Natural Product Synthesis

Total Synthesis and Biological Evaluation of Monorhizopodin and 16-*epi*-Monorhizopodin**

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For many years, polyketide natural products have provided the scientific community with a rich source of novel molecular architectures, many of which have become important therapeutics for clinical use.^[1] In 1993, the polyketide rhizopodin was isolated from the myxobacterium *Myxococccus stipitatus*.^[2] It was shown to display an interesting array of biological properties, including potent antitumor activity against a range of cancer cell lines in the low nanomolar range and the ability to inhibit the polymerization of actin.^[2,3] Despite its original structural assignment as the 19-membered monomeric lactone **1a** (monorhizopodin), recent studies revealed dimeric structure **2** to be the correct architecture of rhizopodin (Scheme 1).^[4] These molecules have started to attract attention from the synthetic community, although no total syntheses have been reported to date.^[5]

We were intrigued as to whether the originally proposed structure for monorhizopodin (1a) might exhibit comparable biological properties to its parent dimer (2), as these molecules contain several common structural elements, including an enamide side chain that is crucial for biological activity. X-ray crystallographic analysis of a rhizopodin–actin complex indicated that the binding of 2 to actin was largely a result of favorable van der Waals interactions between the enamide side chain and certain hydrophobic residues in the binding cleft.^[6] Thus, the construction of 1a might shed light on the effect of the macrocycle itself upon the binding affinity of these compounds. Herein, we describe a highly convergent and enantioselective total synthesis of monorhizopodin (1a)

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Scheme 1. Structures of monorhizopodin (1 a), 16-*epi*-monorhizopodin (1 b), and rhizopodin (2).

and its C16 epimer (1b), as well as preliminary biological evaluation of these compounds.

Scheme 2 depicts, in retrosynthetic format, the outline of the synthetic strategy employed for the construction of monorhizopodin (1a) and 16-epi-monorhizopodin (1b). Thus, rupture of the macrolactone and the C22-C23 and C29-N bonds led, after functional group adjustments, to hydroxy carboxylic acid 3 and ketophosphonate 4 as potential precursors to the target molecules. Disassembly of 3 at the indicated bond (C15-C16) through a SmI₂-mediated Barbier reaction revealed iodoester 5 and aldehyde 6 as the required building blocks for its construction. Finally, disconnection of the diene system of 5 through a Stille coupling traced the origins of this compound to vinyl iodide 7 and oxazolecontaining vinyl stannane 8. The choice for the SmI₂mediated Barbier coupling over the more conventional Grignard reaction was based on the remarkably mild nature of the former reagent.^[7] This characteristic was thought to be essential for the survival of the resulting secondary alcohol,

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Scheme 2. Retrosynthetic analysis of monorhizopodin (1). HWE = Horner–Wadsworth–Emmons, TBDPS = *tert*-butyldiphenylsilyl, TBS = *tert*-butyldimethylsilyl, TES = triethylsilyl.

whose facile elimination toward the oxazole moiety was considered to be potentially problematic.

The synthesis of monorhizopodin (1a) and its 16-*epi*diastereoisomer (1b) commenced with the construction of the requisite building blocks in their enantiomerically pure forms, starting with ketophosphonate 4. Thus, as shown in Scheme 3, enantioselective crotylation of aldehyde $9^{[8]}$ with (–)-(Ipc)₂*trans*-crotyl borane by employing Brown's protocol afforded alcohol 10 in 91 % yield as a > 10:1 mixture of diastereomers and >95 % *ee*. Methylation (NaH, MeI, 76 % yield) and cleavage of the terminal olefin by ozonolysis (O₃; Ph₃P, 84 % yield) of the latter compound led to aldehyde 12 through intermediate 11. Reaction of aldehyde 12 with the lithium species derived from dimethyl methylphosphonate and *n*BuLi, and subsequent DMP oxidation of the resulting mixture of diastereomeric alcohols, furnished ketophosphonate 4 in 98 % overall yield.



Scheme 3. Construction of ketophosphonate 4. Reagents and conditions: a) (-)- $(lpc)_2$ -trans-crotyl borane (1.2 equiv), BF₃·OEt₂ (1.2 equiv), THF, -78 °C, 1 h; 3 N NaOH (aq), H_2O_2 , Et₂O, 25 °C, 10 h, 91%, d.r. > 10:1; b) NaH (1.5 equiv), MeI (1.5 equiv), THF, 0 to 25 °C, 1 h, 76%; c) O_3 , CH₂Cl₂, -78 °C; then PPh₃ (1.5 equiv), 25 °C, 1 h, 84%; d) MeP(O) (OMe)₂ (5.5 equiv), nBuLi (5.5 equiv), THF, -78 °C, 1 h; 12 (1.0 equiv), -78 °C, 2.5 h; DMP (1.4 equiv), CH₂Cl₂, 25 °C, 10 min, 98% for two steps. DMP=Dess-Martin periodinane, lpc=isopino-campheyl, THF = tetrahydrofuran.

Aldehyde 6 was synthesized from known diol $13^{[9]}$ as summarized in Scheme 4. Thus, acetonide formation from 13 $(Me_2C(OMe)_2, (\pm)$ -CSA (cat.), 96% yield) and subsequent ozonolysis of the resulting olefin (14, O₃; Ph₃P, 83 % yield) led to aldehyde 15, which was subjected to reagent-controlled crotylation under Brown's conditions $((+)-(Ipc)_2-cis-crotyl)$ borane) to afford secondary alcohol 16 in 75% yield as a single diastereomer. Methylation of 16 (NaH, MeI) and subsequent ozonolysis (O3; Ph3P) and reduction with NaBH4 gave primary alcohol 18, whose silvlation (TBDPSCl, imidazole, DMAP (cat.)) led to acetonide TBDPS ether 19, via intermediates 17 and 18, in 75% overall yield for the four steps. Liberation of the 1,3-diol system of 19 required mild acid conditions (PPTS) and recycling $(3 \times)$ to avoid significant degradation of the TBDPS ether and resulted in 56% vield of diol 20 (plus 33% recovered starting material). Selective oxidation of the primary hydroxy group of 20 (PhI(OAc)₂, TEMPO (cat.)), and subsequent TES protection (TESOTf, 2,6-lutidine), furnished, via intermediate 21, the required aldehyde 6 (75% overall yield for the 2 steps).

Scheme 5 summarizes the construction of vinyl iodide 7. By following a procedure developed by Denmark et al., α , β unsaturated aldehyde 22^[10] was treated with silvl ketene acetal $23^{[11]}$ in the presence of catalyst 24 and SiCl₄^[12] to afford secondary alcohol 25 in 69% yield and > 90% ee. Exposure of 25 to KHMDS and benzaldehyde afforded benzylidene acetal **26** (71% yield), whose configuration was confirmed by nOe interactions. Subsequent treatment of 26 with (\pm) -CSA in MeOH revealed dihydroxy methyl ester 27 through cleavage of the acetal group and transesterification (54% yield plus 44% recovered starting material). Selective protection of the hydroxy group proximal to the ester moiety was achieved through δ -lactone formation within 27 (K₂CO₃; *p*TsOH, 96%) overall yield) and silvlation (TBSCl, DMAP, 84% yield), thus leading to TBS ether 29 via hydroxy compound 28. Subsequent opening of the δ -lactone moiety of **29** (NaOMe, 86%) yield), and subsequent methylation (Me₃OBF₄, 2,6-di-tert-



Scheme 4. Construction of aldehyde **6.** Reagents and conditions: a) (±)-CSA (0.010 equiv), Me₂C(OMe)₂, DMF, 25 °C, 2 h, 96%; b) O₃, CH₂Cl₂, -78 °C; then PPh₃ (2.0 equiv), 25 °C, 1 h, 83%; c) (+)-(lpc)₂*cis*-crotyl borane (1.2 equiv), BF₃·OEt₂ (1.2 equiv), THF, -78 °C; 1 h; 3 N NaOH (aq), H₂O₂, Et₂O, 25 °C, 10 h, 75%; d) NaH (1.3 equiv), Mel (2.0 equiv), THF, 0 to 25 °C, 1 h, 89%; e) O₃, CH₂Cl₂, -78 °C; then PPh₃ (2.0 equiv), 25 °C, 1 h; NaBH₄ (1.1 equiv), MeOH, 0 °C, 30 min, 86% for two steps; f) TBDPSCl (1.1 equiv), imidazole (1.3 equiv), DMAP (0.10 equiv), CH₂Cl₂, 0 to 25 °C, 1 h, 98%; g) PPTS (1.0 equiv), CH₂Cl₂, MeOH, 25 °C, 16 h, 56% (33% recovered **19**); h) TEMPO (0.30 equiv), PhI(OAc)₂ (3.0 equiv), CH₂Cl₂, 25 °C, 12 h, 85%; i) TESOTf (1.2 equiv), 2.6-lutidine (2.4 equiv), CH₂Cl₂, -78 °C, 30 min, 88%. DMAP = 4-dimethylaminopyridine, DMF = *N*,*N*-dimethylformamide, PPTS = pyridinium *para*-toluenesulfonate, TEMPO = 2,2,6,6-tetramethyl-1-piperidinyloxy.

21: R = H

6: R = TES

i) TESOT

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butyl-4-methyl pyridine, 80% yield) and iodination (NIS, 96% yield), furnished vinyl iodide **7** via intermediates **30** and **31**.

Vinyl stannane 8 was constructed from known oxazole aldehyde $32^{[13]}$ and converted into advanced iodide 5 as summarized in Scheme 6. Thus, treatment of 32 with allenyltri-*n*-butylstannane in the presence of $Ti(OiPr)_4$ and (S)-BINOL afforded alcohol 33 in 60% yield (plus 24% recovered starting material) and >95% ee.[14] Methylation of 33 (NaH, MeI, 93% yield) and subsequent desilylation (aqueous HF, 86% yield) led to primary alcohol 35 via intermediate 34. Regio- and stereoselective addition of tri-nbutyltin hydride to the terminal acetylene unit of 35 was achieved through palladium catalysis $([Pd_2(dba)_3],$ Cy₃P·HBF₄, *i*Pr₂NEt, (*n*Bu)₃SnH, 67% yield) and afforded vinyl stannane 8. Coupling of this stannane with vinyl iodide 7 in the presence of CuTC^[15] led to alcohol 36 (75% yield), which was converted into the desired advanced iodide 5 by sodium iodide through its mesylate derivative (MsCl, Et₃N; NaI, 96% overall yield).

With fragments **4–6** now available, the stage was set for their coupling and elaboration to the targeted molecules



Scheme 5. Construction of vinyl iodide 7. Reagents and conditions: a) **23** (1.2 equiv), cat. (*R*,*R*)-**24** (0.015 equiv), SiCl₄ (1.1 equiv), *i*Pr₂NEt (0.20 equiv), CH₂Cl₂, -78 °C, 2 h, 69%; b) PhCHO (3.3 equiv), KHMDS (0.30 equiv), THF, 0 °C, 1 h, 71%; c) (±)-CSA (0.30 equiv), MeOH, 25 °C, 20 h, 54% (44% recovered **26**); d) K₂CO₃ (2.0 equiv), MeOH, H₂O, 25 °C, 15 min; *p*TsOH (1.0 equiv), THF, 25 °C, 1 h, 96% for two steps; e) TBSCI (2.4 equiv), DMAP (3.9 equiv), CH₂Cl₂, 25 °C, 2 h, 84%; f) NaOMe (2.0 equiv), MeOH, 0 °C, 1 h, 86%; g) Me₃OBF₄ (4.0 equiv), 2,6-di-*tert*-butyl-4-methyl pyridine (5.0 equiv), CH₂Cl₂, 25 °C, 12 h, 80%; h) NIS (3.0 equiv), MeCN, 60 °C, 10 h, 96%. CSA= camphorsulfonic acid, KHMDS = potassium hexamethyldisilazide, NIS = *N*iodosuccinimide, *p*TsOH = *para*-toluenesulfonic acid.

monorhizopodin (1a) and its 16-epi-diastereoisomer (1b). As shown in Scheme 7, a mixture of 5 (1.0 equiv) and 6 (1.5 equiv) was exposed to the action of SmI₂ in THF and afforded a 1:1 diastereomeric mixture of alcohols 37 at C16 (56% yield). Given the complexity of these substrates, the performance of SmI₂ in this coupling reaction is remarkable and provides further testament for the power of this reagent in organic synthesis.^[7] Having achieved coupling of the key fragments, the resulting mixture of alcohols 37 was oxidized through the action of DMP and afforded ketone 38 in 95% yield. Saponification of the methyl ester of **38** (LiOH, 60 °C) and subsequent removal of the TES group (PPTS) then led to hydroxy acid 3 (62% overall yield), which was now primed for macrolactonization. After screening several reaction conditions we found that optimal results could be obtained by employing the protocol developed by Shiina et al.^[16] Thus, slow addition of hydroxy acid 3 to solution of MNBA in toluene and DMAP at 60 °C afforded keto macrolactone 39 in good yield. Owing to its tailing TLC properties, this ketone was difficult to purify and, therefore, was reduced with NaBH₄ in the presence of CeCl₃ in MeOH at -20 °C to give hydroxy compounds 40 a and 40 b as a chromatographically separable mixture of diastereomers (ca. 2:1 in favor of 40 a; under the high dilution conditions employed in the macrolactonization process, no dimeric material was observed).

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Scheme 6. Synthesis of vinyl stannane 8 and advanced iodide 5. Reagents and conditions: a) $Ti(OiPr)_4$ (1.0 equiv), (S)-BINOL (1.0 equiv), M.S. (4 Å), CH_2CI_2 , 40 °C, 1 h; 32 (1.0 equiv), allenyl-tri-*n*butylstannane (1.2 equiv), -24 °C, 72 h, 60% (24% recovered 32); b) NaH (2.0 equiv), Mel (2.0 equiv), THF, 0 to 25 °C, 1 h, 93%; c) 48% aq HF (2.0 equiv), MeCN, 25 °C, 3 h, 86%; d) [Pd₂(dba)₃] (0.0050 equiv), Cy₃P·HBF₄ (0.020 equiv), *i*Pr₂NEt (0.040 equiv), (*n*Bu)₃SnH (1.2 equiv), CH₂CI₂, 25 °C, 10 min, 67%; e) 7 (1.0 equiv), 8 (1.0 equiv), CuTC (10.0 equiv), NMP, 0 °C, 30 min, 75%; f) MsCl (1.2 equiv), Et₃N (1.5 equiv), CH₂CI₂, -78 °C, 30 min; NaI (5.0 equiv), acetone, 25 °C, 1 h, 96% for two steps. BINOL = 1,1'-bi(2-naphthol), CuTC = copper(I) thiophene-2-carboxylate, Cy = cyclohexyl, dba = *tran s*,*trans*-dibenzylideneacetone, Ms = methanesulfonyl, M.S. = molecular seives, NMP = N-methyl-2-pyrrolidinone.

Interestingly, reduction of ketone **39** with NaBH₄ alone delivered alcohol **40b** exclusively. Other reducing agents led to unsatisfactory results. The configurations of diastereomers **40a** and **40b** were determined by the ${}^{13}CNMR$ method

Scheme 7. Fragment coupling and completion of the total synthesis of monorhizopodin (1a). Reagents and conditions: a) 5 (1.0 equiv), 6 (1.5 equiv), Sml₂ (3.0 equiv), THF, 25 °C, 5 min, 56%; b) DMP (3.0 equiv), CH₂Cl₂, 25 °C, 15 min, 95%; c) LiOH·H₂O (6.0 equiv), (CH₃)₂CHOH/THF/H₂O (4:1:1), 60 °C, 1 h; PPTS (1.0 equiv), CH₂Cl₂, MeOH, 25 °C, 12 h, 62% for two steps; d) 3 (1.0 equiv in THF/toluene (1:1), slow addition by syringe pump), MNBA (2.0 equiv), DMAP (6.0 equiv), M.S. (4 Å), toluene, 60 °C, 20 h; NaBH₄ (3.0 equiv), CeCl₃·7 H₂O (10.0 equiv), MeOH, -20 °C, 30 min, 70% for two steps, d.r. = 2:1; e) TBAF (10.0 equiv), AcOH (10.0 equiv), DMF, 25 °C, 12 h, 89%; f) TESOTf (10.0 equiv), 2,6-lutidine (20 equiv), CH₂Cl₂, -78°C, 30 min, 66%; g) PPTS (0.10 equiv), CH₂Cl₂, MeOH, 25 °C, 30 min, 89%; h) SO₃·py (2.0 equiv), *i*Pr₂NEt (6.0 equiv), CH₂Cl₂, DMSO, 25 °C, 30 min, 81 %; i) 4 (2.0 equiv), Ba(OH)₂ (0.5 equiv), THF, H₂O, 25 °C, 2 h, 65 %; j) [{CuH(PPh₃)}₆] (1.0 equiv), benzene, 25 °C, 8 h, 87%; k) TASF (5.0 equiv), DMF, 25 °C, 8 h, 70 %; l) TEMPO (0.10 equiv), PhI(OAc)₂ (2.0 equiv), CH₂Cl₂, 25 °C, 4 h, 74%; m) HN(Me)CHO (20 equiv), PPTS (0.14 equiv), M.S. (4 Å), C₆H₆, 80°C, 8 h, 78%. DMSO = dimethylsulfoxide, MNBA = 2-methyl-6-nitrobenzoic anhydride, py = pyridine, TASF = tris(dimethylamino)sulfonium difluorotrimethylsilicate, TBAF = tetra-*n*-butylammonium fluoride.



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developed by Rychnovsky et al.^[17] Thus, the acetonides obtained from 40a and 40b upon reductive macrolactone opening, selective protection of the primary alcohol with a TBDPS group, and acetonide formation were employed (40 a $(^{13}CNMR (CDCl_3))$: gave anti-acetonide **40** a' $\delta =$ 101.61 ppm), while **40b** gave syn-acetonide **40b**' (¹³C NMR (CDCl₃): $\delta = 99.04$ ppm); see the Supporting Information). In preparation for the side chain attachment, the silvl protecting groups of diastereomer 40a (corresponding to the C16 rhizopodin configuration) were removed (TBAF, AcOH) and afforded triol 41 a in 89% yield. The latter compound was then converted into its tris-TES derivative 42 a (TESOTf, 2,6lutidine, 66 % yield), which underwent selective monodesilylation (PPTS, 89% yield) and oxidation (SO₃·py, 81% yield) and afforded aldehyde 44a through primary alcohol 43a. This aldehyde was then treated with ketophosphonate 4 in the presence of Ba(OH)₂ and afforded the corresponding α,β unsaturated ketone (65% yield), which was reduced with Stryker's reagent and led to its saturated counterpart (45 a) in 87% yield. At this juncture, all that remained to complete the synthesis of monorhizopodin (1a) was global deprotection and installation of the enamide moiety. To this end, trissilvlated ketone 45a was first exposed to TASF (to produce triol 46a, 70% yield) and then to PhI(OAc)₂/TEMPO (cat.) to furnish, through selective oxidation of the primary alcohol, dihydroxy aldehyde 47a (74% yield). The latter was condensed with N-methyl formamide (PPTS, M.S. (4 Å), 80°C) and afforded monorhizopodin (1a) in 78% yield as a mixture of E/Z geometrical isomers (ca. 2:1 in favor of the E isomer). By following the same sequence, 16-epi-monorhizopodin (1b) was also synthesized from diastereoisomer 40b (see the Supporting Information). The ¹H and ¹³C NMR and IR spectroscopic data of 1a and 1b were similar to those reported for rhizopodin (2).^[4] The monomeric structures of these compounds were confirmed by mass spectrometry.

With synthetic samples of monorhizopodin (1a) and 16epi-monorhizopodin (1b) available to us, we were in a position to evaluate their biological properties in actin polymerization and cytotoxicity assays. As shown in Figure 1, monorhizopodin (1a) exhibited potent inhibitory activity of actin polymerization, as expected from its enamide side chain structural motif. This activity, which is mimicked by monorhizopodin's 16-epi-isomer (1b), albeit with somewhat lower potency, is comparable to that of latrunculin A (see Figure 1), which was used as a standard in this assay. However, neither monorhizopodin (1a) nor 16-epi-monorhizopodin (1b) exhibited cytotoxicity against MDA-MB-231 breast cancer cells (up to 100 µM concentrations), thus presenting an interesting dichotomy and a puzzle regarding their divergence from rhizopodin (2). Although further investigations are needed to explain this phenomenon, we hypothesize that either these compounds are unable to displace G-actin binding proteins, such as profilin,^[18] within cells, or that they fail to penetrate the cell membrane to reach their target.

In conclusion, a highly convergent total synthesis of monorhizopodin (1a) and 16-*epi*-monorhizopodin (1b) has been developed, rendering these monomeric homologues of the powerful antitumor agent rhizopodin (2) available for



Figure 1. Inhibition of actin polymerization by monorhizopodin (1 a). The concentration of actin was 5 μ M, that of monorhizopodin (1 a) was as indicated. For the corresponding graphs obtained with 16-*epi*-monorhizopodin (1b) and further details of the assay, see the Supporting Information. LatA = latrunculin A.

biological investigations. Preliminary studies showed these compounds to be endowed with actin-binding properties but devoid of any associated cytotoxicity, thus posing interesting questions regarding the role of the dimeric nature of rhizopodin (2) in its mode of action. Further studies directed toward the elucidation of the mechanism of action and the differences of rhizopodin (2) and its monomeric homologues, (1a) and (1b), as well as the total synthesis of the former are in progress.

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