

CF₃-BOP, CF₃-PyBOP and CF₃-HBTU: New peptide coupling reagents specially suited for α -aminoisobutyric acid condensations^a

Jac C.H.M. Wijkmans, John A.W. Kruijtzter, Gijs A. van der Marel,
Jacques H. van Boom and Wim Bloemhoff

Leiden Institute of Chemistry, Gorlaeus Laboratories, Leiden University, P.O. Box 9502,
2300 RA Leiden, The Netherlands
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Abstract. The peptide-coupling reagents CF₃-BOP (**1b**), CF₃-PyBOP (**1c**) and CF₃-HBTU (**2a**) with built-in 1-hydroxy-6-(trifluoromethyl)benzotriazole proved to be highly efficient for the coupling of α -aminoisobutyric acid residues.

Introduction^b

The isolation of an increasing number of naturally occurring peptides bearing α -aminoisobutyric acid (Aib)¹ residues has stimulated the chemical synthesis of this type of biologically interesting peptides. However, it became apparent that introduction of Aib into peptides using well-established coupling reagents in solid-phase syntheses, e.g. dicyclohexylcarbodiimide/1-hydroxybenzotriazole and BOP² (**1a**), was a slow and low-yielding process³. In order to solve this problem much effort has been directed to the development of more powerful reagents⁴. We now wish to report that the CF₃-OBt-containing

reagents CF₃-BOP (**1b**), CF₃-PyBOP (**1c**) and CF₃-HBTU (**2a**) are promising tools for the coupling of Aib.

Results and discussion

Some years ago,⁵ we presented the bifunctional phosphorylating reagent *O*-(2-chlorophenyl)-*O,O*-bis(benzotriazol-1-yl)phosphate **3a** for the introduction of 3' → 5' internucleotide phosphotriester linkages. The rate of the two-step phosphorylation could be substantially enhanced by application of the CF₃ analogue **3b**⁶. On the basis of this information it was expected that the readily accessible CF₃-substituted reagents **1b,c** and **2a** would exert a beneficial effect on the introduction of the Aib moiety into peptides.

In order to assess this assumption, we first examined the coupling of isoleucine methyl ester with *Z*-protected Aib. It can be seen in entry 1 (Table I) that *in-situ* activation with both BOP (**1a**) and CF₃-BOP (**1b**) proceeded in an excellent yield. On the other hand, the yield of the CF₃-BOP-mediated coupling of the more hindered⁷ proline methyl ester with *Z*-Aib-OH (entry 2) was much higher than with BOP. The difference in efficacy between the latter reagent and CF₃-BOP was even more pronounced in the peptide bond formation between *Z*-Aib-OH and *H*-Aib-OMe (entry 3). In this respect, it is interesting to note that Coste et al.^{3b} used the highly active bromine-containing phosphonium salts BroP⁸ (**1d**) and PyBroP⁹ (**1e**) for the preparation of *Z*-Aib-Aib-OMe (**4c**). However, the coupling efficiency of reagents **1d,e** did not deviate substantially from those with BOP and its pyrrolidine homologue PyBOP⁹ (**1f**). In addition, Coste et al.^{3b} provided evidence that the (Py)BOP-mediated condensation of *Z*-Aib-OH with the sterically hindered Aib methyl ester proceeded via the benzotriazolyl (OBt) ester of the *N*-protected Aib. In order to substantiate the effect of OBt and CF₃-OBt esters on the formation of *Z*-Aib-Aib-OMe (**4c**), coupling was conducted after preactivation of the *N*-protected Aib with BOP and CF₃-BOP. It is evident from the results in entry 4 that the Aib dipeptide **4c**

^a Dedicated to Professor G.I. Tesser on the occasion of his 65th birthday.

^b **Abbreviations** are in accordance with the recommendations of the IUPAC-IUB as set forth in J. Biol. Chem. **260**, 14 (1985). Additional abbreviations:

Aib = α -aminoisobutyric acid

HOBt = 1-hydroxybenzotriazole

BOP = (benzotriazol-1-yloxy)tris(dimethylamino)phosphonium hexafluorophosphate

BroP = bromotris(dimethylamino)phosphonium hexafluorophosphate

Bz = benzoyl

CF₃-BOP = [6(trifluoromethyl)benzotriazol-1-yloxy]-tris(dimethylamino)phosphonium hexafluorophosphate

CF₃-HBTU = 2-[6(trifluoromethyl)benzotriazol-1-yl]-1,1,3,3-tetramethyluronium hexafluorophosphate

CF₃-PyBOP = [6(trifluoromethyl)benzotriazol-1-yloxy]tris(pyrrolidino)phosphonium hexafluorophosphate

DIEA = *N,N*-diisopropylethylamine

Fmoc = fluoren-9-ylmethoxycarbonyl

HATU = 2-(7-azabenzotriazol-1-yl)-1,1,3,3-tetramethyluronium hexafluorophosphate

= 2-(triazolo[4,5-*I*]pyridin-3-yl)-1,1,3,3-tetramethyluronium hexafluorophosphate

MeVal = *N*-methylvaline

PyBOP = (benzotriazol-1-yloxy)tris(pyrrolidino)phosphonium hexafluorophosphate

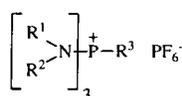
PyBroP = bromotris(pyrrolidino)phosphonium hexafluorophosphate

Z = benzyloxycarbonyl.

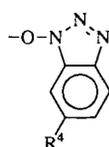
Table I Relevant data on the formation of Aib-containing dipeptides with BOP (1a), CF₃-BOP (1b), CF₃-PyBOP (1c) and CF₃-HBTU (2a)

Entry	Dipeptide		Yield (%) ^{a,b}			
			1a	1b	1c	2a
1	Z-Aib-Ile-OMe	4a	90	95	n.d. ^c	n.d. ^c
2	Z-Aib-Pro-OMe	4b	76	95	n.d. ^c	n.d. ^c
3	Z-Aib-Aib-OMe	4c	47	82	83	66
4	Z-Aib-Aib-OMe	4c	52 ^{c,d}	83 ^c	n.d. ^c	70 ^f
5	Z-Aib-Aib-OMe	4c	67 ^g	94 ^g	n.d. ^c	85 ^g
6	Fmoc-Aib-Aib-OMe	4d	51	86	82	73

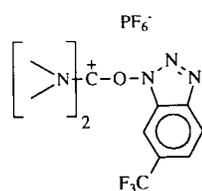
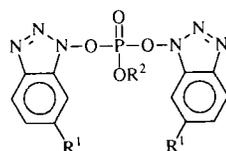
^a The yields were determined after 1h coupling at room temperature (see Experimental section). ^b In case of incomplete reaction (entries 2–6) the sole side-product was the (CF₃-)OBt-ester of the *N*-protected amino acid. ^c Amino acid ester hydrochloride was added after 5 min preactivation. ^d Addition of CF₃-HOBt (0.5 eq), DIEA (0.5 eq) and amino acid hydrochloride (1.0 eq), after 5 min preactivation, furnished 4c in 53% yield. ^e n.d. = not determined. ^f Amino acid ester was added after 20 min preactivation. The long preactivation time was necessary due to the low solubility of CF₃-HBTU in CH₂Cl₂. ^g DMF as solvent



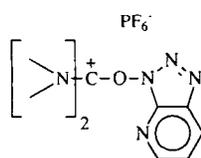
- 1 a: R¹ = R² = CH₃, R³ = OBt (BOP)
 b: R¹ = R² = CH₃, R³ = CF₃-OBt (CF₃-BOP)
 c: R¹, R² = (CH₂)₄, R³ = CF₃-OBt (CF₃-PyBOP)
 d: R¹ = R² = CH₃, R³ = Br (BroP)
 e: R¹, R² = (CH₂)₄, R³ = Br (PyBroP)
 f: R¹, R² = (CH₂)₄, R³ = OBt (PyBOP)



- OBt: R⁴ = H
 CF₃-OBt: R⁴ = CF₃

2 a: CF₃-HBTU

- 3 a: R¹ = H, R² = 2-chlorophenyl
 b: R¹ = CF₃, R² = 2-chlorophenyl



5: HATU

could be isolated in comparable yields with those resulting from *in-situ* activation of Z-Aib-OH with BOP and CF₃-BOP, thus indicating that the acylation of an amine with decreased reactivity proceeds predominantly via the OBt or the more activated CF₃-OBt ester. Apart from this, it has been established¹⁰ that the solvent DMF may significantly enhance the efficiency of peptide bond formation. The latter effect was confirmed by the near quantitative formation of 4c (entry 5) using CF₃-BOP as the activating agent.

At this stage, we were anxious to find out whether the pyrrolidine analogue CF₃-PyBOP (1c) and the uronium derivative CF₃-HBTU (2a) would be as effective as CF₃-BOP. It can be seen in entry 3 and 6 that CF₃-PyBOP gave similar results in the synthesis of 4c and 4d. How-

ever, CF₃-HBTU-assisted condensation of two Aib residues (entries 3–6) gave lower yields than with CF₃-BOP. Interestingly, the recently introduced¹¹ and highly advocated¹² 7-azabenzotriazole-based reagent HATU (5) gave a lower yield (*i.e.* 69%) than CF₃-BOP in the coupling of Z-Aib-OH with H-Aib-OMe. The diminished efficacy of the uronium salts CF₃-HBTU and HATU might be attributed¹³ to the *in-situ* generated strongly basic tetramethylurea.

Stimulated by the results thus far obtained, we turned our attention to the rather troublesome¹⁵ coupling of *N*-methylvaline (MeVal). In accordance with expectation, we found that both BOP- and CF₃-BOP-mediated condensation of valine methyl ester with Z-protected MeVal (entry 1 in Table II) gave the desired dipeptide 4e in high yield. In contrast, reaction of Z-Val-OH with MeVal methyl ester (entry 2) proceeded unsatisfactorily, even at 35°C (entry 3)¹⁶. Nonetheless, it is clear that CF₃-BOP is also superior in this particular case to BOP (*cf.* the high-yielding CF₃-BOP-assisted couplings in entries 2–6 in Table I). On the other hand, acylation of *N*-methylated valine under the agency of BroP and PyBroP afforded higher yields than with (Py)BOP or CF₃-BOP¹⁵.

Finally, it is of interest to note that loss of chiral integrity using CF₃-BOP (see note ^e in Table II) was, as gauged by the Davies test¹⁷, in the same order as for BOP¹⁸.

In conclusion, the results presented in this paper indicate that shelf-stable CF₃-BOP, CF₃-PyBOP and CF₃-HBTU are efficient coupling reagents for the introduction of Aib residues. The application of these promising condensing reagents in the solid-phase synthesis of Aib-containing peptides will be reported in due course.

Experimental

General methods and materials

DMF was stirred with CaH₂ and distilled under reduced pressure. CH₂Cl₂ was distilled from P₂O₅ and stored over molecular sieves (4

Table II Relevant data on the formation of dipeptides with BOP (1a) and CF₃-BOP (1b)

Entry	Dipeptide		Yield (%) ^a	
			1a	1b
1	Z-MeVal-Val-OMe	4e	92	95
2	Z-Val-MeVal-OMe ^{b,c}	4f	4	32
3	Z-Val-MeVal-OMe ^{b,c}	4f	12 ^d	49 ^d
4	Bz-Val-Val-OMe ^e	4g	32	46

^a The yields were determined after 1h coupling at room temperature (see Experimental section). ^b The sole side-product was the (CF₃-)OBt-ester of Z-Val-OH. ^c After 1 h, hydrazine monohydrate (0.1 ml, 2 mmol) was added to facilitate purification. ^d Conducted at 35°C. ^e %DL diastereoisomer: 53% and 50%, for BOP and CF₃-BOP, respectively.

Å). DIEA was distilled from KOH. BOP reagent was purchased from Richelieu Biotechnologies. HATU was purchased from Millipore. Fmoc-Aib-OH was obtained from NovaBiochem. MeVal was prepared according to a procedure described by *McDermott* and *Benoiton*¹⁹. Reactions were run at ambient temperature unless noted otherwise. TLC analysis was performed on Schleicher and Schüll DC Fertigfolien F1500 LS254 employing the following solvent systems: A (ethyl-acetate/light-petroleum-(b.p. 40–60°C), 1/1, v/v), B (ethyl-acetate/light-petroleum-(b.p. 40–60°C), 1/2, v/v). Compounds were visualized by UV (254 nm) and 4,4'-methylenebis(*N,N*-dimethylaniline) reagent²⁰. Column chromatography was performed on Kieselgel 60, 230–100 mesh (Merck). Melting points were measured on a Büchi melting-point apparatus and are uncorrected. Optical rotations were determined with a PROPOL automatic polarimeter at 20°C. Mass spectra were obtained with a Finnigan MAT S50 710 (Finnigan MAT, San José) spectrometer equipped with an electrospray interface. 1D- and 2D-(¹H–¹H COSY) ¹H-NMR spectra were recorded at 300 MHz on a Bruker WM-300 spectrometer interfaced with an ASPECT 2000 computer. ¹³C- and ³¹P-NMR spectra were recorded on a Jeol JNM-FX 200 spectrometer, operating at 50.1 and 80.7 MHz, respectively. Chemical shifts (δ) are given in ppm relative to the signal for internal Me₄Si for ¹H, and to the signal for internal chloroform (δ 77.0) or acetone (δ 29.8) for ¹³C. ³¹P chemical shifts are given relative to 85% H₃PO₄ as external standard.

[(6-Trifluoromethyl)benzotriazol-1-*y*-oxy]tris(dimethylamino)phosphonium hexafluorophosphate, CF₃-BOP (**1b**). A solution of 1-hydroxy-6-(trifluoromethyl)benzotriazole²¹ (1.02 g, 5.0 mmol) and triethylamine (0.70, 5.0 mmol) in CH₂Cl₂ (10 ml) was carefully added to a mixture of chlorotris(dimethylamino)phosphonium hexafluorophosphate²² (1.72 g, 5.0 mmol) in acetone (25 ml) at 0°C. After stirring for 30 min at room temperature, the mixture was concentrated and acetone (40 ml) was added followed by the addition of water (200 ml) upon which the phosphonium salt precipitated. The white precipitate was collected by filtration and redissolved in CH₂Cl₂ (100 ml). The solution was washed with water (2 × 30 ml), dried (MgSO₄) and concentrated to give **1b** (2.25 g, 88%) as a white solid which was used without further purification. The obtained phosphonium salt, recrystallized from CH₂Cl₂/light-petroleum-(b.p. 40–60°C), decomposes at $T > 140^\circ\text{C}$. MS (m/z): 365 [**1b**-PF₆]⁺, 31P NMR (acetone-*d*₆): δ 45.9 (s, P⁺), –141.1 (septet, PF₆[–], $J_{\text{P,F}}$ 710.5 Hz). ¹³C NMR (acetone-*d*₆): δ 144.9, 128.1 (C_q), 132.2 (q, C-CF₃, $J_{\text{C,F}}$ 32.7 Hz), 124.3 (q, C-CF₃, $J_{\text{C,F}}$ 272.6 Hz), 123.4, 123.3, 123.0, 107.8, 107.7 (CH-arom.), 37.7 (CH₃-N, $J_{\text{C,P}}$ 4.4 Hz).

[(6-Trifluoromethyl)benzotriazol-1-*yoxy*]tris(pyrrolidino)phosphonium hexafluorophosphate, CF₃-PyBOP (**1c**). Prepared as described for the preparation of **1b** starting from bromotris(pyrrolidino)-phosphonium hexafluorophosphate²³ (3.60 g, 7.7 mmol); yield 3.13 g (69%). Phosphonium salt **1c**, recrystallized from CH₂Cl₂/light-petroleum-(b.p. 40–60°C), decomposes at $T > 140^\circ\text{C}$. MS (m/z): 258 [O=P-(N(CH₂)₄)₃+H]⁺, 443 [**1c**-PF₆]⁺, 515 [2O=P-(N(CH₂)₄)₃+H]⁺. ³¹P NMR (acetone-*d*₆): δ 33.1 (s, P⁺), –141.1 (septet, PF₆[–], $J_{\text{P,F}}$ 708.8 Hz). ¹³C NMR (acetone-*d*₆): δ 145.0, 128.2 (C_q), 132.1 (q, C-CF₃, $J_{\text{C,F}}$ 32.7 Hz), 124.5 (q, C-CF₃, $J_{\text{C,F}}$ 272.6 Hz), 123.4, 123.1, 108.0 (CH-arom.), 49.0 (CH₂-CH₂-N, $J_{\text{C,P}}$ 4.4 Hz), 26.6 (CH₂-CH₂-N, $J_{\text{C,P}}$ 8.8 Hz).

2-[6-(Trifluoromethyl)benzotriazol-1-yl]-1,1,3,3-tetramethyluronium hexafluorophosphate, CF₃-HBTU (**2a**). Prepared starting from tetramethylchloroformamidinium hexafluorophosphate²⁴ (1.40 g, 5.0 mmol) in an analogous method to that used for **1b**; yield 1.74 g (78%). Uronium salt **2a**, recrystallized from acetone/CH₂Cl₂, decomposes at $T > 160^\circ\text{C}$. MS (m/z): 302 [**2a**-PF₆]⁺. ³¹P NMR (acetone-*d*₆): δ –141.2 (septet, PF₆[–], $J_{\text{P,F}}$ 708.5 Hz). ¹³C NMR (acetone-*d*₆): δ 151 (C⁺), 136.3, 134.2 (C_q), 129.4 (q, C-CF₃, $J_{\text{C,F}}$ 33.7 Hz), 124.3 (q, C-CF₃, $J_{\text{C,F}}$ 272.6 Hz), 130.6, 116.9, 115.0 (CH-arom.), 42.8, 42.4 (CH₃).

General procedure for BOP, CF₃-PyBOP- and CF₃-HBTU-mediated dipeptide formation

To a mixture of Z-amino acid (0.50 mmol), amino acid methyl ester hydrochloride (0.50 mmol) and either BOP (220 mg, 0.50 mmol), CF₃-BOP (255 mg, 0.50 mmol), CF₃-PyBOP (293 mg, 0.50 mmol) or CF₃-HBTU (224 mg, 0.50 mmol) in CH₂Cl₂ (3 ml) was added DIEA (0.26 ml, 1.50 mmol). After 1 h, the mixture was diluted with ethyl acetate (100 ml), washed with water (2 × 30 ml), 1M NaHCO₃ (2 × 30 ml), water (2 × 30 ml), 1M KHSO₄ (2 × 30 ml), brine (2 × 30 ml), dried (MgSO₄) and concentrated. When needed, purification of the residue by column chromatography (eluent: light-petroleum-(b.p. 40–60°C)/ethyl-acetate, 2/1 → 1/1, v/v) gave **4a–g**.

N-[α -(Benzyloxycarbonylamino)isobutyryl]-*L*-isoleucine methyl ester, Z-Aib-Ile-OMe (**4a**). R_f 0.75 (system A); [α]_D +18.2° (c 1, CHCl₃). MS (m/z): 365 [M+H]⁺, 387 [M+Na]⁺. ¹H NMR (CDCl₃): δ 7.4–7.3 (m, 5H, H-arom.), 6.86 (bd, 1H, NH Ile, $J_{\text{H,αNH}}$ 6.4 Hz), 5.41 (s, 1H, NH Aib), 5.09 (AB, 2H, CH₂ Z), 4.56 (dd, 1H, H α Ile, J 4.7 and 8.5 Hz), 3.70 (s, 3H, OCH₃), 1.9 (m, 1H, H β Ile), 1.55 and 1.52 (2 s, each 3H, H β Aib), 1.4–1.3 and 1.2–1.1 (2 m, each 1H, H γ Ile), 0.89 (t, 3H, H δ Ile, $J_{\gamma,\delta}$ 7.3 Hz), 0.87 (d, 3H, H γ Ile, $J_{\beta,\gamma}$ 6.9 Hz). ¹³C NMR (CDCl₃): δ 174.0, 172.0 (C=O Aib, Ile), 154.8 (C=O Z), 136.0 (C_q Z), 128.1, 127.7, 127.6 (CH-arom.), 66.3 (CH₂ Z), 56.6 (C α Aib), 56.2 (C α Ile), 51.6 (OCH₃), 37.4 (C β Ile), 25.5 (C β Aib), 24.7 (C γ Ile), 15.1 (β -CH₃ Ile), 11.2 (C δ Ile).

N-[α -(Benzyloxycarbonylamino)isobutyryl]-*L*-proline methyl ester, Z-Aib-Pro-OMe (**4b**). R_f 0.19 (system A); [α]_D –63.8° (c 1, CHCl₃). MS (m/z): 349 [M+H]⁺, 371 [M+Na]⁺. ¹³C NMR (CDCl₃): δ 172.7, 171.9 (C=O Aib, Pro), 154.1 (C=O Z), 136.3 (C_q Z), 128.1, 127.9, 127.8 (CH-arom.), 66.0 (CH₂ Z), 60.4 (C α Pro), 56.4 (C α Aib), 51.7 (OCH₃), 47.5 (C δ Pro), 27.4 (C β Pro), 25.4 (C γ Pro), 24.7, 24.1 (C β Aib).

N-(*N*-Benzyloxycarbonyl- α -aminoisobutyryl)- α -aminoisobutyric acid methyl ester, Z-Aib-Aib-OMe (**4c**). R_f 0.18 (system B). MS (m/z): 337 [M+H]⁺, 359 [M+Na]⁺. ¹H NMR (CDCl₃): δ 7.4–7.3 (m, 5H, H-arom.), 6.92 and 5.42 (2 s, each 2H, NH), 5.10 (s, 2H, CH₂ Z), 3.71 (s, 3H, OCH₃), 1.50 (s, 12H, H β). ¹³C NMR (CDCl₃): δ 174.8, 173.5 (C=O Aib), 155.0 (C=O Z), 136.2 (C_q Z), 128.3, 127.9, 127.8 (CH-arom.), 66.5 (CH₂ Z), 56.7, 56.2 (C α), 52.3 (OCH₃), 25.2, 24.3 (C β).

N-(*N*-Fluoren-9-ylmethoxycarbonyl- α -aminoisobutyryl)- α -aminoisobutyric acid methyl ester, Fmoc-Aib-Aib-OMe (**4d**). R_f 0.41 (system A). MS (m/z): 425 [M+H]⁺, 447 [M+Na]⁺. ¹H NMR (CDCl₃): δ 7.75 and 7.59 (2 d, 4H, H-arom., J 7.4 and 7.5 Hz), 7.6–7.3 (m, 4H, H-arom.), 6.90 and 5.50 (2 s, each 1H, NH Aib), 4.40 (d, 2H, CH₂ Fmoc, J 6.6 Hz), 4.20 (t, 1H, CH Fmoc, J 6.8 Hz), 3.71 (s, 3H, OCH₃), 1.53 (s, 12H, H β). ¹³C NMR (CDCl₃): δ 174.8, 173.5 (C=O Aib), 155.0 (C=O Fmoc), 143.7, 141.2 (C_q Fmoc), 127.5, 126.9, 124.9, 119.8 (CH-arom.), 66.4 (CH₂ Fmoc), 56.7, 56.3 (C α), 52.4 (OCH₃), 47.1 (CH Fmoc), 25.1, 24.4 (C β).

N-[*N*-(Benzyloxycarbonyl)-*L*-*N*-methylvalyl]valine methyl ester, Z-MeVal-Val-OMe (**4e**). R_f 0.54 (system B). [α]_D –93.4° (c 1, MeOH); lit.¹⁵ [α]_D –95° (c 1, MeOH). MS (m/z): 379 [M+H]⁺, 401 [M+Na]⁺. ¹H NMR (CDCl₃): δ 7.34 (bs, 5H, H-arom.), 6.53 and 6.01 (d, and 'bs', ratio 4/1, 1H, NH, $J_{\text{H,αNH}}$ 8.3 Hz), 5.17 (AB, 2H, CH₂ Z), 4.49 (dd, 1H, H α Val, $J_{\alpha,\beta}$ 5.7 Hz, $J_{\text{H,αNH}}$ 8.7 Hz), 4.15 and 4.05 (d and 'bd', ratio 4/1, 1H, H α MeVal, $J_{\alpha,\beta}$ 11.1 Hz), 3.72 (s, 3H, OCH₃), 2.90 (s, 3H, NCH₃), 2.3 (m, 1H, H β MeVal), 2.1 (m, 1H, H β Val), 0.96 and 0.89 (2 d, each 3H, H γ MeVal, $J_{\beta,\gamma}$ 6.4 Hz), 0.83 and 0.80 (2 d, each 3H, H γ Val, $J_{\beta,\gamma}$ 6.8 Hz). ¹³C NMR (CDCl₃): δ 171.7, 169.9 (C=O MeVal, Val), 157.2 (C=O Z), 136.3 (C_q Z), 128.2, 127.7, 127.4 (CH-arom.), 67.2 (CH₂ Z), 64.9, 56.6 (C α MeVal, Val), 51.7 (OCH₃), 30.6, 29.6 (C β MeVal, Val), 25.8 (NCH₃), 19.2, 18.6, 18.4, 17.3 (C γ MeVal, Val).

N-[*N*-(Benzyloxycarbonyl)-*L*-valyl]-*L*-*N*-methylvaline methyl ester, Z-Val-MeVal-OMe (**4f**). R_f 0.52 (system B). [α]_D –110.0° (c 1, MeOH); lit.¹⁵ [α]_D –63° (c 1, MeOH). MS (m/z): 347 [M+H-CH₃OH]⁺, 379 [M+H]⁺, 401 [M+Na]⁺. ¹³C NMR (CDCl₃): δ 173.0, 171.1 (C=O MeVal, Val), 156.3 (C=O Z), 136.3 (C_q Z), 128.4, 127.9, 127.7 (CH-arom.), 66.7 (CH₂ Z), 65.0, 61.4, 55.8, 55.6 (C α MeVal, Val), 51.7 (OCH₃), 31.4, 31.2 (C β MeVal, Val), 27.4, 26.9 (NCH₃), 19.7, 19.1, 18.6, 17.4 (C γ MeVal, Val).

N-(*N*-Benzyloxyvalyl)valine methyl ester, Bz-Val-Val-OMe (**4g**) (*diastereoisomeric mixture*). MS (m/z): 335 [M+H]⁺, 357 [M+Na]⁺. ¹H NMR (CDCl₃): δ 7.9–7.8 and 7.5–7.4 (2 m, 5H, H-arom.), 7.2–7.1 (m, 2H, NH), 4.8–4.7 and 4.6–4.5 (2 m, each 1H, H α), 3.75 and 3.67 (2 s, 3H, OCH₃ LL and DL form respectively), 2.1–2.3 (m, 2H, H β), 1.0–0.9 (m, 12H, H γ). ¹³C NMR (CDCl₃): δ 172.0, 171.8, 171.7, 171.5 (C=O Val), 167.3 (C=O Bz), 131.5, 128.4, 127.1 (CH-arom.), 58.6, 57.4 (C α), 51.9 (OCH₃), 31.4, 30.9, 30.7 (C β), 19.3, 19.1, 19.0, 18.9, 18.3, 18.1, 17.9, 17.7 (C γ).

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