitative yields. Since the sulphur atom, contained in the oxathiolane ring, poisons the Pd catalyst, debenzylation of the heterocycle base was successfully achieved with the use of iodotrimethylsilane^{10,11} to afford the racemic *cis* nucleoside 17 and the racemic *trans* nucleoside 18 in modest yields.

Evaluation of antiviral activity was done as described previously.¹² The compounds were evaluated against HIV-1 and HIV-2, herpes simplex virus type 1 (HSV-1)



Scheme 3. (a) DIBAL, dry DCM, -90 °C then Ac₂O, pyridine, DMAP, rt, 72 h; (b) silylated 2, dry DCE, TMSOTf, rt, 24 h; (c) TBAF, dry THF, rt, 3 h; (d) NaI, TMSCl, dry CHCl₃, rt, 18 h.

Compd	Anti-HIV activity			Anti-HSV activity			
	EC ₅₀ (μM) HIV-1 in CEM cells	EC ₅₀ (μM) HIV-2 in CEM cells	CC ₅₀ (µM)	EC ₅₀ (μM) HSV-1 (KOS) in HEL cells	EC ₅₀ (μM) HSV-2 (G) in HEL cells	EC ₅₀ (μM) HSV-1 TK neg in HEL cells	CC ₅₀ (µM)
9	> 50	> 50	101 ± 6.2	40	> 900	500	> 900
10	> 50	> 50	> 50	177	500	500	> 900
(±)- 17	> 80	> 80	203 ± 2.5	32	81	80	> 100
(±)-18	> 80	> 80	203 ± 11.9	40	90	95	>400
ACV	> 200	> 200	> 200	0.32	1.3	88	>1700
(acyclovir) DHPG (ganciclovir) HPMPC (cidofovir)	> 200 > 200	> 200 > 200	> 200 > 200	0.57	0.06	_	> 400 > 350

Table 1. Anti-HIV and anti-HSV activities of compounds 9, 10, (\pm) -17, and (\pm) -18

 EC_{50} , effective concentration or concentration required to inhibit 50% of virus induced cytopathicity; CC_{50} , cytotoxic concentration or concentration required to reduce cells viability by 50%; —not determined.

and type 2 (HSV-2), vaccinia virus and vesicular stomatitis virus in HEL cells, parainfluenza-3 virus, reovirus-1, Sindbis virus, coxsackie B4 virus, Punta Toro virus in Vero cells and respiratory syncytial virus in HeLa cell cultures. The anti-HIV and HSV activities of compounds 9, 10, 17 and 18 are represented in Table 1. None of these compounds were active against any of the tested viruses except for herpes simplex virus (Table 1). Compound 10, 17 and 18 displayed moderate activity against both wild-type HSV-1 and HSV-2, as well as against a thymidine kinase deficient strain of HSV-1 that has a 270 fold reduced sensitivity for acyclovir than the wild-type virus. The weak but consistent activity of these compounds against HSV-1 and HSV-2 is encouraging and may suggest that these new nucleosides are metabolised to their 5'-triphosphates or as such recognised by the viral DNA polymerases. The (moderate) activity of an form of a nucleoside (10 and 18) is quite unusual, few such molecules have been reported. Further work is needed both to design analogues with an improved anti-herpes virus activity and to unravel their mechanism of action.

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References and Notes

1. Tilbrook, G. S.; Hider, R. C. Iron chelators for clinical use. In *Metals Ions in Biological Systems*; Sigel, A., Sigel, H., Eds.; Iron Transport and Storage in Microorganisms, Plants and Animals, Vol. 35, Marcel Dekker: New York, 1998; p.691

2. Hider, R. C.; Liu, Z. D.; Piyamongkol, S. Tran. Sci. 2000, 23, 201.

3. Lederman, H. M.; Cohen, A.; Lee, J. W. W.; Freedman, M. H.; Gelfand, E. W. *Blood* **1984**, *64*, 748.

4. Sappey, C.; Boelaert, J. R.; Legrand, P. S.; Forceille, C.; Favier, A.; Piette, J. *AIDS Res. Hum. Retroviruses* **1995**, *11*, 1049. 5. (a) Georgiou, N. A.; van der Bruggen, T.; Hider, R. C.; Marx, J.; Van Asbeck, B. S. *Eur. J. Clin. Invest.* **2002**, *32*, 91. Georgiou, N. A.; van der Bruggen, T.; Oudshoorn, M.; Nottet, H.; Marx, J.; Van Asbeck, B. S. *Trans. Sci.* **2000**, *23*, 249. Georgiou, N. A.; van der Bruggen, T.; Oudshoorn, M.; Nottet, H.; Marx, J.; Van Asbeck, B. S. *J. Clin. Virol.* **2001**, *20*, 141

6. Dobbin, P. S.; Hider, R. C.; Hall, A. D.; Taylor, P. D.; Sarpong, P.; Porter, J. B.; Xiao, G.; van der Helm, D. J. Med. Chem. **1993**, *36*, 2448.

 Niedballa, U.; Vorbrüggen, H. J. Org. Chem. 1974, 39, 3668.
(a) Liu, G.; Bruenger, F. W.; Barrios, A. M.; Miller, S. C. Nucleosides Nucleotides 1995, 14, 1901. (b) Mao, D. T.; Driscoll, J. S.; Marquez, V. E. J. Med. Chem. 1984, 27, 160.

9. Choi, W. B.; Wilson, L. J.; Yeola, S.; Liotta, D. C. J. Am. Chem. Soc. 1991, 113, 9377.

10. Jung, M. E.; Lyster, M. A. J. Org. Chem. 1977, 42, 3761.

11. Huang, J. J.; Rideout, J. L.; Martin, G. E. Nucleosides Nucleotides 1995, 14, 195.

12. Neyts, J.; Reymen, D.; Letourneur, D.; Jozefonvicz, J.; Schols, D.; Esté, J.; Andrei, G.; McKenna, P.; Witvrouw, M.; Ikeda, S.; Clement, J.; De Clercq, E. *Biochem. Pharmacol.* **1995**, *50*, 743.