

# Pyrrolidino-tetrahydroisoquinolines bearing pendant heterocycles as potent dual H<sub>3</sub> antagonist and serotonin transporter inhibitors

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**Abstract**—A series of novel and potent 6-heteroaryl-pyrrolidino-tetrahydroisoquinolines with dual histamine H<sub>3</sub> antagonist/serotonin transporter inhibitor activity is described. In vitro and in vivo data are discussed.

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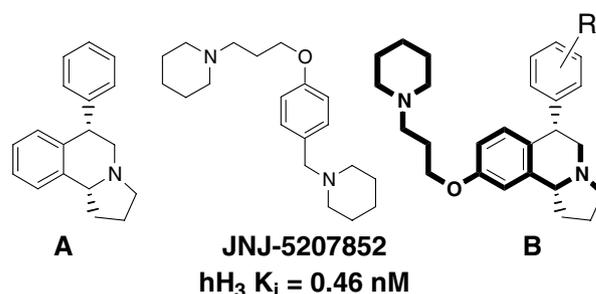
More than 340 million people suffer from depression worldwide, making it a serious global health issue.<sup>1</sup> One of the potentially debilitating symptoms of depression is fatigue.<sup>2</sup> Selective serotonin reuptake inhibitors (SSRIs) are the most frequently prescribed antidepressant drugs, however, these drugs often fail to improve the symptom of fatigue even as mood improves.<sup>3,4</sup> Some SSRIs even induce fatigue and excessive sleepiness.<sup>5,6</sup>

One possible approach to mitigating the fatigue associated with depression and/or its treatment is through the use of a histamine H<sub>3</sub> antagonist. Histamine H<sub>3</sub> receptor antagonists increase wakefulness<sup>7</sup> without showing non-specific stimulant effects such as stimulation of locomotor activity.<sup>8</sup> Thus, the case can be made that H<sub>3</sub> antagonists would be useful adjuncts to antidepressant therapy. In the current paper, we describe a medicinal chemistry effort to synthesize molecules combining H<sub>3</sub> receptor antagonism and blockade of serotonin reuptake.

One of our strategies for the preparation of dual activity H<sub>3</sub> antagonist/serotonin transporter (SERT) inhibitors was to introduce an H<sub>3</sub> pharmacophore to a known SERT inhibitor.<sup>9,10</sup> In one part of these efforts, we

elect to use a pyrrolidino-tetrahydroisoquinoline scaffold, developed as part of an earlier in house antidepressant program,<sup>11</sup> as a template on which to add an H<sub>3</sub> pharmacophore (Fig. 1A). Pharmacophore models for the H<sub>3</sub> receptor suggest a near linear disposition (*m*- or *p*-) of two tertiary amines separated by a phenylene and a hydrophobic chain of at least four atoms would frequently lead to potent H<sub>3</sub> antagonists.<sup>12</sup> We envisioned attachment of a piperidinylpropyloxy side chain to the pyrrolidino-tetrahydroisoquinoline scaffold in such a way as to give a complete H<sub>3</sub> pharmacophore (Fig. 1B).

In a previous report, we disclosed the SAR of pendant phenyl ring substituents (R).<sup>13</sup> In this paper, we further explore the SAR of this region through the introduc-



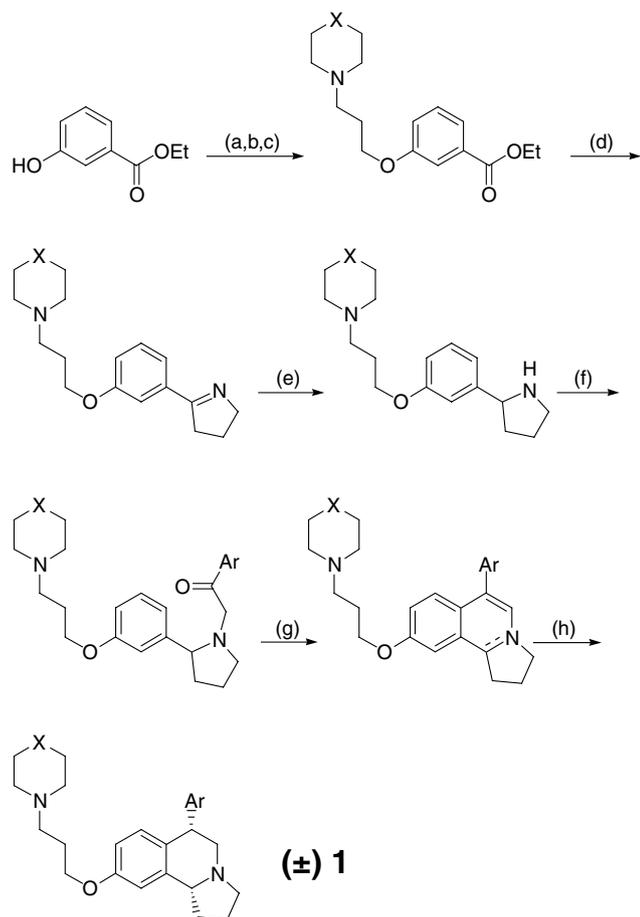
**Figure 1.** (A) SERT inhibitor template. (B) Dual SERT inhibitor and H<sub>3</sub> antagonist template with the H<sub>3</sub> pharmacophore in bold.

**Keywords:** Serotonin transporter; Histamine H<sub>3</sub>; Microdialysis.

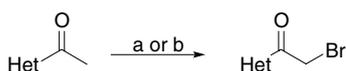
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tion of aromatic heterocycles as either pendant substituents to the phenyl ring or as phenyl ring replacements. Compounds as in Figure 1B were prepared according to Scheme 1 and the bromo ketones utilized in step f were prepared by one of two methods shown in Scheme 2. Heterocyclic ketones with a protonatable nitrogen were first treated with 1 equiv of HBr in acetic acid followed by elemental bromine. Non-basic (to HBr) heterocyclic ketones were simply brominated in chloroform, methylene chloride or methanol.



**Scheme 1.** Reagents and conditions: (a) 1-bromo-3-chloropropane, 3 equiv  $K_2CO_3$ , acetone, reflux, 2 d, 92%; (b) 3.5 equiv NaI, acetone, reflux, 2 d, 98%; (c) 1.3 equiv piperidine or morpholine, 3 equiv  $K_2CO_3$ , EtOH, 60 °C, 2 d, 87%; (d) i—2 equiv *N*-vinylpyrrolidinone, 1.4 equiv NaH, THF rt, reflux; 2 h; ii—6 N HCl reflux, 24 h, 72%; (e)  $LiAlH_4$ , THF, 24 h, rt, 88%; (f) bromo-ketone,  $Et_3N$ , THF, 30 min, 80–99%; (g) MSA, rt–80 °C, 1–6 h (temperature and time depend on R); (h)  $NaCNBH_3$ , HCl, MeOH, bromocresol green, pH 4.5–5.5, 30 min–1 h, 15–70%.



**Scheme 2.** Het, aromatic heterocycle; (a) for basic heterocycles: 1 equiv HBr (48%), 0.9 M HOAc, 1.1 equiv  $Br_2$ , 0–75 °C, 1–4 h; (b) for non-basic heterocycles: 0.14 M in 5:2  $Et_2O/CHCl_3$ , or 0.14 M in MeOH, 1 equiv  $Br_2$ , 0 °C–rt, 18 h, yields typically 70–80% for both methods.

In addition to varying the heteroaryl groups, the side chain was modified to contain either a piperidine or a morpholine. The piperidiny derivatives were generally more potent, but the morpholine compounds had lower  $c \log P$ 's.<sup>14</sup> In vitro data for the final products **1a–p** are found in Table 1.

Perusal of Table 1 reveals that these compounds all have a high affinity for the  $H_3$  receptor.<sup>15</sup> This should not be too surprising as the  $H_3$  pharmacophore portion of these molecules has not been modified beyond interchanging piperidine and morpholine, both of which are known to give high affinity  $H_3$  ligands.<sup>16</sup> The compounds were shown to be potent antagonists in a functional assay with  $pA_2$ 's in the range of 8.5–10.2. However, the nature of the pendant heteroaryl strongly influences SERT binding.<sup>17,18</sup> Generally, these compounds exhibited greater potency against rSERT than the hSERT. Pyrazolo-phenyl and imidazolo-phenyl derivatives (**1a–b**) were reasonably well tolerated by the SERTs. Of the pyridyl derivatives (**1c–g**), the 3-pyridyl compounds were the most potent. Introduction of an additional ring nitrogen to give a pyrazine (**1h**) resulted in a significant loss of SERT potency. The most potent compounds were the thienyl derivatives (**1i–l**), maintaining excellent activity at both the rat and human SERTs. Thiazoles, pyrazoles, and imidazopyridines were all much less potent against the SERTs.

Compound **1d** was studied in a mouse brain to plasma ratio experiment.<sup>19</sup> After ip administration (10 mg/kg), **1d** achieved a maximal plasma concentration of 1.43  $\mu M$  after 15 min, and a maximal brain concentration of 0.92  $\mu M$  after 2 h. Mean residence time in the plasma was 6.57 h and 31.29 h in the brain. The blood–brain barrier (BBB) coefficient was 0.75.<sup>20</sup>

Compound **1d** was screened against a panel of other targets (receptors, ion channels, transporters, etc.) and those revealing significant affinity (>50% at 1  $\mu M$ ) were retested and  $K_i$ 's determined. Other targets of interest included the dopamine transporter (DAT;  $K_i = 107$  nM), norepinephrine transporter (NET;  $K_i = 460$  nM), and the  $\kappa$  opioid receptor ( $K_i = 74$  nM).

The BBB data as well as the DAT activity prompted us to study **1d** via in vivo microdialysis coupled with HPLC-electrochemical detection in an effort to assess the effects of **1d** on the extracellular concentration of serotonin (5-HT) and dopamine (DA) in the frontal cortex of the rat brain.<sup>21</sup> Within 4 h of dosing **1d** (3 mg/kg sc) the 5-HT and DA levels had increased 300% and 500% over basal levels, respectively. The increases in neurotransmitter concentrations were sustained for the duration of the experiment (21 h).

In conclusion, the described compounds are potent brain penetrating dual  $H_3$  antagonist/SERT inhibitors with potential utility for treating depression.

**Table 1.** Binding data for rat and human serotonin reuptake transporters and for the human histamine H<sub>3</sub> receptor for compounds **1a–p**

Compound <b>1</b>	X	Ar	rSERT K <sub>i</sub> (nM)	hSERT K <sub>i</sub> (nM)	hH <sub>3</sub> K <sub>i</sub> (nM)
<b>a</b>	O		6 (±1.2)	12.3 (±2.7)	5.3 (±0.4)
<b>b</b>	O		6.7 (±1.5)	16.7 (±2.0)	5.7 (±1.1)
<b>c</b>	CH <sub>2</sub>		20 (±6.0)	66 (±32)	1 (±0.7)
<b>d</b>	CH <sub>2</sub>		2 (±0.7)	8.3 (±1.8)	0.8 (±0.1)
<b>e</b>	O		4.3 (±0.8)	12.3 (±2.3)	1.6 (±0.4)
<b>f</b>	CH <sub>2</sub>		3.3 (±0.4)	18.7 (±2.2)	0.8 (±0.1)
<b>g</b>	O		8.3 (±1.1)	31.3 (±0.8)	2.7 (±0.4)
<b>h</b>	O		22.7 (±7.6)	88 (±2.6)	4 (±0.7)
<b>i</b>	CH <sub>2</sub>		1.2 (±0.5)	2.7 (±0.8)	1.0 (±0)
<b>j</b>	O		3 (±0.7)	6 (±0.7)	3 (±0.7)
<b>k</b>	CH <sub>2</sub>		0.7 (±0.2)	1.3 (±0.4)	0.7 (±0.2)
<b>l</b>	O		2 (±0)	5 (±0)	3 (±0.7)
<b>m</b>	O		6 (±0)	19.7 (±0.4)	3.7 (±0.4)
<b>n</b>	O		53.3 (±20)	291 (±102)	3.3 (±0.4)
<b>o</b>	O		24.3 (±9.4)	121 (±27)	4 (±0.7)
<b>p</b>	O		39 (±7.8)	119 (±11.4)	2 (±0)

rSERT, rat serotonin transporter; hSERT, human serotonin transporter; hH<sub>3</sub>, human histamine H<sub>3</sub> receptor: numbers in parentheses are the standard error of the mean (SEM) for each data set:  $n \geq 3$  for all in vitro data.

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### References and notes

- Jane-Llopis, E.; Hosman, C.; Jenkins, R.; Anderson, P. *Br. J. Psychiatry; J. Ment. Sci.* **2003**, *183*, 384.
- Tylee, A.; Gastpar, M.; Lepine, J. P.; Mendlewicz, J. *Int. Clin. Psychopharmacol.* **1999**, *14*, 139.

3. Nierenberg, A. A.; Keefe, B. R.; Leslie, V. C.; Alpert, J. E.; Pava, J. A.; Worthington, J. J., 3rd; Rosenbaum, J. F.; Fava, M. *J. Clin. Psychiatry* **1999**, *60*, 221–225.
4. Fava, G. A.; Fabbri, S.; Sonino, N. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **2002**, *26*, 1019.
5. Beasley, C. M., Jr.; Koke, S. C.; Nilsson, M. E.; Gonzales, J. S. *Clin. Ther.* **2000**, *22*, 1319–1330.
6. Zajecka, J. M. *J. Clin. Psychiatry* **2000**, *61*(Suppl. 2), 20.
7. Monti, J. M.; Jantos, H.; Boussard, M.; Altier, H.; Orellana, C.; Olivera, S. *Eur. J. Pharmacol.* **1991**, *205*, 283.
8. Barbier, A. J.; Berridge, C.; Dugovic, C.; Laposky, A. D.; Wilson, S. J.; Boggs, J.; Aluisio, L.; Lord, B.; Mazur, C.; Pudiak, C. M.; Langlois, X.; Xiao, W.; Apodaca, R.; Carruthers, N. I.; Lovenberg, T. W. *Br. J. Pharmacol.* **2004**, *143*, 649.
9. Keith, J. M.; Gomez, L. A.; Letavic, M. A.; Ly, K. S.; Jablonowski, J. A.; Seierstad, M.; Barbier, A. J.; Wilson, S. J.; Boggs, J. D.; Fraser, I. C.; Mazur, C.; Lovenberg, T. W.; Carruthers, N. I. *Bioorg. Med. Chem. Lett.* **2007**, *17*, 702.
10. Letavic, M. A.; Keith, J. M.; Jablonowski, J. A.; Stocking, E. M.; Gomez, L. A.; Ly, K. S.; Miller, J. M.; Barbier, A. J.; Bonaventure, P.; Boggs, J. D.; Wilson, S. J.; Miller, K. L.; Lord, B.; McAllister, H. M.; Tognarelli, D. J.; Wu, J.; Abad, M. C.; Schubert, C.; Lovenberg, T. W.; Carruthers, N. I. *Bioorg. Med. Chem. Lett.* **2007**, *17*, 1047–1051.
11. Maryanoff, B. E.; McComsey, D. F.; Gardocki, J. F.; Shank, R. P.; Costanzo, M. J.; Nortey, S. O.; Schneider, C. R.; Setler, P. E. *J. Med. Chem.* **1987**, *30*, 1433–1454.
12. Apodaca, R.; Dvorak, C. A.; Xiao, W.; Barbier, A. J.; Boggs, J. D.; Wilson, S. J.; Lovenberg, T. W.; Carruthers, N. I. *J. Med. Chem.* **2003**, *46*, 3938.
13. Keith, J. M.; et al., <http://dx.doi.org/10.1016/j.bmcl.2007.01.106>.
14. **1d** has a *c log P* of 3.21, **1e** has a *c log P* of 2.08.
15. For the H<sub>3</sub> assay see Barbier, A. J.; Berridge, C.; Dugovic, C.; Laposky, A. D.; Wilson, S. J.; Boggs, J.; Aluisio, L.; Lord, B.; Mazur, C.; Pudiak, C. M.; Langlois, X.; Xiao, W.; Apodaca, R.; Carruthers, N. I.; Lovenberg, T. W. *Br. J. Pharmacol.* **2004**, *143*, 649.
16. Apodaca, R.; Dvorak, C. A.; Xiao, W.; Barbier, A. J.; Boggs, J. D.; Wilson, S. J.; Lovenberg, T. W.; Carruthers, N. I. *J. Med. Chem.* **2003**, *46*, 3938.
17. Carruthers, N. I.; Gomez, L. A.; Jablonowski, J. A.; Keith, J. M.; Letavic, M. A.; Ly, K.S.; Miller, J.M.B.; Stocking, E.M.; Wolin, R.L. *PCT Int. Appl.* **2006**, WO 2006066197, 192 pp.
18. In a previous report (see Ref. 14) we described differences in potency between enantiomers. Both enantiomers were highly potent at H<sub>3</sub> but only one, the (–)-isomer, possessed significant SERT activity.
19. Also called a BBB experiment.
20. The BBB coefficient is defined as LOG (AUC<sub>brain</sub>/AUC<sub>plasma</sub>).
21. See Ref. 13 for experimental details Paxinos, G.; Watson, C. *The Rat Brain in Stereotaxic Coordinates*, Third ed.; Academic Press: San Diego, CA, 1997.