DOI: 10.1002/ejoc.200600926

New Scalable Asymmetric Aminomethylation Reaction for the Synthesis of β²-Amino Acids

Roba Moumné,^[a,b] Bernard Denise,^{[c][†]} Karine Guitot,^[a] Henri Rudler,^[c] Solange Lavielle,^[a] and Philippe Karoyan*^[a]

Keywords: β²-Amino acids / Aminomethylation / Silyl ketene acetals / Mannich-type iminium electrophiles / Scalable synthesis

Germany, 2007)

β-Amino acids are useful tools in the design of peptidomimetics, and the development of new methods for their syntheses, particularly the synthesis of β^2 -amino acids, remains an important challenge. Here we report a new scalable route

Introduction

The development of scalable syntheses of β -amino acids has constituted an important challenge since oligomers of these compounds were reported to be excellent peptidomimetics.^[1,2] This goal has been reached in the case of β^3 amino acids; indeed, the Arndt-Eistert homologation of natural amino acids allows the large-scale synthesis of all the β^3 -amino acids, which are now commercially available.^[3] This has permitted an extensive study of oligomers of these compounds and their use for the development of interesting peptidomimetics.

On the other hand, it is known that different β -peptide secondary structures,^[4] especially the design of β -turn-type structures,^[5] require the introduction of β^2 -amino acid isomers. Use of these compounds, however, is still highly limited for practical reasons, as no general methodology for the multigram preparation of these compounds has yet been reported (especially those bearing suitably protected functionalities on their side chains), even though it is a prerequisite for peptide synthesis. Although Seebach et al. succeeded in preparing all of the β^2 -amino acids bearing proteinogenic side chains, the syntheses of some are not straightforward, and are limited to laboratory scales.^[6] Many other methods

[a] Chemistry, Université Pierre et Marie Curie-Paris 6, FR 2769, CNRS/UMR 7613, Synthèse, Structure et Fonction de Molécules Bioactives, Université P. et M. Curie, Case courrier 45, 4 Place Jussieu, 75252 Paris Cedex 05, France Fax: +33-1-44273843 E-mail: karoyan@ccr.jussieu.fr [b] Senn Chemicals AG

- Industriestrasse 12, P. O. Box 267, 8157 Dielsdorf, Switzerland Chemistry, Université Pierre et Marie Curie-Paris 6, FR 2769, CNRS/UMR 7611, Laboratoire de Synthèse Organique, Université P. et M. Curie,

4 Place Jussieu, 75252 Paris Cedex 05, France

[†] Deceased November 6, 2005; this article is dedicated to his memory.

have been reported,^[7] including a recent elegant organocatalytic one developed by Gellman et al.,^[8] but none of them seems amenable to large-scale synthesis, especially in view of the purification steps required. The most widely used route involves aminomethylation of an enolate equivalent bearing the side chain of the amino acid. Many aminomethylating agents for this purpose have been reported, but their use presents severe limitations that hinder up-scaling, such as their low stabilities and/or reactivities and the harsh conditions required to deprotect the amine moieties.^[9] We recently reported our preliminary results on the homologation of α -amino acids to β^2 -amino acids by means of Reformatsky-type reactions using highly reactive preformed iminium ions as aminomethylating agents.^[10] Here we disclose the scope and limitations of this approach, together with its extension to the aminomethylation of silvl enol ethers, which has allowed the development of a highly efficient and scalable synthesis of a wide range of optically pure β^2 -amino acids.

based on the aminomethylation of silvl ketene N,O-acetals

(© Wiley-VCH Verlag GmbH & Co. KGaA, 69451 Weinheim,

by Mannich-type iminium electrophiles.

Results and Discussion

The classical Reformatsky reaction involves nucleophilic species produced through zinc insertion into the carbonhalogen bonds in a-haloacetate esters.^[11] This reaction is usually conducted under very mild conditions, the major proviso being that highly reactive electrophiles are usually needed. The addition of Reformatsky reagents to reactive imine derivatives has been widely used for the preparation of β^3 - or disubstituted $\beta^{2,3}$ -amino acids (Gilman-Speeter reaction),^[12] but the aminomethylation of such reagents is limited by the inaccessibility of stable and reactive formylimine equivalents. Interestingly, Mannich-type iminium ions are very reactive aminomethylating agents: Knochel et al. have reported the aminomethylation of organozinc and

1912

Grignard reagents through the use of iminium trifluoroacetates generated in situ.^[13] We have thus explored the scope of these iminium trifluoroacetate salts with regard to the Reformatsky reaction.

Aminal **1** was generated by heating dibenzylamine and aqueous formaldehyde at reflux, as reported by Knochel et al.,^[13] and isolated by crystallization as a stable solid compound (Scheme 1). Treatment of this aminal with trifluoroacetic anhydride produced the corresponding trifluoroacetate iminium salt, which was directly used in the Reformatsky reaction without further purification.

$$NBn_{2}H \xrightarrow{CH_{2}O, 37\% \text{ aq}} Bn_{2}N \bigvee NBn_{2} \xrightarrow{(CF_{3}CO)_{2}O} N \underset{\parallel}{\overset{Bn}{\sim}} \overset{+,Bn}{\underset{\parallel}{}} CF_{3}CO_{2}^{-}$$

Scheme 1. Synthesis of the iminium trifluoroacetate salt.

α-Bromo esters **4a–e**, bearing proteinogenic side chains, if not commercially available, were easily obtained from the corresponding α-amino acids by diazotization,^[14] and subsequent esterification under conditions compatible with classical protective groups for peptide synthesis.^[15] The zinc intermediate was generated, as reported by Dardoize et al.,^[16] in formaldehyde dimethylacetal at room temperature and then treated with dibenzylideneiminium trifluoroacetate, providing compounds **5a–e** in good yields (Scheme 2 and Table 1). Subsequent debenzylation with ammonium formate in the presence of palladium on charcoal,^[17] followed by ester saponification and *N*-Boc protection, provided racemic β²-amino acids suitably protected for peptide synthesis.



Scheme 2. Synthesis of racemic β^2 -amino acids through Reformatsky reactions with the iminium trifluoroacetate salt.

Table 1. Yields for the Reformatsky reactions with iminium trifluo-roacetate.

Entry	R	rac-5 (%)
a	CH ₃ (4a)	5a (43)
b	<i>i</i> Bu (4b)	5b (66)
с	CH_2Ph (4c)	5c (74)
d	CH_2 -(3-indolyl-N-Boc) (4d)	5d (73)
e	CH_2CO_2 - tBu (4e)	5e (72)

The ease of this reaction prompted us to explore its asymmetric version for the synthesis of β^2 -homoleucine, which was needed for our structure/activity relationship studies of biologically active peptides. In classical Reformatsky reactions, the attachment of chiral auxiliaries

(derived from menthol, oxazolidines or oxazinanes) to halo precursors had previously given rise to some diastereoselectivity and enantiofacial differentiation.^[11] We chose Oppolzer's sultam, easily obtained on multigram scales for laboratory needs and used on kilogram scales for industrial purposes (Scheme 3).^[18] Good to excellent diastereoselectivities are usually observed with this chiral auxiliary. Sulfonamide **6a** was thus obtained by coupling 2-bromo-3-methylbutanoic acid with (+)-camphorsultam as shown in Scheme 3.



(*R*)– β^2 -homoleucine

Scheme 3. Synthesis of optically pure β^2 -homoleucine through a Reformatsky reaction using the iminium trifluoroacetate salt. S* = (+)-camphorsultam.

Treatment of sulfonamide **6a** under the conditions described above resulted in the corresponding β^2 -amino acid derivative **7a** as a single diastereoisomer. The stereochemistry of compound **7a** was deduced by comparison of the optical rotation of the *N*-Boc- β^2 -homoleucine (obtained through deprotection/reprotection steps) with that of an authentic sample.^[19] The optical purity was confirmed by NMR analysis by the methodology reported by Seebach et al. (Scheme 4).^[20] It should be noted that the stereochemical course of this aminomethylation reaction is identical to that usually observed with Oppolzer's sultam for glycinate-derived enolates.^[21]



Scheme 4. Enantiomeric excess.

We then studied the scope of this aminomethylation reaction with different α -bromosulfonamides (Scheme 5 and Table 2). The mild conditions required for the Reformatsky reaction are compatible with many of the sensitive protecting groups typically used in peptide synthesis. This could allow for the direct synthesis of β^2 -amino acids bearing functional groups on their side chains, avoiding many deprotection/reprotection steps usually required for their preparation.

Table 2. Levels of conversion and dr values for asymmetric Reformatsky reactions with the iminium trifluoroacetate salt.

Entry	R	Product	Conversion (yield) ^[a]	$dr^{[a]}$
a	<i>i</i> Bu (6a)	7a	69% (64%)	>98:2
b	CH ₃ (6b)	7b	71% (nd)	72:28
с	CH_2Ph (6c)	7c	71% (57%)	87:13
d	$CH_2(N-Boc-indolyl)$ (6d)	7d	nd ^[b]	nd ^[c]
e	(CH ₂) ₄ NHBoc (6e)	7e	69% (47%)	nd ^[c]

[a] Calculated from the NMR spectrum of the crude mixture. [b] A complex mixture that contained the product was obtained. [c] Complex mixture of diastereoisomers with *cis/trans-N*-Boc isomerism.



Scheme 5. Asymmetric synthesis of β^2 -amino acids through Reformatsky reactions using the iminium trifluoroacetate salt. S* = (+)-camphorsultam.

Unfortunately, use of α -bromo sulfonamides bearing less hindered side chains gave disappointing results. Indeed, with all the compounds tested, inseparable mixtures of diastereoisomers were obtained. Screening of different reaction conditions (solvents, temperature, reagent loading) and use of other chiral auxiliaries (Evans oxazolidinone, menthol) did not improve the diastereoselectivities of the aminomethylations.^[22]

We anticipated that the use of classical enolate chemistry with this aminomethylating reagent should solve the problem, but all the attempts to achieve reactions with ester enolates generated from various bases with the iminium trifluoroacetate ion failed. Silyl enol ethers have been reported as an alternative to ester enolates,^[23] so we examined the reactivity of the chiral silyl ketene N,O-acetals, generated in situ from Oppolzer's sultam derivatives, toward the iminium trifluoroacetate salt (Scheme 6 and Table 3).



Scheme 6. Aminomethylation of chiral silyl ketene N,O-acetal with the iminium trifluoroacetate salt. $S^* = (+)$ -camphorsultam.

In a first attempt the silvl ketene acetal was generated from the corresponding alkanoylsultam sulfonamide^[24] by treatment with tert-butyldimethylsilyl triflate and triethylamine at room temperature.^[25] After addition of the iminium trifluoroacetate salt, the protected β^2 -homoalanine derivative 11a was obtained in moderate yield (60%) and with virtually complete diastereoselectivity. In the case of derivatives bearing hindered side chains (11b-d, 11f-g), the formation of the corresponding ketene acetal was not observed. Replacement of tert-butyldimethylsilyl triflate with the cheaper trimethylsilyl triflate allowed the formation of the silvlenol intermediates of all other alkanoylsultam sulfonamides. Indeed, tert-butyldimethylsilyl triflate was inefficient, probably because of the steric hindrance developed in the transition state, preventing the formation of the (Z)silvlenol ether. These intermediates reacted with the iminium salt to give the corresponding protected β^2 -amino acids as single diastereoisomers, with all compounds being obtained in excellent yields as solids. The stereochemical

	Table 3.	Yields	of	the	asymme	tric	aminon	nethy	lations	of	silyl	ketene	N.	O-acetal	S.
--	----------	--------	----	-----	--------	------	--------	-------	---------	----	-------	--------	----	----------	----

Entry	R	R'Me ₂ SiOTf	Product (yield)	dr ^[b]
а	CH ₃ (9a)	TBDMSOTf	11a (60%)	>98:2
b		TMSOTf	(91%)	>98:2
с	<i>i</i> Pr (9b)	TBDMSOTf	11b (0%)	
d		TMSOTf	(81%)	>98:2
e	<i>i</i> Bu (9c)	TBDMSOTf	11c (0%)	
f		TMSOTf	(82%)	>98:2
g	Bn (9d)	TBDMSOTf	11d (0%)	
ĥ		TMSOTf	(72%)	>98:2
i	hexyl (9e)	TBDMSOTf	11e (61%)	>98:2
j		TMSOTf	(74%)	>98:2
k	Br-(CH ₂) ₄ (9f)	TMSOTf	11f (76%)	>98:2
1	<i>N</i> -benzylindolyl-3-methylene (9g) ^[a]	TMSOTf	11g (88%)	>98:2

[a] Use of 2 equiv. of TMSOTf. [b] Calculated from NMR data of the crude mixture.

course of this aminomethylation reaction is identical to that observed in the Reformatsky strategy,^[10] and also to that usually observed with glycine enolates derived from Oppol-zer's sultam.^[21]

The obtained derivatives were then debenzylated with ammonium formate in the presence of palladium on charcoal, with the chiral auxiliary being removed by saponification. All compounds were isolated by simple crystallization after Boc protection of their amine moieties (Scheme 7). The optical purities were confirmed by measurement of the optical rotations and comparison with the literature data.



Scheme 7. Deprotection and Boc protection.

Conclusions

We have developed a straightforward synthesis of β^2 amino acids, which has been easily scaled up for the preparation of the β^2 -homophenylalanine derivative in 100 g quantities in the laboratory by use of the same procedure as described for smaller scales. All reactions may be performed at room temperature and the compounds can be isolated by simple precipitation or crystallization. With the strategy described here we have shown that β^2 -amino acids with various side chains can be obtained, establishing the versatility of this aminomethylation reaction. This route opens a new avenue to explore the potential of β^2 -amino acids in various fields.

Experimental Section

General Considerations: Tetrahydrofuran (THF) was distilled from sodium/benzophenone and kept over molecular sieves (4 Å). Dichloromethane (CH₂Cl₂) and triethylamine (NEt₃) were distilled from CaH₂. Anhydrous methylal, anhydrous DMF and other reagents were obtained from commercial suppliers and used without further purification. Compound 4a is commercially available. Oppolzer's sultams were a gift from Senn Chemicals. Anhydrous reactions were performed under argon, and glassware was dried in an oven prior to use. Prior to preparation of the HBr solution, the commercial concentrated HBr was washed with a solution of tributyl phosphate in CHCl₃ (5%) to remove contaminating Br₂. Reaction progress was monitored by TLC on precoated silica plates (Merck 60 F₂₅₄ 0.25 µm). Spots were detected by UV, molybdophosphoric acid oxidation or ninhydrin reagent. Flash column chromatography was carried out with Merck 60 F254 0.040-0.063 µm silica gel. ¹H and ¹³C NMR spectra were recorded with Bruker Avance 250 or 400 MHz spectrometers. Chemical shifts (δ) are reported in ppm relative to internal TMS. Optical rotations were determined at 20 °C with a Perkin-Elmer Model 141 polarimeter with a 10 cm path-length cell. Microanalyses were obtained from CNRS central service. HR mass spectra were recorded at the Université Pierre et Marie Curie with an LTQ Orbitrap™ (Thermoelectron Corporation, Bremen, Germany). Cy = cyclohexane; EA = ethyl acetate.

General Procedure for the Conversion of α -Amino Acids with Nonfunctionalized Side Chains into α -Bromo Acids: Sodium nitrite (1.25 equiv.) in H₂O was added dropwise at 0 °C to a stirred solution of L-amino acid and sodium bromide (3.5 equiv.) in H₂SO₄ (2.5 M, 1.3 mL per mmol). After stirring at 0 °C for 1 h, the reaction mixture was brought to room temperature and stirred for 6 h. The mixture was extracted with ethyl acetate, and the combined organic solutions were washed with brine, dried (MgSO₄) and concentrated in vacuo.

General Procedure for the Conversion of Amino Acids with Functionalized Side Chains into the Corresponding *a*-Bromo Acids: NaBr (3.7 equiv.) was dissolved in an aqueous solution of HBr (0.75 M) in a three-necked flask with an overhead stirrer. The solution was cooled to -15 °C and sodium nitrite (1.3 equiv.) was added in one portion. The amino acid (1 equiv.) was then added and the reaction mixture was kept between -15 °C and -10 °C for 1.5 h. The mixture was extracted twice with precooled EtOAc (0 °C) and the combined organic layers were washed with saturated aqueous solution of NH₄Cl, dried (MgSO₄) and concentrated in vacuo.

Protection of the Acid Function for Compounds with Nonfunctionalized Side Chains: The α -bromo acid was treated with a solution of concentrated sulfuric acid (30 μ L per mmol) in methanol (2 mL per mmol) and the mixture was heated at reflux for 1 h. The solution was allowed to cool to room temperature and concentrated in vacuo, Et₂O was added, and the organic layer was washed with aqueous NaHCO₃ (5%) followed by brine, dried with MgSO₄ and concentrated in vacuo.

Protection of the Acid Function for Compounds with Functionalized Side Chains: The α -bromo acid was treated with a solution of acetyl chloride (30 μ L per mmol) in methanol (2 mL per mmol) for 1 h. The solution was allowed to cool to room temperature and concentrated in vacuo.

General Procedure for the Acylation of Oppolzer's Sultam Starting from Acid Chloride Derivatives: A solution of 10,2-camphorsultam (1 equiv.) in dimethoxyethane (3 mL per mmol) was added slowly at room temperature to a stirred suspension of NaH (1.1 equiv.) in the same solvent. After the mixture had been stirred at room temperature for 1 h, the acid chloride derivative (1.1 equiv.) was added. The mixture was stirred at room temperature for an additional 1 h, and was then quenched with water and concentrated. After addition of ethyl acetate, the solution was washed with saturated aqueous NH₄Cl, and the organic layer was dried with anhydrous MgSO₄ and concentrated. The acylsultam derivatives were obtained by precipitation from diethyl ether/pentane.

General Procedure for the Acylation of Oppolzer's Sultam Starting from Carboxylic Acid Derivatives: A solution of 10,2-camphorsultam (1 equiv.) in dimethoxyethane (3 mL per mmol) was added slowly at room temperature to a stirred suspension of NaH (1.1 equiv.) in the same solvent. The solution was stirred for 1 h. In parallel, isobutyl chloroformate (1.1 equiv.) and N-methylmorpholine (1.1 equiv.) were added at -15 °C to a solution of the carboxylic acid (1 equiv.) in dimethoxyethane (3 mL per mmol). After 15 min of stirring at -15 °C, this preformed mixed anhydride was rapidly introduced, after filtration through a Celite pad, into the sultam sodium salt solution. After 1 h of stirring at -15 °C and 1 h at room temperature, the crude mixture was quenched with water and concentrated. Ethyl acetate was added and the organic layer was washed with saturated aqueous NH₄Cl, dried with anhydrous MgSO₄ and concentrated. Acylsultam derivatives were obtained by precipitation from diethyl ether/pentane.

General Procedure for the Reformatsky Reaction: Dibromoethane (20 μ L per mmol) was added to a suspension of zinc dust (1.5 equiv.) in methylal (0.5 mL per mmol) and the mixture was heated at reflux for a few minutes. A solution of the α -bromo acid (1 equiv.) in methylal (1 mL per mmol) was added dropwise, the reaction mixture was stirred at room temperature for 15 min, and a solution of the iminium trifluoroacetate, freshly prepared from *N*,*N*,*N'*,*N'*-tetrabenzylmethanediamine,^[13] was added. The reaction mixture was stirred at room temperature for 1.5 h, and was then quenched with a saturated solution of NH₄Cl and extracted with Et₂O. The combined organic layers were washed with saturated aqueous NaHCO₃, aqueous Na₂S₂O₃ (5%) and saturated aqueous NH₄Cl, and were then dried (MgSO₄) and concentrated.

General Procedure for the Aminomethylation: Trimethylsilyl triflate (1.5 equiv.) and triethylamine (1.1 equiv.) were added successively to a stirred solution of the sulfonamide derivative (1 equiv.) in CH_2Cl_2 (1 mL per mmol) and the mixture was stirred at room temperature for 6 h. A solution of *N*,*N*-dibenzyliminium trifluoroacetate salt (1.1 equiv.) in CH_2Cl_2 (1 mL per mmol) freshly prepared from *N*,*N*,*N'*,*N'*-tetrabenzylmethanediamine^[13] was added and the mixture was stirred for 15 min. TBAF in THF (1 M, 1 equiv.) was added, the reaction was quenched with water, and CH_2Cl_2 was removed in vacuo. The aqueous layer was extracted with diethyl ether and the combined organic solutions were washed with a saturated solution of NH₄Cl, dried (MgSO₄) and concentrated in vacuo.

General Procedure for the Deprotection and Boc Protection: Pd/C (10%, 100 mg per mmol) and ammonium formate (5 equiv.) were added to a solution of the amine in a THF/MeOH mixture (1:4, 5 mL per mmol) and the reaction mixture was stirred at room temperature. The solution was filtered through Celite and concentrated to give the corresponding amine, which was dissolved in a THF/ water mixture (1:1, 5 mL per mmol). After addition of LiOH (2 equiv.), the reaction mixture was stirred at room temperature overnight, the THF was then evaporated, and dioxane (2 mL per mmol) was added. After addition of Boc₂O (1.1 equiv.) and K₂CO₃ (1.1 equiv.), the solution was stirred for 5 h and then concentrated to remove organic solvents. After addition of water and extraction with Et₂O, the aqueous layer was cooled to 0 °C, acidified to pH = 2 with HCl (1 M), and extracted with ethyl acetate. The combined organic layers were dried and concentrated in vacuo to afford the Boc-protected amino acid, which was purified by recrystallization from diethyl ether/pentane.

(*S*)-2-Bromopropionic Acid (3a):^[26] This compound was prepared from L-alanine (50 g, 562 mmol), the crude product being purified by distillation under reduced pressure (P = 0.06 Torr, T = 63-64 °C) to give an oil (58.7 g, 68%); $R_{\rm f} = 0.24$ (Cy/EA/AcOH, 8:2:0.1). $[a]_{\rm D}^{20} = -28.5$ (c = 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 11.54$ (br. s, 1 H), 4.42 (q, J = 6.8 Hz, 1 H), 1.85 (d, J = 6.8 Hz, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 176.6$, 39.4, 21.4 ppm. C₃H₅BrO₂ (152.97): calcd. C 23.55, H 3.29; found C 23.60, H 3.55.

(*S*)-2-Bromo-4-methylpentanoic Acid (3b):^[27] This compound was prepared from L-leucine (20 g, 153 mmol), the crude product being purified by distillation under reduced pressure (P = 0.03 Torr, T = 96 °C) to give an oil (20.8 g, 70%); $R_{\rm f} = 0.60$ (CH₂Cl₂/EtOH, 9:1). $[a]_{\rm D}^{20} = -39.9$ (c = 2, MeOH). ¹H NMR (400 MHz, CDCl₃): $\delta = 9.80-10.50$ (br. s, 1 H), 4.30 (t, J = 7.5 Hz, 1 H), 1.93 (t, J = 7.5 Hz, 2 H), 1.76–1.90 (m, 1 H), 0.98 (d, J = 6.5 Hz, 3 H), 0.94 (d, J = 6.25 Hz, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 176.4$, 44.0, 43.2, 26.3, 22.3, 21.5 ppm.

(*S*)-2-Bromo-3-phenylpropionic Acid (3c):^[28] This compound was prepared from L-phenylalanine (20 g, 121.2 mmol), the crude prod-

uct being purified by flash column chromatography (Cy/EA/AcOH, 99:1) to give an oil (19 g, 68%), $R_{\rm f} = 0.33$ (CH₂Cl₂/EtOH/AcOH, 9:1:0.1). [*a*]_D²⁰ = -11.4 (*c* = 2, MeOH). ¹H NMR (400 MHz, CDCl₃): δ = 7.18–7.36 (m, 5 H), 4.42 (t, *J* = 7.7 Hz, 1 H), 3.47 (dd, *J* = 8 Hz, 14.2 Hz, 1 H), 3.24 (dd, *J* = 7.2 Hz, 14.2 Hz, 1 H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 175.5, 136.3, 129.4, 129.0, 128.5, 127.5, 44.8, 40.7 ppm. C₉H₉BrO₂ (229.07): calcd. C 47.19, H 3.96; found C 47.21, H 3.60.

(*S*)-2-Bromo-3-[*N*-(*tert*-butoxycarbonyl)indol-3-yl]propionic Acid (3d):^[14b] This compound was prepared from H-Trp(Boc)-OH (5 g, 16.4 mmol), the crude product being purified by flash column chromatography (Cy/EA/AcOH, 7:3:0.1) to give compound **3d** as an oil (4 g, 67%); $R_f = 0.21$ (Cy/EA/AcOH, 7:3:0.1). $[a]_D^{20} = -13.2$ (c = 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 8.87$ (br. s, 1 H), 8.10 (br. s, 1 H), 7.51–7.53 (m, 2 H), 7.32 (t, J = 7.3 Hz, 1 H), 7.24 (t, J = 6.8 Hz, 1 H), 4.52 (t, J = 7.6 Hz, 1 H), 3.58 (dd, J = 8.1 Hz, 14.9 Hz, 1 H), 2.93 (dd, J = 6.8 Hz, 14.9 Hz, 1 H), 1.65 (s, 9 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 174.5$, 149.6, 135.3, 129.7, 124.7, 124.5, 122.7, 118.5, 115.5, 115.4, 84.0, 43.3, 30.5, 28.2 ppm.

(*S*)-2-Bromo-3-(*tert*-butoxycarbonyl)propionic Acid (3e);^[14b] This compound was prepared from H-Asp(O/Bu)-OH (5 g, 26.4 mmol), the crude product being purified by flash column chromatography (Cy/EA/AcOH, 7:3:0.1) to give compound **3e** as an oil (11.5 g, 86%); $R_{\rm f} = 0.48$ (Cy/EA/AcOH, 6:4:0.1). $[a]_{\rm D}^{20} = -45.4$ (c = 2, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 10.60$ (br. s, 1 H), 4.55 (dd, J = 9.1 Hz, 6.1 Hz, 1 H), 3.18 (dd, J = 9.1 Hz, 16.9 Hz, 1 H), 2.93 (dd, J = 6.1 Hz, 16.9 Hz, 1 H), 1.40 (s, 9 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 174.8$, 168.7, 82.5, 40.7, 38.1, 27.9 ppm. C₉H₁₃BrO₂ (233.10): calcd. C 37.96, H 5.18; found C 37.70, H 5.33.

Methyl (S)-2-Bromo-4-methylpentanoate (4b): Ester **4b** was purified by flash column chromatography (Cy/EA, 99:1) to give an oil (2.5 g, 79%), $R_{\rm f} = 0.67$ (Cy/EA, 8:2). $[a]_{\rm D}^{20} = -49.6$ (c = 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 4.29$ (t, J = 7.6 Hz, 1 H), 3.78 (s, 3 H), 1.86–2.05 (m, 2 H), 1.71–1.81 (m, 1 H), 0.96 (d, J = 6.8 Hz, 3 H), 0.91 (d, J = 6.8 Hz, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 170.6$, 52.9, 44.3, 43.5, 26.3, 22.3, 21.5 ppm.

Methyl (S)-2-Bromo-3-phenylpropanoate (4c):^[29] Ester **4c** was purified by flash column chromatography (Cy/EA, 99:1) to give an oil (6.8 g, 94%); $R_{\rm f} = 0.33$ (Cy/EA, 95:5). $[a]_{\rm D}^{20} = -11.1$ (c = 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.17-7.29$ (m, 5 H), 4.39 (t, J = 8.3 Hz, 1 H), 3.36 (s, 3 H), 3.45 (dd, J = 8.3 Hz, 14.1 Hz, 1 H), 3.22 (dd, J = 7.1 Hz, 13.9 Hz, 1 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 169.8$, 136.7, 129.1, 128.6, 127.3, 52.8, 45.1, 41.0 ppm.

Methyl (*S*)-2-Bromo-3-[*N*-(*tert*-butoxycarbonyl)indol-3-yl]propionate (4d): The crude product was purified by flash column chromatography (Cy/EA, 97:3) to give compound 4d as an oil (1 g, 59%); $R_f = 0.58$ (Cy/EA, 8:2). $[a]_D^{20} = -18.6$ (c = 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 8.12-8.13$ (m, 1 H), 7.47–7.53 (m, 2 H), 7.31–7.34 (m, 2 H), 4.50 (dd, J = 8.6 Hz, 6.6 Hz, 1 H), 3.73 (s, 3 H), 3.58 (dd, J = 8.6 Hz, 14.9 Hz, 1 H), 3.35 (dd, J = 6.6 Hz, 14.9 Hz, 1 H), 1.66 (s, 9 H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 170.0, 150.0, 135.5, 129.7, 124.7, 124.5, 122.7, 118.6, 115.8, 115.4, 83.8, 53.0, 43.5, 30.8, 20.2 ppm. C₁₇H₂₀BrNO₄ (382.25): calcd. C 53.42, H 5.27, N 3.66; found C 53.00, H 5.43, N 3.27.

Methyl (*S*)-2-Bromo-4-(*tert*-butoxycarbonyl)propionate (4e): The crude product was purified by flash column chromatography (Cy/ EA, 97:3) to give compound 4e as an oil (2.9 g, 51%); $R_{\rm f} = 0.53$ (Cy/EA, 8:2). $[a]_{\rm D}^{20} = -53.5$ (c = 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 4.53$ (dd, J = 9.1 Hz, 6.1 Hz, 1 H), 3.80 (s, 3 H), 3.19 (dd, J = 9.1 Hz, 16.9 Hz, 1 H), 2.91 (dd, J = 6.1 Hz, 16.9 Hz, 1 H), 1.44 (s, 9 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 169.7$, 168.6,

82.0, 53.1, 40.9, 38.3, 27.9 ppm. $C_9H_{15}BrO_4$ (267.12): calcd. C 40.47, H 5.66; found C 40.89, H 5.68.

Methyl 2-[(Dibenzylamino)methyl]propionate (5a): The crude product was treated with a solution of TFA in EA (1%) and purified by flash column chromatography (Cy/EA, 8:2; then Cy/EA/NEt₃, 8:2:0.1) to give **5a** as an oil (3 g, 43%); $R_f = 0.61$ (Cy/EA, 8:2). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.19-7.30$ (m, 10 H), 3.62 (d, J = 13.4 Hz, 2 H), 3.61 (s, 3 H), 3.46 (d, J = 13.4 Hz, 2 H), 2.73–2.70 (m, 2 H), 2.37–2.44 (m, 1 H), 1.07 (d, J = 6.3 Hz, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 176.1$, 139.2, 128.9, 128.1, 126.9, 58.4, 57.3, 51.4, 38.6, 15.3 ppm. C₁₉H₂₃NO₂ (297.39): calcd. C 76.73, H 7.80, N 4.71; found C 76.87, H 7.99, N 4.65.

Methyl 2-[(Dibenzylamino)methyl]-4-methylpentanoate (5b): The crude product was treated with a solution of TFA in EA (1%) and purified by flash column chromatography (Cy/EA, 8:2; then Cy/ EA/NEt₃, 8:2:0.1) to give **5b** as an oil (4.8 g, 66%); $R_f = 0.69$ (Cy/ EA, 8:2). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.20-7.30$ (m, 10 H), 3.72 (d, J = 13.6 Hz, 2 H), 3.63 (s, 3 H), 3.36 (d, J = 13.6 Hz, 2 H), 2.73–2.80 (m, 2 H), 2.36–2.39 (m, 1 H), 1.39–1.49 (m, 2 H), 1.15–1.22 (m, 1 H), 0.85 (d, J = 6.6 Hz, 3 H), 0.82 (d, J = 6.6 Hz, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 175.9$, 139.2, 128.9, 128.1, 126.9, 58.4, 56.8, 51.3, 42.9, 39.6, 26.3, 23.0, 22.0 ppm. C₂₂H₂₉NO₂ (339.47): calcd. C 77.84, H 8.61, N 4.13; found C 77.61, H 8.73, N 4.04.

Methyl 2-I(Dibenzylamino)methyl]-3-phenylpropionate (5c): Compound 5c was obtained after precipitation in MeOH/CHCl₃ as a white powder (14.6 g, 74%); $R_{\rm f} = 0.65$ (Cy/EA, 8:2); m.p. 83 °C. ¹H NMR (400 MHz, CDCl₃): $\delta = 7.1-7.3$ (m, 15 H), 3.66 (d, J = 13.6 Hz, 2 H), 3.42 (d, J = 13.6 Hz, 2 H), 3.55 (s, 3 H), 2.95–3.00 (m, 3 H), 2.79–2.85 (m, 1 H), 2.75–2.76 (m, 2 H), 2.50–2.54 (m, 1 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 174.9$, 139.2, 139.0, 129.0, 128.7, 128.4, 128.2, 127.0, 126.3, 58.5, 55.9, 51.3, 46.9, 36.4 ppm. C₂₅H₂₇NO₂ (373.49): calcd. C 80.40, H 7.29, N 3.75; found C 80.16, H 7.34, N 3.67.

Methyl 3-[*N*-(*tert*-Butoxycarbonyl)indol-3-yl]-2-[(dibenzylamino)methyl]propionate (5d): Compound 5d was obtained after flash column chromatography (Cy/EA, 98:2) as an oil (14.6 g, 73%); $R_{\rm f}$ = 0.53 (Cy/EA, 8:2). ¹H NMR (400 MHz, CDCl₃): δ = 8.1 (br. s, 1 H), 7.43 (d, *J* = 7.6 Hz, 1 H), 7.19–7.31 (m, 14 H), 3.67 (d, *J* = 13.6 Hz, 2 H), 3.45 (d, *J* = 13.6 Hz, 2 H), 3.59 (s, 3 H), 3.07–3.13 (m, 1 H), 2.80–2.90 (m, 1 H), 2.59 (dd, 1 H, 5.8 Hz, 12.6 Hz), 1.65 (s, 9 H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 175.1, 149.7, 138.0, 130.3, 128.9, 128.1, 127.0, 124.3, 123.1, 122.4, 118.8, 115.2, 83.4, 58.4, 56.0, 51.5, 44.8, 28.2, 25.7 ppm. MS (CI; NH₃): *m*/*z* = 513 [M + H]⁺, 413 [M – Boc]⁺.

Methyl 3-(*tert*-Butoxycarbonyl)-2-[(dibenzylamino)methyl]propionate (5e): Compound 5e was obtained after flash column chromatography (Cy/EA, 97:3) as an oil (1.3 g, 72%); $R_{\rm f}$ = 0.49 (Cy/EA, 8:2). ¹H NMR (400 MHz, CDCl₃): δ = 7.2–7.4 (m, 10 H), 3.59 (d, J = 13.6 Hz, 2 H), 3.51 (d, J = 13.6 Hz, 2 H), 3.3 (s, 3 H), 3.0–3.1 (m, 1 H), 2.7 (dd, J = 7.3 Hz, 10.2 Hz, 1 H), 2.4–2.6 (m, 3 H), 1.4 (s, 9 H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 174.4, 171.2, 138.9, 128.9, 128.2, 127.0, 126.3, 80.7, 58.3, 55.1, 51.7, 40.5, 35.6, 28.0 ppm. C₂₄H₃₁NO₄ (397.51): calcd. C 72.52, H 7.86, N 3.52; found C 72.72, H 7.89, N 3.46.

(2.5)-*N*-(2-Bromo-4-methylpentanoyl)camphorsultam 6a:^[30] 61% yield (8 g); $R_{\rm f} = 0.23$ (Cy/EA, 9:1); m.p. 132 °C. $[a]_{\rm D}^{20} = 59.5$ (c = 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 4.91$ (t, J = 7.3 Hz, 1 H), 3.93 (dd, J = 4.8 Hz, 7.3 Hz, 1 H), 3.49 (d, J = 13.9 Hz, 2 H), 3.39 (d, J = 13.9 Hz, 2 H), 1.94–2.09 (m, 3 H), 1.88–1.94 (m, 4 H), 1.74 (m, 1 H), 1.36–1.45 (m, 2 H), 1.20 (s, 3 H), 0.98 (s, 3

H), 0.97 (d, J = 6.8 Hz, 3 H), 0.92 (d, J = 6.6 Hz, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 168.3$, 64.9, 53.0, 48.7, 47.9, 44.5, 43.7, 42.4, 37.5, 32.7, 26.5, 26.4, 22.4, 22.0, 20.7 19.9 ppm. C₁₆H₂₆BrNO₃S (392.35): calcd. C 48.98, H 6.68, N 3.57; found C 48.60, H 6.72, N 3.53.

(25)-*N*-(2-Bromopropionyl)camphorsultam 6b: 68% yield (12.3 g); $R_{\rm f} = 0.26$ (Cy/EA, 9:1); m.p. 132 °C. $[a]_{\rm D}^{20} = 81.0$ (c = 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 4.98$ (q, J = 6.6 Hz, 1 H), 3.93 (dd, J = 5.3 Hz, 7.3 Hz, 1 H), 3.44 (d, J = 13.9 Hz, 1 H), 3.56 (d, J = 13.9 Hz, 1 H), 1.98–2.11 (m, 2 H), 1.86–1.95 (m, 3 H), 1.82 (d, J = 6.8 Hz, 3 H), 1.34–1.46 (m, 2 H), 1.20 (s, 3 H), 0.99 (s, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 168.7$, 64.9, 52.9, 48.8, 47.8, 44.5, 39.4, 37.5, 32.8, 26.5, 20.7, 20.5, 19.9 ppm. C₁₃H₂₀BrNO₃S (350.27): calcd. C 44.58, H 5.76, N 4.00; found C 44.59, H 5.83, N 3.95.

(2*S*)-*N*-(2-Bromo-3-phenylpropionyl)camphorsultam 6c:^[30] 82% yield (6.7 g); $R_{\rm f} = 0.23$ (Cy/EA, 9:1); m.p. 156–157 °C. $[a]_{20}^{2D} = 84.1$ (c = 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.32-7.23$ (m, 5 H), 5.07 (t, J = 7.1 Hz, 1 H), 3.88 (dd, J = 7.6 Hz, 5 Hz, 1 H), 3.53 (dd, J = 14.2 Hz, 7.0 Hz, 1 H), 3.39 (d, J = 13.9 Hz, 1 H), 3.49 (d, J = 13.9 Hz, 1 H), 3.25 (dd, J = 7.3 Hz, 14.2 Hz, 1 H), 2.01–2.09 (m, 2 H), 1.89–1.94 (m, 3 H), 1.30–1.41 (m, 2 H), 1.18 (s, 3 H), 0.96 (s, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 167.7$, 136.5, 129.6, 128.5, 127.2, 64.8, 52.9, 48.7, 47.8, 44.8, 44.5, 39.8, 37.5, 32.7, 26.4, 20.7, 19.9 ppm. C₁₉H₂₄BrNO₃S (426.37): calcd. C 53.52, H 5.67, N 3.29; found C 53.31, H 5.80, N 3.35.

(2*S*)-*N*-{2-Bromo-3-[*N*-(*tert*-butoxycarbonyl)indol-3-yl]propionyl}camphorsultam 6d: Compound 6d was obtained after purification by flash column chromatography (Cy/EA, 85:15) as a white powder (2.3 g, 44%); $R_{\rm f} = 0.18$ (Cy/EA, 9:1); m.p. 79 °C. $[a]_{\rm D}^{20} = 53.8$ (c =1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 8.1$ (m, 1 H), 7.54– 7.60 (m, 2 H), 7.24–7.33 (m, 2 H), 5.15–5.20 (m, 1 H), 3.91 (dd, *J* = 5.3 Hz, 7.1 Hz, 1 H), 3.63 (dd, *J* = 15.2 Hz, 7.1 Hz, 1 H), 3.50 (d, *J* = 13.6 Hz, 1 H), 3.44 (d, *J* = 13.6 Hz, 1 H), $\delta = 3.35$ (dd, *J* = 15.4 Hz, 7.6 Hz, 1 H), 2.01–2.10 (m, 2 H), 1.82–1.94 (m, 3 H), 1.66 (s, 9 H), 1.30–1.42 (m, 2 H), 1.18 (s, 3 H), 0.96 (s, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 167.7$, 149.6, 135.3, 129.9, 124.7, 124.4, 122.6, 118.9, 115.6, 115.2, 83.6, 64.9, 52.9, 48.8, 47.8, 44.5, 43.2, 37.5, 32.7, 29.6, 28.2, 26.4, 20.7, 19.8 ppm.

(2*S*)-*N*-{2-Bromo-6-[(*tert*-butoxycarbonyl)amino]hexanoyl}camphorsultam 6e: Compound 6e was obtained after purification by flash column chromatography (Cy/EA, 9:1) as a powder (430 mg, 70%); $R_{\rm f} = 0.37$ (Cy/EA, 8:2); m.p. 110–112 °C. $[a]_{\rm D}^{20} =$ 42.2 (*c* = 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 4.79$ (t, *J* = 7.1 Hz, 1 H), 3.93 (dd, *J* = 5.3 Hz, 7.3 Hz, 1 H), 3.69 (t, *J* = 7.3 Hz, 2 H), 3.53 (d, *J* = 13.6 Hz, 1 H), 3.43 (d, *J* = 13.6 Hz, 1 H), 2.04–2.09 (m, 3 H), 1.90–1.99 (m, 4 H), 1.65 (s, 9 H), 1.19–1.44 (m, 6 H), 1.19 (s, 3 H), 0.98 (s, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 168.1$, 152.1, 85.2, 64.9, 53.0, 48.8, 47.9, 44.5, 44.3, 40.2, 37.6, 33.2, 32.8, 28.1, 26.4, 24.6, 20.7, 19.9 ppm.

(2*S*)-*N*-{(2*R*)-2-[(Dibenzylamino)methyl]-4-methylpentanoyl}camphorsultam 7a: Compound 7a was obtained by precipitation from methanol/chloroform as a single diastereoisomer (7.2 g, 64%); $R_{\rm f} = 0.21$ (Cy/EA, 9:1); m.p. 108 °C. $[a]_{\rm D}^{20} = 69.9$ (c = 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.18-7.33$ (m, 10 H), 3.87 (dd, J = 5 Hz, 7.6 Hz, 1 H), 3.68 (d, J = 13.9 Hz, 2 H), 3.41–3.52 (m, 5 H), 2.76 (dd, J = 6.3 Hz, 12.4 Hz, 1 H), 2.50 (dd, J = 12.4 Hz, 7.3 Hz, 1 H), 2.00–2.11 (m, 2 H), 1.86–1.92 (m, 4 H), 1.46–1.56 (m, 3 H), 1.31–1.42 (m, 2 H), 1.22 (s, 3 H), 0.97 (s, 3 H), 0.87 (d, J = 5.8 Hz, 6 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 174.9$, 139.1, 129.1, 128.0, 126.8, 65.5, 58.1, 56.5, 53.2, 48.2, 47.8, 44.6, 42.6, 38.8, 38.6, 32.9, 26.5, 26.0, 23.0, 22.7, 21.0, 19.9 ppm.

 $C_{31}H_{42}N_2O_3S$ (522.74): calcd. C 71.23, H 8.10, N 5.36; found C 71.05, H 8.30, N 5.11.

(2*S*)-*N*-{2-[(Dibenzylamino)methyl]propionyl}camphorsultam 7b: Compound 7b was obtained as a mixture of diastereoisomers (72:28); $R_{\rm f} = 0.39$ (Cy/EA, 8:2); major diastereoisomer. ¹H NMR (400 MHz, CDCl₃): $\delta = 7.2$ -7.3 (m, 10 H), 3.9 (dd, J = 7.6 Hz, 5.1 Hz, 1 H), 3.6 (s, 4 H), 3.4–3.5 (m, 3 H), 2.9 (dd, J = 7.6 Hz, 12.6 Hz, 1 H), 2.4 (dd, J = 6.3 Hz, 12.6 Hz, 1 H), 2.0–2.1 (m, 1 H), 2.0 (dd, J = 7.6 Hz, 13.9 Hz, 1 H), 2.02–2.11 (m, 2 H), 1.86–1.91 (m, 3 H), 1.3–1.4 (m, 2 H), 1.2 (s, 3 H), 1.1 (s, 3 H), 1.0 (s, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 175.3$, 138.9, 128.0, 126.8, 65.4, 57.9, 57.8, 53.1, 48.2, 47.7, 44.7, 38.6, 38.4, 32.9, 26.4, 21.0, 19.9, 15.5 ppm.

(2*S*)-*N*-{2-[(Dibenzylamino)methyl]-3-phenylpropionyl}camphorsultam 7c: Compound 7c was obtained after purification by flash column chromatography (Cy/EA, 9:1) as a mixture of diastereoisomers (87:13, 5.1 g, 57%), $R_f = 0.41$ (Cy/EA, 8:2). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.16-7.26$ (m, 15 H), 3.86 (dd, J = 7.6 Hz, 5.1 Hz, 1 H), 3.73 (m, 1 H), 3.41–3.54 (m, 6 H), 3.04 (dd, J = 13.6 Hz, 5.6 Hz, 1 H), 2.92 (dd, J = 12.6 Hz, 8.1 Hz, 1 H), 2.65 (dd, J = 13.6 Hz, 8.6 Hz, 1 H), 2.49 (dd, J = 12.6 Hz, 5.3 Hz, 1 H), 2.02–2.11 (m, 2 H), 1.87–1.91 (m, 3 H), 1.31–1.38 (m, 2 H), 1.24 (s, 3 H), 0.98 (s, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 174.1$, 139.3, 138.7, 129.4, 129.1, 128.3, 128.0, 126.8, 126.2, 65.4, 57.7, 55.7, 53.2, 48.3, 47.8, 44.9, 44.8, 38.6, 36.0, 32.9, 26.5, 20.1, 19.9 ppm.

(2*S*)-*N*-{6-[(*tert*-Butoxycarbonyl)amino]-2-[(dibenzylamino)methyl]hexanoyl}camphorsultam 7e: Compound 7e was obtained after purification by flash column chromatography (Cy/EA, 9:1) as a mixture of diastereoisomers (251 mg, 47%); $R_f = 0.41$ (Cy/EA, 8:2). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.18-7.34$ (m, 10 H), 3.86 (t, *J* = 7.2 Hz, 1 H), 3.54–3.80 (m, 4 H), 3.45–3.53 (m, 3 H), 2.29–3.34 (m, 2 H), 2.75 and 2.69 (dd, *J* = 12.2 Hz, 6.2 Hz and dd, *J* = 13.5 Hz, 4.7 Hz, 1 H), 2.57 and 2.48 (dd, *J* = 13 Hz, 8.7 Hz and dd, *J* = 12.5 Hz, 7.2 Hz, 1 H), 2.03–2.06 (m, 2 H), 1.86–1.89 (m, 3 H), 1.52–1.64 (m, 13 H), 1.13–1.37 (m, 7 H), 0.96 and 0.98 (s and s, 3 H) ppm.

(2*S*)-*N*-Propionylcamphorsultam 9a:^[31] 91% yield (5.7 g); $R_{\rm f} = 0.43$ (Cy/EA, 8:2); m.p. 145 °C. $[a]_{\rm D}^{20} = 116.5$ (c = 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 3.86$ (dd, J = 7.6 Hz, 5.1 Hz, 1 H), 3.49 (d, J = 13.9 Hz, 1 H), 3.43 (d, J = 13.9 Hz, 1 H), 2.69–2.81 (m, 2 H), 2.04–2.16 (m, 1 H), 2.07 (dd, J = 13.6 Hz, 7.6 Hz, 1 H), 1.85–1.96 (m, 3 H), 1.32–1.44 (m, 2 H), 1.16 (t, 3 H, 7.3 Hz), 1.16 (s, 3 H), 0.97 (s, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 172.6$, 65.2, 52.9, 48.5, 47.7, 44.6, 38.5, 32.8, 28.9, 26.4, 20.8, 19.9, 8.39 ppm. $C_{13}H_{21}NO_{3}S$ (271.38): calcd. C 57.54, H 7.80, N 5.16; found C 57.60, H 7.83, N 5.07.

(2*R*)-*N*-(3-Methylbutanoyl)camphorsultam 9b:^[31] 93% yield (6.4 g), $R_{\rm f} = 0.45$ (Cy/EA, 8:2), m.p. 114 °C. $[a]_{\rm D}^{20} = -88.4$ (c = 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 3.88$ (t, J = 6.3 Hz, 1 H), 3.50 (d, J = 13.6 Hz, 1 H), 3.43 (d, J = 13.6 Hz, 1 H), 2.66 (dd, J = 15.6 Hz, 7.0 Hz, 1 H), 2.51 (dd, J = 15.6 Hz, 6.8 Hz, 1 H), 2.18– 2.28 (m, 1 H), 2.08–2.1 (m, 2 H), 1.87–1.96 (m, 3 H), 1.32–1.44 (m, 2 H), 1.16 (s, 3 H), 0.97 (s and t, 9 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 171.5$, 65.2, 53.0, 48.2, 47.7, 44.6, 44.2, 38.6, 32.8, 26.4, 25.5, 22.3, 22.2, 20.8, 19.8 ppm. C₁₅H₂₅NO₃S (299.43): calcd. C 60.17, H 8.42, N 4.68; found C 60.15, H 8.48, N 4.61.

(2*S*)-*N*-(4-Methylpentanoyl)camphorsultam 9c:^[31] 81 % yield (5.84 g); $R_{\rm f} = 0.44$ (Cy/EA, 8:2); m.p. 44 °C. $[a]_{20}^{20} = 97.6$ (c = 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 3.85-3.88$ (m, 1 H), 3.50 (d, J = 13.9 Hz, 1 H), 3.43 (d, J = 13.9 Hz, 1 H), 2.66–2.76 (m, 2 H), 2.04–2.13 (m, 2 H), 1.86–1.96 (m, 3 H), 1.54–1.63 (m, 2 H), 1.16 (s, 3 H), 0.97 (s, 3 H), 0.90 (d, J = 6.3 Hz, 3 H), 0.89 (d, J = 6.3 Hz, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 172.3$, 65.2, 53.0, 48.4, 47.7, 44.6, 38.5, 33.6, 33.2, 32.8, 27.6, 26.5, 22.3, 22.2, 20.8, 19.9 ppm. C₁₆H₂₇NO₃S (313.46): calcd. C 61.31, H 8.68, N 4.47; found C 61.15, H 8.85, N 4.41.

(2*S*)-*N*-(3-Phenylpropanoyl)camphorsultam 9d:^[31] 92% yield (11.9 g); $R_{\rm f} = 0.37$; (Cy/EA, 8:2); m.p. 149 °C. [*a*]₂₀^D = 82.2 (*c* = 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.2-7.3$ (m, 5 H), 3.86 (t, *J* = 6.3 Hz, 1 H), 3.48 (d, *J* = 13.6 Hz, 1 H), 3.41 (d, *J* = 13.6 Hz, 1 H), 3.0–3.1 (m, 4 H), 2.0–2.1 (m, 2 H), 1.8–1.9 (m, 3 H), 1.3–1.4 (m, 2 H), 1.08 (s, 3 H), 0.95 (s, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 171.1$, 140.1, 128.5, 128.4, 126.2, 65.2, 52.9, 48.4, 47.7, 44.6, 38.4, 36.9, 32.8, 30.4, 26.4, 20.8, 19.9 ppm. C₁₉H₂₅NO₃S (347.47) calcd. C 65.68, H 7.25, N 4.03; found C 65.40, H 7.35, N 3.99.

(2*S*)-*N*-Octanoylcamphorsultam 9e: This compound, obtained as an oil, was purified by flash column chromatography (Cy/EA, 95:5), 92% yield (7.32 g); $R_{\rm f} = 0.61$ (Cy/EA, 8:2). $[a]_{\rm D}^{20} = 88.4$ (c = 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 3.86$ (dd, J = 7.3 Hz, 5.3 Hz, 1 H), 3.49 (d, J = 13.9 Hz, 1 H), 3.43 (d, J = 13.9 Hz, 1 H), 2.68–2.73 (m, 2 H), 2.08–2.12 (m, 2 H), 1.86–1.91 (m, 2 H), 1.63–1.69 (m, 2 H), 1.21–1.41 (m, 12 H), 1.16 (s, 3 H), 0.97 (s, 3 H), 0.87 (t, J = 7.1 Hz, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 65.2$, 52.9, 48.3, 47.7, 44.6, 38.5, 35.4, 32.8, 31.6, 28.97, 28.92, 26.9, 26.4, 24.4, 22.5, 20.8, 19.8, 14.0 ppm. $C_{18}H_{31}NO_{3}S$ (341.51): calcd. C 63.31, H 9.15, N 4.10; found C 63.40, H 9.28, N 4.07.

(2*S*)-*N*-(6-Bromohexanoyl)camphorsultam 9f: Compound 9f was obtained by crystallization from ethanol, 82% yield (7.56 g); $R_{\rm f} = 0.49$ (Cy/EA, 8:2); m.p. 54–55 °C. $[a]_{\rm D}^{20} = 87.8$ (c = 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 3.86$ (dd, J = 7.4 Hz, 5.3 Hz, 1 H), 3.50 (d, J = 13.6 Hz, 1 H), 3.43 (d, J = 13.6 Hz, 1 H), 3.40 (t, J = 6.8 Hz, 2 H), 2.71–2.75 (m, 2 H), 2.04–2.12 (m, 2 H), 1.85–1.92 (m, 5 H), 1.65–1.74 (m, 2 H), 1.33–1.53 (m, 4 H), 1.15 (s, 3 H), 0.97 (s, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 171.6$, 65.2, 52.9, 48.4, 47.7, 44.6, 38.5, 35.1, 33.5, 32.8, 32.4, 27.5, 26.4, 23.5, 20.8, 19.9 ppm. C₁₆H₂₆NO₄SBr (408.35): calcd. C 48.98, H 6.68, N 3.57; found C 48.93, H 6.75, N 3.57.

(2S)-N-[3-(N-Benzylindol-3-yl)propanoyl]camphorsultam 9g: Compound 9g was obtained from 3-(N-benzylindol-3-yl)propionic acid. A stirred solution of 3-(indol-3-yl)propionic acid (10 g, 52.9 mmol) in DMF (55 mL) was cooled to 0 °C and NaH (4.66 g, 116.4 mmol) was added in small portions. The mixture was stirred at 0 °C for 30 min and benzyl bromide (6.6 mL, 55.54 mmol) was added. After the mixture had been stirred for 1 h, the reaction was quenched with water, the mixture acidified to pH = 2, and extracted with diethyl ether. The combined organic layers were washed with saturated aqueous NH₄Cl, dried and concentrated in vacuo. Purification by flash column chromatography (Cy/EA/AcOH, 7:3:0.1) gave 3-(N-benzylindol-3-yl)propionic acid as a white solid, 75% yield (11 g); $R_{\rm f} = 0.25$ (Cy/EA, 8:2); m.p. 117 °C. ¹H NMR (400 MHz, CDCl₃): δ = 7.60 (d, J = 7.6 Hz, 1 H), 7.22–7.29 (m, 5 H), 7.17 (t, J = 7.1 Hz, 1 H), 7.11 (t, J = 7.6 Hz, 1 H), 7.07 (d, J = 7.1 Hz, 1 H), 6.93 (s, 1 H), 5.24 (s, 2 H), 3.71 (t, J = 7.6 Hz, 2 H), 2.76 (t, J = 7.6 Hz, 2 H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 179.5, 137.6, 136.7, 128.7, 127.7, 127.5, 126.7, 125.6, 121.9, 119.1, 118.8, 113.8, 109.7, 49.9, 34.8, 20.4 ppm. C₁₈H₁₇NO₂: calcd. C 77.40, H 6.13, N 5.01; found C 77.12, H 6.37, N 4.89. Compound 9g was obtained from this compound according to the general procedure and purified by flash column chromatography (Cy/EA, 8:2), white solid, 70% yield (4.7 g), $R_{\rm f} = 0.42$ (Cy/EA/NEt₃, 8:2:0.1), m.p. 123 °C. $[a]_D^{20} = 57.4 (c = 1, CHCl_3)$. ¹H NMR (400 MHz, CDCl₃): δ = 7.63–7.65 (m, 1 H), 7.20–7.28 (m, 4 H), 7.07–7.16 (m, 4 H), 6.97 (s, 1 H), 5.23 (s, 2 H), 3.85 (t, *J* = 6.3 Hz, 1 H), 3.45 (d, *J* = 13.9 Hz, 1 H), 3.42 (d, *J* = 13.9 Hz, 1 H), 3.06–3.19 (m, 4 H), 2.03– 2.05 (m, 2 H), 1.81–1.87 (m, 3 H), 1.29–1.39 (m, 2 H), 1.05 (s, 3 H), 0.92 (s, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 171.5, 137.7, 136.6, 128.7, 127.9, 127.4, 126.7, 125.8, 121.7, 119.1, 119.0, 113.8, 109.5, 65.2, 52.9, 49.8, 48.4, 47.7, 44.6, 38.5, 36.1, 32.8, 26.4, 20.7, 20.2, 19.8 ppm. C₂₈H₃₂N₂O₃S (476.63): calcd. C 70.56, H 6.77, N 5.88; found C 70.31, H 6.98, *N*, 5.67.

(2*S*)-*N*-{(*2R*)-2-[(Dibenzylamino)methyl]propanoyl}camphorsultam 11a: Crystallization from CHCl₃/MeOH, 91% yield (16.7 g); $R_f =$ 0.39 (Cy/EA, 8:2); m.p. 143 °C. $[a]_{D}^{2D} = 50.2$ (c = 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.2$ -7.3 (m, 10 H), 3.9 (dd, J =7.6 Hz, 5.1 Hz, 1 H), 3.6 (s, 4 H), 3.4–3.5 (m, 3 H), 2.9 (dd, J =7.6 Hz, 12.6 Hz, 1 H), 2.4 (dd, J = 6.3 Hz, 12.6 Hz, 1 H), 2.0–2.1 (m, 1 H), 2.0 (dd, J = 7.6 Hz, 13.9 Hz, 1 H), 2.02–2.11 (m, 2 H), 1.86–1.91 (m, 3 H), 1.3–1.4 (m, 2 H), 1.2 (s, 3 H), 1.1 (s, 3 H), 1.0 (s, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 175.3$, 138.9, 128.0, 126.8, 65.4, 57.9, 57.8, 53.1, 48.2, 47.7, 44.7, 38.6, 38.4, 32.9, 26.4, 21.0, 19.9, 15.5 ppm. C₂₈H₃₆N₂O₃S (480.66): calcd. C 69.96, H 7.55, N 5.83; found C 69.85, H 7.67, N 5.92.

(2*R*)-*N*-{(2*S*)-2-[(Dibenzylamino)methyl]-3-methylbutanoyl}camphorsultam 11b: Purification by flash column chromatography (Cy/EA, 92:8), 81% yield (8.3 g); $R_f = 0.45$ (Cy/EA, 8:2); m.p. 96 °C. [a]₂₀²⁰ = -41.4 (c = 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.19-7.31$ (m, 10 H), 3.88 (dd, J = 5.0 Hz, 7.6 Hz, 1 H), 3.64 (d, J = 13.6 Hz, 1 H), 3.51 (d, J = 13.6 Hz, 1 H), 3.49 (d, J = 13.9 Hz, 1 H), 3.43 (d, J = 13.6 Hz, 1 H), 3.27–3.29 (m, 1 H), 2.93 (dd, J = 9.1 Hz, 12.6 Hz, 1 H), 2.59 (dd, J = 3.8 Hz, 12.9 Hz, 1 H), 2.16–2.20 (m, 1 H), 1.98–2.08 (m, 2 H), 1.87–1.94 (m, 3 H), 1.31– 1.41 (m, 2 H), 1.25 (s, 3 H), 0.97 (s, 3 H), 0.91 (d, J = 7.1 Hz, 3 H), 0.88 (d, J = 7.1 Hz, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 174.3$, 138.9, 129.3, 128.0, 126.8, 65.7, 57.8, 53.4, 53.3, 48.5, 48.0, 47.7, 44.7, 38.7, 32.9, 29.1, 26.5, 21.1, 20.9, 19.9, 18.5 ppm. C₃₀H₄₀N₂O₃S (508.72): calcd. C 70.83, H 7.93, N 5.51; found C 70.22, H 7.99, N 5.45. MS (ESI⁺): m/z = 509 [MH⁺].

(2*S*)-*N*-{(2*R*)-2-[(Dibenzylamino)methyl]-4-methylpentanoyl}camphorsultam 11c: Purification by flash column chromatography (Cy/EA, 9:1), 82% yield (2.7 g); analytical data are identical to those of compound 7a obtained by the Reformatsky reaction.

(2*S*)-*N*-{(2*R*)-2-[(Dibenzylamino)methyl]-3-phenylpropanoyl}camphorsultam 11d: Purification by flash column chromatography (Cy/EA, 9:1), 76% yield (9.3 g); $R_f = 0.41$ (Cy/EA, 8:2); m.p. 59 °C. [*a*]₂₀²⁰ = 45.6 (*c* = 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ = 7.16– 7.26 (m, 15 H), 3.86 (dd, *J* = 7.6 Hz, 5.1 Hz, 1 H), 3.73 (m, 1 H), 3.41–3.54 (m, 6 H), 3.04 (dd, *J* = 13.6 Hz, 5.6 Hz, 1 H), 2.92 (dd, *J* = 12.6 Hz, 8.1 Hz, 1 H), 2.65 (dd, *J* = 13.6 Hz, 8.6 Hz, 1 H), 2.49 (dd, *J* = 12.6 Hz, 5.3 Hz, 1 H), 2.02–2.11 (m, 2 H), 1.87–1.91 (m, 3 H), 1.31–1.38 (m, 2 H), 1.24 (s, 3 H), 0.98 (s, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 174.1, 139.3, 138.7, 129.4, 129.1, 128.3, 128.0, 126.8, 126.2, 65.4, 57.7, 55.7, 53.2, 48.3, 47.8, 44.9, 44.8, 38.6, 36.0, 32.9, 26.5, 20.1, 19.9 ppm. C₃₄H₄₀N₂O₃S (556.76): calcd. C 73.35, H 7.24, N 5.03; found C 73.01, H 7.31, N 4.90.

(2*S*)-*N*-{(*2R*)-2-[(Dibenzylamino)methyl]octanoyl}camphorsultam 11e: Purification by flash column chromatography (Cy/EA, 95:5), 74% yield (12.98 g); $R_f = 0.5$ (Cy/EA, 8:2); m.p. 60 °C. $[a]_D^{20} = 13.7$ (c = 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.21-7.32$ (m, 10 H), 3.88 (dd, J = 7.6 Hz, 5.3 Hz, 1 H), 3.65 (d, J = 13.6 Hz, 1 H), 3.51 (d, J = 13.6 Hz, 1 H), 3.50 (d, J = 13.9 Hz, 1 H), 3.43 (d, J = 13.9 Hz, 1 H), 3.34–3.38 (m, 1 H), 2.80 (dd, J = 12.6 Hz, 7.1 Hz, 1 H), 2.52 (dd, J = 12.6 Hz, 6.8 Hz, 1 H), 2.01–2.12 (m, 2 H), 1.86–1.93 (m, 3 H), 1.55 (m, 2 H), 1.27–1.42 (m, 2 H), 1.22 (m, 11 H), 0.97 (s, 3 H), 0.86 (t, J = 7.1 Hz, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 174.8$, 139.0, 129.1, 128.0, 126.8, 65.4, 58.0, 56.4, 53.3, 48.2, 47.8, 44.7, 44.2, 38.6, 38.6, 32.9, 31.6, 30.1, 29.4, 27.2, 26.5, 21.0, 19.9, 14.1 ppm. C₃₃H₄₆N₂O₃S (550.79): calcd. C 71.96, H 8.42, N 5.09; found C 71.67, H 8.62, N 4.94.

(2*S*)-*N*-{(2*R*)-6-Bromo-2-[(dibenzylamino)methyl]hexanoyl}camphorsultam 11f: Purification by flash column chromatography (Cy/EA, 9:1), 76% yield (14.6 g); $R_{\rm f} = 0.47$ (Cy/EA, 8:2); m.p. 90 °C. [a]_D²⁰ = 57.3 (c = 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ = 7.20–7.40 (m, 10 H), 3.87 (dd, J = 7.3 Hz, 5.3 Hz, 1 H), 3.68 (d, J = 13.6 Hz, 2 H), 3.51 (d, J = 13.9 Hz, 1 H), 3.47 (d, J = 13.6 Hz, 2 H), 3.43 (d, J = 13.9 Hz, 1 H), 3.31–3.37 (m, 3 H), 2.77 (dd, J = 12.6 Hz, 6.6 Hz, 1 H), 2.52 (dd, J = 12.6 Hz, 7.3 Hz, 1 H), 2.03–2.07 (m, 2 H), 1.86–1.90 (m, 3 H), 1.77–1.81 (m, 2 H), 1.56– 1.60 (m, 2 H), 1.33–1.42 (m, 4 H), 1.21 (s, 3 H), 0.97 (s, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 174.4, 139.0, 129.0, 128.1, 126.9, 65.4, 58.2, 56.3, 53.2, 48.2, 47.7, 44.6, 44.0, 38.5, 33.5, 32.9, 32.8, 28.9, 26.4, 25.7, 21.0, 19.9 ppm. MS (ESI⁺): m/z = 603 [M + H]⁺.

(2*S*)-*N*-{(*2R*)-3-(*N*-Benzylindol-3-yl)-2-[(dibenzylamino)methyl]propanoyl}camphorsultam 11g: Purification by flash column chromatography (Cy/EA, 9:1), 88% yield (5.8 g); $R_{\rm f} = 0.35$ (Cy/ EA, 8:2); m.p. 80–81 °C. $[a]_{120}^{2D} = 25.0$ (c = 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.78-7.80$ (m, 1 H), 7.09–7.34 (m, 16 H), 6.99–7.01 (m, 2 H), 6.85 (s, 1 H), 5.20 (s, 2 H), 3.88 (m, 2 H), 3.42– 3.52 (m, 6 H), 3.21 (dd, J = 14.4 Hz, 3.8 Hz, 1 H), 2.94 (dd, J =12.4 Hz, 10.3 Hz, 1 H), 2.86 (dd, J = 14.4 Hz, 9.1 Hz, 1 H), 2.56 (dd, J = 12.6 Hz, 4.5 Hz, 1 H), 2.06–2.15 (m, 2 H), 1.85–1.87 (m, 3 H), 1.30–1.37 (m, 2 H), 1.23 (s, 3 H), 0.96 (s, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 174.7$, 138.7, 137.9, 136.6, 129.1, 128.6, 128.4, 127.9, 127.0, 126.7, 126.6, 121.6, 119.6, 112.4, 109.4, 65.4, 57.5, 55.8, 53.2, 49.8, 48.3, 47.7, 44.7, 38.6, 32.8, 26.5, 25.8, 21.0, 19.9 ppm. C₄₃H₄₇N₃O₃S (685.92): calcd. C 75.29, H 6.91, N 6.13; found C 75.45, H 7.20, N 5.81.

(*R*)-2-{[(*tert*-Butoxycarbonyl)amino]methyl}propionic Acid (12a): 68% yield (4.6 g), $R_{\rm f} = 0.20$ (CH₂Cl₂/MeOH/AcOH, 99:1:0.1); m.p. 89–90 °C. $[a]_{\rm D}^{20} = -17.7$ (*c* = 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 10.6$ (br. s, 1 H), 6.43 and 5.08 (br. s, 1 H), 3.21–3.39 (m, 2 H), 2.65–2.74 (m, 1 H), 1.47 and 1.44 (s, 9 H), 1.2 (d, *J* = 7.5 Hz, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 180.9$, 179.6, 157.8, 156.1, 81.1, 79.6, 44.1, 42.7, 40.3, 40.0, 28.4, 14.6 ppm. C₉H₁₇NO₄ (203.24): calcd. C 53.19, H 8.43, N 6.89; found C 53.41, H 8.42, N 6.85.

(*S*)-2-{[(*tert*-Butoxycarbonyl)amino]methyl}-4-methylbutanoic Acid (12b): 59% yield (1.7 g), $R_{\rm f} = 0.30$ (Cy/EA/AcOH, 7:3:0.1); m.p. 78 °C. $[a]_{\rm D}^{20} = 6.9$ (c = 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ = 6.89 and 5.05 (br. s, 1 H), 3.37–3.47 (m, 1 H), 3.20–3.27 (m, 0.3 H), 3.06–3.17 (m, 0.6 H), 2.42–2.49 (m, 1 H), 1.87–2.03 (m, 1 H), 1.48 and 1.44 (s, 9 H), 0.97 and 0.99 (d, J = 6.6 Hz, 6 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 178.2$, 158.3, 81.1, 52.7, 52.0, 40.6, 39.5, 28.9, 28.4, 26.4, 20.4, 20.3, 19.8 ppm. C₁₀H₁₉NO₄ (217.26): calcd. C 57.12, H 9.15, N 6.06; found C 56.98, H 9.29, N 5.98.

(*R*)-2-{[(*tert*-Butoxycarbonyl)amino]methyl}-4-methylpentanoic Acid (12c): 80% yield (2.3 g), $R_{\rm f} = 0.21$ (Cy/EA/AcOH, 8:2:0.1); m.p. 74 °C. [*a*]_D²⁰ = -3.1 (*c* = 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 6.76$ and 5.02 (br. s, 1 H), 3.32–3.41 (m, 1 H), 3.19–3.26 and 3.00–3.07 (m, 1 H), 2.68–2.70 (m, 1 H), 1.64–1.72 (m, 1 H), 1.52– 1.60 (m, 1 H), 1.44–1.48 (s, 9 H), 1.19–1.35 (m, 3 H), 0.93 (d, *J* = 6.1 Hz, 3 H), 0.92 (d, *J* = 6.6 Hz, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 180.8$, 179.4, 155.9, 158.1, 79.6, 81.1, 43.7, 44.1, 41.7, 43.0, 38.6, 39.0, 28.4, 25.8, 22.4 ppm. C₁₂H₂₃NO₄ (245.32): calcd. C 58.75, H 9.45, N 5.71; found C 58.84, H 9.59, N 5.61. (*R*)-2-{[(*tert*-Butoxycarbonyl)amino]methyl}-3-phenylpropionic acid (12d): 73 % yield (1.6 g); $R_{\rm f} = 0.21$ (Cy/EA/AcOH, 8:2:0.1); m.p. 74 °C. $[a]_{\rm D}^{20} = 16.9$ (c = 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): $\delta = 9.65$ (br. s, 1 H), 7.18–7.30 (m, 5 H), 6.67 and 5.01 (br. s, 1 H), 3.37–3.41 (m, 0.5 H), 3.22–3.27 (m, 1 H), 2.80–3.11 (m, 3 H), 2.63– 2.72 (m, 0.5 H), 1.37–1.42 (s, 9 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 179.01$, 179.28, 155.91, 157.98, 138.19, 138.06, 128.91, 128.77, 126.62, 79.73, 81.19, 47.17, 47.53, 41.25, 42.01, 35.66, 35.82 ppm. C₁₅H₂₁NO₄ (279.33): calcd. C 64.50, H 7.58, N 5.01; found C 64.19, H 7.62, N 4.90.

(R)-2-{[N-(tert-Butoxycarbonyl)amino]methyl}octanoic Acid (12e): Purification by flash column chromatography (Cy/EA/AcOH, 85:15:1), 44% yield (0.87 g). ¹H NMR (400 MHz, CDCl₃): δ = 6.63 and 5.01 (br. s, 1 H), 3.34-3.40 (m, 2 H), 3.23-3.28 (m, 2 H), 3.10-3.12 (m, 2 H), 2.61 (m, 1 H), 1.27–1.64 (m, 19 H), 0.88 (t, J =6.8 Hz, 6 H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 180.7, 180.0, 158.2, 156.1, 81.3, 79.7, 45.8, 42.8, 41.5, 31.7, 29.8, 29.3, 28.5, 27.1, 22.7, 14.2 ppm. The amino acid was crystallized as its dicyclohexylamine (DCHA) salt from diethyl ether (700 mg, 24%); m.p. 103 °C. $[a]_{D}^{20} = -3.73$ (c = 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ = 8.10 (br. s, 2 H), 5.51 and 5.20 (br. s, 1 H), 3.29–3.35 (m, 1 H), 3.11-3.18 (m, 1 H), 2.89-2.96 (m, 2 H), 2.62-2.31 (m, 1 H), 1.99-2.02 (m, 4 H), 1.78-1.81 (m, 4 H), 1.58-1.66 (m, 3 H), 1.12-1.42 (m, 28 H), 0.86 (t, J = 6.8 Hz, 3 H) ppm. ¹³C NMR (100 MHz, $CDCl_3$): $\delta = 180.02, 156.1, 78.5, 52.5, 47.6, 42.5, 31.8, 30.8, 29.6,$ 28.5, 27.7, 25.3, 24.8, 22.6, 14.1 ppm. C₂₆H₅₀N₂O₄ (454.69): calcd. C 68.68, H 11.48, N 6.13; found C 68.44, H 11.63, N 6.05.)

Acknowledgments

This work is part of the PhD Thesis of R. M., who is grateful to Senn Chemicals for the financial support.

- [1] For a review, see: D. Seebach, A. K. Beck, D. J. Bierbaum, *Chem. Biodiversity* **2004**, *1*, 1111.
- [2] For recent studies, see: E. P. English, R. S. Chumanov, S. H. Gellman, T. Compton, J. Biol. Chem. 2006, 281, 2661; J. A. Kritzer, O. M. Stephens, D. A. Guarracino, S. K. Reznik, A. Schepartz, Bioorg. Med. Chem. 2005, 13, 11; O. M. Stephens, S. Kim, B. D. Welch, M. E. Hodsdon, M. S. Kay, A. Schepartz, J. Am. Chem. Soc. 2005, 127, 13126; J. X. Qiu, E. J. Petersson, E. E. Matthews, A. Schepartz, J. Am. Chem. Soc. 2006, 128, 11338.
- [3] E. P. Ellmerer-Müller, D. Brössner, N. Maslouh, A. Tako, *Helv. Chim. Acta* 1998, 81, 59.
- [4] D. Seebach, K. Gademann, J. V. Schreiber, J. L. Matthews, T. Hintermann, B. Jaun, L. Oberer, U. Hommel, H. Widmer, *Helv. Chim. Acta* 1997, 80, 2033; D. Seebach, S. Abele, K. Gademann, G. Guichard, T. Hintermann, B. Jaun, J. L. Matthews, J. V. Schreiber, *Helv. Chim. Acta* 1998, 81, 932; M. Rueping, J. V. Schreiber, G. Lelais, B. Jaun, D. Seebach, *Helv. Chim. Acta* 2002, 85, 2577.
- [5] D. Seebach, S. Abele, K. Gademann, B. Jaun, Angew. Chem. Int. Ed. 1999, 38, 1595; Angew. Chem. 1999, 111, 1700; X.

Daura, K. Gademann, H. Schäfer, B. Jaun, D. Seebach, W. F. Van Gunsteren, J. Am. Chem. Soc. 2001, 123, 2393.

- [6] T. Hintermann, D. Seebach, *Helv. Chim. Acta* 1998, 81, 2093;
 R. Sebesta, D. Seebach, *Helv. Chim. Acta* 2003, 86, 4061; D. Seebach, K. Namoto, Y. R. Mahajan, P. Bindschädler, *Chem. Biodiversity* 2004, 1, 65 and references cited therein.
- [7] E. Juaristi, V. A. Soloshonok in *Enantioselective Synthesis of Beta-Amino Acids*, 2nd ed., John Wiley & Sons, New York, 2005; G. Lelais, D. Seebach, *Biopolymers (Peptides Science)* 2004, 76, 206.
- [8] Y. Chi, S. H. Gellman, J. Am. Chem. Soc. 2006, 128, 6804.
- [9] a) N-(Chloromethyl)benzamide, see: D. A. Evans, F. Urpi, T. C. Somers, J. S. Clark, M. T. Bilodeau, J. Am. Chem. Soc. 1990, 112, 8215; b) 1-[(benzoylamino)methyl]benzotriazole, see A. R. Katritzky, X. Lan, W.-Q. Fan, Synthesis 1994, 445; c) N-(bromomethyl)phthalimide, see: M. Calmes, F. Escale, C. Glot, M. Rolland, J. Martinez, Eur. J. Org. Chem. 2000, 2459.
- [10] P. Karoyan, S. Lavielle, R. Moumne, H. Rudler, B. Denise, Patent No. 0508890, August 30th, **2005**; R. Moumne, S. Lavielle, P. Karoyan, *J. Org. Chem.* **2006**, *71*, 3332.
- [11] A. Fürstner, Synthesis 1989, 571; R. Ocampo, W. R. Dolbier Jr, Tetrahedron 2004, 60, 9325.
- [12] D. J. Hart, H. Deok-Chang, Chem. Rev. 1989, 89, 1447.
- [13] N. Millot, C. Piazza, S. Avolio, P. Knochel, *Synthesis* 2000, 7, 941.
- [14] a) N. Izumiya, A. Nagamtsu, *Bull. Chem. Soc. Jpn.* 1952, 52, 265; b) A. J. Souers, S. Schürer, H. Kwack, A. A. Virgilio, J. A. Ellman, *Synthesis* 1999, 4, 583; c) I. Shin, M. Lee, J. Lee, M. Jung, W. Lee, J. Yoon, *J. Org. Chem.* 2000, 65, 7667.
- [15] See Experimental Section.
- [16] F. Dardoize, J.-L. Moreau, M. Gaudemar, Bull. Soc. Chim. Fr. 1973, 5, 1668 and references cited therein.
- [17] M. K. Anwer, A. F. Spatola, Synthesis 1980, 929.
- [18] V. Farina, J. T. Reeves, C. Senanayake, J. J. Song, *Chem. Rev.* 2006, 106, 2734.
- [19] R. Ponsinet, G. Chassaing, J. Vaissermann, S. Lavielle, Eur. J. Org. Chem. 2000, 83.
- [20] A. Böhm, D. Seebach, Helv. Chim. Acta 2000, 83, 3262.
- [21] W. Oppolzer, R. Moretti, S. Thomi, *Tetrahedron Lett.* **1989**, *30*, 6009.
- [22] R. Moumne, Ph.D. Thesis, University Paris 6, France, 2005.
- [23] M. Arend, B. Westermann, N. Risch, Angew. Chem. Int. Ed. 1998, 37, 1044; Angew. Chem. 1998, 110, 1096.
- [24] W. Oppolzer, O. Tamura, J. Deerberg, *Helv. Chim. Acta* 1992, 75, 1965.
- [25] W. Oppolzer, C. Starkemann, I. Rodriguez, G. Bernardinelli, *Tetrahedron Lett.* 1991, 32, 61.
- [26] C. A. Archer, N. R. Thomas, D. Giani, *Tetrahedron: Asym*metry **1993**, 4, 1141.
- [27] C. Campion, A. H. Davidson, J. P. Dickens, M. J. Crimmin, WO9005716, 1990.
- [28] F. S. Arno, A. L. Bettag, J. Org. Chem. 1981, 46, 2393.
- [29] D. N. Harpp, L. Q. Bao, C. J. Balck, J. Q. Gleason, R. A. Smith, J. Org. Chem. 1975, 40, 3420.
- [30] B. R. Ward, A. Pelter, D. Goubet, M. C. Pritchard, *Tetrahe*dron: Asymmetry 1995, 6, 469.
- [31] W. Oppolzer, O. Tamura, J. Deerberg, *Helv. Chim. Acta* 1992, 75, 1965.

Received: October 23, 2006 Published Online: February 27, 2007