Tetrahedron Letters,Vol.23,No.41,pp 4191-4194,1982 0040-4039/82/414191-04\$03.00/0 Printed in Great Britain ©1982 Pergamon Press Ltd.

SYNTHESIS OF 6-ALKYL AND 6-ARYL SUBSTITUTED 9-B-D-RIBOFURANOSYL PURINES VIA THE NICKEL CATALYZED COUPLING OF GRIGNARD REAGENTS TO 2',3',5'-Tris-0-(t-BUTYLDIMETHYLSILYL)-9-B-D-RIBOFURANOSYL-6-CHLOROPURINE

Donald E. Bergstrom and P. Anantha Reday

Department of Chemistry, University of North Dakota, Grand Forks, North Dakota, 58202, USA

Summary: A series of 6-substituted purine nucleosides have been synthesized in moderate yield by the nickel catalyzed cross coupling reaction between alkyl- and aryl- Grignard reagents and 2',3',5'-tris-0-(t-butyldimethylsilyl)-9-8-D-ribofuranosyl-6-chloropurine.

Adenosine analogues with alkyl and aryl group linked to the amino group at C-6 show a considerable range of biological activity.¹ For example, N^6 -cyclohexyladenosine strongly binds A_1 -adenosine receptors² and N^6 -(4-hydroxy-3-methyl-trans-butenyl)adenosine(zeatin riboside) is an active cytokinin.³ The amino group at C-6 is not always necessary for activity. The most potent inhibitors of nucleoside transport in erythrocytes are purine nucleosides to which a substituted benzyl group is attached via sulfur to C-6. 4 A number of 6-alkylated purines⁵ and their nucleoside analogues⁶ show interesting biological activity. However, despite efforts to synthesize 6-alkyl and 6-aryl nucleoside analogues for biological testing no direct broadly useful synthetic route to this class of compounds has been developed. Until recently attempts to form carbon-carbon bonds at the 6-position through nucleophilic displacement of the corresponding chloropurine nucleosides were generally unsuccessful. A notable exception is the preparation of 2,6-dialkylpurine nucleosides $\frac{8}{5}$ by the phosphorane alkylation method of Taylor and Martin.⁹ Recently, a number of papers have appeared on the reactions of 9-(2',3',5'-tri-O-benzyl)-B-D-ribofurancsides of 6-chloropurine and 6-methylsulfonylpurine with the carbanions from the active methylene compounds diethyl malonate, ethyl cyanoacetate, malononitrile and nitromethane, 10^{10} as a route to 6-alkylpurine ribonucleosides. Two rearrangement reactions, a photo-Claisen¹¹ and an Eschenmoser contraction¹² have also been used to create carbon-carbon bonds at C-6. None of these routes are of guite as broad scope, however, as the coupling reaction described here.

As part of our overall effort to synthesize modified nucleosides via organotransition metal chemistry we now report a short efficient synthesis of 6-substituted purine nucleosides. One other report of modification of purine nucleosides by organotransition metal intermediates has appeared recently. A palladium catalyzed reaction of Grignard reagents with the trimethylsilyl protected derivative of 8-bromoadenosine was used to prepare 8-alkyl- and aryl-adenosines.¹³ Following the independent reports of Corriu and Masse¹⁴ and Kumada <u>et al</u>¹⁵ in 1972, that metal phosphine complexes catalyse the selective cross coupling of Grignard reagents with aryl and alkenyl halides, a series of papers¹⁶ have appeared in the literature

on the synthetic utility of this reaction. Derivatives of 6-chloropurine would seem to be ideal candidates for substituion by this procedure.

 $2',3',5'-Tris-O-(t-buty]dimethy]sily])-9-\beta-D-ribofuranosyl-6-chloropurine (1) was$ $prepared from the known 6-chloro-9-<math>\beta$ -D-ribofuranosylpurine¹⁷ by silylation with an excess of t-butyldimethylsilyl chloride, imidazole and DMF by the literature procedure.¹⁸ Nucleoside <u>1</u> reacts readily with Grignard reagents via a nickel complex generated from dichloro[1,3-bis(diphenylphosphine)propane]nickel(II)¹⁹ [Ni(dppp)Cl₂] (Scheme I). The products, following deprotection with tetra-n-butylammonium fluoride in THF and chromatographic purification, were isolated in 40-50% yield (Table I).²⁰ The scope of the reaction extends to Grignard reagents generated from 1° and 2° alkyl and aryl halides. Consequently this method has somewhat greater scope than other methods. The primary limitation is undoubtedly that imposed by the presence of functional groups reactive towards Grignard reagents.

In a typical procedure: To a mixture of 2', 3', 5'-tris-O-(t-butyldimethylsilyl)-9-B-Dribofuranosyl-6-chloropurine (0.565 g, 1 mmole) and Ni(dppp)Cl₂ (0.075 g, 0.15 mmole) in anhydrous ether (75 ml) was added freshly prepared Grignard reagent (10 equivalents) at 0° under nitrogen atmosphere. The brown colored reaction mixture was allowed to stand at room temperature, with stirring, overnight. Hydrolysis with saturated aqueous NH₄Cl (25 ml) and extraction with ether followed by washing with water (2 x 50 ml), drying over anhydrous Na₂SO₄ and removal of the solvent at reduced pressure yielded, in all cases, a yellow viscous liquid.

The foregoing crude product, without purification, was treated with tetra-n-butylammonium fluoride (10 ml) 1 M in THF at 0° with the exclusion of moisture for 4 hr. THF was removed at reduced pressure and the black residue purified by chromatography on silica gel (MeOH-CHCl₃, 15:85 v/v) followed by Bio-gel P-2 (water). Lyophilization of the solution yielded colorless fluffy material in 40-50% overall yield. The actual yield of the coupling reaction is some what higher, but complete separation of the product from tetra-n-butylammonium salts and nickel catalyst requires chromatographic conditions that results in some loss of product. Studies are currently underway to assess the biological activity of these analogues. ACKNOWLEDGMENT

Mass spectra were run at the Midwest Center for Mass Spectrometry funded by the National Science Foundation. The authors express their appreciation to John Orwin for preliminary studies. This work was supported by Grant No. CA 30050 awarded by the National Cancer Institute, whom we gratefully acknowledge.

TABLE I

Products from the Reaction of Nucleoside 1 with Grignard Reagents

Grignard	Overall			Calcd. Mass	Deviation	
Reagent	Product ²¹	Yield (%)	<u>M.P.</u>	<u>(M + H⁺)</u>	(ppm)	
EthylMgI	3a	45	105	281.124981(C ₁₂ H ₁₇ N ₄ 0 ₄)	1.8	
Cyclohexy1MgBr	3Ъ	47	99-100	335.171931(C16H23N404)	1.0	
PhenylMgBr	3c	40	93	329.124981(C ₁₆ H ₁₇ N ₄ O ₄)	0.8	
2-PhenylethylMgBr	3d	50	67-70	10 17 4 1		
3-PhenylpropylMgBr	3e	48	62-63	371.17931(C ₁₉ H ₂₃ N ₄ O ₄)	1.6	
4-Methy1-3-penten-1-y1MgBr	Зf	42	48-49	15 25 4 4		

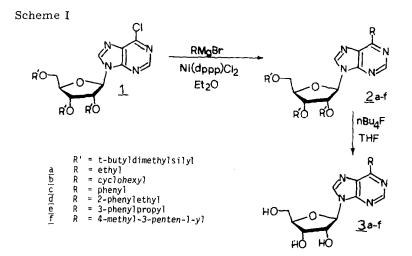


TABLE II	¹³ C NMR Spectra ^a								
Carbon									
Purine	<u>3a</u>	<u>3b</u>	<u>3c</u>	<u>3d</u>	<u>3e</u>	3f			
2 4 5 6 8	151.78 150.28 132.29 162.76 143.98	151.45 149.11 132.68 167.57 143.59	152.23 151.91 135.31 153.01 144.89	151.32 149.24 133.78 162.95 143.98	151.19 148.98 133.59 163.64 143.91	151.26 149.18 133.07 163.41 144.17			
β-D-ribose									
1' 2' 3' 4' 5'	87.78 73.75 70.43 85.77 61.40	90.77 73.62 71.73 87.00 62.63	87.65 73.68 70.17 85.64 61.14	90.90 73.62 71.86 87.00 62.70	90.77 73.49 71.79 87.00 62.70	90.70 73.75 71.57 87.00 62.83			
C-6 Substituent	1 / 2	$3 \frac{4}{2}$	3 4	2 3 4 5	3 4 5 6	3 4 6			
1 2 3 4 5 6 7	25.73 12.21	41.65 30.93 25.92 25.92	131.12 129.36 128.65 130.86	34.63 33.79 140.54 128.26 128.26 126.11	35.41 24.45 32.55 141.25 128.19 128.19 125.72	33.14 26.60 122.41 133.59 26.70 17.67			

 $^{\rm a}{\rm Spectra}$ were run in ${\rm CDCl}_{\rm 3}$ and are referenced to internal TMS.

REFERENCES

- 1. N.H. Fleysher, J. Med. Chem., 1972, 15, 187-91.
- 2. J.W. Daly, <u>J. Med. Chem.</u>, 1982, <u>25</u>, 197-207.
- 3. S.M. Hecht, N.J. Leonard, R.Y. Schmitz, and F. Skoog, Phytochemistry, 1970, 9, 1173.
- 4. A.R.P. Paterson and A.I. Simpson, Canada J. Biochem., 1966, 44, 1423-33.
- T.R. Henderson, C.R. Frihart, N.J. Leonard, R.Y. Schmitz and F. Skoog, <u>Phytochemistry</u>, 1975, <u>14</u>, 1687; F.S. Phillips, S.S. Stenberg, L. Hamilton, and D.A. Clark, <u>Ann. NY Acad.</u> <u>Sci.</u>, 1954, <u>60</u>, 283; D.A. Clark, F.S. Phillips, S.S. Stenberg, and C.C. Stock, ibid, 1954, <u>60</u>, 235.
- H.P. Schnebli, D.L. Hill, and L.L. Bennett, Jr., <u>J. Biol. Chem.</u>, 1967, <u>242</u>, 1997; J.A. Montgomery and K. Hewson, J. Med. Chem., 1968, 11, 48.
- J.D. Westover, G.R. Revanker, R.K. Robins, R.D. Madsen, J.R. Ogden, J.A. North, R.W. Mancuso, R.J. Rousseau, and E.L. Stephen, J. Med. Chem., 1981, 24, 941.
- 8. L.F. Christensen, P.D. Cook, R.K. Robins, and R.B. Meyer, Jr., <u>J. Carbohydr. Nucleosides</u>, <u>Nucleotides</u>, 1977, 4, 175-88.
- 9. E.C. Taylor and S.F. Martin, J. Am. Chem. Soc., 1974, 96, 8095.
- T. Miyasaka, H. Suemune, and K. Arakawa, <u>Nucleic Acids Res. Special Publ.</u>, 1978, <u>5</u>, 273-6; A. Yamane, Y. Nomota, A. Matsuda, and T. Ueda, <u>ibid</u>, 209-312; A. Yamane, A. Matsuda, and T. Ueda, <u>Chem. Pharm. Bull.</u>, 1980, <u>28</u>, 150-6.
- 11. E. Takeshi and J. Zemlicka, Nucleic Acids Symp. Ser., 1981, 9, 33-6.
- 12. H. Vorbrüggen and K. Krolikiewicz, Angew. Chem. Int. Ed. Engl., 1976, 15, 689.
- 13. N. Cong-Danh, J-P. Beaucourt, and L. Pichat, Tetrahedron Lett., 1979, 3159-62.
- 14. R.J.P. Corriu and J.P. Masse, J. Chem. Soc. Chem. Comm., 1972, 144.
- 15. K. Tamao, K. Sumitani, and M. Kumada, J. Am. Chem. Soc., 1972, 94, 4374.
- K. Tamao, K. Sumitani, Y. Kiso, M. Zembayashi, A. Fujioka, S. Kodama, I. Nakajima, A. Minato, and M. Kumada, Bull. Chem. Soc., Japan., 1976, 49, 1958 and references therein.
- J. Zemlicka and J. Owens in "Nucleic Acid Chemistry", Part 2, L.B. Townsend and R.S. Tipson, Eds., John Wiley and Sons, NY, 1976, p. 611.
- K.K. Ogilvie, K.L. Sadana, E.A. Thompson, M.A. Quilliam and J.B. Westmore, <u>Tetrahedron</u> <u>Lett.</u>, 1974, 2861.
- 19. M. Kumada, K. Tamao, and K. Sumitani, Org. Synth., 1973, 58, 127.
- 20. A shorter route, devised with J. Orwin, required only four steps. Inosine was protected at the three hydroxyl groups with t-butyldimethylsilyl and the protected derivative converted to nucleoside $\underline{1}$ with SOCl₂ in DMF. However, this latter reaction leads to size products which makes purification difficult and overall yields are low as a consequence.
- 21. All compounds were characterized by ¹H NMR, ¹³C NMR, UV, and Fast Atom Bombardment (FAB) low and high resolution Mass Spectrometry. Cf. D.H. Williams, C. Bradley, G. Bojesen, S. Santikarn, and L.C.E. Taylor, J. Am. Chem. Soc., 1981, 103, 5700.

(Received in USA 21 June 1982)