

Novel Inhibitors of AP-1 and NF- κ B Mediated Gene Expression: Structure–Activity Relationship Studies of Ethyl 4-[(3-Methyl-2,5-Dioxo(3-pyrrolinyl))amino]-2-(trifluoromethyl)pyrimidine-5-carboxylate

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Abstract—In an effort to identify novel inhibitors of AP-1 and NF- κ B mediated transcriptional activation, several analogues of ethyl 4-[(3-methyl-2,5-dioxo(3-pyrrolinyl))amino]-2-(trifluoromethyl)pyrimidine-5-carboxylate (**1**) were synthesized and tested in two in vitro assays. The 2-(2'-thienyl) substituted compound (**11**) was identified as the most potent in this series. © 2000 Elsevier Science Ltd. All rights reserved.

Introduction

There is now abundant evidence that T-lymphocytes orchestrate both the initiation and propagation of immune responses through the secretion of protein mediators termed cytokines.¹ These cytokines play a very important role in a number of inflammatory diseases.^{2,3} In allergies and autoimmune diseases—such as asthma, psoriasis, rheumatoid arthritis, and transplant rejection—T-cell driven immune responses appear to overreact. In activated T cells, transcription factors such as the activator protein-1 (AP-1), regulate IL-2 production and production of matrix metalloproteinases, while the nuclear factor- κ B (NF- κ B), is essential for the transcriptional regulation of the proinflammatory cytokines IL-1, IL-6, IL-8, and TNF α .⁴ Based on these observations, it appears that inhibition of AP-1 and NF- κ B transcriptional activation in T cells may represent an attractive target in the development of novel antiinflammatory drugs (Fig. 1).⁵

Using automated high-throughput assays with stably transfected human jurkat T-cells, we identified a compound (**1**) from a diversity library that inhibited both AP-1 and NF- κ B mediated transcriptional activation (IC_{50} = 1 μ M) without blocking basal transcription driven by the β -actin promoter. In addition, **1** had a similar inhibitory

effect on the production of IL-2 and IL-8 levels in stimulated cells. Our goal was to improve potency by exploring different substituents around the pyrimidine ring. We introduced various groups on the pyrimidine ring at 2, 4 and 6-positions of **1**. This paper describes the synthesis and the structure–activity relationship of this series of compounds.

Chemistry

The synthesis of 2- and 4-substituted analogues is shown in Scheme 1. An appropriately substituted amidine was cyclized with diethyl 2-(ethoxymethylene)propane-1,3-dioate in ethanol and sodium ethoxide to give **4**. The amidines that are not available commercially were prepared either from the corresponding nitrile or acid as shown.⁶ The hydroxy group in **4** was converted to a chloro group⁷ and then treated with hydrazine or methylhydrazine to give the appropriate intermediate, **5**. Treatment of the hydrazine intermediates (**5**) with appropriately substituted maleic anhydride resulted in the final products (**6–44**). The treatment of **10** with acetic anhydride resulted in **38**. Table 1 shows the list of compounds that were prepared with modifications at R₁, R₂, R₃, and R₄.

We also examined the importance of the carboxylate group at the 5-position. Several ester bioisosteres were introduced in place of the ethyl ester of **11**, which were

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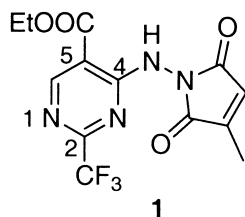


Figure 1.

synthesized as shown in Schemes 2 and 3. The synthesis of **46** and **49** was initiated from the chloro derivative **44** (Scheme 2). The chlorine at the 4-position was replaced with a benzyl thiol followed by conversion of the ester to the isoxazole, **45**.⁸

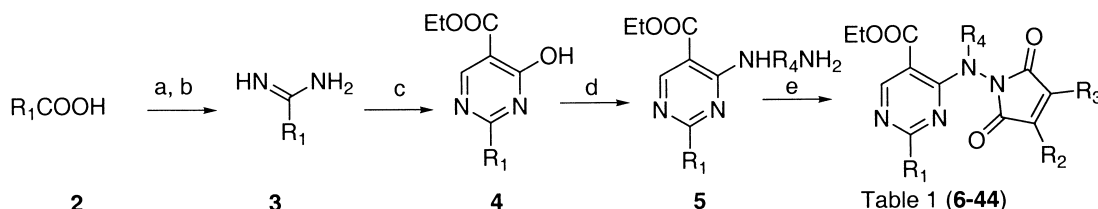
The *S*-benzyl group in **45** was oxidized with *m*-CPBA to give a sulfone. The sulfone was treated with hydrazine followed by citraconic anhydride to give **46**. The ester group in **44** was hydrolyzed and converted to cyano analogue, **48**. The treatment of **48** with sodium azide and ammonium chloride in DMF resulted in a tetrazole ring. The tetrazole ring was methylated and *S*-benzyl group

was converted to *N*-aminocitraconamide as discussed earlier to give **49**.

The synthesis of oxazole analogue **51** started from the free acid **47** (Scheme 3). The acid group was converted to an ester using a chloroacetone and to methyl oxazole **50** using acetamide and $\text{BF}_3\text{-Et}_2\text{O}$.⁸ Compound **50** was converted to **51** using a reaction sequence for the conversion of **45** to **46**. The carboxylic acid group in **47** was converted to 2-methyl-1,3,4-oxadiazole.⁹ The oxadiazole intermediate was converted to **52** using the reaction sequence for the conversion of **45** to **46**. The treatment of acid **47** with oxalyl chloride, followed by 2-aminoethanol, and then thionyl chloride resulted in dihydro oxazole **53**. The citraconamido group was introduced on to **54** as described earlier.

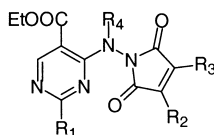
Results and Discussion

The analogues synthesized as part of this study were evaluated in Jurkat T-cells stably transfected with promoter-reporter gene constructs driven by either an AP-1

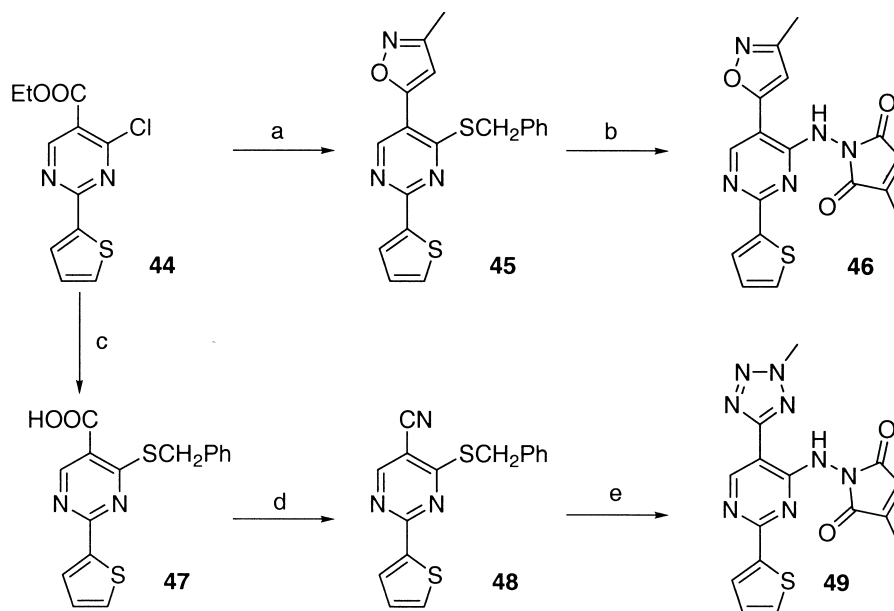


Scheme 1. (a) (i) $(\text{COCl})_2$, DMF; (ii) NH_3 , 3. SOCl_2 ; (b) (i) HCl (gas), EtOH; (ii) NH_3 (gas), EtOH; (c) Diethylethoxymethylene malonate, EtONa, EtOH; (d) (i) POCl_3 ; (ii) R_4NHNH_2 ; (e) Citraconic anhydride, CHCl_3 , Δ .

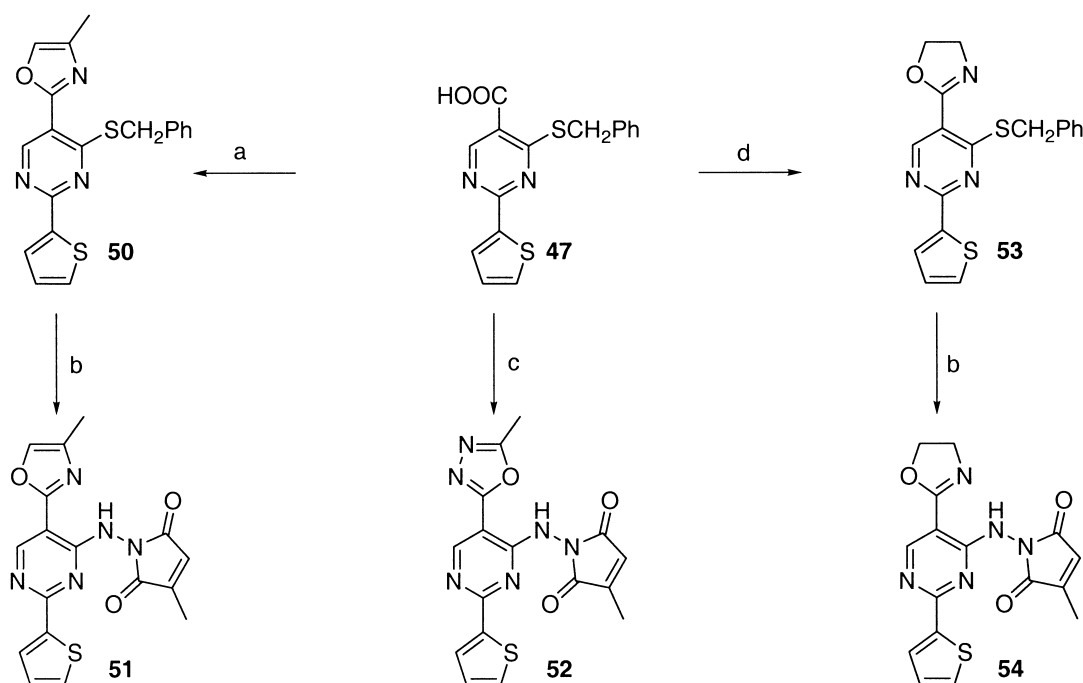
Table 1. In vitro evaluation in Jurkat T-cells of compounds with R_1 through R_4 modifications on the pyrimidine ring. IC_{50} values for the both AP-1 and NF- κB mediated transcriptional activation were the same.



No.	R_1	R_2	R_3	R_4	IC_{50} , μM	No.	R_1	R_2	R_3	R_4	IC_{50} , μM
1	CF_3	CH_3	H	H	1	26	5-Methyl-2-thienyl	CH_3	H	H	0.2
6	CH_3	CH_3	H	H	16	27	5-Chloro-2-thienyl	CH_3	H	H	0.3
7	CH_3CH_2	CH_3	H	H	10	28	2-Benzo-thienyl	CH_3	H	H	4
8	<i>t</i> -Bu	CH_3	H	H	1	29	2-Furanoyl	CH_3	H	H	0.2
9	SCH_3	CH_3	H	H	0.2	30	Cyclopropyl	CH_3	H	H	1
10	Ph	CH_3	H	H	0.1	31	2-(Cyclo-hex-2-enyl-methyl	CH_3	H	H	2
11	2-Thienyl	CH_3	H	H	0.02	32	3,5-Dichloro-phenyl	CH_3	H	H	0.3
12	4-Pyridyl	CH_3	H	H	6	33	Benzyl	CH_3	H	H	7
13	2,6-Dichloro-4-pyridyl	CH_3	H	H	2	34	Phenoxy	CH_3	H	H	15
14	3-Quinoliny	CH_3	H	H	6	35	2-Phenyl-thio	CH_3	H	H	1
15	4-(2-Methyl-thiazolyl)	CH_3	H	H	8	36	2-Phenyl-sulfonyl	CH_3	H	H	10
16	1-(3, 5-Dimethyl-pyrazolyl)	CH_3	H	H	10	37	Phenyl	CH_3	H	CH_3	2
17	4-Methoxy-phenyl	CH_3	H	H	0.9	38	Phenyl	CH_3	H	Ac	4
18	3-Methoxy-phenyl	CH_3	H	H	5	39	2-Thienyl	CH_3	H	CH_3	1
19	4-Fluoro-phenyl	CH_3	H	H	0.7	40	4-Fluoro-phenyl	CH_3	H	Me	3
20	4-Chloro-phenyl	CH_3	H	H	0.3	41	CH_3CH_2	CH_3	H	Me	10
21	4-Trifluoro-methyl-phenyl	CH_3	H	H	0.4	42	CF_3	CH_3	CH_3	H	10
23	3-Bromo-phenyl	CH_3	H	H	0.7	43	CF_3	Ph	H	H	20
24	3-Nitro-phenyl	CH_3	H	H	7	44	CF_3	H	H	H	10
25	3-Thienyl	CH_3	H	H	0.1						



Scheme 2. (a) (i) PhCH₂SH, THF; (ii) Acetone oxime, *n*-BuLi, THF; (iii) H₂SO₄, THF; (b) (i) *m*-CPBA; (ii) NH₂NH₂; (iii) Citraconic anhydride, CHCl₃, reflux; (c) (i) PhCH₂SH, THF; (ii) NaOH; (d) (i) (COCl)₂; (ii) NH₃ (gas); (iii) SOCl₂, DMF; (e) (i) NaN₃, NH₄Cl, DMF; (ii) MeI, K₂CO₃; (iii) *m*-CPBA; (iv) NH₂NH₂; (v) Citraconic anhydride, CHCl₃, δ .



Scheme 3. (a) (i) ClCH₂COCH₃; (ii) Acetamide, BF₃-Et₂O, xylenes; (b) (i) *m*-CPBA; (ii) NH₂NH₂; (iii) citraconic anhydride; (c) (i) CH₃CONHNH₂, POCl₃, δ ; (ii) *m*-CPBA; (iii) NH₂NH₂; (iv) Citraconic anhydride, CHCl₃, δ ; (d) (i) (COCl)₂; (ii) NH₂CH₂CH₂OH; (iii) SOCl₂, EtOAc-CHCl₃.

binding site or a NF- κ B binding site.¹⁰ All the compounds were tested in both assays. The IC₅₀ values for these compounds are shown below. Since all the compounds had similar IC₅₀ values in the both AP-1 and NF- κ B assays, average values are shown (Table 1).

Compound 1 had IC₅₀ activity in both the cell based assays at 1 μ M. The substitution of a methyl (6) and ethyl (7) in the place of trifluoromethyl group resulted in the loss of activity. However, the introduction of a *t*-butyl

(8) group resulted in a compound with comparable activity. The *S*-methyl (9) and phenyl (10) substituents improved activity. The 2-thienyl (11) substituent resulted in a 50-fold improved activity. The substituted and unsubstituted heterocyclic rings (12–16) at 2-position resulted in loss of activity. Any substitution on the phenyl ring of 10 (compounds 17–24) resulted in decreased activity. Similarly, introduction of a group on the thienyl ring of 11 (compounds 26–28) also resulted in decreased activity. Introduction of a methyl group at R₃ (42), a

Table 2. In vitro evaluation in Jurkat T-cells of compounds with modifications of R₅ of the pyrimidine ring. IC₅₀ values for the both AP-1 and NF-κB mediated transcriptional activation were the same.

No	R ₅	IC ₅₀ , μM	No	R ₅	IC ₅₀ , μM
46		0.3	51		0.2
48	CN	5	52		0.3
49		0.2	53		0.2

phenyl group at R₂ (**43**) or a hydrogen at R₂ (**44**) resulted in the decreased activity. Similarly, substitution of a methyl group at R₄ (**37**, **39**, **40** and **41**) resulted in the decreased activity. The substitution of an acetyl group at R₄ (**38**) also resulted in decreased activity (Table 2).

Five ester bioisoster modifications were introduced in place of the ester of **11**. A cyano analogue (**48**) was also

tested. The bioisosteres **49**, **51** and **53** were 10-fold less active in the cell based assays. Analogues **46** and **52** were 15-fold less active and cyano analogue **48** was 250-fold less active. Based on the above structural activity relationship studies, a thienyl group at 2-position ring, an *N*-aminocitraconamido group at 4-position, and an ethyl ester at 5-position of the pyrimidine ring **1** are optimum substituents for the biological activity. To date we have identified a potent and novel inhibitor (**11**) of AP-1 and NF-κB mediated transcriptional activation.

References

- Palanki, M. S. S.; Manning, A. M. *Exp. Opin. Ther. Patents* **1999**, *9*, 27.
- Manning, A. M. *Drug Discovery Today* **1996**, *1*, 151.
- Lewis, A. J. *Emerging Drugs: The Prospect for Improved Medicines*; Annual Executive Briefing, Ashley Publications Ltd., 1996, 31.
- Vossen, A. C. T. M.; Savelkoul, H. F. J. *Mediat. Inflamm.* **1994**, *3*, 403.
- Manning, A. M.; Lewis, A. J. *Rheumatoid Arthritis* **1997**, *1*, 65.
- Dox, W. In *Organic Synthesis*; Gilman, H., Ed.; John Wiley and Sons: New York, 1941, Vol 1, p 5.
- Church, T.; Albright, P. J. *Org. Chem.* **1995**, *60*, 3750.
- Diana, G. D.; Oglesby, R. C.; Akullian, V.; Carabateas, P. M.; Cutcliffe, D.; Mallamo, J. P.; Otto, M. J.; McKinlay, M. A.; Maliski, E. G.; Michalec, S. J. *J. Med. Chem.* **1987**, *30*, 383.
- Kotian, P.; Mascarella, W.; Abraham, P.; Lewin, A.; Boja, J.; Kuhar, M.; Carroll, I. *J. Med. Chem.* **1996**, *39*, 2753.
- Sullivan, R. W.; Bigam, C. G.; Erdman, P. E.; Palanki, M. S. S.; Anderson, D. W.; Goldman, M.; Ransone, L. J.; Suto, M. J. *J. Med. Chem.* **1998**, *41*, 413.