Design and Synthesis of a Tag-Free Chemical Probe for Photoaffinity Labeling

Timo Mayer^[a] and Martin E. Maier^{*[a]}

Keywords: Photoaffinity labeling / Click chemistry / Triazoles / 1,3-Dipoles / Diazirines / Nitrogen heterocycles

The novel aromatic diazirine-containing benzoic acid 22 was prepared via the diazirine $\mathbf{11}$ as the key intermediate. After formylation of the aryl ring and cleavage of the methyl ether function of aldehyde 12, the phenolic hydroxy group was converted into the ether 21 terminating in an alkyne function. Oxidation of the aldehyde to the carboxylic acid provided the chemical probe 22 designed for tag-free photoaffinity labeling. In a proof-of-concept study it could be shown that irradiation of the simple ester 23 indeed yields the methanol insertion product 24. A subsequent click reaction with benzyl azide 20 led to the triazole 25. A more complicated example was realized with the esterification of bafilomycin A₁ (27) with acid 22.

(© Wiley-VCH Verlag GmbH & Co. KGaA, 69451 Weinheim, Germany, 2007)

Introduction

A timeless challenge in biology and drug discovery involves finding a matching pair between one entity and a large number of possible partners. Thus, libraries of small molecules have been screened for binding to or inhibition of an enzyme.^[1,2] Also the target of an active compound, frequently a natural product, needs to be found. Depending on the nature of the ligand, activity-based profiling^[3] or affinity chromatography can be used. However, in some cases like low-abundance proteins, recourse to photoaffinity labeling (PAL) has to be made.^[4] Besides target fishing, PAL can be used to elucidate the structural and functional properties of biological systems. In PAL a ligand is connected to a photoreactive group. Irradiation of the bound ligand-label conjugate generates a reactive intermediate that hooks onto the protein of interest by forming a covalent bond. Among various photophores, aryl(trifluoromethvl)diazirines^[5] have found widespread applications. They are readily photolyzed to reactive carbenes at around 360 nm.^[6] Furthermore, they can be kept small enough that the biological activity of the modified ligand does not suffer too much. In order to identify the PAL products after binding and photolysis, a reporter group should be contained within the photophore or somewhere else in the ligand.^[7] Historically, radioisotopes have been employed in this context (cf. compounds 1,^[8,9] Figure 1). As an alternative to radioisotopes, photoaffinity biotinylation can be used (cf. compound $2^{[10-12]}$ Figure 1). However, the polar and big biotin-anchored tag often renders the ligand-affinity conju-

[a] Institut für Organische Chemie, Universität Tübingen, Auf der Morgenstelle 18, 72076 Tübingen, Germany Fax: +49-7071-295137 E-mail: martin.e.maier@uni-tuebingen.de

gate substantially less active than the parent ligand. In order to circumvent these shortcomings compact bifunctional probes carrying a small functional group for selective modification after photoaffinity labeling (post-PAL modification, PPALM) have recently been described (cf. 3, 4, and 5). These are formed by catalyzed azide-alkyne [3+2] cycloaddition^[13-16] or Staudinger-Bertozzi ligation reactions.^[17,18] In fact, Suzuki and co-workers showed that the azidoalkyl group of an ether derivative of diazirine 3 largely survives the conditions used for photoactivation of the 3-(trifluoromethyl)-3H-diazirin-3-yl group.^[19] They also showed that the sequence of photoreaction followed by cycloaddition with benzyl azide worked in vitro on a derivative of **4**.^[20]



Figure 1. Some important examples of photoaffinity probes based on 3-(trifluoromethyl)-3H-diazirines and azides.

In connection with the task of localizing the binding partner of a natural product a tag-free 4-[3-(trifluoromethyl)-3H-diazirinyl]benzoic acid was sought. The idea was to

Supporting information for this article is available on the WWW under http://www.eurjoc.org or from the author.

shorten the side-chain of benzoic acid 2 and terminate it with an azide or alkyne. This should allow selective fishing of a labeled protein by click chemistry. In this paper we detail the synthesis of the bifunctional benzoic acid 22 and illustrate the sequence of carbene trapping followed by click chemistry on a simple model system.

Results and Discussion

Starting from 3-bromoanisole (6) the key intermediate 3-(3-methoxyphenyl)diazirine 11 was prepared using the classical sequence^[21] for introduction of the 3-(trifluoromethyl)diazirin-3-yl group (Scheme 1). Compared with the literature methods some steps were slightly modified. Thus, metalation of bromide 6 with *n*-butyllithium followed by acylation of the aryllithium intermediate with methyl trifluoroacetate gave ketone 7 in excellent yield. Ketone 7 was converted into the oxime 8 using hydroxylamine hydrochloride in the presence of pyridine. Treatment of the oxime 8 (E/Zmixture) with tosyl chloride led to the *p*-tolylsulfonyloxime 9. All these steps proceeded in almost quantitative yields. Formation of the diaziridine 10 using ammonia followed by oxidation of **10** with silver oxide^[21a,21b] furnished diazirine 11. The most convenient way to introduce the formyl group involves Friedel-Crafts alkylation of anisole 11 with dichloromethyl methyl ether in the presence of TiCl₄ followed by aqueous work-up.^[22] These conditions gave the desired aldehyde 12 (41%) along with the regioisomer 13 (9%). The two aldehydes could be separated by flash chromatography. In order to prepare for the attachment of the side-chain, the aryl methyl ether was cleaved using boron tribromide in dichloromethane which led to hydroxybenzaldehyde 14.

It was then planned to form an ether bond between phenol **14** and a suitable alkynol derivative. As shown in Scheme 2 the tosylate **16** was prepared in two steps by monoalkylation of ethanediol with propargyl bromide^[23] followed by tosylation of the alkynol **15**. In order to demonstrate a click reaction of the alkyne functionality, we chose the simple benzyl azide **20**. This compound was easily accessible by routine steps from 3-methoxybenzaldehyde (**17**). Thus, reduction of the aldehyde to the benzyl alcohol **18** was followed by bromination using PBr₃ leading to benzyl bromide **19**. A final nucleophilic substitution of the bromide with sodium azide in DMSO provided the azide **20**. The methoxy group would later facilitate the identification of the click product by NMR spectroscopy.

Alkylation of the 2-hydroxybenzaldehyde 14 with the tosylate 16 was performed at a slightly elevated temperature in the presence of potassium carbonate and catalytic amounts of tetrabutylammonium iodide in DMF (Scheme 3). These conditions gave the ether 21 in 75% yield. The etherification was also possible under Mitsunobu conditions.^[24] However, the yield was lower and separation of the byproducts turned out to be problematic. The final oxidation of the aldehyde 21 to the acid 22 was rather problematic. Typical reagents for related oxidation reactions such as permanganate did not work.^[21] Finally, we found the combination



Scheme 1. Synthesis of the 4-[3-(trifluoromethyl)-3*H*-diazirin-3-yl] benzaldehyde **14**.





Scheme 2. Preparation of the alkyne 16 and benzyl azide 20.

of sodium chlorite with sulfamic acid to be suitable.^[25] This way the benzoic acid **22** could be obtained in 94% yield. In large-scale reactions, simple extraction of the reaction mixture followed by concentration of the organic layer pro-

vided the pure acid **22** as an amorphous solid without any further purification. As a model the acid **22** was esterified with 2-propanol using the condensing agent 1-ethyl-3-(dimethylaminopropyl)carbodiimide (EDC) providing ester **23** in reasonable yield.





Scheme 3. Synthesis of the photoaffinity probe 22 by alkylation of phenol 14 and oxidation of the benzaldehyde 21.

By irradiating a solution of diazirine 23 in methanol with a UV lamp (8 W, 366 nm), the corresponding carbene was generated, as evidenced by the isolation of the methanol adduct 24 (Scheme 4). This adduct is characterized by the methoxy signal ($\delta = 3.41$ ppm) and the methine hydrogen (δ = 4.48 ppm). Owing to the CF₃ group the latter appears as a quartet ($^{2}J = 6.6$ Hz). Although the yield of adduct 24 was moderate, its formation shows that the side-chain does not react with the intermediate carbene. According to LC-MS analysis, irradiation on a larger scale produces the reduced derivative (2,2,2-trifluoroethyl substituent) as a sideproduct. In a subsequent click reaction between alkyne 24 and the azide 20 the triazole 25 was formed. Remarkably, the order of events could also be reversed. Thus, the diazirine function seems to be compatible with the conditions of the click reaction, as indicated by the successful formation of triazole 26. This shows that the alkyne function might be used as a handle for conjugation with other small groups prior to irradiation. Subjection of diazirine 26 to irradiation in methanol gave compound 25 as well.

In order to illustrate the potential use of acid **22** we treated it with the natural product bafilomycin A_1 (**27**). This macrolactone, isolated from *Streptomyces griseus*, is a well-known inhibitor of V-ATPase.^[26,27] In this regard it is closely related to concanamycin A.^[28] As has been described before, the hydroxy group at C-21 can be selectively acylated, alkylated, and oxidized without affecting the V-ATPase activity too much.^[29] On the other hand, modifications to 7-OH cause a significant reduction in activity. Ac-

Scheme 4. Photoreaction of the alkynyl-functionalized phenyl(trifluoromethyl)diazirine 23 in methanol followed by a click reaction with benzyl azide 20.

cordingly, reaction of bafilomycin A₁ (27) with acid 22 (3 equiv.) in the presence of the condensing agent EDC and DMAP provided the ester 28 in good yield (Scheme 5). Even though an excess of acid 22 was used, besides ester 28 some unreacted 27 was recovered. In the NMR spectrum the signal of the methine 21-H resonates at $\delta = 5.11-5.20$ ppm as opposed to 3.53 ppm in 27. The carbon atom C-21 appears at $\delta = 75.6$ ppm ([D₆]acetone) as opposed to 70.4 ppm.



Scheme 5. Esterification of bafilomycin A1 (27) at 21-OH with acid 22 yielding ester 28.

We also demonstrated a click reaction with the ester 28. It seemed that the azido-biotin derivative 32 would be a suitable reagent (Scheme 6).^[30] In order to prepare this reagent, 2-bromoethylamine was converted into azidoethylamine (29) by treating it with sodium azide.^[31] In a separate operation biotin (30) was activated as its N-hydroxysuccinimide derivative 31[32] in a condensation reaction performed in the presence of the reagent EDC. Thereafter, treatment of N-hydroxysuccinimidobiotin (31) with azidoethylamine (29) in DMF in the presence of triethylamine furnished the azide^[30] 32 in good yield. When the azidobiotin derivative 32 was treated with the bafilomycin A_1 derivative 28 under typical click conditions (copper sulfate, sodium ascorbate as reducing agent) in a mixture of tBuOH and H₂O the ligation product 33 could be isolated in a moderate yield as a colorless amorphous solid. The two



Scheme 6. Synthesis of the biotin azide **32** and its click reaction with the alkyne **28**.

bafilomycin A_1 derivatives **28** and **33** are currently being tested for inhibition of V-ATPase. Owing to the smaller appendage one would expect a higher activity for **28** over **33**.

Conclusions

In this paper we have described the synthesis of the novel bifunctional probe **22** featuring the 3-(trifluoromethyl)-3*H*-diazirin-3-yl group as well as an alkynyl side-chain. In a model study we have shown that the carbene insertion product can be evidenced by a Cu^I-catalyzed Huisgen alkyne-azide 1,3-dipolar cycloaddition reaction. Esterification of bafilomycin A₁ (**27**) with the acid **22** gave the ester **28**. In a click reaction with the biotinyl azide **32** the triazole **33** was obtained. With the availability of biotinyl azides such as **32** or fluorescence dyes containing an azide group,^[13] applications of **22** in chemical biology present itself. Studies along these lines are currently being pursued.

Experimental Section

General: ¹H and ¹³C NMR: Bruker Avance 400 spectrometer, Bruker AMX 600 spectrometer; spectra were recorded at 295 K in CDCl₃, [D₆]acetone, or [D₆]DMSO; chemical shifts are calibrated to the residual proton and carbon resonances of the solvent: CDCl₃ $(\delta_{\rm H} = 7.25, \delta_{\rm C} = 77.0 \text{ ppm}), [D_6]$ acetone $(\delta_{\rm H} = 2.04, \delta_{\rm C} = 29.8 \text{ ppm}),$ $[D_6]DMSO$ (δ_H = 2.49, δ_C = 39.5 ppm). Melting points: Büchi Melting Point B-540, uncorrected. HRMS (FT-ICR): Bruker Daltonic APEX 2 with electron-spray ionization (ESI). The minimal resolution of this machine is 1 ppm ($\Delta m/m \times 10^6$). Flash chromatography: J. T. Baker silica gel, 43-60 µm. Thin-layer chromatography: Macherey-Nagel Polygram Sil G/UV₂₅₄ plates. Analytical LC-MS: HP 1100 Series connected with an ESI-MS Agilent G1946C detector, positive mode with a fragmentor voltage of 40 eV; column: Nucleosil 100-5, C-18 HD, 5 mm, 70 × 3 mm Macherey-Nagel; eluent: NaCl solution (5 mM)/acetonitrile; gradient: 0-10-15-17-20 min with 20-80-80-99-99% acetonitrile; flow: 0.5 mLmin⁻¹. All solvents used in the reactions were distilled before use. Dry tetrahydrofuran was distilled from sodium and benzophenone, whereas dry CH2Cl2, dimethylformamide, pyridine, and DMSO were distilled from CaH₂. Petroleum ether with a boiling range of 40-60 °C was used. Reactions were generally run under argon. All commercially available compounds were used as received unless stated otherwise. Reactions with diazirine-containing compounds were protected from light. Bafilomycin A1 was a gift from Dr. Carlo Farina, Brane Discovery Srl, via Zambeletti 25, 20021 Baranzate (MI), Italy.

2,2,2-Trifluoro-1-(3-methoxyphenyl)ethanone (7):^[21b,33] *n*BuLi (2.5 M in hexane, 28 mL, 1.5 equiv.) was slowly added to a solution of 1-bromo-3-methoxybenzene (8.9 g, 6.0 mL, 47.6 mmol) in dry THF (100 mL) at -78 °C. The reaction mixture was stirred at this temperature for 2 h before a solution of methyl trifluoroacetate (8.1 g, 6.3 mL, 63.3 mmol, 1.3 equiv.) in dry THF (40 mL) was added over a period of 30 min. After the mixture had been stirred for an additional 2 h, a solution of conc. HCl (10.5 mL) in methanol (25 mL) was added. The resulting solution was warmed up to 0 °C, diluted with Et₂O (60 mL), washed with water (2 × 10 mL), dried with MgSO₄, and filtered. The solvent was evaporated in vacuo and the crude product purified by kugelrohr distillation, 60 °C/10⁻¹ mbar (ref.^[33] 84–85 °C/12 Torr) to give the title com-



pound (9.38 g, 46.0 mmol, 96%) as a colorless liquid. $R_{\rm f} = 0.21$ (hexane/EtOAc, 5:1). ¹H NMR (400 MHz, CDCl₃): $\delta = 3.86$ (s, 3 H, CH₃), 7.24 (dd, J = 8.1, 2.0 Hz, 1 H, 4-H), 7.44 (t, J = 8.0 Hz, 1 H, 5-H), 7.55 (s, 1 H, 2-H), 7.65 (d, J = 7.6 Hz, 1 H, 6-H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 55.5$ (CH₃), 113.9 (d, J = 1.5 Hz, C-2), 116.6 (q, ¹ $J_{\rm CF} = 291.3$ Hz, CF₃), 122.3 (C-4), 122.7 (q, J = 3.0 Hz, CCF₃) ppm.

2,2,2-Trifluoro-1-(3-methoxyphenyl)ethanone Oxime (8):^[33] A solution of 2,2,2-trifluoro-1-(3-methoxyphenyl)ethanone (7) (9.38 g, 46.0 mmol) and hydroxylamine hydrochloride (4.2 g, 60.4 mmol, 1.3 equiv.) in pyridine (100 mL) and dry ethanol (50 mL) was refluxed at an oil-bath temperature of 85 °C for 14 h. The solvent was removed under reduced pressure and the resulting residue partitioned between Et₂O (80 mL) and water (60 mL). The organic layer was washed with 1 N HCl $(3 \times 10 \text{ mL})$ and water $(1 \times 10 \text{ mL})$, dried with MgSO₄, and filtered. Evaporation of the solvent afforded the pure oxime (9.8 g, 44.72 mmol, 97%) as a colorless liquid (*E*/*Z* mixture, 6:4 ratio). $R_{\rm f} = 0.63$ (CHCl₃/EtOAc, 10:1). ¹H NMR (400 MHz, CDCl₃): δ = 3.83 (s, 3 H, CH₃), 6.98–7.11 (m, 3 H, 2-H, 4-H, 6-H), 7.33 (t, J = 8.3 Hz, 0.4 H, 5-H), 7.40 (t, J =7.8 Hz, 0.6 H, 5-H), 9.19 (s, 0.6 H, NOH), 9.34 (s, 0.4 H, NOH) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 55.36 (CH₃), 55.38 (CH₃) 113.9 (C-2), 114.2 (C-2), 116.1 (C-4), 116.3 (C-4), 118.2 (q, ${}^{1}J_{CF}$ = 282.5 Hz, CF₃), 120.5 (q, ${}^{1}J_{CF}$ = 275.2 Hz, CF₃), 120.7 (C-6), 120.8 (C-6), 127.0 (C-1) 129.65 (C-5), 129.73 (C-5), 131.1 (C-1), 147.5 (q, ${}^{2}J_{CF}$ = 32.9 Hz, CCF₃), 147.7 (q, ${}^{2}J_{CF}$ = 30.7 Hz, CCF₃), 159.40 (C-3), 159.54 (C-3) ppm.

2,2,2-Trifluoro-1-(3-methoxyphenyl)-N-{[(4-methylphenyl)sulfonyl]oxy}ethanimine (9): Toluenesulfonyl chloride (4.3 g, 22.5 mmol, 1.5 equiv.) was added to a solution of 2,2,2-trifluoro-1-(3-methoxyphenyl)ethanone oxime (8) (3.3 g, 15.0 mmol) in pyridine (50 mL) and the resulting solution was refluxed for 2 h at an oilbath temperature of 125 °C. After cooling, the solvent was evaporated and the residue partitioned between Et₂O (250 mL) and water (60 mL). The organic layer was washed with 1 ${\rm N}$ HCl (3 $\times 15$ mL) and water $(1 \times 15 \text{ mL})$, dried with MgSO₄, and filtered. Evaporation of the solvent afforded the pure title compound (5.5 g, 14.7 mmol, 98%) as a colorless solid, m.p. 111 °C. $R_{\rm f} = 0.17$ (toluene). ¹H NMR (400 MHz, CDCl₃): δ = 2.47 (s, 3 H, CH₃), 3.81 (s, 3 H, OCH₃), 6.87 (s, 1 H, 2-H), 6.93 (d, J = 7.6 Hz, 1 H, 4-H), 7.04 (dd, J = 8.5, 1.9 Hz, 1 H, 6-H), 7.37 (t, J = 7.3 Hz, 3 H, 5-H, 3'-H, 5'-H), 7.87 (d, J = 8.3 Hz, 2 H, 2'-H, 6'-H) ppm. ¹³C NMR $(100 \text{ MHz}, \text{CDCl}_3): \delta = 21.7 \text{ (CH}_3), 55.4 \text{ (OCH}_3), 113.9 \text{ (C-2)},$ 117.1 (C-4), 119.5 (q, ${}^{1}J_{CF}$ = 277.4 Hz, CF₃), 120.4 (C-6), 125.6 (C-5), 129.2 (C-3', C-5'), 129.9 (C-2', C-6'), 130.0 (aryl C), 131.1 (aryl C), 146.1 (C-4'), 153.9 (q, ${}^{2}J_{CF}$ = 33.6 Hz, CCF₃), 159.5 (C-3) ppm.

3-(3-Methoxyphenyl)-3-(trifluoromethyl)diaziridine (10):^[21b,33] A solution of 2,2,2-trifluoro-1-(3-methoxyphenyl)-*N*-{[(4-meth-ylphenyl)sulfonyl]oxy}ethanimine (**9**) (5.5 g, 14.73 mmol) in dry CH₂Cl₂ (65 mL) was added to liquid ammonia (50 mL) at -78 °C over a period of 1 h. The solution was stirred at -78 °C for 12 h and then warmed up to room temperature overnight which led to evaporation of the excess ammonia and some of the CH₂Cl₂. The colorless solid was removed by filtration and the filtrate was washed with water (3×5 mL). The organic layer was dried with MgSO₄, filtered, and concentrated in vacuo. The resulting crude product was purified by flash chromatography (CHCl₃/EtOAc, 20:1) to give the diaziridine **10** (2.39 g, 11.0 mmol, 74%) as a slightly yellow liquid. *R*_f = 0.30 (CHCl₃/EtOAc, 20:1). ¹H NMR (400 MHz, CDCl₃): δ = 2.24 (d, *J* = 8.6 Hz, 1 H, NH), 2.78 (d, *J* = 8.3 Hz, 1 H, NH), 3.81 (s, 3 H, CH₃), 6.97 (d, *J* = 8.3 Hz, 1 H,

4-H), 7.14 (s, 1 H, 2-H), 7.19 (d, J = 7.6 Hz, 1 H, 6-H), 7.32 (t, J = 8.0 Hz, 1 H, 5-H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 55.3$ (CH₃), 58.0 (q, ² $J_{CF} = 35.9$ Hz, CCF_3), 113.6 (C-2), 115.8 (C-4), 120.3 (C-6), 123.5 (q, ¹ $J_{CF} = 278.8$ Hz, CF₃), 129.9 (C-5), 133.0 (C-1), 159.7 (C-3) ppm.

3-(3-Methoxyphenyl)-3-(trifluoromethyl)-3H-diazirine (11): A solution of sodium hydroxide (2.4 g, 60.0 mmol) in water (30 mL) was added slowly to a boiling solution of silver(I) nitrate (10.2 g, 60.0 mmol) in water (60 mL). The precipitate was filtered and washed with water (100 mL), acetone (100 mL), and Et₂O (100 mL) to give Ag₂O as a brown solid. The freshly prepared Ag₂O (9.5 g, 41.0 mmol, 3.8 equiv.) was added to a solution of diaziridine 10 (2.38 g, 10.9 mmol) in dry Et₂O (100 mL) and stirred at room temperature for 3.5 h. The dispersion was filtered, the filter cake rinsed with Et₂O, and the organic layer dried with MgSO₄. Filtration and evaporation of the solvent gave the pure diazirine 11 (2.17 g, 10.04 mmol, 92%) as a colorless liquid. $R_{\rm f} = 0.84$ (CHCl₃/EtOAc, 20:1). ¹H NMR (400 MHz, CDCl₃): δ = 3.80 (s, 3 H, CH₃), 6.69 (s, 1 H, 2-H), 6.77 (d, J = 7.6 Hz, 1 H, 4-H), 6.91–6.97 (m, 1 H, 6-H), 7.30 (t, J = 8.1 Hz, 1 H, 5-H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 28.4 (q, ²*J*_{CF} = 40.3 Hz, *C*CF₃), 55.3 (CH₃), 112.2 (d, J = 1.5 Hz, C-2), 115.2 (C-4), 118.7 (C-6), 122.1 (q, ${}^{1}J_{CF} =$ 274.4 Hz, CF₃), 130.0 (C-5), 130.5 (C-1), 159.8 (C-3) ppm.

2-Methoxy-4-[3-(trifluoromethyl)-3*H*-diazirin-3-yl]benzaldehyde (12) and 4-Methoxy-2-[3-(trifluoromethyl)-3*H*-diazirin-3-yl]benzaldehyde (13):^[22a] TiCl₄ (460 mg, 2.4 mmol, 1.5 equiv.) at 0 °C, followed by dichloromethyl methyl ether (276 mg, 2.4 mmol, 1.5 equiv.) was slowly added to a stirred solution of diazirine 11 (350 mg, 1.62 mmol) in dry CH₂Cl₂ (15 mL). The reaction mixture was warmed up to room temperature and stirred at this temperature for 1 h before it was cooled again to 0 °C and quenched with water (10 mL). The organic phase was diluted with CH₂Cl₂ (15 mL) and washed with water (2×5 mL), saturated NaHCO₃ solution (2×5 mL), and water (2×5 mL). The organic layer was dried with MgSO₄, filtered, and concentrated in vacuo. The resulting crude product was purified by flash chromatography (hexane/EtOAc, 5:1) to afford aldehyde 12 (161 mg, 0.66 mmol, 41%) and the isomeric aldehyde 13 (34 mg, 0.14 mmol, 9%) as yellow oils.

Aldehyde 12: $R_{\rm f} = 0.40$ (hexane/EtOAc, 5:1). ¹H NMR (400 MHz, CDCl₃): $\delta = 3.92$ (s, 3 H, CH₃), 6.68 (s, 1 H, 3-H), 6.83 (d, J = 8.1 Hz, 1 H, 5-H), 7.82 (d, J = 8.1 Hz, 1 H, 6-H), 10.42 (s, 1 H, CHO) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 28.6$ (q, ² $J_{\rm CF} = 40.3$ Hz, CCF₃), 55.8 (CH₃), 109.5 (C-3), 118.6 (C-5), 121.8 (q, ¹ $J_{\rm CF} = 274.4$ Hz, CF₃), 125.4 (C-1), 129.0 (C-6), 136.6 (C-4), 161.5 (C-2), 188.7 (CHO) ppm.

Aldehyde 13: $R_{\rm f} = 0.28$ (hexane/EtOAc, 5:1). ¹H NMR (400 MHz, CDCl₃): $\delta = 3.88$ (s, 3 H, CH₃), 7.05 (dd, J = 8.7, 2.2 Hz, 1 H, 5-H), 7.16 (s, 1 H, 3-H), 7.89 (d, J = 8.6 Hz, 1 H, 6-H), 10.38 (s, 1 H, CHO) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 27.2$ (q, ² $J_{\rm CF} = 42.5$ Hz, *C*CF₃), 55.8 (CH₃), 116.2 (aryl C), 116.8 (aryl C), 121.6 (q, ¹ $J_{\rm CF} = 275.2$ Hz, CF₃), 129.1 (aryl C), 130.8 (aryl C), 133.8 (aryl C), 164.6 (C-4), 188.2 (CHO) ppm.

2-Hydroxy-4-[3-(trifluoromethyl)-3*H***-diazirin-3-yl]benzaldehyde** (14): Tribromoborane (1 $mathbb{m}$ in CH₂Cl₂, 5.5 mL, 5.5 mmol, 1.1 equiv.) was added slowly to a solution of 2-methoxy-4-[3-(trifluoromethyl)-3*H*-diazirin-3-yl]benzaldehyde (12) (1.2 g, 4.91 mmol) in dry CH₂Cl₂ (15 mL) at -20 °C. The reaction was warmed up to 0 °C, stirred at this temperature for 1 h, and then quenched with water (5 mL). The solution was diluted with CH₂Cl₂ (60 mL), the layers were separated, and the organic layer washed with saturated NaCl solution (3 × 10 mL), dried with MgSO₄, filtered, and concentrated in vacuo. The crude product was purified by flash chromatography (hexane/CH₂Cl₂, 2:1) to give the hydroxy aldehyde **14** (870 mg, 3.78 mmol, 77%) as a colorless solid, m.p. 44 °C. $R_{\rm f}$ = 0.32 (hexane/CH₂Cl₂, 2:1). ¹H NMR (400 MHz, CDCl₃): δ = 6.75– 6.80 (m, 2 H, 3-H, 5-H), 7.59 (d, J = 7.8 Hz, 1 H, 6-H), 9.91 (s, 1 H, CHO), 11.04 (s, 1 H, OH) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 28.4 (q, ²J_{CF} = 41.0 Hz, *C*CF₃), 115.8 (C-3), 117.3 (d, J = 1.5 Hz, C-5), 120.8 (C-1), 121.6 (q, ¹J_{CF} = 275.2 Hz, CF₃), 133.9 (C-6), 138.0 (C-4), 161.3 (C-2), 195.9 (CHO) ppm. HRMS (ESI): calcd. for C₉H₃F₃N₂O₂ [M + H]⁺ 229.02304; found 229.02302.

2-(Prop-2-ynyloxy)ethanol (15): Finely pulverized NaOH (2.16 g, 54 mmol, 1.2 equiv.) was added to a vigorously stirred solution of 3-bromoprop-1-yne (5.34 g, 44.9 mmol) and ethylene glycol (5.57 g, 89.8 mmol, 2 equiv.) at 0 °C. The resulting suspension was refluxed at 45 °C for 4 h, filtered, and the filtrate extracted with CHCl₃ (3 × 25 mL). The combined organic layers were washed with water (3 × 10 mL), dried with MgSO₄, filtered, and concentrated in vacuo. The crude product was purified by flash chromatography (hexane/EtOAc, 2:1) to afford alcohol **15** (1.62 g, 16.18 mmol, 30%) as a colorless liquid. $R_f = 0.18$ (CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 2.43$ (t, J = 2.3 Hz, 1 H, C≡CH), 2.71 (s, 1 H, OH), 3.56–3.61 (m, 2 H, CH₂), 3.68–3.72 (m, 2 H, CH₂), 4.15 (d, J = 2.3 Hz, 2 H, CH₂C≡C) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 58.2$ (CH₂C≡C), 61.4 (CH₂OH), 71.1 (CH₂CH₂OH), 74.7 (C≡CH), 79.4 (C≡CH) ppm.

2-(Prop-2-ynyloxy)ethyl Toluenesulfonate (16): Solid p-toluenesulfonyl chloride (1.14 g, 6 mmol, 1.1 equiv.) was added to a stirred solution of 2-(prop-2-ynyloxy)ethanol (15) (546 mg, 5.46 mmol) in pyridine (10 mL) at 0 °C. After 17 h saturated NaHCO₃ solution (20 mL) was added and the reaction mixture was stirred at the same temperature for another 10 min before it was poured into a saturated NaHCO₃ solution (20 mL) at room temperature. The resulting mixture was extracted with Et_2O (3 × 20 mL). The combined organic layers were washed with 1 N HCl ($3 \times 20 \text{ mL}$), water $(1 \times 20 \text{ mL})$, dried with MgSO₄, filtered, and concentrated in vacuo. The crude product was purified by flash chromatography (hexane/EtOAc, 3:1) to give the tosylate 16 (1.13 g, 4.45 mmol, 82%) as a colorless liquid. $R_f = 0.62$ (hexane/EtOAc, 3:2). ¹H NMR (400 MHz, CDCl₃): $\delta = 2.41$ (t, J = 2.4 Hz, 1 H, C=CH), 2.42 (s, 3 H, CH₃), 3.68-3.72 (m, 2 H, pTsOCH₂CH₂), 4.09 (d, J = 2.3 Hz, 2 H, CH₂C≡C), 4.14–4.18 (m, 2 H, *p*TsOCH₂), 7.33 (d, J = 8.1 Hz, 2 H, 2-H, 6-H), 7.79 (d, J = 8.3 Hz, 2 H, 3-H, 5-H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 21.6 (CH₃), 58.3 $(CH_2C\equiv C)$, 67.1 (*p*TsOCH₂), 68.8 (*p*TsOCH₂*C*H₂), 75.0 (C=*C*H), 78.9 (C≡CH), 128.0 (C-2, C-6), 129.8 (C-3, C-5), 132.9 (C-1), 144.9 (C-4) ppm

(3-Methoxyphenyl)methanol (18): Sodium borohydride (777 mg, 20.6 mmol, 0.5 equiv.) was added to a solution of 3-methoxybenzaldehyde (17) (5.6 g, 41.1 mmol) in THF (120 mL) and H₂O (4 mL) and the mixture stirred at room temperature for 20 min. Then, water (120 mL) was added and the solution stirred for another 5 min. The mixture was extracted with CH_2Cl_2 (3×75 mL). The combined organic layers were dried with MgSO₄, filtered, and concentrated in vacuo. This way the benzylic alcohol 18 (5.58 g, 40.39 mmol, 98%) was obtained as a colorless liquid, sufficiently pure for the subsequent step. $R_{\rm f} = 0.18$ (CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ = 1.96 (t, J = 5.2 Hz, 1 H, OH), 3.80 (s, 3 H, OCH₃), 4.65 (d, J = 5.5 Hz, 2 H, CH₂), 6.80–6.85 (m, 1 H, 4-H), 6.90–6.94 (m, 2 H, 2-H, 6-H), 7.26 (t, J = 8.1 Hz, 1 H, 5-H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 55.2 (CH₃), 65.2 (CH₂), 112.2 (C-2), 113.2 (C-4), 119.1 (C-6), 129.5 (C-5), 142.5 (C-1), 159.8 (C-3) ppm.

1-(Bromomethyl)-3-methoxybenzene (19): Phosphorous tribromide (705 mg, 2.6 mmol, 0.36 equiv.) was added to a solution of (3-meth-

oxyphenyl)methanol (18) (1.0 g, 7.24 mmol) in dry CH₂Cl₂ (25 mL) at 0 °C. The mixture was stirred at room temperature for 90 min before it was treated with cold water (5 mL). The layers were separated and the water phase extracted with CH₂Cl₂ (3×5 mL). The combined organic layers were washed with water (2×10 mL), a saturated NaHCO₃ solution (1×10 mL), a saturated NaHCO₃ solution (1×10 mL), a saturated NaCl solution (1×10 mL), dried with MgSO₄, filtered, and concentrated in vacuo to afford the pure bromide 19 (1.1 g, 5.47 mmol, 76%) as a colorless liquid. $R_{\rm f}$ = 0.25 (petroleum ether). ¹H NMR (400 MHz, CDCl₃): δ = 3.81 (s, 3 H, OCH₃), 4.46 (s, 2 H, CH₂), 6.84 (dd, *J* = 8.2, 2.2 Hz, 1 H, 4-H), 6.93 (s, 1 H, 2-H), 6.97 (d, *J* = 7.6 Hz, 1 H, 6-H), 7.25 (t, *J* = 7.8 Hz, 1 H, 5-H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 33.5 (CH₂), 55.3 (CH₃), 114.2 (C-4), 114.4 (C-2), 121.3 (C-6), 129.8 (C-5), 139.1 (C-1), 159.7 (C-3) ppm.

1-(Azidomethyl)-3-methoxybenzene (20): A solution of sodium azide (0.5 M, 190.3 mg, 2.93 mmol, 1.1 equiv.) in DMSO (5.86 mL), which was prepared by stirring the sodium azide in DMSO at room temperature for 24 h, was added to 1-(bromomethyl)-3-methoxybenzene (19) (535 mg, 2.66 mmol). The mixture was stirred for 1 h at room temperature before it was quenched with water (10 mL). The mixture was extracted with Et_2O (3×10 mL). The combined organic layers were washed with water $(2 \times 10 \text{ mL})$ and a saturated NaCl solution $(1 \times 10 \text{ mL})$, dried with MgSO₄, filtered, and concentrated in vacuo. The pure azide 20 (372 mg, 2.28 mmol, 86%) was obtained as a colorless liquid. $R_{\rm f} = 0.16$ (petroleum ether). ¹H NMR (400 MHz, CDCl₃): δ = 3.82 (s, 3 H, CH₃), 4.30 (s, 2 H, CH₂), 6.84–6.92 (m, 3 H, 2-H, 4-H, 6-H), 7.29 (t, J = 7.8 Hz, 1 H, 5-H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 54.7 (CH₂), 55.2 (CH₃), 113.6 (aryl C), 113.8 (aryl C), 120.4 (C-6), 129.8 (C-5), 136.8 (C-1), 159.9 (C-3) ppm.

2-[2-(Prop-2-ynyloxy)ethoxy]-4-[3-(trifluoromethyl)-3H-diazirin-3yllbenzaldehyde (21): Potassium carbonate (710 mg, 5.14 mmol, 1.7 equiv.) was added to a cooled solution (0 °C) of 2-hydroxybenzaldehyde 14 (695 mg, 3.02 mmol) in dry DMF (60 mL) followed by stirring of the mixture for 30 min. The cooling bath was removed, then the tosylate 16 (870 mg, 3.42 mmol, 1.1 equiv.) dissolved in DMF (10 mL) was added, followed by Bu₄NI (120 mg, 0.3 mmol, 0.1 equiv.). The mixture was stirred at 55 °C for 16 h. It was then diluted with EtOAc (100 mL), washed with 1 N HCl ($1 \times 40 \text{ mL}$) and a saturated NaCl solution (1×40 mL), dried with MgSO₄, filtered, and concentrated in vacuo. The crude product was purified by flash chromatography (petroleum ether/ Et_2O , 6:1) to give the slightly yellow aldehyde 21 (706 mg, 2.2 mmol, 75%), m.p. 40 °C. $R_{\rm f} = 0.45$ (toluene/EtOAc, 25:1). ¹H NMR (400 MHz, CDCl₃): δ = 2.47 (t, J = 2.2 Hz, 1 H, C \equiv CH), 3.94–3.96 (m, 2 H, Ar-OCH₂CH₂), 4.24–4.27 (m, 4 H, ArOCH₂, CH₂C≡C), 6.73 (s, 1 H, 3-H), 6.83 (d, J = 8.3 Hz, 1 H, 5-H), 7.83 (d, J = 8.1 Hz, 1 H, 6-H), 10.47 (s, 1 H, CHO) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 28.5 (q, ${}^{2}J_{CF} = 41.0 \text{ Hz}$, CCF₃), 58.6 (CH₂C=C), 67.7 (Ar- OCH_2CH_2), 68.3 (ArOCH₂), 75.1 (C=CH), 79.0 (C=CH), 110.8 (C-3), 118.9 (C-5), 121.7 (q, ${}^{1}J_{CF} = 275.2 \text{ Hz}, \text{ CF}_{3}$), 125.6 (C-1), 128.9 (C-6), 136.6 (C-4), 160.9 (C-2), 188.8 (CHO) ppm. HRMS (ESI): calcd. for $C_{14}H_{11}F_3N_2O_3$ [M + Na]⁺ 335.0614; found 335.0612.

2-[2-(Prop-2-ynyloxy)ethoxy]-4-[3-(trifluoromethyl)-3*H*-diazirin-3yl]benzoic Acid (22): Sulfamic acid (400 mg, 4.13 mmol, 1.85 equiv.) was added to a solution of aldehyde 21 (695 mg, 2.23 mmol) in THF (20 mL) and water (10 mL). The mixture was stirred for 5 min at room temperature before sodium chlorite (365 mg, 4.04 mmol, 1.81 equiv.) dissolved in water (4 mL) was added. The reaction mixture was stirred for 3.5 h at room temperature and then extracted with CH_2Cl_2 (3 × 20 mL). The combined organic layers were dried



with MgSO₄, filtered, and concentrated in vacuo to afford the pure benzoic acid **22** (685 mg, 2.09 mmol, 94%) as a colorless solid, m.p. 70 °C. Purification by flash chromatography was accompanied by some loss of product. $R_{\rm f} = 0.16$ (CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 2.48$ (t, J = 2.3 Hz, 1 H, C=CH), 3.95–3.99 (m, 2 H, ArOCH₂CH₂), 4.26 (d, J = 2.3 Hz, 2 H, CH₂C=C), 4.37–4.41 (m, 2 H, ArOCH₂), 6.78 (s, 1 H, 3-H), 6.92 (d, J = 8.3 Hz, 1 H, 5-H), 8.16 (d, J = 8.3 Hz, 1 H, 6-H), 10.58 (br. s, 1 H, CO₂H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 28.3$ (q, ${}^{2}J_{\rm CF} = 41.0$ Hz, CCF₃), 58.5 (CH₂C=C), 66.7 (ArOCH₂CH₂), 69.4 (ArOCH₂), 75.6 (C=CH), 78.5 (C=CH), 111.3 (C-3), 119.6 (C-1), 119.9 (d, J = 1.5 Hz, C-5), 121.7 (q, ${}^{1}J_{\rm CF} = 275.2$ Hz), 134.3 (C-6), 135.9 (C-4), 157.4 (C-2), 164.5 (CO₂H) ppm. HRMS (ESI): calcd. for C₁₄H₁₁F₃N₂O₄ [M + Na]⁺ 351.0563; found 351.0560.

Isopropyl 2-[2-(Prop-2-ynyloxy)ethoxy]-4-[3-(trifluoromethyl)-3Hdiazirin-3-yl]benzoate (23): To a solution of benzoic acid 22 (100.0 mg, 0.3 mmol) in dry CH₂Cl₂ (4 mL) was added DMAP (73.3 mg, 0.6 mmol, 2 equiv.), followed by propan-2-ol (0.05 mL, 0.6 mmol, 2 equiv.), and EDC (0.06 mL, 51.2 mg, 0.33 mmol, 1.1 equiv.). The resulting solution was stirred at room temperature for 16 h and then diluted with CH_2Cl_2 (35 mL). The mixture was washed with water $(3 \times 5 \text{ mL})$, dried with MgSO₄, filtered, and concentrated in vacuo. The crude product was purified by flash chromatography (petroleum ether/ Et_2O , 5:1) to give the ester 23 (61.0 mg, 0.17 mmol, 55%) as a colorless viscous oil. $R_{\rm f} = 0.32$ (petroleum ether/Et₂O, 5:1). ¹H NMR (400 MHz, CDCl₃): δ = 1.34 $[d, J = 6.3 \text{ Hz}, 6 \text{ H}, CH(CH_3)_2], 2.44 (t, J = 2.3 \text{ Hz}, 1 \text{ H}, C \equiv CH),$ 3.89-3.94 (m, 2 H, ArOCH₂CH₂), 4.16-4.21 (m, 2 H, ArOCH₂), 4.27 (d, J = 2.3 Hz, 2 H, CH₂C=C), 5.16–5.28 [m, 1 H, CH(CH₃) 2], 6.70 (s, 1 H, 3-H), 6.80 (d, J = 8.1 Hz, 1 H, 5-H), 7.74 (d, J = 8.1 Hz, 1 H, 6-H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 21.9 $[CH(CH_3)_2]$, 28.4 (q, ${}^{2}J_{CF}$ = 41.0 Hz, CCF₃), 58.6 (CH₂C=C), 67.9 (ArOCH₂CH₂), 68.72 [CH(CH₃)₂], 68.74 (ArOCH₂), 74.8 $(C \equiv CH)$, 79.4 ($C \equiv CH$), 111.5 (C-3), 118.6 (C-5), 121.8 (q, ${}^{1}J_{CF} =$ 275.2 Hz, CF₃), 123.0 (C-1), 131.8 (C-6), 133.8 (C-4), 158.0 (C-2), 165.0 (CO_2iPr) ppm. HRMS (ESI): calcd. for $C_{17}H_{17}F_3N_2O_4$ [M + Na]⁺ 393.1033; found 393.1034.

Isopropyl 2-[2-(Prop-2-ynyloxy)ethoxy]-4-(2,2,2-trifluoro-1-methoxyethyl)benzoate (24): A stirred 1 mm solution of diazirine 23 (5.5 mg, 14.9 mmol) in MeOH (15 mL) was irradiated at 366 nm with a UV lamp (8 W, Waldmann, type 600352) at a distance of 1 cm for 1 h in a Duran Schlenk tube under nitrogen. Thereafter, the reaction mixture was concentrated at reduced pressure and the resulting crude product was purified by flash chromatography (petroleum ether/EtOAc, 8:1) to afford the insertion product 24 (2.1 mg, 5.61 mmol, 38%) as a colorless oil. $R_{\rm f} = 0.16$ (petroleum ether/EtOAc, 8:1). ¹H NMR (400 MHz, CDCl₃): δ = 1.35 [d, J = 6.3 Hz, 6 H, CH(CH₃)₂], 2.44 (t, J = 2.2 Hz, 1 H, C=CH), 3.41 (s, 3 H, OCH₃), 3.91–3.95 (m, 2 H, ArOCH₂CH₂), 4.21–4.25 (m, 2 H, ArOCH₂), 4.28 (d, J = 2.3 Hz, 2 H, CH₂C=C), 4.48 (q, J = 6.6 Hz, 1 H, CHCF₃), 5.19–5.30 [m, 1 H, CH(CH₃)₂], 7.02 (d, J = 8.1 Hz, 1 H, 5-H), 7.05 (s, 1 H, 3-H), 7.75 (d, J = 8.1 Hz, 1 H, 6-H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 21.9 [CH(CH₃)₂], 58.4 (OCH₃), 58.7 (CH₂C≡C), 68.0 (ArOCH₂CH₂), 68.5 [CH(CH₃)₂], 68.6 (Ar-OCH₂), 74.7 (C=*C*H), 79.5 (*C*=*C*H), 81.0 (${}^{2}J_{CF}$ = 31.5 Hz, *C*CF₃), 112.9 (C-3), 120.6 (C-5), 123.0 (C-1), 123.4 (${}^{1}J_{CF}$ = 281.8 Hz, CF₃), 131.4 (C-6), 137.4 (C-4), 158.1 (C-2), 165.6 (CO2iPr) ppm. HRMS (ESI): calcd. for $C_{18}H_{21}F_3O_5$ [M + Na]⁺ 397.1233; found 397.1236.

Isopropyl 2-(2-{[1-(3-Methoxybenzyl)-1*H***-1,2,3-triazol-4-yl]methoxy}ethoxy)-4-(2,2,2-trifluoro-1-methoxyethyl)benzoate (25):** a) From alkyne **24**: The alkyne **24** (6.9 mg, 18.4 μmol) and the azide **20** (3.6 mg, 22.1 μmol, 1.2 equiv.) were suspended in *t*BuOH (1 mL) and H₂O (0.75 mL) at room temperature under nitrogen. Then, a 0.01 M aqueous solution of sodium ascorbate (0.37 mg, 0.19 mL, 1.84 µmol) was added followed by a 0.005 M aqueous solution of copper(II) sulfate pentahydrate (0.05 mg, 0.04 mL, 0.18 µmol). After being stirred for 20 h, water (2 mL) was added and the aqueous phase extracted with CH₂Cl₂ (3×5 mL). The combined organic layers were washed with water, dried with MgSO₄, filtered, and concentrated in vacuo. The residue was purified by flash chromatography (petroleum ether/EtOAc, 1:1) to give the triazole **25** (5.7 mg, 10.6 µmol, 58%) as a colorless oil.

b) From diazirine **26**: A stirred 0.75 mM solution of diazirine **26** (2 mg, 3.7 μ mol) in MeOH (5 mL) was irradiated at 366 nm with a UV lamp (8 W, Waldmann, type 600352) at a distance of 1 cm for 1 h in a Duran Schlenk tube under nitrogen. The reaction mixture was concentrated at reduced pressure and the resulting crude product was purified by flash chromatography (petroleum ether/EtOAc, 1:1) to afford insertion product **25** (1.14 mg, 2.1 μ mol, 57%) as a colorless oil. This material was identical in all respects (TLC, LC-MS and ¹H NMR) to the triazole prepared by the other route.

 $R_{\rm f} = 0.25$ (petroleum ether/EtOAc, 1:1). ¹H NMR (400 MHz, CDCl₃): $\delta = 1.30$ [d, J = 6.1 Hz, 6 H, CH(CH₃)₂], 3.40 (s, 3 H, OCH₃), 3.76 (s, 3 H, ArOCH₃), 3.89–3.94 (m, 2 H, ArOCH₂CH₂), 4.17-4.23 (m, 2 H, ArOCH₂), 4.47 (q, J = 6.5 Hz, 1 H, CHCF₃), 4.75 (s, 2 H, OCH2-triazole), 5.12-5.23 [m, 1 H, CH(CH3)2], 5.48 (s, 2 H, NCH₂), 6.80 (s, 1 H, 2'-H), 6.83-6.88 (m, 2 H, 4'-H, 6'-H), 7.01 (d, J = 8.1 Hz, 1 H, 5-H), 7.03 (s, 1 H, 3-H), 7.26 (t, J = 8.0 Hz, 1 H, 5'-H), 7.64 (s, 1 H, 5''-H), 7.74 (d, J = 7.8 Hz, 1 H, 6-H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 21.9$ [CH(CH₃)₂], 54.1 (NCH₂), 55.3 (ArOCH₃), 58.4 (OCH₃), 64.9 (OCH₂-triazole), 68.4 [*C*H(CH₃)₂], 68.6 (OCH₂CH₂O), 81.0 (q, ${}^{2}J_{CF}$ = 31.0 Hz, CCF₃), 112.8 (C-3), 113.7 (C-2'), 114.1 (C-4'), 120.3 (C-6'), 120.5 (C-5), 122.7 (C-1, C-5''), 123.4 (q, ${}^{1}J_{CF} = 282.0 \text{ Hz}, \text{ CF}_{3}$), 130.1 (C-5'), 131.4 (C-6), 136.0 (C-1'), 137.5 (C-4), 145.5 (C-4''), 158.2 (C-2), 160.1 (C-3'), 165.4 (CO2iPr) ppm. HRMS (ESI): calcd. for $C_{26}H_{30}F_{3}N_{3}O_{6}$ [M + H]⁺ 538.21593; found 538.21562; Δ (rel.) = 0.58 ppm.

Isopropyl 2-(2-{[1-(3-Methoxybenzyl)-1H-1,2,3-triazol-4-yl]methoxy}ethoxy)-4-[3-(trifluoromethyl)-3H-diazirin-3-yl]benzoate (26): The alkyne 23 (24.0 mg, 64.8 µmol) and the azide 20 (12.7 mg, 77.8 µmol, 1.2 equiv.) were suspended in tBuOH (1 mL) and H₂O (0.2 mL) at room temperature under nitrogen. Then, a 0.01 м aqueous solution of sodium ascorbate (1.3 mg, 0.66 mL, 6.48 µmol) was added, followed by a 0.005 M aqueous solution of copper(II) sulfate pentahydrate (0.16 mg, 0.13 mL, 0.65 µmol). After being stirred for 20 h, water (3 mL) was added and the aqueous phase was extracted with CH_2Cl_2 (3 × 10 mL). The combined organic layers were washed with water, dried with MgSO4, filtered, and concentrated in vacuo. The crude product was purified by flash chromatography (petroleum ether/EtOAc, 1:1) to give the triazole 26 (12.4 mg, 23.2 μ mol, 36%) as a colorless oil. $R_{\rm f} = 0.34$ (petroleum ether/ EtOAc, 1:1). ¹H NMR (400 MHz, CDCl₃): δ = 1.29 [d, J = 6.1 Hz, 6 H, CH(CH₃)₂], 3.76 (s, 3 H, OCH₃), 3.88-3.93 (m, 2 H, Ar-OCH₂CH₂), 4.13-4.19 (m, 2 H, ArOCH₂), 4.74 (s, 2 H, OCH₂triazole), 5.11-5.21 [m, 1 H, CH(CH₃)₂], 5.48 (s, 2 H, NCH₂), 6.66 (s, 1 H, 3-H), 6.79 (d, J = 7.1 Hz, 2 H, 5-H, 2'-H), 6.83–6.89 (m, 2 H, 4'-H, 6'-H), 7.26 (t, J = 7.8 Hz, 1 H, 5'-H), 7.61 (s, 1 H, 5''-H), 7.73 (d, J = 8.1 Hz, 1 H, 6-H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 21.8 [CH(CH₃)₂], 28.3 (q, ²J_{CF} = 41.0 Hz, CCF₃), 54.1 (NCH₂), 55.3 (ArOCH₃), 64.9 (OCH₂-triazole), 68.5 (Ar-OCH₂CH₂), 68.6 [CH(CH₃)₂], 68.7 (ArOCH₂), 111.4 (C-3), 113.7 (C-2'), 114.1 (C-4'), 118.5 (C-5), 120.3 (C-6'), 121.8 (q, ${}^{1}J_{CF} =$ 275.2 Hz, CF₃), 122.7 (C-5''), 122.8 (C-1), 130.1 (C-5'), 131.8 (C-

6), 133.8 (C-4), 136.0 (C-1'), 145.4 (C-4''), 158.1 (C-2), 160.1 (C-3'), 164.9 (COO) ppm. HRMS (ESI): calcd. for $C_{25}H_{26}F_3N_5O_5$ [M + H]⁺ 534.19585; found 534.19543; Δ (rel.) = 0.79 ppm.

NMR Spectroscopic Data for Bafilomycin A₁ (27): ¹H NMR (400 MHz, [D₆]acetone): δ = 0.76 (d, J = 6.8 Hz, 3 H, Me-33), 0.86 (d, J = 6.8 Hz, 3 H, Me-30), 0.91 (m, 9 H, Me-25, Me-28, Me-32), 0.98 (d, J = 7.1 Hz, 3 H, Me-31), 1.03 (d, J = 7.1 Hz, 3 H, Me-27), 1.08-1.13 (m, 2 H, 20-H), 1.22-1.30 (m, 1 H, 22-H), 1.77 (q, J =7.0 Hz, 1 H, 18-H), 1.83–1.90 (m, 2 H, 8-H, 24-H), 1.92 (s, 3 H, Me-29), 1.97 (d, J = 0.8 Hz, 3 H, Me-26), 1.99–2.03 (m, 9-H), 2.11– 2.20 (m, 2 H, 16-H, 20-H), 2.49-2.58 (m, 1 H, 6-H), 3.22 (s, 3 H, 14-OMe), 3.27–3.31 (m, 1 H, 7-H), 3.44 (dd, *J* = 10.4, 2.0 Hz, 1 H, 23-H), 3.53 (ddd, J = 15.4, 10.3, 5.4 Hz, 1 H, 21-H), 3.62 (s, 3 H, 2-OMe), 4.05 (t, J = 8.8 Hz, 1 H, 14-H), 4.11 (d, J = 5.6 Hz, 1 H, 7-OH), 4.17 (ddd, J = 10.7, 4.2, 1.5 Hz, 1 H, 17-H), 4.71 (d, J =4.3 Hz, 1 H, 17-OH), 5.0 (dd, J = 8.3, 1.0 Hz, 1 H, 15-H), 5.13 (dd, J = 14.9, 9.1 Hz, 1 H, 13-H), 5.24 (d, J = 2.0 Hz, 1 H, 19-OH), 5.78 (d, J = 10.6 Hz, 1 H, 11-H), 5.95 (d, J = 8.8 Hz, 1 H, 5-H), 6.66 (dd, J = 10.9, 4.0 Hz, 1 H, 12-H), 6.70 (s, 1 H, 3-H) ppm. ¹³C NMR (100 MHz, [D₆]acetone): δ = 7.3 (Me-31), 10.3 (Me-30), 12.6 (Me-32), 14.1 (Me-26), 14.7 (Me-33), 17.7 (Me-27), 20.3 (Me-29), 21.7 (Me-25), 22.2 (Me-28), 28.7 (C-24), 37.9 (C-6), 38.2 (C-16), 41.6 (C-8), 41.9 (C-22), 42.2 (C-9), 43.0 (C-18), 44.5 (C-20), 55.6 (14-OMe), 60.1 (2-OMe), 70.4 (C-21), 71.5 (C-17), 76.9 (C-23), 77.4 (C-15), 80.3 (C-7), 83.3 (C-14), 99.7 (C-19), 125.2 (C-11), 126.9 (C-13), 132.7 (C-4), 134.1 (C-3), 134.4 (C-12), 141.9 (C-2), 144.8 (C-10), 145.7 (C-5), 167.7 (C-1) ppm.

21-O-{2-[2-(Prop-2-ynyloxy)ethoxy]-4-[3-(trifluoromethyl)-3H-diaziren-3-yl]benzoyl}bafilomycin A1 (28): Benzoic acid 22 (10.5 mg, 32.1 µmol, 2 equiv.) followed by DMAP (4.1 mg, 33.7 µmol, 2.1 equiv.), and EDC (6.0 µL, 33.7 µmol, 2.1 equiv.) were added to a stirred solution of bafilomycin A1 (10.0 mg, 16.1 µmol) in CH2Cl2 (2 mL). After stirring for 24 h at room temperature, additional acid 22 (5.3 mg, 16.1 µmol, 1 equiv.), DMAP (2.3 mg, 19.3 µmol, 1 equiv.), and EDC (3.4 µL, 19.3 µmol, 1.2 equiv.) were added and stirring was continued for a further 24 h. The solvent was evaporated in vacuo and the resulting crude product purified by flash chromatography (hexane/EtOAc, 3:2) to afford ester 28 (11.9 mg, 12.7 µmol, 79%) as a colorless solid as well as unreacted bafilomycin A₁ (2.0 mg, 3.2 μ mol). R_f = 0.56 (hexane/EtOAc, 3:2). ¹H NMR (600 MHz, [D₆]acetone): δ = 0.81 (d, J = 6.9 Hz, 3 H, Me-33), 0.87 (d, J = 6.9 Hz, 3 H, Me-30), 0.91 (d, J = 6.5 Hz, 6 H, Me-25, Me-32), 0.96 (d, J = 6.9 Hz, 3 H, Me-28), 1.01 (d, J = 7.1 Hz, 3 H, Me-31), 1.04 (d, J = 6.9 Hz, 3 H, Me-27), 1.35–1.40 (m, 1 H, 20-H), 1.68 (ddq, J = 16.9, 10.5, 6.5 Hz, 1 H, 22-H), 1.84–1.89 (m, 2 H, 8-H, 18-H), 1.92 (s, 3 H, Me-29), 1.93-1.96 (m, 1 H, 24-H), 1.97 (d, J = 1.2 Hz, 3 H, Me-26), 2.01 (d, J = 10.8 Hz, 1 H, 9-H), 2.16 (ddq, J = 17.6, 6.9, 0.6 Hz, 1 H, 16-H), 2.40 (dd, J = 11.8, 4.9 Hz, 1 H, 20-H), 2.51–2.57 (m, 1 H, 6-H), 2.94 (t, J = 2.4 Hz, 1 H, C=CH), 3.23 (s, 3 H, 14-OMe), 3.28-3.31 (m, 1 H, 7-H), 3.63 (s, 3 H, 2-OMe), 3.63-3.65 (m, 1 H, 23-H), 3.88-3.90 (m, 2 H, Ar- OCH_2CH_2), 4.05 (app. t, J = 8.9 Hz, 1 H, 14-H), 4.11 (d, J =5.5 Hz, 1 H, 7-OH), 4.19 (ddd, J = 10.7, 4.3, 1.6 Hz, 1 H, 17-H), 4.25 (d, J = 2.4 Hz, 2 H, $CH_2C \equiv CH$), 4.27–4.30 (m, 2 H, Ar-OCH₂), 4.78 (d, J = 3.7 Hz, 1 H, 17-OH), 4.97 (dd, J = 8.6, 1.2 Hz, 1 H, 15-H), 5.11–5.20 (m, 2 H, 13-H, 21-H), 5.45 (d, J = 1.8 Hz, 1 H, 19-OH), 5.79 (d, J = 10.8 Hz, 1 H, 11-H), 5.95 (d, J = 8.8 Hz, 1 H, 5-H), 6.66 (dd, J = 14.9, 10.8 Hz, 1 H, 12-H), 6.70 (d, J =0.6 Hz, 1 H, 3-H), 6.90 (d, J = 0.6 Hz, 1 H, 3-ArH), 7.01 (dd, J = 8.1, 0.8 Hz, 1 H, 5-ArH), 7.78 (d, J = 8.1 Hz, 1 H, 6-ArH) ppm. ¹³C NMR (100 MHz, [D₆]acetone): $\delta = 7.4$ (Me-31), 10.3 (Me-30), 12.8 (Me-32), 14.1 (Me-26), 14.6 (Me-33), 17.7 (Me-27), 20.3 (Me-29), 21.6 (Me-25), 22.2 (Me-28), 28.7 (C-24), 29.1 (q, ${}^{2}J_{CF}$ =

41.0 Hz, CCF₃), 37.9 (C-6), 38.3 (C-16), 39.0 (C-22), 40.6 (C-20), 41.6 (C-8), 42.2 (C-9), 42.9 (C-18), 55.6 (14-OMe), 58.8 (CH₂C=C), 60.1 (2-OMe), 68.7 (ArOCH₂CH₂), 69.6 (ArOCH₂), 71.5 (C-17), 75.6 (C-21), 76.1 (C=CH), 76.6 (C-23), 77.3 (C-15), 80.3 (C-7), 80.6 (C=CH), 83.3 (C-14), 99.8 (C-19), 112.3 (Ar-3), 119.4 (Ar-5), 122.9 (q, ${}^{1}J_{CF}$ = 273.7 Hz, CF₃), 124.7 (Ar-1), 125.1 (C-11), 126.9 (C-13), 132.4 (Ar-6), 132.7 (C-4), 133.7 (Ar-4), 134.2 (C-3), 134.4 (C-12), 141.9 (C-2), 144.8 (C-10), 145.7 (C-5), 158.7 (Ar-2), 166.0 (ArCO₂), 167.7 (C-1) ppm. HRMS (ESI): calcd. for C₄₉H₆₇F₃N₂O₁₂ [M + Na]⁺ 955.45383; found 955.45387; Δ(rel.) = 0.04 ppm.

2-Azidoethylamine (29): 2-Bromoethylamine hydrobromide (500 mg, 2.44 mmol) was added to a solution of sodium azide (475.9 mg, 7.32 mmol, 3 equiv.) in H₂O (2 mL). The stirred solution was heated to 75 °C for 21 h before it was cooled to 0 °C. Et₂O (2 mL) was added followed by solid KOH (800 mg). The organic phase was separated and the aqueous layer extracted with HgSO₄, filtered, and the solvent removed carefully by rotary evaporation (35 °C, 750 mbar) to afford the azide **29** (171 mg, 1.99 mmol, 82%) as a colorless liquid. $R_f = 0.39$ (EtOAc/MeOH, 3:1). ¹H NMR (400 MHz, CDCl₃): $\delta = 1.27$ (s, 2 H, NH₂), 2.80–2.84 (m, 2 H, CH₂N₃), 3.30 (t, J = 5.7 Hz, 2 H, CH_2 NH₂) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 41.2$ (CH₂NH₂), 54.6 (CH₂N₃) ppm.

1-{[5-(2-Oxohexahydro-1H-thieno[3,4-d]imidazol-4-yl)pentanoyl]oxy}pyrrolidine-2,5-dione (31): N-Hydroxysuccinimide (56.5 mg, 0.49 mmol, 1.2 equiv.) followed by EDC (86.9 µL, 0.49 mmol, 1.2 equiv.) was added to a solution of D-biotin (100.0 mg, 0.41 mmol) in DMF (10 mL). The solution was stirred for 21 h at room temperature. The solvent was evaporated in vacuo and the resulting residue dissolved in CH₂Cl₂ (200 mL). The organic phase was washed with NaHCO3 solution (2 \times 10 mL) and a saturated NaCl solution $(1 \times 10 \text{ mL})$, dried with MgSO₄, and filtered. The solvent was removed under vacuo to give biotin-NHS **31** (71.0 mg, 0.21 mmol, 51%) as a colorless solid. ¹H NMR (400 MHz, [D₆]DMSO): δ = 1.35–1.54 (m, 3 H, 4'-H, 5'-H), 1.58– 1.68 (m, 3 H, 3'-H, 5'-H), 2.57 (d, J = 11.4 Hz, 1 H, SCH₂), 2.66 $(t, J = 7.3 \text{ Hz}, 2 \text{ H}, 2'-\text{H}), 2.78-2.85 \text{ [m, 5 H, CH}_2\text{CH}_2 \text{ (succinyl)},$ SCH2], 3.06-3.12 (m, 1 H, SCH), 4.11-4.16 (m, 1 H, 3a-H), 4.27-4.32 (m, 1 H, 6a-H), 6.36 (s, 1 H, 1-NH), 6.42 (s, 1 H, 3-NH) ppm. ¹³C NMR (100 MHz, [D₆]DMSO): δ = 24.3 (C-3'), 25.4 [CH₂CH₂ (succinyl)], 27.6 (C-5'), 27.8 (C-4'), 30.0 (C-2'), 39.9 (SCH₂), 55.2 (SCH), 59.2 (C-6a), 61.0 (C-3a), 162.7 [(HN)₂CO], 168.9 (CO₂), 170.3 [N(CO)₂] ppm.

N-(2-Azidoethyl)-5-(2-oxohexahydro-1H-thieno[3,4-d]imidazol-4-yl)pentanamide (32): Et₃N (0.03 mL, 0.20 mmol, 1 equiv.) was added to a solution of 2-azidoethylamine (29) (24.1 mg, 0.28 mmol, 1.4 equiv.) in DMF (3 mL), followed by the addition of biotin-NHS 31 (70.0 mg, 0.20 mmol) in DMF (2 mL). The resulting solution was stirred at room temperature for 24 h. The solvent was evaporated in vacuo and the crude product purified by flash chromatography (acetone/MeOH, 10:1) to give the desired azido-biotin 32 (47.7 mg, 0.15 mmol, 75%) as a colorless solid. $R_f = 0.22$ (acetone/ MeOH, 10:1). ¹H NMR (400 MHz, $[D_6]DMSO$): $\delta = 1.20-1.38$ (m, 2 H, 4'-H), 1.39-1.55 (m, 3 H, 3'-H, 5'-H), 1.55-1.65 (m, 1 H, 5'-H), 2.06 (t, J = 7.3 Hz, 2 H, 2'-H), 2.56 (d, J = 12.9 Hz, 1 H, SCH₂), 2.80 (dd, J = 12.4, 5.1 Hz, 1 H, SCH₂), 3.05–3.11 (m, 1 H, SCH), 3.19-3.24 (m, 2 H, $CH_2CH_2N_3$), 3.31 (d, J = 7.6 Hz, 2 H, CH₂N₃), 4.08–4.14 (m, 1 H, 3a-H), 4.26–4.32 (m, 1 H, 6a-H), 6.35 (s, 1 H, 1-NH), 6.42 (s, 1 H, 3-NH), 8.03 (t, J = 5.3 Hz, 1 H, CONH) ppm. ¹³C NMR (100 MHz, [D₆]DMSO): $\delta = 25.2$ (C-3'), 28.0 (C-5'), 28.2 (C-4'), 35.1 (C-2'), 38.1 (CH₂CH₂N₃), 39.9



(SCH₂), 50.0 (CH₂N₃), 55.4 (SCH), 59.2 (C-6a), 61.0 (C-3a), 162.7 [(HN)₂CO], 172.4 (CONH) ppm. HRMS (ESI): calcd. for $C_{12}H_{20}N_6O_2S$ [M + H]⁺ 313.14412; found 313.14419; Δ (rel.) = 0.22 ppm.

21-O-{2-(2-{[1-(2-{[5-(2-Oxohexahydro-1H-thieno[3,4-d]imidazol-4yl)pentanoyl]amino}ethyl)-1H-1,2,3-triazol-4-yl]methoxy}ethoxy)-4-[3-(trifluoromethyl)-3*H*-diaziren-3-yl]benzoyl}bafilomycin A₁ (33): Azido-biotin 32 (2.0 mg, 6.4 µmol, 1.2 equiv.) was added to a stirred solution of bafilomycin A_1 derivative 28 (5.0 mg, 5.1 μ mol) in H₂O (1 mL) and tBuOH (1 mL) at room temperature. An aqueous sodium ascorbate solution (0.01 M, 55 µL, 0.5 µmol) was added followed by an aqueous solution of copper(II) sulfate pentahydrate (0.005 M, 10 µL, 0.05 µmol). The reaction mixture was stirred for 24 h at 35 °C before further sodium ascorbate solution (55 µL, 0.5 μ mol) and copper(II) sulfate pentahydrate solution (10 μ L, 0.05 µmol) were added. After another 24 h of stirring the solvent was removed under vacuum and the crude product purified by flash chromatography (CH₂Cl₂/MeOH, 10:1) to afford the ligation product 33 (2.0 mg, 1.6 µmol, 31%) as a colorless solid. Owing to the small amount a ¹³C spectrum could not be acquired. According to the ¹H NMR spectrum, the purified 33 contains a small amount of unreacted azide **32**. $R_{\rm f} = 0.18$ (CH₂Cl₂/MeOH, 10:1). ¹H NMR (600 MHz, $[D_6]$ acetone): $\delta = 0.81$ (d, J = 7.0 Hz, 3 H, Me-33), 0.87 (d, J = 6.6 Hz, 3 H, Me-30), 0.91 (m, 6 H, Me-25, Me-32), 0.96 (d, J = 6.7 Hz, 3 H, Me-28), 0.99 (d, J = 7.0 Hz, 3 H, Me-31), 1.04 (d, J = 7.0 Hz, 3 H, Me-27), 1.35–1.48 (m, 3 H, 20-H, 4'-H), 1.54– 1.72 (m, 4 H, 22-H, 3'-H, 5'-H), 1.72-1.80 (m, 1 H, 5'-H), 1.84-1.89 (m, 2 H, 8-H, 18-H), 1.92 (s, 3 H, Me-29), 1.93-1.96 (m, 1 H, 24-H), 1.98 (m, 3 H, Me-26), 2.00-2.02 (m, 1 H, 9-H), 2.13-2.16 (m, 1 H, 16-H), 2.19 (t, J = 7.2 Hz, 2 H, NHCOCH₂), 2.40 (dd, J = 11.8, 4.4 Hz, 1 H, 20-H), 2.51-2.57 (m, 1 H, 6-H), 2.67-2.71 (m, 2 H, SCH₂), 2.90-2.94 (m, 3 H), 3.17-3.22 (m, 1 H, SCH), 3.23 (s, 3 H, 14-OMe), 3.28-3.31 (m, 1 H, 7-H), 3.35-3.42 (m, 4 H, CH₂CH₂triazole), 3.63 (s, 3 H, 2-OMe), 3.65–3.68 (m, 1 H, 23-H), 3.88-3.91 (m, 2 H, ArOCH₂CH₂), 4.03-4.11 (m, 2 H, 7-OH, 14-H), 4.16-4.20 (m, 1 H, 17-H), 4.26-4.29 (m, 2 H, ArOCH₂), 4.30-4.34 (m, 1 H, 3a-H), 4.48–4.52 (m, 1 H, 6a-H), 4.68 (s, 2 H, OCH₂₋ triazole), 4.77 (d, J = 4.2 Hz, 1 H, 17-OH), 4.98 (m, 1 H, 15-H), 5.17 (m, 2 H, 13-H, 21-H), 5.50 (s, 1 H, 19-OH), 5.72 (s, 1 H, NH), 5.80 (d, J = 10.4 Hz, 1 H, 11-H), 5.92 (s, 1 H, NH), 5.94–5.97 (m, 1 H, 5-H), 6.66 (dd, J = 14.9, 11.0 Hz, 1 H, 12-H), 6.71 (s, 1 H, 3-H), 6.89 (s, 1 H, 3-ArH), 7.02 (d, J = 8.9 Hz, 1 H, 5-ArH), 7.29– 7.37 (m, 1 H, triazole), 7.81 (d, J = 8.1 Hz, 1 H, 6-ArH), 7.96 (s, 1 H, CONH) ppm. HRMS (ESI): calcd. for C₆₁H₈₇F₃N₈O₁₄S [M + Na]⁺ 1267.59068; found 1267.58951; Δ (rel.) = 0.90 ppm.

Supporting Information (see also the footnote on the first page of this article): Copies of the 1 H and 13 C NMR spectra of the described compounds.

Acknowledgments

Financial support by the Deutsche Forschungsgemeinschaft and the Fonds der Chemischen Industrie is gratefully acknowledged. We also thank Paul Schuler (Institute of Organic Chemistry) for measuring capillary NMR spectra of the bafilomycin derivatives.

- [1] G. Tochtrop, R. W. King, *Comb. Chem. High Throughput Screening* **2004**, *7*, 677–688.
- [2] K. H. Bleicher, H.-J. Böhm, K. Müller, A. I. Alanine, *Nature Rev. Drug Discovery* 2003, 2, 369–378.
- [3] a) C. Drahl, B. F. Cravatt, E. J. Sorensen, Angew. Chem. 2005, 117, 5936–5958; Angew. Chem. Int. Ed. 2005, 44, 5788–5809;

b) M. J. Evans, A. Saghatelian, E. J. Sorensen, B. F. Cravatt, *Nat. Biotechnol.* **2005**, *23*, 1303–1307; c) S. A. Sieber, S. Niessen, H. S. Hoover, B. F. Cravatt, *Nat. Chem. Biol.* **2006**, *2*, 274–281.

- [4] For some reviews, see: a) F. Kotzyba-Hibert, I. Kapfer, M. Goeldner, Angew. Chem. 1995, 107, 1391–1408; Angew. Chem. Int. Ed. Engl. 1995, 34, 1296–1312; b) S. A. Fleming, Tetrahedron 1995, 51, 12479–12520; c) Y. Hatanaka, Y. Sadakane, Curr. Top. Med. Chem. 2002, 2, 271–288; d) A. Blencowe, W. Hayes, Soft Matter 2005, 1, 178–205; e) Y. Sadakane, Y. Hatanaka, Anal. Sci. 2006, 22, 209–218; f) E. L. Vodovozova, Biochemistry (Moscow) 2007, 72, 1–20.
- [5] J. Brunner, H. Senn, F. M. Richards, J. Biol. Chem. 1980, 255, 3313–3318.
- [6] M. Hashimoto, Y. Hatanaka, Anal. Biochem. 2006, 348, 154– 156.
- [7] See, for example: G. Ingenhorst, K. U. Bindseil, C. Boddien, S. Dröse, M. Gaßel, K. Altendorf, A. Zeeck, *Eur. J. Org. Chem.* 2001, 4525–4532.
- [8] Y. Ambroise, F. Pillon, C. Mioskowski, A. Valleix, B. Rousseau, Eur. J. Org. Chem. 2001, 3961–3964.
- [9] T. Weber, J. Brunner, J. Am. Chem. Soc. 1995, 117, 3084–3095.
- [10] Y. Hatanaka, M. Hashimoto, Y. Kanaoka, *Bioorg. Med. Chem.* 1994, 2, 1367–1373.
- [11] M. Daghish, L. Hennig, M. Findeisen, S. Giesa, F. Schumer, H. Hennig, A. G. Beck-Sickinger, P. Welzel, *Angew. Chem.* **2002**, *114*, 2404–2408; *Angew. Chem. Int. Ed.* **2002**, *41*, 2293– 2297.
- [12] J.-j. Park, Y. Sadakane, K. Masuda, T. Tomohiro, T. Nakano, Y. Hatanaka, *ChemBioChem* 2005, 6, 814–818.
- [13] L. Ballell, M. van Scherpenzeel, K. Buchalova, R. M. J. Liskamp, R. J. Pieters, Org. Biomol. Chem. 2006, 4, 4387–4394.
- [14] For the uncatalyzed reaction, see: a) R. Huisgen, Angew. Chem.
 1963, 75, 604–637; Angew. Chem. Int. Ed. Engl. 1963, 2, 565–598; b) R. Huisgen, L. Moebius, G. Müller, H. Stangl, G. Szeimies, J. M. Vernon, Chem. Ber. 1965, 98, 3992–4013.
- [15] For the copper(I)-catalyzed reaction, see: a) C. W. Tornøe, C. Christensen, M. Meldal, J. Org. Chem. 2002, 67, 3057–3064;
 b) V. V. Rostovtsev, L. G. Green, V. V. Fokin, K. B. Sharpless, Angew. Chem. 2002, 114, 2708–2711; Angew. Chem. Int. Ed. 2002, 41, 2596–2599.
- [16] For some reviews, see: a) H. C. Kolb, M. G. Finn, K. B. Sharpless, Angew. Chem. 2001, 113, 2056–2075; Angew. Chem. Int. Ed. 2001, 40, 2004–2021; b) V. D. Bock, H. Hiemstra, J. H. van Maarseveen, Eur. J. Org. Chem. 2006, 51–68; c) J.-F. Lutz, Angew. Chem. 2007, 119, 1036–1043; Angew. Chem. Int. Ed. 2007, 46, 1018–1025.
- [17] a) E. Saxon, C. R. Bertozzi, *Science* 2000, 287, 2007–2010; b)
 K. L. Kiick, E. Saxon, D. A. Tirrell, C. R. Bertozzi, *Proc. Natl. Acad. Sci. U.S.A.* 2002, 99, 19–24; c) N. J. Agard, J. M. Baskin, J. A. Prescher, A. Lo, C. R. Bertozzi, *ACS Chem. Biol.* 2006, 1, 644–648.
- [18] M. Hashimoto, Y. Hatanaka, Chem. Pharm. Bull. 2005, 53, 1510–1512.
- [19] T. Hosoya, T. Hiramatsu, T. Ikemoto, M. Nakanishi, H. Aoyama, A. Hosoya, T. Iwata, K. Maruyama, M. Endo, M. Suzuki, Org. Biomol. Chem. 2004, 2, 637–641.
- [20] T. Hosoya, T. Hiramatsu, T. Ikemoto, H. Aoyama, T. Ohmae, M. Endo, M. Suzuki, *Bioorg. Med. Chem. Lett.* 2005, 15, 1289– 1294.
- [21] a) M. Nassal, *Liebigs Ann. Chem.* 1983, 1510–1523; b) J. E. Baldwin, A. J. Pratt, M. G. Moloney, *Tetrahedron* 1987, 43, 2565–2575; c) Y. Hatanaka, H. Nakayama, Y. Kanaoka, *Heterocycles* 1993, 35, 997–1004.
- [22] a) M. Hashimoto, Y. Kanaoka, Y. Hatanaka, *Heterocycles* 1997, 46, 119–122; b) E. L. Bentz, H. Gibson, C. Hudson, M. G. Moloney, D. A. Seldon, E. S. Wearmouth, *Synlett* 2006, 247–250; c) H. Nakashima, M. Hashimoto, Y. Sadakane, T. Tomohiro, Y. Hatanaka, *J. Am. Chem. Soc.* 2006, 128, 15092–15093.

- [23] E. Hernandez, A. Galan, C. Rovira, J. Veciana, Synthesis 1992, 1164-1169
- [24] T. Shigenari, T. Hakogi, S. Katsumura, Chem. Lett. 2004, 33, 594-595.
- [25] F. M. Hauser, S. R. Ellenberger, Synthesis 1987, 723-724.
- [26] G. Werner, H. Hagenmaier, H. Drautz, A. Baumgartner, H. Zähner, J. Antibiot. 1984, 37, 110-117.
- [27] For recent reviews, see: a) T. Nishi, M. Forgac, Nat. Rev. Mol. Cell Biol. 2002, 3, 94-103; b) C. Farina, S. Gagliardi, Curr. Pharm. Des. 2002, 8, 2033-2048; c) W. Junge, N. Nelson, Science 2005, 308, 642-644; d) K. W. Beyenbach, H. Wieczorek, J. Exp. Biol. 2006, 209, 577-589.
- [28] M. Huss, G. Ingenhorst, S. Koenig, M. Gassel, S. Dröse, A. Zeeck, K. Altendorf, H. Wieczorek, J. Biol. Chem. 2002, 277, 40544-40548.
- [29] S. Gagliardi, P. A. Gatti, P. Belfiore, A. Zocchetti, G. D. Clarke,
- C. Farina, J. Med. Chem. 1998, 41, 1883–1893. [30] a) P.-C. Lin, S.-H. Ueng, M.-C. Tseng, J.-L. Ko, K.-T. Huang, S.-C. Yu, A. K. Adak, Y.-J. Chen, C.-C. Lin, Angew. Chem.

2006, 118, 4392-4396; Angew. Chem. Int. Ed. 2006, 45, 4286-4290; b) J. L. Meier, A. C. Mercer, H. Rivera Jr, M. D. Burkart, J. Am. Chem. Soc. 2006, 128, 12174-12184; c) X.-L. Sun, C. L. Stabler, C. S. Cazalis, E. L. Chaikof, Bioconjugate Chem. 2006, 17, 52-57.

- [31] a) B. Carboni, A. Benalil, M. Vaultier, J. Org. Chem. 1993, 58, 3736-3741; b) W. G. Lewis, F. G. Magallon, V. V. Fokin, M. G. Finn, J. Am. Chem. Soc. 2004, 126, 9152-9153; c) F. E. Hahn, V. Langenhahn, T. Pape, Chem. Commun. 2005, 5390-5392; d) S. C. Ritter, B. König, Chem. Commun. 2006, 4694-4696.
- [32] a) E. W. S. Chan, S. Chattopadhaya, R. C. Panicker, X. Huang, S. Q. Yao, J. Am. Chem. Soc. 2004, 126, 14435-14446; b) L. An, Y. Tang, S. Wang, Y. Li, D. Zhu, Macromol. Rapid Commun. 2006, 27, 993-997; c) J. Wang, M. Uttamchandani, L. P. Sun, S. Q. Yao, Chem. Commun. 2006, 717-719.
- [33] Y. Hatanaka, M. Hashimoto, H. Kurihara, H. Nakayama, Y. Kanaoka, J. Org. Chem. 1994, 59, 383-387.

Received: March 2, 2007 Published Online: July 23, 2007