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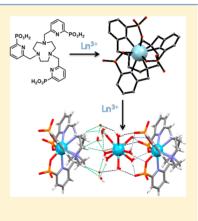
# Formation of Mono- and Polynuclear Luminescent Lanthanide Complexes based on the Coordination of Preorganized **Phosphonated Pyridines**

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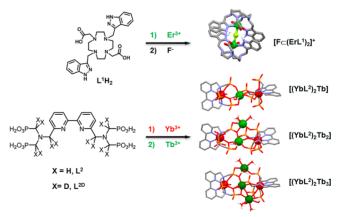
Supporting Information

**ABSTRACT:** A series of polynuclear assemblies based on ligand L (1,4,7-tris[hydrogen (6methylpyridin-2-yl)phosphonate]-1,4,7-triazacyclononane) has been developed. The coordination properties of ligand L with  $Ln^{III}$  (Ln = La, Eu, Tb, Yb, Lu) have been studied in water (pH = 7.0) and in  $D_2O$  (pD = 7.0) by UV-absorption spectrometry, spectrofluorimetry, <sup>1</sup>H and <sup>31</sup>P NMR, DOSY, ESI-mass spectrometry, and X-ray diffraction. This nonadentate ligand forms highly stable mononuclear complexes in water and provides a very efficient shielding of the Ln cations, as emphasized by the very good luminescence properties of the Yb complex in D<sub>2</sub>O, especially regarding its lifetime ( $\tau_{D_2O} = 10.2 \ \mu s$ ) and quantum yield ( $\phi_{D,O}$  = 0.42%). In the presence of excess Ln<sup>III</sup> cation, polynuclar complexes of  $[(LnL)_2Ln_x]$  stoichiometry (x = 1 and x = 2) are observed in solution. In the solid state, a dinuclear complex of La could be isolated and structurally characterized by X-ray diffraction, unraveling the presence of strong hydrogen bonding interactions between a  $La(H_2O)_{q^3}$ cation and the  $[LaL]^{3-}$  complex.



# INTRODUCTION

Photonic up conversion (UC) is very appealing for bioanalytical assays and theranostic applications since it allows for the conversion of low energy photons in the near-infrared to visible emission.<sup>1</sup> Although this phenomenon is usually observed in solid state materials<sup>2</sup> and nanomaterials,<sup>3,4</sup> a few discrete molecular systems have proven their efficiency.<sup>5–9</sup> In particular, we have recently demonstrated that UC was also possible with supramolecular systems in solution at room temperature using lanthanide coordination complexes.<sup>7,8</sup> In the first example,<sup>7</sup> visible up converted emission of  $Er^{3+}$  was observed upon laser excitation of a sandwich  $[(L^1Er)-F$ - $(L^{1}Er)$ <sup>+</sup> assembly, in which the two Er centers were embedded in a macrocyclic DOTA derivative (L<sup>1</sup>, Scheme 1), which holds the dimeric structure by  $\pi - \pi$  stacking interactions and Hbonding.<sup>10</sup> In the second strategy, heteropolynuclear lanthanide (Ln) assemblies have been developed in order to promote Yb  $\rightarrow Tb$  energy transfers. In this case, the tetraphosphonated octadentate ligand  $L^2$  (Scheme 1) formed highly stable mononuclear  $[LnL^2]$  complexes which, in the presence of excess Ln<sup>3+</sup> salts, self-assembled into polynuclear [(LnL<sup>2</sup>)<sub>2</sub>Ln],  $[(LnL^2)_2Ln_2]$ , and  $[(LnL^2)_2Ln_3]$  species. The assembly is driven by the strong polarization of the ligand due to the electron-rich phosphonated pendant arms.<sup>11</sup> Such engineering Scheme 1. Polynuclear Lanthanide Assemblies with Ligands  $L^1$ ,  $L^2$ , and  $L^{2D}$ 

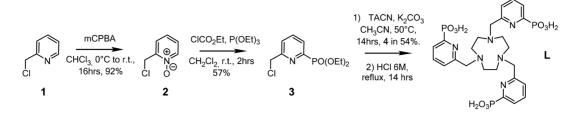


offers a very good shielding of the Ln cations by the ligands and very interesting photophysical properties in water and D<sub>2</sub>O. In particular, the deuterated analogue YbL<sup>2D</sup> was used in the

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#### Scheme 2. Synthesis and Chemical Structure of Ligand L



formation of  $[(YbL^{2D})_2Tb_x]$  complexes (x = 1-3), which afforded characteristic visible UC emission of Tb, upon NIR excitation of the Yb at 980 nm.

In our search for new systems with improved efficiency, we turned our interest toward the development of chelates based on the 1,4,7-triazacyclononane (tacn) macrocycle appended with pyridine phosphonate units (Scheme 2). Lanthanide complexes with ligands based on tacn substituted with bipyridines,<sup>12</sup> picolinate,<sup>13</sup> 8-hydroxyquinoline,<sup>14</sup> and pyridyl phosphinate<sup>15–17</sup> have proven to be very efficient in the design of highly stable mononuclear lanthanide complexes with very interesting photophysical properties in the solid-state, in solution, and even *in cellulo*<sup>18,19</sup> or on cell membranes<sup>20</sup> and are ideal candidates for the development of biomedical imaging probes. Derivatives with two-photon antenna are also appealing for two-photon microscopy experiments in both the visible and near-infrared spectral ranges.<sup>21</sup>

Introduction of 6-phosphonated-2-methylene bridged pyridines on the nitrogen atoms is expected to lead to the formation of two five-membered chelate rings per arm; a nonadentate coordination suited to the protection of the first coordination sphere of the Ln cations and a  $C_3$  symmetry with the negatively charged phosphonate functions on one side of the complexes, with the possibility to further develop electrostatic interactions.<sup>11,22</sup> Furthermore, phosphonated pyridines are expected to provide highly stable lanthanide complexes compared to their carboxylated analogues<sup>11,13,23</sup> and efficient sensitization of Tb<sup>3+</sup>, Eu<sup>3+</sup>, and, more interestingly, Yb<sup>3+</sup>.

In this study, we report the synthesis of the new ligand L (Scheme 2) and a detailed analysis of its coordination properties with  $Ln^{3+}$  ions by UV-absorption spectrometry, spectrofluorimetry, <sup>1</sup>H and <sup>31</sup>P NMR, DOSY, ESI-mass spectrometry, and X-ray diffraction.

# EXPERIMENTAL SECTION

General Methods. Solvents and starting materials were purchased from Aldrich, Acros, and Alfa Aesar and used without further purification. Flash column chromatography was performed with silica gel (10-63  $\mu$ m, Macherey Nagel) or on C18 reversed phase silica (Macherey Nagel). IR spectra were recorded on a PerkinElmer Spectrum One spectrophotometer as solid samples, and only the most significant absorption bands are given in cm<sup>-1</sup>. Elemental analysis and mass spectrometry analysis were carried out by the Service Commun d'Analyses of the University of Strasbourg. <sup>1</sup>H, <sup>31</sup>P, and <sup>13</sup>C NMR spectra and 2D COSY, NOESY, HSQC, and HMBC experiments were recorded on Avance 300 and Avance 400 spectrometers operating at 300 and 400 MHz for <sup>1</sup>H, respectively. Chemical shifts are reported in ppm, with residual protonated solvent as internal reference.<sup>24</sup> When unspecified, coupling constants refer to H - H coupling. The given pH values are corrected for the deuterium isotopic effects (pD = apparent pH + 0.4).<sup>2</sup>

Synthesis of Diethyl (6-(Chloromethyl)pyridin-2-yl)phosphonate (3). In a 250 mL round-bottom flask, 2-(chloromethyl)pyridine-1-

oxide<sup>26</sup> (4.00 g, 27.9 mmol) was dissolved in dichloromethane (100 mL) under an argon atmosphere. Ethyl chloroformate (8.0 mL, 83.7 mmol) was added, and the solution was stirred at r.t. for 30 min. Triethyl phosphite (14.8 mL, 86.3 mmol) was added dropwise, and the resulting solution was stirred at r.t. during 2 h. The resulting mixture was washed with saturated NaHCO2, water, and brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated to dryness under vacuum. Impurities were removed by distillation under reduced pressure. Purification of the crude product was performed by column chromatography over silica gel (CH2Cl2/MeOH gradient from 100:0 to 97:3), yielding 3 (4.17 g, 57%) as an oil. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$  7.78 (m, 2H), 7.57 (td,  $J_1 = 7.7$  Hz,  $J_2 = 1.6$  Hz, 1H), 4.65 (s, 2H), 4.15 (m, 4H), 1.27 (t, J = 7.2 Hz, 6H) ppm. <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz):  $\delta$  157.7 (d, J = 22.8 Hz), 151.6 (d, J = 226.7 Hz), 137.2 (d, J = 12.1 Hz), 127.2 (d, J = 24.9 Hz), 125.2 (d, J = 3.6 Hz), 63.2 (d, J = 5.9 Hz), 46.4, 16.4 (d, J = 5.9 Hz) ppm. <sup>31</sup>P NMR (CDCl<sub>3</sub>, 162 MHz):  $\delta$  10.0 ppm. IR (cm<sup>-1</sup>, ATR):  $\nu$  2983 (m), 1582 (m), 1478 (w), 1447 (m), 1392 (w), 1249 (s,  $\nu_{P=0}$ ), 1182 (m), 1016 (s), 961 (s), 881 (m).  $ESI^+/MS: m/z = 270.064$  ([M+Li]<sup>+</sup>, 100%). Anal. Calcd for C10H15ClNO3P: C, 45.55; H, 5.73; N, 5.31. Found: C, 44.91; H, 5.71; N, 5.28.

Synthesis of Compound 4. A Schlenk tube containing 3 (2.00 g, 7.6 mmol), tacn (325 mg, 2.5 mmol), and flame-dried K<sub>2</sub>CO<sub>3</sub> (1.75 g, 12.7 mmol), dissolved in anhydrous CH<sub>3</sub>CN, was stirred for 16 h at 50 °C. The remaining K<sub>2</sub>CO<sub>3</sub> was removed by filtration, and the solvent was removed under reduced pressure. The resulting residue was dissolved in dichloromethane, washed with water and brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated to dryness under vacuum. Purification of the crude product was performed by column chromatography over alumina (CH<sub>2</sub>Cl<sub>2</sub>/MeOH gradient from 100:00 to 95:5), yielding compound 4 (1.18 g, 56%) as pale yellow oil. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta$  7.74 (t, J = 6.8 Hz, 3H), 7.68 (td,  $J_1 = 7.6 \text{ Hz}, J_2 = 5.9 \text{ Hz}, 3\text{H}), 7.60 \text{ (d, } J = 7.6 \text{ Hz}, 3\text{H}), 4.23-4.11 \text{ (m,}$ 12 H), 3.89 (s, 6H), 2.85 (s, 12H), 1.28 (t, J = 7.1 Hz, 18H) ppm. <sup>13</sup>C NMR (CDCl<sub>3</sub>, 125 MHz): δ 161.6 (br), 151.3 (d, *J* = 227.5 Hz), 136.5 (d, J = 12.3 Hz), 126.5 (d, J = 25.4 Hz), 125.7, 64.4, 63.0 (d, J = 5.9 Hz), 55.9, 16.5 (d, J = 5.9 Hz) ppm. <sup>31</sup>P NMR (CDCl<sub>3</sub>, 202 MHz):  $\delta$ 11.0 ppm. IR (cm<sup>-1</sup>, ATR):  $\nu$  2982 (w), 2906 (w), 1672 (w), 1582 (w), 1443 (m), 1391 (w), 1367 (w), 1254 (s,  $\nu_{P=O}$ ), 1163 (m), 1016 (s), 961 (s), 790 (m), 760 (m), 564 (m), 532 (m). ESI<sup>+</sup>/MS: m/z =811.34 ([M + H]<sup>+</sup>, 100%). Anal. Calcd for C<sub>36</sub>H<sub>57</sub>N<sub>6</sub>O<sub>9</sub>P<sub>3</sub>•H<sub>2</sub>O: C, 52.17; H, 7.15; N, 10.14. Found: C, 51.79; H, 6.96; N, 9.64.

Synthesis of Ligand L. Compound 4 (200 mg, 0.24 mmol) was dissolved in 6 N HCl solution (30 mL) and was refluxed during 16 h. The solvent was removed under vacuum, and ligand L (175 mg, 0.20 mmol, 90%) was obtained as a white powder. <sup>1</sup>H NMR (D<sub>2</sub>O, 500 MHz):  $\delta$  8.21 (td,  $J_1$  = 7.9 Hz,  $J_2$  = 3.7 Hz, 3 H), 7.96 (t, J = 7.4 Hz, 3 H), 7.79 (d, J = 8.0 Hz, 3 H), 4.51 (s, 6 H), 3.27 (s, 12 H) ppm. <sup>13</sup>C NMR (D<sub>2</sub>O, 125 MHz):  $\delta$  153.1 (d, J = 194.4 Hz), 152.7 (d, J = 13.7 Hz), 143.3 (d, J = 10.0 Hz), 127.8 (d, J = 16.4 Hz), 127.5, 57.5, 50.1 ppm. <sup>31</sup>P NMR (D<sub>2</sub>O, 202 MHz):  $\delta$  1.84 ppm. IR (cm<sup>-1</sup>, ATR):  $\nu$  2638 (m, br), 1628 (w), 1403 (w), 1147 (m), 925 (s), 812 (m), 692 (m), 532 (s). ESI<sup>+</sup>/MS: m/z = 643.15 ([M + H]<sup>+</sup>, 100%). Anal. Calcd for C<sub>24</sub>H<sub>33</sub>N<sub>6</sub>O<sub>9</sub>P<sub>3</sub>•3HCl•3H<sub>2</sub>O: C, 35.77; H, 5.25; N, 10.43. Found: C, 35.38; H, 5.08; N, 10.45.

**Synthesis and Characterization of the Ln<sup>3+</sup> Complexes.** *General Method.* In order to control the strict 1:1 stoichiometry of the complexes and to avoid excess of Ln cations in the formed complexes, the complexes were prepared in  $D_2O$  and the formations of the complexes were monitored by <sup>1</sup>H NMR. To a solution of ligand L (*ca.* 40 mg) in  $D_2O$ , a volume corresponding to half an equivalent of Ln cation of a stock solution of  $LnCl_3 \cdot xH_2O$  in  $D_2O$  was added. The <sup>1</sup>H NMR spectrum of the solution was recorded, and the exact volume corresponding to the missing half equivalent was added. A 0.1 M NaOD solution was added to raise the pD to 7 and the complete formation of the mononuclear complex was verified by <sup>1</sup>H NMR. Addition of HCl until pH = 3.5 resulted in the formation of a precipitate. Complete precipitation was achieved by addition of a THF/methanol mixture. The precipitate was isolated by centrifugation, dried under reduced pressure (2h, 40 °C), and purified (except for the Lu and Tb complexes) by flash column chromatography on a reverse phase C18 column (H<sub>2</sub>O/MeOH/TFA 0.1%) to give [LnL] complex as a white powder.

[**EuL**]. From 456 μL of EuCl<sub>3</sub>·6H<sub>2</sub>O at 0.11 M (50 mmol) and 40.0 mg (50 mmol) of ligand L, one obtains 35.1 mg of [EuL]· (CF<sub>3</sub>CO<sub>2</sub>H)<sub>2</sub>·2H<sub>2</sub>O (70%). <sup>1</sup>H NMR (D<sub>2</sub>O, pD = 7.0, 400 MHz):  $\delta$  8.42 (s, br, 3H), 7.53 (s, 3H), 7.30 (tr, br, 3H), 6.03 (d, br, <sup>3</sup>J = 7.0 Hz), 3.72 (s, br, 3H), 1.03 (s, br, 3H), -1.72 (s, br, 3H), -2.82 (s, br, 3H), -5.61 (s, br, 3H) ppm. ESI<sup>+</sup>/MS: *m/z* = 791.1 ([(EuL)+H]<sup>+</sup>, 85%), 793.1 ([(EuL)+H]<sup>+</sup>, 100%). <sup>19</sup>F NMR (D<sub>2</sub>O, 282 MHz):  $\delta$  -75.60 (s). Anal. Calcd for C<sub>24</sub>H<sub>30</sub>EuN<sub>6</sub>O<sub>9</sub>P<sub>3</sub>•2TFA•2H<sub>2</sub>O (C<sub>28</sub>H<sub>36</sub>EuF<sub>6</sub>N<sub>6</sub>O<sub>15</sub>P<sub>3</sub>): C, 31.86; H, 3.44; N, 7.96. Found: C, 31.67; H, 3.54; N, 8.21.

[YbL]. From 415  $\mu$ L (50 mmol) of a 0.11 M solution of YbCl<sub>3</sub>.6H<sub>2</sub>O and 40.0 mg (50 mmol) of ligand L, one obtains 22.6 mg (32%) of [YbL]·5(CF<sub>3</sub>CO<sub>2</sub>H)·2H<sub>2</sub>O. <sup>1</sup>H NMR (D<sub>2</sub>O, 300 MHz, pD = 7.87):  $\delta$  23.00 (s, br, 3H), 11.69 (s, br, 3H), 10.94 (s, br, 6H), 5.90 (s, br, 6H), -5.45 (s, br, 3H), -6.60 (s, br, 3H), -17.18 (s, br, 3H) ppm. <sup>19</sup>F NMR (D<sub>2</sub>O, 282 MHz):  $\delta$  -75.60 (s). Anal. Calcd for C<sub>24</sub>H<sub>30</sub>N<sub>6</sub>O<sub>9</sub>P<sub>3</sub>Yb•5TFA•2H<sub>2</sub>O (C<sub>34</sub>H<sub>39</sub>F<sub>15</sub>N<sub>6</sub>O<sub>21</sub>P<sub>3</sub>Yb): C, 28.79; H, 2.77; N, 5.92. Found: C, 29.20; H, 3.08; N, 5.79. ESI<sup>+</sup>/MS: *m*/*z* = 814.1 ([(YbL)+H]<sup>+</sup>, 100%).

**[LuL]**. From 944  $\mu$ L (51 mmol) of a 0.054 M solution of LuCl<sub>3</sub>.6H<sub>2</sub>O and 40.9 mg (51 mmol) of ligand L, one obtains 27.9 mg (44%) of [LuL]. <sup>1</sup>H NMR (D<sub>2</sub>O, 400 MHz, pD = 6.3):  $\delta$  7.98 (td, <sup>3</sup>J<sub>H-H</sub> = 7.6 Hz, <sup>4</sup>J<sub>P-H</sub> = 3.7 Hz, 3H, H<sub>4Pyr</sub>), 7.73 (t, <sup>3</sup>J<sub>H-H</sub> = 6.5 Hz, <sup>3</sup>J<sub>P-H</sub> = 6.5 Hz, 3H, H<sub>2Pyr</sub>), 7.52 (d, <sup>3</sup>J<sub>H-H</sub> = 7.5 Hz, 3H, H<sub>5pyc</sub>), 4.25 (AB system,  $\delta_{\rm A}$  = 4.70 ppm,  $\delta_{\rm B}$  = 3.81 ppm,  $J_{\rm AB}$  = 13.9 Hz, 6H), 3.37 (td, J = 12.8 Hz, J = 4.2 Hz, 3H), 2.75 (dd, J = 16.2 Hz, J = 5.2 Hz, 3H), 2.41 (dd, J = 12.9 Hz, J = 4.4 Hz, 3H), 2.27 (td, J = 13.8 Hz, J = 5.0 Hz, 3 H) ppm. ESI<sup>+</sup>/MS: m/z = 814.1 ([LuL-2H]<sup>+</sup>,100%); 837.1 ([LuL-3H+Na]<sup>+</sup>, 40%). <sup>31</sup>P NMR (D<sub>2</sub>O, 202 MHz, pD = 6.3):  $\delta$  12.86 ppm. Anal. Calcd for C<sub>24</sub>H<sub>30</sub>N<sub>6</sub>O<sub>9</sub>P<sub>3</sub>Lu·7NaCl·4H<sub>2</sub>O (C<sub>24</sub>H<sub>38</sub>Cl<sub>7</sub>LuN<sub>6</sub>Na<sub>7</sub>O<sub>13</sub>P<sub>3</sub>): C, 22.25; H, 2.96; N, 6.49. Found: C, 22.04; H, 3.05; N, 6.32.

[TbL]. 46.2 mg (123.7 mmol) of TbCl<sub>3</sub>.6H<sub>2</sub>O in D<sub>2</sub>O (2 mL) were added to 97.9 mg (122 mmol) of ligand. <sup>1</sup>H NMR (D<sub>2</sub>O, 400 MHz, pD = 7.96):  $\delta$  84.56 (s, br, 3 H), 32.84 (s, br, 3 H), 17.11 (s, br, 3 H), -5.48 (s, br, 3 H), -8.40 (s, br, 3 H), -9.77 (s, br, 6 H), -14.53 (s, br, 3 H), -70.05 (s, br, 3H). <sup>31</sup>P NMR (D<sub>2</sub>O, 202 MHz, pD = 7.96):  $\delta$  -94.37 (br). ESI<sup>-</sup>/MS: m/z = 398.02 ([M-2H]<sup>2-</sup>, 100%); 797.04 ([M-H]<sup>-</sup>, 74%). Anal. Calcd for C<sub>24</sub>H<sub>30</sub>N<sub>6</sub>O<sub>9</sub>P<sub>3</sub>Tb-3NaCl·7H<sub>2</sub>O (C<sub>24</sub>H<sub>44</sub>Cl<sub>3</sub>TbN<sub>6</sub>Na<sub>3</sub>O<sub>16</sub>P<sub>3</sub>): C, 26.65; H, 3.91; N, 7.77. Found: C, 26.68; H, 4.10; N, 7.78.

X-ray Crystallography. Crystals suitable for X-ray diffraction were obtained by slow evaporation of aqueous solutions. The crystals were placed in oil, and a single crystal was selected, mounted on a nylon loop, and placed in a low-temperature N<sub>2</sub> stream. X-ray diffraction data collection was carried out on a Bruker APEX II Kappa-CCD or a D8 VENTURE Bruker AXS diffractometer equipped with an Oxford Cryosystem liquid N<sub>2</sub> device, using Mo K $\alpha$  radiation ( $\lambda = 0.71073$  Å) at 150(2) K (Centre de diffractométrie X, Université de Rennes 1, France). The Bruker SMART program was used to refine the values of the cell parameters. Data reduction and correction absorption (SADABS) were carried out using the Bruker SAINT programs. The structures were solved by direct methods using the SIR97 program<sup>27</sup> and then refined with full-matrix least-squares methods based on F<sup>2</sup> (SHELX-97)<sup>28</sup> with the aid of the WINGX program.<sup>29</sup> All non-

hydrogen atoms were refined with anisotropic displacement parameters. For the [EuL] structure, the H atoms of phosphonate groups have been found in the electron density maps and refined as idealized hydroxyl group with torsion from electron density (AFIX 147 instruction). H atoms on the less agitated water molecules have been found in electron density map and refined with restraints on O---H (DFIX 0.84 0.01) and H.H (DANG 1.34 0.01). The four most agitated water molecules were squeezed. Their contribution to the calculated structure factors was estimated following the BYPASS algorithm,<sup>30</sup> implemented as the SQUEEZE option in PLATON.<sup>31</sup> The new data set, free of the four highly agitated water molecules contribution, was then used in the final refinement. For the [LaL] structure, H atoms on water molecules are found in electron density map and refined with restraints on O…H (DFIX 0.84 0.01) and H…H (DANG 1.34 0.01). All the other hydrogen atoms localized on carbon atoms were included in their calculated positions. Crystallographic data for structural analysis of the mononuclear Eu and of the La dinuclear complexes have been deposited with the Cambridge Crystallographic Data Centre under CCDC nos. 1589355 and 1589357, respectively. Copies of this information are available free of charge from the Web site (www.ccdc.cam.ac.uk).

Physicochemical and Spectrophotometric Studies. Materials. Distilled water was purified by passing through a mixed bed of ionexchanger (Bioblock Scientific R3-83002, M3-83006) and activated carbon (Bioblock Scientific ORC-83005). All the stock solutions were prepared by weighing solid products using an AG 245 Mettler Toledo analytical balance (precision 0.01 mg). Metal cation solutions were prepared from their chloride (LaCl<sub>3</sub>·7H<sub>2</sub>O, EuCl<sub>3</sub>·6H<sub>2</sub>O, TbCl<sub>3</sub>·6H<sub>2</sub>O, YbCl<sub>3</sub>·6H<sub>2</sub>O, LuCl<sub>3</sub>·6H<sub>2</sub>O, 99.9%, Sigma-Aldrich), and their concentrations were determined by colorimetric titrations with EDTA ( $10^{-2}$ M, Merck, Titriplex III) according to standard procedures, using xylenol orange as indicator.<sup>32</sup> Sodium hydroxide (NaOH) and hydrochloric acid (HCl) were used to adjust pH during titrations. All the experiments described were repeated at least two times. Spectroscopic measurements were performed with  $10 \times 10 \text{ mm}^2$ quartz Suprasil certified cells (Helma Analytics). UV/vis absorption spectra were recorded on a PerkinElmer lambda 950 or on a Specord spectrometer from Jena Analytics. Steady state emission spectra were recorded on an Edinburgh Instrument FLP920 spectrometer working with a continuous 450 W Xe Lamp and a red sensitive R928 photomultiplier from Hamamatsu in Pelletier housing for visible detection (230 to 900 nm) or a Hamamatsu R5 509-72 photomultiplier for the Vis-NIR part. All spectra were corrected for the instrumental functions. For emission spectra upon UV excitation, a 399 nm cutoff filter was used to eliminate second order artifacts. Phosphorescence lifetimes were measured on the same instrument working in the Multi-Channel Spectroscopy (MCS) mode, using a Xenon flash lamp as the excitation source. For short millisecond lifetimes, the intensity decay was corrected from the lamp intensity decay profile using a scattering solution of Ludox in water. Luminescence quantum yields were measured according to conventional procedures,33 with optically diluted solutions (optical density <0.05), using [Ru(bipy)<sub>3</sub>Cl<sub>3</sub>] in water ( $\Phi = 0.04$ )<sup>34</sup> as reference for Eu, a bipyridine Tb complex, [TbL(H<sub>2</sub>O)] in water ( $\Phi = 0.31$ )<sup>35</sup> as reference for Tb, and cardiogreen (IR125) in MeOH ( $\Phi = 0.078$ ) for Yb.<sup>36</sup> The errors are estimated to be 10% on the lifetimes and 15% on the luminescence quantum yields.

<sup>1</sup>*H* and <sup>31</sup>*P* M/*R* Titrations. Aliquots of a stock solution of the lanthanide trichloride salt in  $D_2O([Lu^{3+}] = 5.4 \times 10^{-2} \text{ M}, [Yb^{3+}] = 0.1 \text{ M}, [Eu^{3+}] = 0.1 \text{ M})$  were added to a ligand solution in  $D_2O$  at pD = 7 ([L] = 17 mM for Lu^{3+}, [L] = 6.9 mM for Yb^{3+} and 110 mM for Eu^{3+}). After each addition, pD was adjusted to a value of 7.0 with 0.1 M NaOD solution in  $D_2O$ . Changes after addition from 0 to 2.5 equiv of metal were monitored by recording the <sup>1</sup>H (400 MHz) and <sup>31</sup>P spectra (162 MHz), respectively. In the case of Lu^{3+}, diffusion spectroscopy (DOSY) NMR studies were carried out in order to measure the diffusion coefficient of the species present in solution. To this aim, a second titration was performed by adding 0, 0.5, 1.2, 1.6, and 2.0 equiv of a LuCl<sub>3</sub>·6H<sub>2</sub>O solution in  $D_2O$  (5.4 × 10<sup>-2</sup> M) to a ligand stock solution (0.021 M) at pD = 7.0.

Mass Spectrometry and lon Mobility Mass Spectrometry. Mass spectrometry and ion mobility experiments were performed on a QTOF mass spectrometer (Synapt G2, Waters, Manchester, UK), coupled to an automated chip-based nanoelectrospray device (Triversa Nanomate, Advion, Ithaca, U.S.A.) operating in the positive ion mode. The sampling cone and the extraction cone voltages were set to 40 and 4 V, respectively. Ions were efficiently thermalized with a constant trapping voltage of 7 V and subsequently transferred into the TOF analyzer.

Ion mobility (IM) experiments were performed under a constant pressure of  $N_2$  of 3.3 mbar in the traveling wave IM cell. A constant pressure of  $1.4 \times 10^{-3}$  mbar of He was applied to focus the ions prior to IM separation. The IM wave height and velocity were 40 V and 950 m/s, respectively. Transfer collision energy was set to 10 V to extract the ions from the IM cell to the TOF analyzer.

Spectrophotometric and Spectrofluorimetric Titrations. Titrations were performed by stepwise addition of microvolumes of a  $10^{-3}$  M stock solution of the lanthanide salt (LnCl<sub>3</sub>·6H<sub>2</sub>O, Ln = Eu, Tb, Yb) at pH 7.0 (Tris-HCl 0.1 M) to 2 mL of ligand solution (5 ×  $10^{-5}$  M in Tris-HCl 0.1 M at pH = 7.0) in 1 cm quartz Suprasil cell. After each addition, the UV–visible absorption spectra were recorded, together with the emission spectra upon excitation of the ligand at  $\lambda_{ex}$ = 267 nm. The software Hypspec V1.1.33 was used to determine the coordination model and calculate the stability constants (log  $\beta$ ) of the formed species.<sup>37</sup>

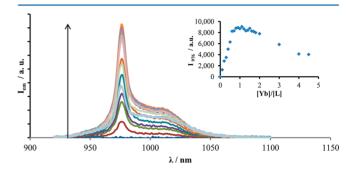
# RESULTS AND DISCUSSION

**Synthesis of Ligand L.** Ligand L was obtained in four synthetic steps starting from the commercially available 2-chloromethylpyridine 1 as depicted in Scheme 2.

The pyridine-*N*-oxide **2** was first obtained according to a literature procedure.<sup>26</sup> Diethyl (6-(chloromethyl)pyridin-2-yl)-phosphonate **3** was obtained by an adaptation of the procedure described for the parent pyridine,<sup>38</sup> using ethyl chloroformate and triethylphosphite in CH<sub>2</sub>Cl<sub>2</sub>. Alkylation of 1,4,7-triaza-cyclonane (TACN) in acetonitrile affords precursor **4**, which is hydrolyzed with 6 M hydrochloric acid to give ligand L as a hydrated trihydrochloride salt.

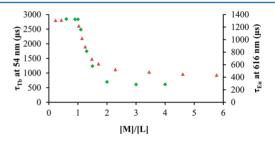
Studies of the Coordination Properties of Ligand L with  $Ln^{3+}$  lons (Ln = La, Eu, Tb, Yb, Lu). UV-Visible Absorption and Spectrofluorimetry. The coordination properties of L with Ln cations in water at pH = 7.0 (0.01 M Tris/ HCl) were investigated by monitoring the changes in UVvisible absorption and steady-state emission spectra upon addition of an increasing amount of metal. Considering that the various Ln complexes formed during the experiments can be present under a very large variety of protonation states but that we have not been able to unambiguously ascertain these later (for example by potentiometry experiments for which precipitation was observed), the protonation states of the complexes will not be specified, unless they are clearly defined (see for example the X-ray crystal structure of the Eu complex below). The absorption spectra of the ligand displayed broad bands in the UV region characteristic of  $\pi \to \pi^*$  transitions of the pyridinephosphonate moieties, with a maximum at 267 nm  $(\varepsilon_{267} = 14660 \text{ M}^{-1} \text{ cm}^{-1})$ . Addition of metal salts caused a hypochromic shift of the absorption spectra concomitant to a 3 nm bathochromic shift (see Figures S1-S3 for Ln = Yb, Tb, and Eu). Similar evolutions were observed along the Ln<sup>3+</sup> series, and in all cases, the presence of an inflection point near one equivalent of the added Ln<sup>3+</sup> suggested the formation of a mononuclear complex together with other polynuclear species. In the case of Yb<sup>3+</sup>, excitation in the ligand at 267 nm led to efficient sensitization of NIR Yb<sup>3+</sup>-centered luminescence by an antenna effect and observation of the characteristic  ${}^{2}F_{5/2} \rightarrow$ 

 ${}^{2}F_{7/2}$  emission band of Yb.<sup>39</sup> The luminescence intensity gradually increases up to *ca.* 1 equiv of metal added and then slowly decreases (Figure 1), which supports the formation of a mononuclear complex together with polynuclear species under these conditions.



**Figure 1.** Evolution of the emission spectra of a solution of L in 0.01 M Tris-HCl ( $4.0 \times 10^{-5}$  M, pH = 7.0,  $\lambda_{ex} = 267$  nm) upon addition of aliquots of YbCl<sub>3</sub>·6H<sub>2</sub>O ( $1.14 \times 10^{-3}$  M). Inset: evolution of the intensity at 976 nm as a function of the [Yb]/[L] ratio.

These observations are also supported by measurements recorded with  $Eu^{3+}$  and  $Tb^{3+}$  cations (see Figures S4–S5) and in particular by the evolution of the average luminescence lifetimes recorded upon addition of  $Eu^{3+}$  and  $Tb^{3+}$ , which significantly shorten for [M]/[L] ratios above unity (Figure 2).



**Figure 2.** Evolution of the average luminescence lifetimes ( $\lambda_{ex} = 267$  nm) measured for the titrations of a ligand solution by EuCl<sub>3</sub>·6H<sub>2</sub>O (red triangles,  $\lambda_{em} = 616$  nm) and TbCl<sub>3</sub>·6H<sub>2</sub>O (green diamonds,  $\lambda_{em} = 545$  nm) (in water, TRIS/HCl 0.01 M, pH = 7.0, [L] =  $5.32 \times 10^{-5}$  M for Eu and  $4.76 \times 10^{-5}$  M for Tb).

<sup>1</sup>H and <sup>31</sup>P NMR Studies. In order to elucidate the structure of the formed species, NMR titrations of ligand L with Ln =  $La^{3+}$ ,  $Eu^{3+}$ ,  $Yb^{3+}$ , and  $Lu^{3+}$  in  $D_2O$  (pD = 7.0, 25 °C) were performed. The <sup>1</sup>H NMR spectra of the ligand were recorded in the presence of 0 to 2.7 equiv of  $LuCl_3$ , and they are shown in Figure 3 (see Figure S6 for the full spectra). Between 0.1 and 0.9 equiv of  $Lu^{3+}$ , two sets of signals are clearly observed, which correspond to the free ligand and to the mononuclear [LuL] complex, respectively. After addition of 1 equiv of Lu<sup>3+</sup>, complete disappearing of the ligand peaks occurred, leaving a single set of 8 signals in which 3 multiplets in the 7.4-8.0 ppm region account for the aromatic protons, one signal is observed at 3.84 ppm for one of the two diastereotopic CH<sub>2</sub> protons of the pendant arm, and four spin systems in the 2.22-3.38 ppm region are assigned to the protons of the tacn skeleton. This pattern is in very good agreement with a rigid mononuclear complex with  $C_3$  symmetry. Evidence of the second methylene proton atom at 4.76 ppm, hidden by the peak of residual nondeuterated water, can be found by recording a  ${}^{1}H-{}^{1}H$ COSY experiment (Figure S7). Between 1.2 equiv and 1.8

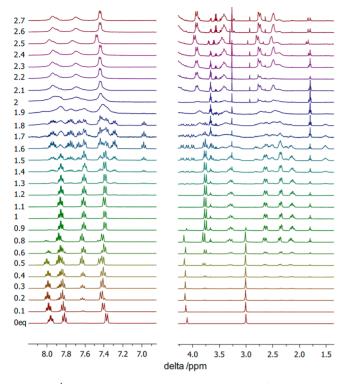
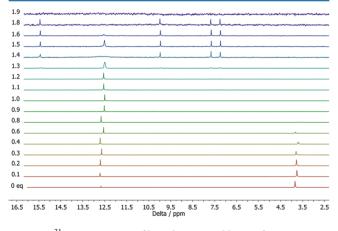


Figure 3. <sup>1</sup>H NMR spectra of ligand L upon addition of 0 to 2.7 equiv of LuCl<sub>3</sub>· $6H_2O$  (D<sub>2</sub>O, pD = 7, 400 MHz).

equiv of added  $Lu^{3+}$  in the solution, the peaks assigned to [LuL] disappeared and four sets of nine new protons were observed. Above 1.8 equiv, a broadening of the signals was observed, which was concomitant to the formation of a precipitate.

Further insights into the structure of the species formed were obtained by monitoring the same sample by  $^{31}$ P (Figure 4).



**Figure 4.** <sup>31</sup>P NMR spectra of ligand L upon addition of 0 to 1.9 equiv of  $LuCl_3 \cdot 6H_2O$  ( $D_2O$ , pD = 7.0, 162 MHz).

Upon addition of 0.1 to 0.9 equiv of LuCl<sub>3</sub>, a strong deshielding by *ca.* 9 ppm of the <sup>31</sup>P singlet was observed, due to the coordination of the phosphonate units. No changes occurred between 0.9 and 1.2 equiv of added Lu<sup>3+</sup> which, in line with previous observations by <sup>1</sup>H NMR, confirmed the large predominance of the monuclear [LuL] species at these stoichiometric ratios. Between 1.2 and 1.8 equiv of metal, the peak assigned to [LuL] progressively broadened and four new signals were observed, all having the same integral within experimental error (see Figure S8 for the integration of the spectra recorded after addition of 1.6 equiv), therefore suggesting either a unique species with four nonequivalent phosphorus atoms or alternatively an equimolar mixture of two species with two nonequivalent phosphorus atoms each, possibly two diastereoisomeric species. At 1.9 equiv and above, precipitation was observed together with the complete disappearing of the NMR peaks.

Further details on the intermediate species were obtained by recording the <sup>1</sup>H and <sup>31</sup>P DOSY (Diffusion Ordered Spectroscopy) spectra in the presence of 0.5, 1.2, 1.6, and 2.0 equiv of LuCl<sub>3</sub>, respectively. The corresponding figures are displayed in the Supporting Information. For each species, the diffusion coefficient was determined by using <sup>t</sup>BuOH as an internal reference. In the presence of 0.5 equiv of metal salt, two species were clearly identified and the diffusion coefficients amount to 283  $\mu$ m<sup>2</sup> s<sup>-1</sup> and 301  $\mu$ m<sup>2</sup> s<sup>-1</sup> for L and [LuL], respectively (Figure S9). Species with fast diffusion coefficients were also observed at 732  $\mu$ m<sup>2</sup> s<sup>-1</sup> and 1870  $\mu$ m<sup>2</sup> s<sup>-1</sup>, with a chemical shift corresponding to water protons. The faster species is in good agreement with the diffusion coefficient reported for HOD in D<sub>2</sub>O (1902  $\mu m^2 s^{-1}$ ),<sup>40</sup> and the other peak may correspond to water molecules in interaction with the second coordination sphere of the complex. As expected from <sup>1</sup>H NMR spectra, <sup>1</sup>H DOSY data in the presence of 1.2 and 1.6 equiv were rather complex; however, neat information can be obtained from <sup>31</sup>P NMR DOSY (Figure 5). In these conditions,

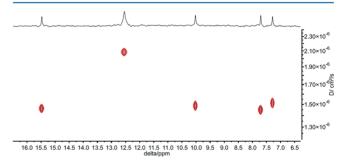


Figure 5. <sup>31</sup>P DOSY of a solution of L in the presence of 1.6 equiv of LuCl<sub>3</sub> in  $D_2O$  (25 °C, pD = 7).

three species could clearly be observed. The first species (with a singlet at 12.64 ppm) had a diffusion coefficient of 205  $\mu$ m<sup>2</sup> s<sup>-1</sup>, which is in good agreement with the value determined for [LuL] by <sup>1</sup>H DOSY. The two other species display very close diffusion coefficients at 145 and 150  $\mu$ m<sup>2</sup> s<sup>-1</sup>, respectively, which was indicative of the formation of adducts with larger sizes, each adduct giving rise to a pair of singlets in <sup>31</sup>P NMR. Hence, they were expected to have similar size and composition. One is characterized by two singlets at 10.00 and 7.26 ppm, respectively, and  $D = 150 \ \mu m^2 \ s^{-1}$ . The other gave rise to two singlets at 15.45 ppm and 7.68 pm and a diffusion coefficient  $D = 145 \ \mu m^2 \ s^{-1}$  (Figure 5). The hydrodynamic radii and volumes of the three species could be obtained from the Stockes-Einstein equation and are summarized in Table 1. From these data, the hydrodynamic volumes calculated for the larger species were 2.5 and 2.8 times larger than the volume determined for the [LuL] monomer. On the basis of the stoichiometric proportions of the compounds in solution  $([Lu^{3+}]/[L] = 1.5)$  and their hydrodynamic volumes, we postulated that the two sets of <sup>31</sup>P signals could be

Table 1. Diffusion Coefficients (*D*), Hydrodynamic Radii ( $R_{\rm H}$ ), Hydrodynamic Volumes ( $V_{\rm H}$ ), the Corresponding Chemical Shifts ( $\delta$ ), and the Postulated Composition of the Species as Obtained from <sup>31</sup>P NMR DOSY for a Solution of L in the Presence of 1.6 equiv of LuCl<sub>3</sub> in D<sub>2</sub>O (25 °C, pD = 7)

$\delta$ $^{31}$ P (ppm)	$D (\mu m^2/s)$	$R_{\rm H}$ (Å)	$V_{\rm H}$ (Å <sup>3</sup> )	Species (M:L)
3.75	301	5.80	817	0:1
12.54	205	8.51	2582	1:1
7.29/10.2	150	11.6	6538	3:2
7.69/15.48	145	12.0	7328	3:2

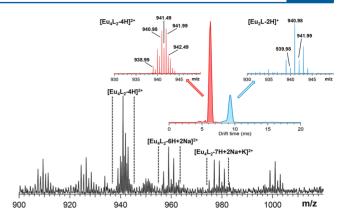
attributed to two species with a  $[Lu_3L_2]$  composition in agreement with the possibility of two diastereoisomers.

At 2.0 equiv, broadening of the signal occurred and the determination of the diffusion coefficients was no longer possible, presumably because of the formation of aggregates of higher M:L stoichiometry.

Variable temperature NMR experiments were also performed on a sample with [M]/[L] = 1.6. <sup>1</sup>H and <sup>31</sup>P spectra from 25 to 80 °C are displayed in Figure S10 and S11, respectively. Significant sharpening of the <sup>1</sup>H NMR signals was clearly observed upon increasing the temperature, thereby suggesting the occurrence of dynamic phenomena such as chemical exchange and/or rearrangement leading to the progressive loss of the four <sup>31</sup>P signals.

<sup>1</sup>H and <sup>31</sup>P NMR titrations were also performed with LaCl<sub>3</sub>, YbCl<sub>3</sub>, and EuCl<sub>3</sub>, and the corresponding spectra are displayed in Figures S12-S17. In all cases, the formation of a mononuclear [LnL] complex with rigid  $C_3$  symmetry was clearly observed and reached its maximum concentration upon addition of 1 equiv of metal salt. However, different behaviors were observed in excess of lanthanide ion. Addition of 1.2 equiv of YbCl<sub>3</sub> resulted in the formation of a nonsymmetrical  $[Ln_3L_2]$ secondary species which became predominant at 1.6 equiv of added metal and which was characterized by two sets of 9 peaks in the <sup>1</sup>H NMR spectrum (Figure S12). At 2.0 equiv, a third species started to grow and above, broadening of the signals was observed concomitantly to the formation of a precipitate in the tube. In the case of  $Eu^{3+}$  and  $La^{3+}$ , precipitation started to occur as soon as 1.5 equiv of metal salts were added, also suggesting the formation of new species with lower solubility.

Mass Spectrometry Analysis. In order to enlighten the composition of the polynuclear species, a similar titration of the ligand with 0 to 5 equiv of EuCl<sub>3</sub> in water (ammonium acetate 0.01 M, pH = 7.0) was monitored by ESI-mass spectrometry. Mass spectra (in positive mode) are presented in the Supporting Information. From 0 to 1 equiv (Figure S18), the ligand peaks at 643.16 m/z ([M + H]<sup>+</sup>), 665.15 m/z ([M + Na]<sup>+</sup>), and 681.11 m/z ([M+K]<sup>+</sup>) decreased to form a pattern with an isotopic peak at 791.07 m/z, which corresponds to the [EuL-2H]<sup>+</sup> species with the corresponding isotopic distribution related to <sup>151</sup>Eu(48%) and <sup>153</sup>Eu(52%). Above 1 equiv and under the same instrumental conditions, the mononuclear complex remains the major species (Figure S19), possibly as a result of the fragmentation of the polynuclear adducts in the source, which renders the titration little informative. However, a detailed examination of the spectra recorded in the presence of 1.2 equiv and above, indicated the presence of a series of peaks which could be attributed to polymetallic species (Figure 6).



**Figure 6.** ESI-MS spectrum of a solution of L in water with 5 equiv of  $EuCl_3 \cdot 6H_2O$  (AcONH<sub>4</sub> 0.01 M, pH = 7). *Inset.* Arrival time distributions obtained by ion mobility for the peak at 938.99 m/z and the corresponding isotopic patterns of each peak.

Ion mobility mass spectrometry experiments on the same sample were used to separate the ions at 940.97, 958.98, and 978.95 m/z as a function of their size. Ion mobility arrival time distributions are presented in Figure 6 (inset) and in Figures S20–S21. For each peak, two contributions could be clearly evidenced, which correspond to  $[Eu_4L_2]^{2+}$  ( $[Eu_4L_2-4H]^{2+}$  at 938.99 m/z,  $[Eu_4L_2-6H+2Na]^{2+}$  at 957.00 m/z,  $[Eu_4L_2-7H +2Na+K]^{2+}$  at 976.97 m/z) and  $[Eu_2L]^+$  ( $[Eu_2L-2H]^+$  at 938.99 m/z,  $[Eu_2L-2H+H_2O]^+$  at 957.00 m/z,  $[Eu_2L-3H+Na+H_2O]^+$  at 976.97 m/z) adducts. However, at this stage it was difficult to conclude whether both species were simultaneously present in solution or if aggregation of the  $[Eu_2L]^+$  species/or dissociation of the  $[Eu_4L_2]^{2+}$  species occurred in the source.

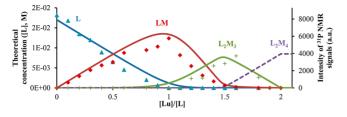
Speciation and Thermodynamic Stability Constants. In light of these observations, a global analysis of the spectrofluorimetric titration experiments was performed with the Hypspec program by implementing a model with three species with M:L stoichiometric ratios of 1:1, 3:2, and 4:2. In both cases, such a model can nicely reproduce the evolution of the emitted luminescence intensity as a function of the M/L ratios (see Figures S22–S24). The overall stability constants determined for Eu<sup>3+</sup> and Tb<sup>3+</sup> complexes are summarized in Table 2. The calculated emission spectra of the ML,  $M_3L_2$ , and

Table 2. Overall Stability Constants of Eu<sup>3+</sup> and Tb<sup>3+</sup> Complexes with Ligand L ([L]  $\approx 5 \times 10^{-5}$  M, TRIS/HCl 0.01 M, pH = 7.0),  $\lambda_{ex}$ = 267 nm

Species (M <sub>x</sub> L <sub>y</sub> )	$\log \beta_{Eu}$	$\log \beta_{Tb}$
1:1	12.9	12.9
3:2	38.6	36.6
4:2	46	42

 $M_4L_2$  species (M = Eu, Tb), together with the evolution of the concentrations of the species formed are displayed in Figures S23–S25.

Assuming a minor variation of the thermodynamic stability constant along the lanthanide series, this model was used to analyze NMR data recorded with Lu<sup>3+</sup>. In particular, Figure 7 represents a comparison between the theoretical distribution species (full lines) and the intensity of the experimental <sup>31</sup>P NMR signals (points) and shows a striking correlation between <sup>31</sup>P NMR data and the previous model obtained from spectrofluorimetric titrations.



**Figure** 7. Theoretical representation of the concentration of the species formed (with log  $\beta_{1:1} = 12.9$ , log  $\beta_{3:2} = 38.6$ , log  $\beta_{4:2} = 46$ , full lines) and evolution of the intensity measured by <sup>31</sup>P NMR for the three species: L (blue triangles), LM (red diamonds), L<sub>2</sub>M<sub>3</sub> (green plus signs) upon titration of ligand L with LuCl<sub>3</sub>.6H<sub>2</sub>O (D<sub>2</sub>O, pD = 7.0, 162 MHz). The M<sub>4</sub>L<sub>2</sub> species was observed by ESI-MS and spectrofluorimetric titrations.

This analysis also endorses our initial attribution of the two sets of two peaks observed in the <sup>31</sup>P NMR spectra between 1.3 and 1.8 equiv of Lu added, to two diastereoisomers with a 3:2 M:L stoichiometry and  $C_3$  symmetry. In conclusion, although the identification of the polynuclear species was not straightforward, a global analysis by NMR, mass spectrometry, and spectrofluorimetric experiments converged toward the evidence of the formation [Ln<sub>3</sub>L<sub>2</sub>], [Ln<sub>4</sub>L<sub>2</sub>] polynuclear species.

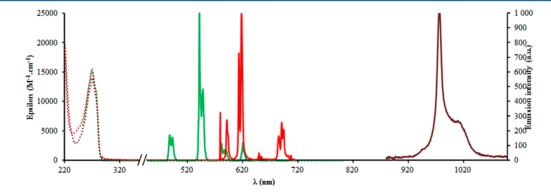
Synthesis and Spectroscopic Characterization of the [LnL] Complexes. Considering the high stability of the [LnL] complexes, it was envisaged to isolate the mononuclear complexes of Eu<sup>3+</sup>, Tb<sup>3+</sup>, Yb<sup>3+</sup>, and Lu<sup>3+</sup>. In order to determine the optimal conditions for their synthesis and purification, the influence of the pH on a solution containing stoichiometric amounts of L and EuCl<sub>3</sub>·6H<sub>2</sub>O was first monitored by recording the Eu<sup>3+</sup> emission spectra upon excitation in the ligand absorption band (Figure S26). This titration indicated that complete formation of the complex occurred above pH =4.45 and once formed, the [EuL] complex remained stable in the pH range from 12.04 to 1.98. Hence, the formation of the desired mononuclear complexes was achieved at neutral pH whereas their precipitation and purification was accomplished under slightly acidic conditions (pH = 3.5), in order to avoid the formation of polynuclear complexes. The complexes were further purified by reverse phase chromatography on C18 and analyzed by elemental analysis and ESI mass spectrometry, showing expected isotopic patterns for [LnL-2H]<sup>+</sup> species (Figures S27-S29). The complexes were further characterized by <sup>1</sup>H and <sup>31</sup>P NMR spectroscopy, except for Tb<sup>3+</sup>, each one giving a set of 9 signals in <sup>1</sup>H NMR (Figures S30-S32) and a

singlet at 12.51 ppm, -8.71 ppm, and 2.51 ppm, for the <sup>31</sup>P NMR spectra for Lu, Eu, and Yb, respectively.

Finally, the spectroscopic properties of the complexes were measured in water and D<sub>2</sub>O and the corresponding absorption and emission spectra are presented in Figure 8. Additional parameters are presented in Table 3, and three points have attracted our attention. At first, strictly monoexponential luminescence decays confirm the presence of unique species in solution. Second, the absence of inner-sphere water molecule endorses the nonadentate coordination of the ligand and the good protection of the metal inside the cavity. Third, the luminescence properties of the Yb complex are rather interesting in terms of lifetimes and quantum yields. In particular, the lifetime measured in deuterated water is rather long in comparison to typical values obtained for efficient Yb<sup>3+</sup> emitters<sup>41-46</sup> and longer than the one previously reported for the tacn-based [Yb(thqtcn-SO<sub>3</sub>)] (8.63  $\mu$ s).<sup>14</sup> However, this value remains significantly shorter than the values obtained for YbL<sup>2</sup> and YbL<sup>2D</sup> (>30  $\mu$ s in D<sub>2</sub>O)<sup>8</sup> and other per-deuterated complexes in organic solvents.47-49

Surprisingly, despite the excellent shielding of the cations from the solvent molecules (q  $\approx 0$  for all complexes),<sup>50</sup> the luminescence quantum yields of the Eu complex were rather modest compared to those obtained with ligands possessing carboxy pyridyl arms (picolinates)<sup>51,52</sup> or phosphinate ones.<sup>53</sup> Considering the good shielding and the long luminescence lifetimes, it is surmised that these poor luminescence quantum yields are to be linked with a modest ligand to metal energy transfer.

Structural Studies of the Mono- and Polynuclear **Species.** While isolating the mononuclear complexes, single crystals suitable for X-ray diffraction analysis were obtained for [EuL], which was isolated in acidic conditions as a mixture of [EuL] and [EuLH]Cl species. The ORTEP diagram is represented in Figure 9. Selected parameters, distances, and angles are presented in Tables S1-S2. The metal ion is ninecoordinated by three nitrogen atoms from the tacn ring, three nitrogen atoms from the aromatic groups, and three oxygen atoms from the phosphonate functions with a pseudo- $C_3$ symmetry. The pyridinephosphonate pendant arms bind the lanthanide in a helical fashion and both enantiomers ( $\Delta$  and  $\Lambda$ ) are present in the solid state structures, therefore leading to centrosymmetric space groups  $(P2_1/a)$ . Similar structures have previously been collected for tacn-based ligands with pyridinecarboxylate,<sup>13</sup> 8-hydroxyquinoline pendant arms,<sup>14</sup> and 6-carboxy-2,2'-bipyridine arms.<sup>12</sup>



**Figure 8.** UV/vis absorption (dotted lines) and normalized emission spectra ( $\lambda_{ex} = 265$  nm, straight lines) of a D<sub>2</sub>O solution of [LnL] with Ln = Yb (brown), Tb (green), and Eu (red).

Table 3. Main Spectroscopic	Parameters of the YbL	, TbL, EuL, and LuL	Complexes in Water and	nd $D_{3}O$ at pH = 7.0
1 1		, , ,	1	2 1

	$\varepsilon_{\mathrm{H_2O}}~(\mathrm{M^{-1}cm^{-1}})^a$	$\varepsilon_{\mathrm{D_2O}} \ (\mathrm{M^{-1}cm^{-1}})^a$	$\phi_{\mathrm{H_{2}O}}~(\%)^{b}$	$\phi_{ ext{D}_2 ext{O}} \ (\%)^{m{b}}$	$ au_{ m H_2O}~(\mu  m s)$	$ au_{\mathrm{D_2O}}~(\mu\mathrm{s})$	$q^{c}$
YbL	13050	13300	0.15	0.42	2.8	10.2	0.0
TbL	14880	15450	51	52	2890	2980	-0.2
EuL	12260	13870	2	8	1252	1375	-0.2
LuL	12610						

 ${}^{a}\lambda$  = 265 nm.  ${}^{b}\phi$  measured relative to [Ru(bipy)<sub>3</sub>Cl<sub>3</sub>]<sup>34</sup> for Eu, to [TbL(H<sub>2</sub>O)]<sup>35</sup> for Tb, and to IR125<sup>36</sup> for Yb. <sup>c</sup>Calculated according to ref 50.

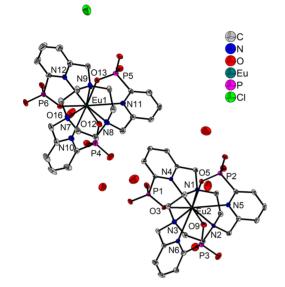


Figure 9. ORTEP diagram of the complexes [EuL], [EuLH]Cl with thermal ellipsoids at 50% probability.

Interestingly, single crystals could be obtained upon cooling a water solution of ligand L containing 1.5 equiv of lanthanum chloride, originally heated at 80 °C. The crystal structure of the  $[(LaL)La(H_2O)_9]$  species is represented in Figure 10.

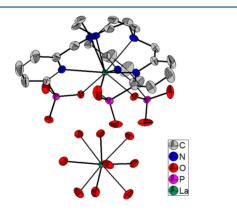


Figure 10. Solid state structure of the  $[(LaL)La(H_2O)_9]$  complex (ellipsoids drawn at 50% probability).

Structural parameters and selected distances and angles are summarized in Tables S1 and S3. The structure is composed of a nonacoordinated  $[LaL]^{3-}$  mononuclear complex in which the  $La^{3+}$  cation is coordinated inside the cavity of the ligand as for the mononuclear [EuL] complex (noted La1) together with a nonahydrated  $[La(H_2O)]^{3+}$  complex (noted La2). Considering the  $C_3$  symmetry of [La1L], the position of the phosphonate allows us to define two kind of interactions between La1 and La2, a head to head (HH) one in which the phosphonate functions of [La1L] point toward La2 and a head to tail (HT) interaction in which La2 faces the triazacyclononane macrocycles. The two motives are repeated along the *c* axis, at distances of 5.716 Å (HH) and 8.461 Å (HT) between the two lanthanum ions (Figure 11). In a column, all [La1L] complexes

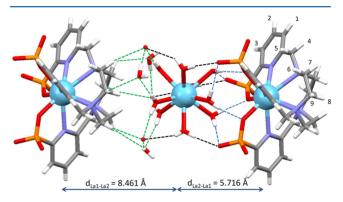


Figure 11. Representation of the hydrogen bonding interactions observed in the crystal structure of the  $[(LaL)La(H_2O)_9]$  complexes (see text for the color code of H bonding interactions) and numbering scheme of the H atoms used for the interpretation of the NMR spectrum.

display the same chirality ( $\Delta$  or  $\Lambda$ ). This chirality is inverted between two adjacent columns. Within the two entities in a HH conformation, the three water molecules coordinated to the hydrated La2 atom and pointing toward the [La1L] complex are held by a H-bond network in which the two proton atoms of a water molecule are linked to one oxygen atom of a phosphonate function coordinated to La1 and to an oxygen atom not directly coordinated to La1 of an adjacent phosphonate function (blue dashed lines in Figure 11).

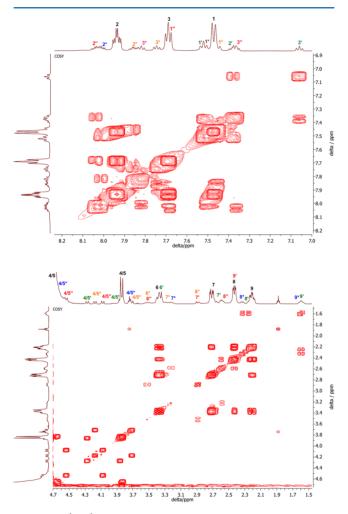
The three water molecules coordinated in the equatorial plane of La2 (relative to the pseudo- $C_3$  axis) are also hydrogen bonded to the three oxygen atoms of the phosphonate functions of the neighboring [La1L] complex (black dashed lines in Figure 11). The other H atoms of these water molecules are bonded to an oxygen atom of a phosphonate function of a [La1L] complex from an adjacent column (isolated oxygen atoms in red in Figure 11).

Weak interactions are also observed between  $[La(H_2O)_9]$ and a neighboring [La1L] complex in the HT configuration. These interactions are relayed by three water molecules or by the oxygen atom of a phosphonate function of a [La1L]complex in a neighboring column (green dashed lines in Figure 11).

We hypothesized that this strong H bonding network may be responsible for the high stability of the  $[Lu_3L_2]$  species observed by NMR and spectroscopy in solution. For a trinuclear complex in the THH configuration ([(LuL)Lu-(LuL)] as schematized in Figure 11), one would expect the observation of two peaks in the <sup>31</sup>P NMR, depending on the HH or HT interaction between the central Lu atom and the

LuL complex. If one now assumes that the chirality of the LuL complexes in the trinuclear complexes can be exchanged for one of the LuL complexes, two pairs of peaks would be expected for the  $[\Delta(LuL)Lu\Delta(LuL)]$  and  $[\Delta(LuL)Lu\Lambda(LuL)]$  diastereoisomeric species.

To further confirm this hypothesis, a mixture containing 1.4 equiv of LuCl<sub>3</sub>·6H<sub>2</sub>O per ligand L was analyzed by <sup>1</sup>H, COSY, NOESY, and ROESY NMR experiments. The attribution of the signals is presented in Figure 12. In this spectrum, five families



**Figure 12.**  ${}^{1}\text{H}-{}^{1}\text{H}$  COSY spectra of ligand L upon addition of 1.4 equiv of LuCl<sub>3</sub>.6H<sub>2</sub>O (D<sub>2</sub>O, pD = 7.0, 400 MHz) and peak assignment (see numbering in Figure 11).

of 9 signals were observed and their attribution could be partially performed by looking at  ${}^{1}H{-}{}^{1}H$  correlations in the COSY experiment (Figure 12). The first family (1–9, in black) corresponds to the mononuclear [LuL] complex. A nuclear Overhauser effect was observed between H1 and H4/5 (see Figure 11 for H numbering); however, dipole–dipole interactions between other species could not be observed. Their attribution was possible with the examination of the ROESY spectrum, based on the 1<sup>x</sup>, 4/5<sup>x</sup> correlations (x = ', '', °, Figure S33). No correlation could clearly be distinguished for the fourth family (1\*–9\*, in blue). Another striking point in the examination of the NOESY spectrum (Figure S34) is the observation of exchange correlations between, on the one hand, the two protons 4 and 5 corresponding to the AB system and on the other hand, between proton atoms from the

triazacyclononane backbone such as 7\* and 7', 7° and 7, or 8" and 8. Such exchanges are typical of  $\Delta \leftrightarrow \Lambda$  interconversion processes associated with the concerted rotation of the pendant arms in  $C_3$  symmetric LnL complexes. However, this observation is particularly surprising for two reasons. The  $\Delta$  $\leftrightarrow \Lambda$  interconversion process could not be observed in the isolated [LuL] complex, even by heating the solution up to 80 °C in D<sub>2</sub>O and is not observed in the NOESY spectrum for the AB spin system of H4 and H5 protons of the remaining [LuL] complex. Then, if such an isomerization has already been observed for lanthanide complexes with 1,4,7-triazacyclononane derivatives such as H4nota<sup>54</sup> and H3bpatcn,<sup>55</sup> they always correspond to ligands with decreased denticity (CN = 6 for H4nota and 8 for H3bpatcn), whereas isomerization was not reported for tacn ligands with functionalized pyridines displaying CN = 9. On the opposite, enantiopure Eu and Tb complexes with trispyridylphosphinate triazacyclononane could be isolated, either using chiral HPLC<sup>17</sup> or directly synthesized thanks to the inclusion of a stereogenic center on the tcn ring.<sup>56</sup>

A possible explanation of the observed exchange is that the interconversion occurs between the diastereomeric pairs, exchanging the  $[\Delta(LuL)Lu\Delta(LuL)]$  into the  $[\Lambda(LuL)Lu\Lambda$ -(LuL)] (and similarly with  $[\Lambda(LuL)Lu\Delta(LuL)]$  and  $[\Delta(LuL)-$ LuA(LuL)]). Considering that the  $\Delta \leftrightarrow \Lambda$  interconversion of [LuL] is slow on the NMR time scale, the exchange between the diastereomeric pairs would require a dissociation process. Unfortunately, within a dyad such as depicted in Figure 11, distances between the two mononuclear LuL complexes are too high to expect any correlations by 2D NOESY NMR. Interestingly, <sup>1</sup>H NMR spectra of mixed species have been recorded by addition of 1 equiv of LuCl<sub>3</sub> on the mononulclear [YbL] complex and by addition of 1 equiv of YbCl<sub>3</sub> and [LuL] (Figure S35), and they indicate no decoordination and no ion exchange at room temperature and upon heating up to 80 °C, therefore confirming the formation of kinetically inert complexes.

# CONCLUSION

In conclusion, the assembly of pyridinephosphonate units into the nonadentate tripodal ligand L allows the synthesis of highly stable monomeric lanthanide complexes that are isostructural along the series. Moreover, such complexes are highly stable in water and display luminescent properties, due to an efficient sensitization of Eu, Tb, and Yb ions by the pyridinephosphonate antenna. More importantly, such a scaffold is responsible for relatively low vibrational quenching, as emphasized by the very good luminescence properties of the Yb complex in D<sub>2</sub>O  $(\tau_{D2O} = 10.2 \ \mu s \text{ and } \phi_{D2O} = 0.42\%)$ . Furthermore, using a combination of UV-absorption spectrometry, spectrofluorimetry, <sup>1</sup>H and <sup>31</sup>P NMR, DOSY, ESI-mass spectrometry, and Xray diffraction, we demonstrated that this nonadentate ligand is a good building block for the formation of highly stable polynuclar complexes of  $[(LnL)_2Ln_x]$  stoichiometry (x = 1 and x = 2). Such architectures are very interesting for studying intermetallic communication and energy transfer within heteropolynuclear species and can potentially lead to very appealing photon up conversion properties. Future works are engaged in this direction.

# ASSOCIATED CONTENT

#### **S** Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.inorg-chem.8b00666.

Evolution of the UV-visible absorption and emission spectra of a solution of L upon addition of Yb<sup>3+</sup>, Tb<sup>3+</sup>, and Eu<sup>3+</sup> (Figures S1-S5); <sup>1</sup>H NMR, <sup>31</sup>P NMR, and <sup>1</sup>H DOSY experiments with Lu:L mixtures (Figures S6-S11); <sup>1</sup>H and <sup>31</sup>P NMR titrations of ligand L with Yb<sup>3+</sup> (Figures S12-S13), Eu<sup>3+</sup> (Figures S14-S15), and La<sup>3+</sup> (Figures S16–S17); ESI-MS titrations of L with Eu<sup>3+</sup> and ionic mobility chromatograms (Figures S18-S21); calculated spectra and species distribution diagrams for the spectrofluorimetric titrations of L with Eu<sup>3+</sup> (Figures S22-S23) and Tb<sup>3+</sup> (Figures S24-S25); pH titration of a solution of Eu:L 1:1 (Figure S26); ESI-MS and <sup>1</sup>H NMR spectra of isolated [EuL] (Figures S27-S31); [LuL] (Figures S28-S30); [YbL] (Figures S29-S32); crystallographic data for [EuL] and [LaL] structures (Tables S1-S3); 2D <sup>1H</sup> NMR spectra of a L:Lu 1:1.4 mixture (Figures S33-S34) (PDF)

# **Accession Codes**

CCDC 1589355 and 1589357 contain the supplementary crystallographic data for this paper. These data can be obtained free of charge via www.ccdc.cam.ac.uk/data\_request/cif, or by emailing data\_request@ccdc.cam.ac.uk, or by contacting The Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge CB2 1EZ, UK; fax: +44 1223 336033.

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#### Notes

The authors declare no competing financial interest.

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