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5-Substituted 4-anilinoquinazolines as potent, selective and orally active inhibitors of erbB2 receptor tyrosine kinase

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Abstract—Starting from a 6,7-substituted quinazoline lead 4, optimisation of 5-substituted quinazolines containing an extended aniline motif led to potent and selective inhibitors of erbB2 receptor tyrosine kinase, and a representative compound 12a inhibited tumour growth in a mouse xenograft model. © 2005 Elsevier Ltd. All rights reserved.

Following the discovery that over-expression of the EGFR (erbB1) and erbB2 receptor tyrosine kinases is found in a number of cancers and is associated with poor prognosis in patients, blockade of this signalling pathway has emerged as a promising approach to selective targeting of tumour cells.¹ The clinical utility of the EGFR inhibitor gefitinib 1 (Fig. 1) in non-small cell lung cancer has been demonstrated,² while the monoclonal antibody trastuzumab has been shown to increase survival time in patients with metastatic breast tumours that over-express erbB2.³ Among compounds currently undergoing clinical trials are the mixed EGFR-erbB2 receptor tyrosine kinase inhibitor lapatinib (GW2016) 2^{4} , the erbB2-selective inhibitor CP-724714 3^{5} and the irreversible inhibitors canertinib (CI-1033),⁶ EKB-569⁷ and HKI-272.8

X-ray crystal structures of EGFR enzyme-inhibitor complexes provide clear evidence that the mechanism of action of receptor tyrosine kinase inhibitors such as 1-3 involves reversible displacement of ATP at the kinase active site, through binding to either the active⁹ or the inactive¹⁰ form of the kinase. As part of the programme of work that led to the discovery of gefitinib, we found 6,7-substituted quinazoline 4^{11} (Fig. 1) to be a

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Figure 1. Screening lead **4** and examples of EGFR and erbB2 receptor tyrosine kinase inhibitors in clinical use or undergoing trials: gefitinib **1**; lapatinib **2**; CP-724714 **3**.

moderately potent inhibitor of erbB2 receptor tyrosine kinase. Like other documented quinazoline-based erbB2 inhibitors, compound **4** contains an extended aniline motif, which we assumed to interact with the selectivity pocket at the erbB2 active site.¹⁰ From consideration of an in-house homology model¹² and related work on inhibitors of c-Src,¹³ we reasoned that switching of the quinazoline substituent from the 6- to the 5-position could improve erbB2 affinity through occupation of the kinase sugar pocket. In this publication, we describe

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how this work led to potent, selective and orally active inhibitors of erbB2.

Representative compounds prepared during the course of this work are listed in Table 1, and synthetic routes are outlined in Scheme 1–4.¹⁴ Initial variant 7 was prepared (Scheme 1) from the appropriate extended aniline¹¹ and chloroquinazoline **6**, in turn obtained from 5,7-dimethoxy-4(3*H*)-quinazolone **5**^{14b} via a sequence involving as the key steps selective demethylation,¹³ protection of quinazolone nitrogen and Mitsunobu alkylation.

Compounds **10a**–c were likewise prepared (Scheme 2) from the appropriate anilines^{11,15} and the corresponding chloroquinazolines **9a–b**, readily accessible in two steps from commercially available 5-fluoro-4(3*H*)-quinazolone **8** via alkoxide displacement and chlorination.

More versatile routes enabled introduction of the aniline para substituent or the quinazoline 5-substituent at the last stage of the synthesis. Reaction of chloroquinazoline 9a with the appropriate amino phenol gave intermediates 11a-b, which were then alkylated to provide

Table 1. erbB2 and EGFR kinase inhibition data¹⁸ for compounds 4, 7, 10a-c, 12a-d, 14

Entry	$IC_{50} (\mu M)^a$			
	erbB2	EGFR		
4	0.056 (±0.003)	0.059 (±0.003)		
7	0.005 (±0)	0.005 (±0.003)		
10a	< 0.002	0.005 (±0.002)		
10b	1.1 (±0.085)	1.8 (±0.25)		
10c	0.024 (±0.001)	2.4 (±0.41)		
12a	<0.002 ^b	0.14 (±0.048)		
12b	< 0.002	0.69 (±0.18)		
12c	< 0.002	$0.28(\pm 0.08)$		
12d	0.070 (±0.014)	2.1 (±0.11)		
14	0.024 (±0.010)	0.44 (±0.026)		

^a $n \ge 3$, standard error is given in parentheses.

^b IC₅₀ 7.6 μM (Kdr), 23 μM (Src).



Scheme 1. Synthesis of compound 7. Reagents and conditions: (a) MgBr₂, pyridine, reflux; (b) NaH, chloromethyl pivalate, DMF, 0–20 °C; (c) 4-hydroxy-*N*-methylpiperidine, Ph₃P, (t-BuO₂C)₂N₂, CH₂Cl₂, 20 °C; (d) NH₃, MeOH, 20 °C; (e) POCl₃, *i*-Pr₂NEt, ClCH₂CH₂Cl, reflux; (f) H₂NAr,¹¹ *i*-Pr₂NEt, *i*-PrOH, reflux.



Scheme 2. Synthesis of compounds 10a–c. Reagents and conditions: (a) NaH, 4-hydroxy-*N*-methylpiperidine or 4-hydroxytetrahydropyran, MeCONMe₂, 80 °C (work up: DOWEX H+/MeOH/NH₃, 20 °C); (b) POCl₃, *i*-Pr₂NEt, CH₂Cl₂, reflux (work up: aq NaHCO₃, 0 °C); (c) H₂NAr,^{11,15} *i*-PrOH, reflux.



Scheme 3. Synthesis of compounds 12a–d. Reagents and conditions: (a) 4-amino-2-chlorophenol or 4-aminophenol, *i*-PrOH, reflux; (b) $ClCH_2Y$,¹⁶ K₂CO₃, 18-crown-6, MeCN, reflux.



Scheme 4. Synthesis of compound 14. Reagents and conditions: (a) SOCl₂, cat HCONMe₂, reflux (work up: aq NaHCO₃, 0 °C);¹⁷ (b) H₂NAr,¹¹ *i*-PrOH, reflux; (c) NaOMe, 15-crown-5, MeOH, dioxan, reflux.

extended aniline variants 12a-d (Scheme 3). Alternatively (Scheme 4) the fluoroquinazoline 13, readily obtained from fluoroquinazolone 8,¹⁷ underwent microwave assisted displacement by alkoxide to give 5-methoxy quinazoline derivative 14.

The compounds listed in Table 1 were evaluated in erbB2 and EGFR kinase assays measuring inhibition of phosphorylation of a synthetic peptide substrate at $K_{\rm m}$ ATP concentration for each enzyme.¹⁸ Lead compound **4** showed moderate inhibition in the erbB2 kinase assay (IC₅₀ 0.056 μ M). Transposition of the quinazoline substituent from the 6- to the 5-position, introduction of a cyclic amine and changing the meta substituent in the extended aniline from methyl to chlorine led to

compound 7 with significantly improved affinity for erbB2 and EGFR. Removal of the 7-methoxy substituent then provided compound **12a** with favourable erbB2 kinase selectivity versus EGFR (IC₅₀ 0.002 μ M vs 0.11 μ M). Compound **12a** also showed a clean selectivity profile versus in-house and external kinase panels (data not shown).¹⁹ Thus, for example, **12a** gave selectivity ratios of >3800 and >11,500, respectively, versus the receptor tyrosine kinases Kdr and Src (Table 1, footnote b).

The remaining compounds included in Table 1 illustrate SAR for the series of 5-substituted quinazolines. Alternative extended anilines (**10a**, **12b–c**) gave comparable ErbB2 activity but differing selectivity versus EGFR. More drastic aniline modifications (**10b**, **12d**) gave reduced affinity, consistent with interaction of the extended aniline with the selectivity pocket at the erbB2 active site¹⁰ and with published work highlighting the importance of the meta substituent.²⁰ Alternative 5-substituted quinazolines (**10c**, **14**) also showed reduced affinity relative to **12a**.

A number of assays^{18,21-24} were used to determine the cellular profile of selected compounds (Table 2). ErbB2-selective inhibitor 12a and mixed erbB2-EGFR inhibitor 10a were shown to inhibit erbB2 autophosphorylation in a MCF7 breast carcinoma cell line engineered to overexpress erbB2 ('Clone 24' assay).²¹ The cellular activity of compounds 10a and 12a was confirmed in a proliferation assay using the BT474C cell line.^{22,23} Compounds **10a** and **12a** were shown to be only weak to moderate inhibitors of growth of the KB cell line when stimulated with EGF,¹⁸ and good separation was also seen between BT474C antiproliferative activity and effect on basal growth in KB cells,¹⁸ consistent with a cellular mode of action involving inhibition of erbB2. The moderate EGFR cellular potency of compound 10a was confirmed using an EGFR autophosphorylation assay,²⁴ reflecting contrasting enzyme and cellular profiles for this compound.

Table 3 summarises pharmacokinetic parameters for compound **12a** determined following dosing to immunocompetent Alderley Park mice at 2 mg/kg iv and 5 mg/kg po. Exposure of the compound scaled well on

Table 2. Cell assay data^{18,21-23} for compounds 10a and 12a

Entry	$IC_{50} (\mu M)^a$					
	Clone 24	BT474C	KB	KB Basal		
10a 12a	0.056 (±0.007) 0.061 (±0.009)	0.027 (±0.007) 0.041 (±0.004)	0.46 ^b (±0.10) 2.6 (±0.81)	1.7 (±0.46) 11.9 (±4.1)		

^a $n \ge 3$, standard error is given in parentheses.

 $^{b}\,IC_{50}$ 2.5 (±0.75) μM for inhibition of phospho-EGFR in KB cells. 24

Table 3. Pharmacokinetic parameters for compound 12a, determined by dosing to immunocompetent mice at 2 mg/kg iv and 5 mg/kg po

Entry	V _{dss} (l/kg)	Cl (ml/min/kg)	po AUC (µM.h)	Bioavailability %
12a	2.6	15	5.5	50

dosing at 100 mg/kg po, and calculation of plasma free concentrations showed the free levels of **12a** to compare favourably with the IC₅₀ value in the Clone 24 phosphorylation assay (Table 4).²⁵

Encouraged by these pharmacokinetic data, we evaluated compound **12a** at 100 mg/kg po q.d. and b.i.d. and at 200 mg/kg po q.d. for inhibition of growth of a BT474C tumour xenograft in a 28-day study in athymic mice.²² No significant reduction in body weight was observed for any animal group in this study. As shown in Figure 2, compound **12a** gave statistically significant inhibition of tumour growth with all dosing schedules. Analysis of excised tumour samples confirmed significant inhibition of phospho-erbB2 (e.g., >90% inhibition 4 and 24 h after the last dose at 100 mg/kg b.i.d.).

Although the BT474C mouse xenograft model responds to both erbB2²² and EGFR inhibitions,²⁶ the weak activity of compound **12a** in the KB cell assay (Table 2) is consistent with xenograft activity reflecting inhibition of erbB2. Further support for this hypothesis is provided by the lack of activity of **12a** at 100 mg/kg po in the EGFR-driven mouse LoVo xenograft model¹⁸ (data not shown).

In summary, optimisation of 5-substituted quinazolines containing an extended aniline motif led to potent and selective inhibitors of erbB2 receptor tyrosine kinase, and a representative compound **12a** inhibited tumour growth in the mouse BT474C xenograft model. Further work describing a wider range of 5-substituted quinazolines as selective inhibitors of erbB2 will be reported in due course.

Table 4. Plasma concentrations for compound **12a** after dosing at 100 mg/kg po,^a and ratios of free plasma levels^b to IC_{50} value in Clone 24 cellular assay

	Time (h)				
	2	4	6	16	24
Plasma concentration 12a (μ M) Ratio free 12a /Clone 24 IC ₅₀		7.2 3.2	6.9 3.1	2.2 0.97	1.1 0.48

^a AUC 98.5 μM.h.

^b Mouse free drug 2.72%.



Figure 2. Inhibition of growth of BT474C xenograft²² in athymic mice dosed po with compound 12a.

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References and notes

- Gross, M. E.; Shazer, R. L.; Agus, D. B. Semin. Oncol. 2004, 31(Suppl 3), 9.
- 2. Herbst, R. S.; Fakuoka, M.; Baselga, J. *Nature Rev. Cancer* 2004, *4*, 956.
- Cobleigh, M. A.; Charles, L.; Vogel, C. L.; Tripathy, D.; Robert, N. J.; Scholl, S.; Fehrenbacher, L.; Wolter, J. M.; Paton, V.; Shak, S.; Lieberman, G.; Slamon, D. J. J. Clin. Oncol. 1999, 17, 2639.
- Wenle, X.; Mullin, R. J.; Keith, B. R.; Liu, L. H.; Ma, H.; Rusnak, D. W.; Owens, G.; Alligood, K. J.; Spector, N. L. Oncogene 2002, 21, 6255.
- Kath, J. C.; Atherton, J.; Barbacci-Tobin, G.; Bhattacharya, S. K.; Boos, C. A.; Boscoe, B. P.; Campbell, M.; Coleman, K. G.; Cox, E. D.; Currier, N. V.; Emerson, E. O.; Gerdin, K.; Goodwin, P.; Harriman, S.; Jani, J. P.; Kwan, T. A.; Liu, Z.; Mairs, E. N.; Mathiowetz, A. M.; Miller, P. E.; Morris, J.; Moyer, J. D.; Pustilnik, L. R.; Rafidi, K.; Richter, D. T.; Rouch, A.; Soderstrom, E. A.; Thompson, C. B.; Tom, N. J.; Wessel, M. D.; Winter, S. M.; Xiao, J.; Zhao, X.; Iwata, K. K.. Abstracts of Papers In 226th National Meeting of the American Chemical Society. New York, NY, September 7–11, 2003; American Chemical Society: Washington, DC, 2003.
- Smaill, J. B.; Showalter, H. D. H.; Zhou, H.; Bridges, A. J.; McNamara, D. J.; Fry, D. W.; Nelson, J. M.; Sherwood, V.; Vincent, P. W.; Roberts, B. J.; Elliott, W. L.; Denny, W. A. J. Med. Chem. 2001, 44, 429.
- Wissner, A.; Overbeek, E.; Reich, M. F.; Floyd, M. B.; Johnson, B. D.; Mamuya, N.; Rosfjord, E. C.; Discafani, C.; Davis, R.; Shi, X.; Rabindran, S. K.; Gruber, B. C.; Ye, F.; Hallett, W. A.; Nilakantan, R.; Shen, R.; Wang, Y. F.; Greenberger, L. M.; Tsou, H. R. J. Med. Chem. 2003, 46, 49.
- Tsou, H. R.; Overbeek-Klumpers, E. G.; Hallett, W. A.; Reich, M. F.; Floyd, M. B.; Johnson, B. D.; Michalak, R. S.; Nilakantan, R.; Discafani, C.; Golas, J.; Rabindran, S. K.; Shen, R.; Shi, X.; Wang, Y. F.; Upeslacis, J.; Wissner, A. J. Med. Chem. 2005, 48, 1107.
- Stamos, J.; Sliwkowski, M. X.; Eigenbrot, C. J. Biol. Chem. 2002, 277, 46265.
- Wood, E. R.; Truesdale, A. T.; McDonald, O. B.; Yuan, D.; Hassell, A.; Dickerson, S. H.; Ellis, B.; Pennisi, C.; Horne, E.; Lackey, K.; Alligood, K. J.; Rusnak, D. W.; Gilmer, T. M.; Shewchuk, L. *Cancer Res.* 2004, 64, 6652.
- Brown, D. S.; Morris, J. J.; Thomas, A.P. PCT Int. Appl. WO 9615118, 1996.
- See also: Schewchuck, L.; Hassell, A.; Wisely, B.; Rocque, W.; Holmes, W.; Veal, J.; Kuyper, L. F. *J. Med. Chem.* 2000, 43, 133.

- Olivier, A.; Costello, G. F.; Fennell, M.; Green, T. P.; Hennequin, L. F.; Jacobs, V.; Lambert-Van der Brempt, C.; Morgentin, R.; Ple, P. A.. Abstracts of Papers In 227th National Meeting of the American Chemical Society, Anaheim, CA, March 28-April 1, 2004; American Chemical Society: Washington, DC, 2004.
- (a) For experimental procedures see: Hennequin, L. F. A.; Kettle, J. G; Pass, M.; Bradbury, R. H. PCT Int. Appl. WO 2003040108; (b) Hennequin, L. F. A.; Kettle, J. G; Pass, M.; Bradbury, R. H. PCT Int. Appl. WO 2003040109.
- Cockerill, G. S.; Carter, M. C.; Guntrip, S. B.; Smith, K. J. PCT Int. Appl. WO 9802434, 1998.
- 16. Newkome, G. R.; Kiefer, G. E.; Xia, Y. J.; Gupta, V. K. Synthesis 1984, 676.
- 17. A reproducible procedure for isolation of the sensitive intermediate 4-chloro-5-fluoro fluoroquinazoline is as follows: DMF (0.2 ml) was added to a suspension of 8 (1.64 g) in thionyl chloride (10 ml) and the mixture was stirred and heated at 80 °C for 6 h. Volatile material was removed by evaporation and the residue was azeotroped with toluene (20 ml). The resulting solid was added portionwise to a vigorously stirred mixture of saturated sodium bicarbonate (50 ml), crushed ice (50 g) and DCM (50 ml) such that the temperature was kept below 5 °C. The organic phase was separated, dried and concentrated to give 4-chloro-5-fluoroquinazoline as a solid, which was used without purification (1.82 g, 99%); NMR (CDCl₃): 7.3–7.45 (m, 1H), 7.85–7.95 (m, 2H), 9.0 (s, 1H).
- Wakeling, A. E.; Guy, S. P.; Woodburn, J. R.; Ashton, S. E.; Curry, B. J.; Barker, A. J.; Gibson, K. H. *Cancer Res.* 2002, 62, 5749.
- 19. The activity of compound **12a** was assessed at a concentration of $10 \,\mu\text{M}$ against a panel of 58 recombinant protein kinases at $K_{\rm m}$ ATP concentration for each enzyme.
- Zhang, Y.-M.; Cockerill, S.; Guntrip, S. B.; Rusnak, D.; Smith, K.; Vanderwall, D.; Wood, E.; Lackey, K. *Bioorg. Med. Chem. Lett.* 2004, 14, 111.
- Bradbury, R. H.; Hennequin, L. F. A.; Kettle, J. G.; McCabe, J.; Turner, A. PCT Int. Appl. WO 2005012290, 2005.
- Baselga, J.; Norton, L.; Albanell, J.; Kim, Y. M.; Mendelsohn, J. *Cancer Res.* 1998, 58, 2825.
 BT474C cells²² (seeded at 1×10⁴ cells/well in 96-well
- 23. BT474C cells²² (seeded at 1×10^4 cells/well in 96-well plates) were exposed to 0.004–22 μ M of compound for 96 h and cell growth/viability was assessed using a standard MTT colorimetric endpoint.¹⁸
- 24. KB cells¹⁸ (seeded at 1.6×10^5 cells/well in 6-well plates in serum containing medium for 72 h) were starved with serum free medium for 48 h. Compound was added for 90 min before addition of EGF (1000 ng/ml) for 5 min. Lysates were generated and a sandwich ELISA was used to measure total EGFR phosphorylation.
- Simeoni, M.; Magni, P.; Cammia, C.; De Nicolao, G.; Croci, V.; Pesenti, E.; Germani, M.; Poggesi, I.; Rocchetti, M. *Cancer Res.* 2004, 64, 1094.
- 26. Unpublished data from dosing an EGFR-selective inhibitor in the BT474C mouse xenograft model.