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A critical crosscatalytic relationship determines the outcome of competition in a replicator network

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Abstract

A network of two synthetic replicators exhibits a critical unidirectional crosscatalytic relationship that directs competing replication processes. In this network, nitrene **N** bearing a 6-methylamidopyridine recognition site can participate in two 1,3-dipolar cycloaddition reactions with two maleimides that differ in the relative position of their carboxylic acid recognition site — either *para* (**M^p**) or *meta* (**M^m**) relative to the maleimide ring. These two cycloaddition reactions create replicators *trans*-**T^p** and *trans*-**T^m**. In isolation, *trans*-**T^p** templates its own formation with an efficiency that is markedly greater than that of *trans*-**T^m**. Kinetic fitting and simulations reveal that this efficiency arises from a higher template-mediated rate constant for the cycloaddition and lower stability of the *trans*-**T^p** template duplex, compared to *trans*-**T^m**. By contrast, in a situation where **M^p** and **M^m** compete for a limited quantity of **N**, the normally less efficient *trans*-**T^m** outcompetes *trans*-**T^p**. Through a series of comprehensive kinetic ¹⁹F{¹H} NMR spectroscopy experiments, this system-level outcome is traced to a critical crosscatalytic pathway, whereby the presence of *trans*-**T^p** templates the formation of *trans*-**T^m**, but not *vice versa*. Replicator *trans*-**T^m** also reduces the efficiency of its competitor *trans*-**T^p** by sequestering *trans*-**T^p** in a heteroduplex that is more stable than homoduplex [**T^p**•**T^p**]. The addition of different templates as instructions reveals that, while the outcome of competition between replicators can be altered selectively, it is limited by the reaction environment employed. These results represent a conceptual and practical framework for the examination of selectivity in replication networks operating outside well-stirred batch reactor conditions.

Introduction

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4 Networks are the building blocks of the world around us, and their inherent interconnectedness¹ often
5 exerts an influence over our everyday lives that can be challenging to predict. The all-pervasive
6 presence of networks in nature² is reflected in the number of disciplines currently engaged^{1c,3} in the
7 study of complex systems^{3a,4} and the properties that emerge through the interactions of their
8 components. Chemistry experienced a paradigm shift around twenty years ago as it moved away from
9 the study of molecular matter in isolation and, instead, began to embrace the notion of complexity and
10 networks. By embracing the nascent field⁵ of systems chemistry, bottom-up approaches for the design,
11 development, and investigation of synthetic chemical models for complex systems have started to
12 appear. A phenomenon of particular interest within the field of systems chemistry is replication. In the
13 context of complex systems, synthetic replicators present a unique opportunity to study these processes
14 using networks constructed from molecules with well-defined structures and with catalytic and
15 recognition properties that can be probed and characterized in detail experimentally. The developments
16 in the field of systems chemistry to date have produced a remarkable variety of replicating systems
17 based on oligonucleotides,⁶ peptides,⁷ and small synthetic molecules,⁸ highlighting and demonstrating
18 that the ability to replicate is not exclusive to biological systems based on nucleic acids and equipped
19 with extensive enzymatic machinery. In addition to self-replication, these systems were shown to
20 express a number of system-level properties such as error-correction,⁹ stereo-specific replication,¹⁰ and
21 Boolean logic.¹¹ Peptide-based replicating systems have, in particular, achieved a significant level of
22 sophistication, examining networks^{9,10,11c,12} comprising more than a single replicator—a feature
23 significantly less well explored in replicating systems based on oligonucleotides¹³ and small-organic
24 molecules.¹⁴ Although the requirements for the operation of minimal self-replicating systems in
25 isolation have been established¹⁵ firmly, the processes in complex networks in the real world do not
26 operate in isolation. For this reason, in this work, we build on our experience in developing self-
27 replicating systems based on small organic molecules^{8a,8c,8d,8f,8g,14a-c} to design and analyze a network of
28 two replicators, connected directly through a shared building block. This network presents a model
29 system that allows us to study how reaction and recognition-mediated processes impact and govern the
30 preference, *i.e.*, the selectivity, of the network for one replicator over another.
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Design and methodology

In order to create a minimal self-replicating system, two components, bearing complementary recognition sites, must be connected together by one or more covalent bonds to afford a template that can act as a specific autocatalyst for its own formation. Once formed, this autocatalytic template can pre-organize the building blocks required for its formation in a ternary complex co-conformation that renders their reaction pseudointramolecular. The bond-forming step produces a template dimer, or duplex, which exist in equilibrium with the catalytically active monomeric template.

Here, a network of two replicators is constructed from two maleimides, \mathbf{M}^p and \mathbf{M}^m , and a single nitron $\mathbf{N}^{[Ref. 14c]}$ (**Figure 1a**, Building blocks). The main distinguishing feature of these two maleimides and the replicators¹⁶ $\mathbf{T}^p^{[Ref. 14c]}$ and \mathbf{T}^m (**Figure 1a**, Replicators) arising from their 1,3-dipolar cycloaddition reactions with \mathbf{N} , is in the location of the carboxylic acid recognition site on the maleimide component (**Figure 1**). In particular, \mathbf{M}^p integrates^{8a,8c,14a-c} a phenylacetic acid in position *para* relative to the maleimide ring, while \mathbf{M}^m incorporates a benzoic acid functionality, in position *meta* with respect to the maleimide ring. For this reason, we use superscript labels *p* and *m* to represent the *para* and *meta* positions of the carboxylic acid recognition site in maleimides (\mathbf{M}) and templates (\mathbf{T}) throughout this work. Initially, these two templates are formed *via* slow, template-independent bimolecular pathways (**Figure 1c**), typically furnishing two diastereoisomeric products, labeled¹⁶ *trans* and *cis*, in a 3:1 ratio in the absence effects originating from molecular recognition. Once the quantity of template formed is sufficient to allow its association with the unreacted components in a catalytically active ternary complex, $[\mathbf{N}\cdot\mathbf{M}^p\cdot\mathbf{T}^p]$ or $[\mathbf{N}\cdot\mathbf{M}^m\cdot\mathbf{T}^m]$ (**Figure 1b**), the template formation proceeds with significant acceleration *via* the autocatalytic cycle. The ability of a template to initiate an autocatalytic cycle depends on the strength of the non-covalent recognition-mediated processes (**Figure 1d**) that govern the recognition between the network components. For example, a replicator with a stronger¹⁷ recognition process would typically require a lower template concentration to enable self-replication to proceed.

Replicators \mathbf{T}^p and \mathbf{T}^m possess a recognition unit complementary to the carboxylic acid, namely, a 6-methyl-2-amidopyridine ring that is originally associated with nitron \mathbf{N} (**Figure 1a**). The relative positions and identities of recognition sites are conserved in the replicator templates. We envisaged that the slight differences in the type and location of the carboxylic acid recognition elements in each system would produce a reaction network of two replicators, referred to as the \mathbf{T}^p - \mathbf{T}^m network, with the potential for crosscatalytic behavior. Hence, reaction of the three individual components together allows the formation of a network where both templates can form simultaneously (**Figure 1b**) through two auto- and two crosscatalytic pathways. The ability of the templates \mathbf{T}^p and \mathbf{T}^m

to act as either auto- or crosscatalytic templates allows the network to be instructed to form one replicator over another by the addition of preformed T^p or T^m . Ultimately, the ability of our replicators to compete for the shared nitron building block will depend on their ability to take part in these auto- and crosscatalytic cycles and for this reason, we wished to undertake a comprehensive kinetic analysis of this small network constructed from two self-replicators to develop an understanding of how structural changes alter the selectivity for one replicator over another and their role in driving system-level behavior.

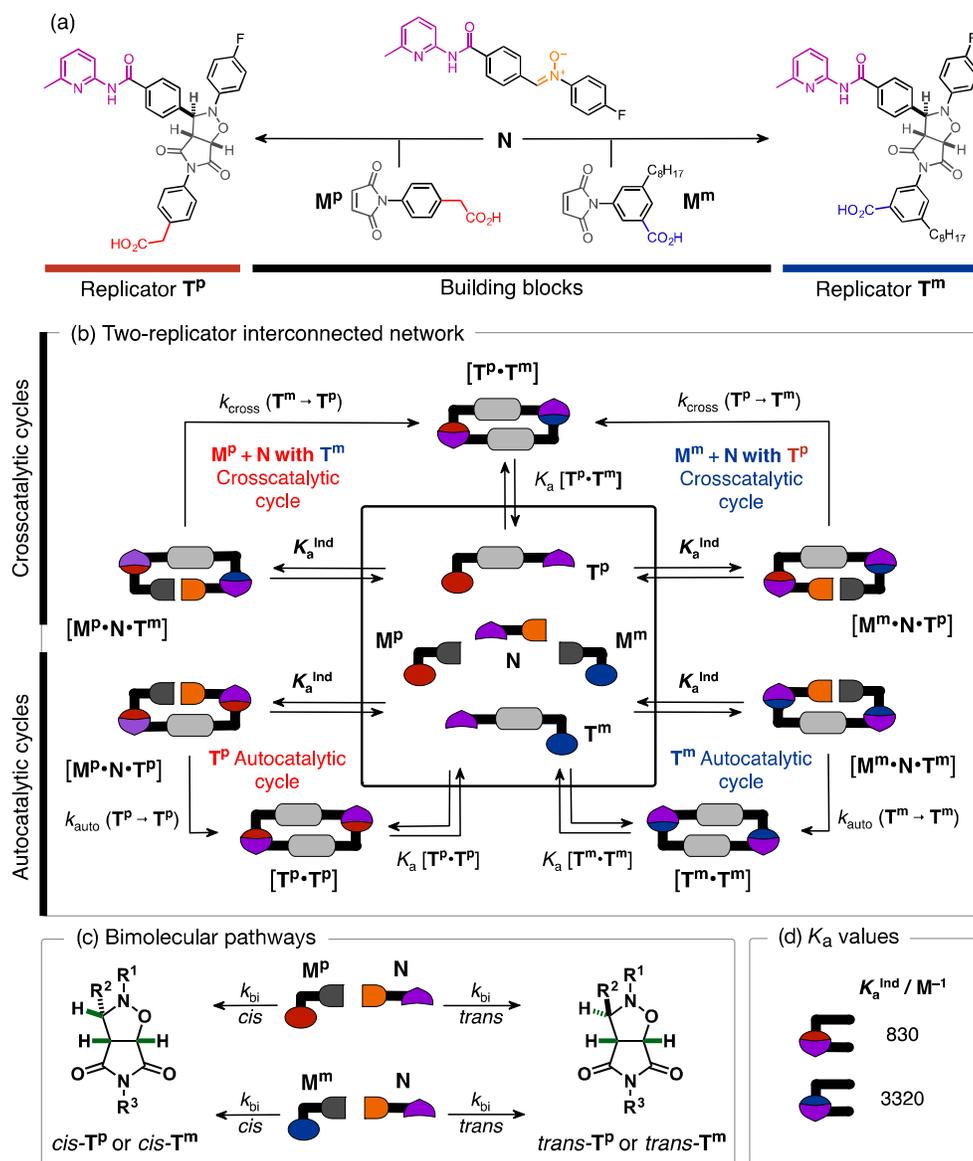


Figure 1. (a) A network of two replicators, T^p and T^m , can be constructed using the 1,3-dipolar cycloaddition reactions of two maleimides, M^p and M^m , and nitron N . Superscript labels p and m represent the *para* and *meta* location of the carboxylic acid recognition site. (b) Maleimides M^p and M^m can react with nitron N to form two replicators, T^p and T^m (central square). Each of these templates can theoretically take part in auto- and crosscatalytic pathways. (c) Maleimides M^p and M^m can react with nitron N also *via* template-independent pathways, resulting in the formation of catalytically active *trans* diastereoisomers and catalytically inert *cis* diastereoisomers of T^p and T^m . (d) The recognition between the network components is governed by specific association constants.

Results and discussion

The first step in the analysis of the T^P - T^m network is the examination of individual bimolecular recognition-independent pathways leading to T^P and T^m . Accordingly, we examined (Figure 2) the reaction of nitrone **N** with two control, recognition-disabled maleimides, M^{PC} and M^{mC} . Details of the synthetic procedures utilized in the syntheses of all reaction components are provided in the Supporting Information. The kinetic analyses of all reaction pathways, facilitated by the presence of the aryl fluorine atom present in nitrone **N**, and all **N**-derived cycloadducts, were performed using 470.3 MHz $^{19}\text{F}\{^1\text{H}\}$ NMR spectroscopy. In all kinetic experiments reported in this study, one or both of the maleimides and the nitrone ($[\text{M}] = [\text{N}] = 5 \text{ mM}$) were allowed to react in CDCl_3 at 5°C , and the reaction progress was monitored by recording 470.3 MHz $^{19}\text{F}\{^1\text{H}\}$ NMR spectra every 30 minutes over a period of 16 h. The concentrations of the reaction components were determined at each time point relative to 1-bromo-2-fluoro-4-nitrobenzene, which was present as an internal standard.

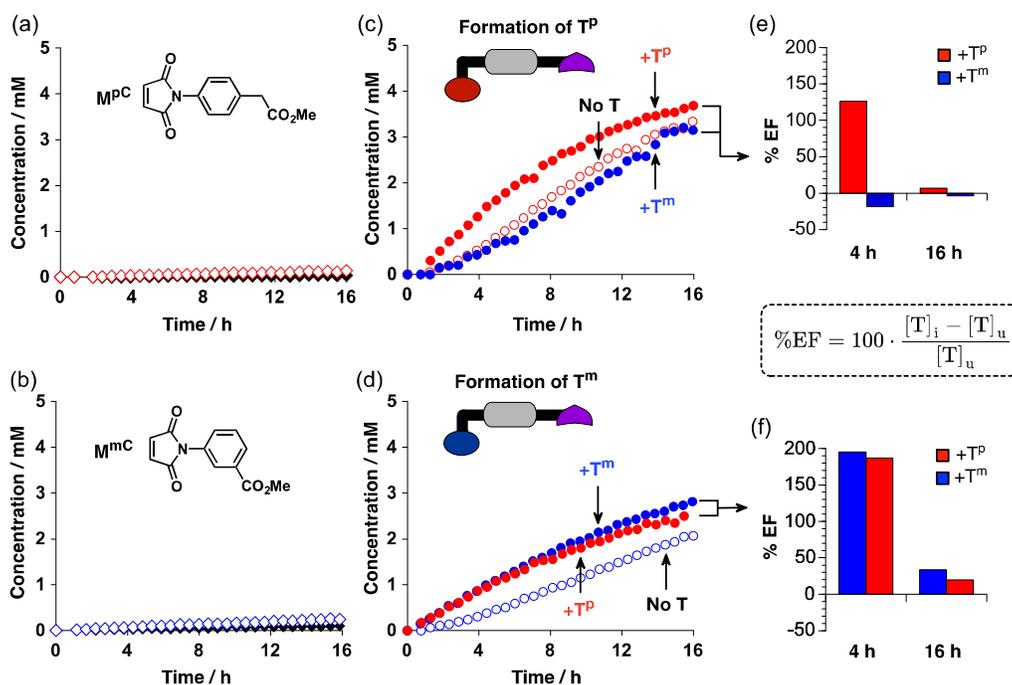


Figure 2. Kinetic experiments examining the reaction of **N** with recognition-disabled maleimides (a) M^{PC} , and (b) M^{mC} , and recognition-enabled maleimides (c) M^P , and (d) M^m , as determined by 470.3 MHz $^{19}\text{F}\{^1\text{H}\}$ NMR spectroscopy relative to 1-bromo-2-fluoro-4-nitrobenzene as an internal standard. Formation of recognition-disabled cycloadducts *trans*- T^{PC} (empty red diamonds), *trans*- T^{mC} (empty blue diamonds) and the corresponding *cis* cycloadducts (filled black diamonds) in (a) and (b) proceeds slowly and unselectively. (c) Formation of replicator *trans*- T^P (no *cis* product was detected) was examined in the absence of template (empty red circles), and in the presence of *trans*- T^P (filled red circles) and *trans*- T^m (filled blue circles). (d) Formation of replicator *trans*- T^m (*cis*- T^m formed $<0.5 \text{ mM}$ in each case, data omitted for clarity) was examined in the absence of template (empty blue circles) and in the presence of *trans*- T^m (filled blue circles) and *trans*- T^P (filled red circles). Graphs in (e) and (f) show the percentage enhancement factors (%EFs) determined in template-instructed ($[T]_i$) experiments (filled symbols in (c) and (d)) relative to the corresponding uninstructed experiments ($[T]_u$) (empty symbols in (c) and (d)). Reaction conditions: [all reactants] = 5 mM, 5°C , CDCl_3 , 16 h. In each case, instructing template was added at 20 mol% (1 mM). Note that the vertical scale in (a) to (d) is identical in order to facilitate direct comparison of reaction conversions.

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3 The control maleimides do not possess the carboxylic acid recognition site required for successful
4 recognition of the amidopyridine unit on nitron component **N** and the absence of recognition-mediated
5 reactivity is clearly evident in the reaction profiles determined for the reaction of **M^{pC}** with **N** (**Figure**
6 **2a**), and **M^{mC}** with **N** (**Figure 2b**). The formation of both recognition-disabled *trans* and *cis*
7 diastereoisomers of **T^{pC}** and **T^{mC}** proceeded very slowly. After 4 h, *trans*-**T^{pC}** reached 0.03 mM
8 (**Figure 3a**), while *cis*-**T^{pC}** was below the limit of detection at this time. After 16 h, the concentration
9 of these two diastereoisomers increased to 0.14 mM and 0.04 mM for *trans* and *cis*, respectively
10 ([*trans*]/[*cis*] = 3.8). By contrast, the formation of **T^{mC}** proceeded slightly more efficiently, and the
11 *trans* and *cis* cycloadducts reached concentrations of 0.07 mM and 0.02 mM, respectively, after 4 h. At
12 $t = 16$ h, their concentration increased further to 0.24 mM and 0.10 mM ([*trans*]/[*cis*] = 2.5)—almost
13 twice as high as those determined for **T^{pC}**. These results demonstrated that (i) in the absence of the
14 carboxylic acid recognition site, the formation of templates can proceed only *via* the slow template-
15 independent pathways, and (ii) formation of **T^{mC}** proceeds noticeably faster than **T^{pC}**, suggesting that
16 replicator **T^m** might be able to exploit the template-mediated cycle at an earlier time point in the
17 reaction than **T^p**.
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Next, we examined the formation of replicators **T^p** (**Figure 2c**, empty red symbols) and **T^m** (**Figure 2d**, empty blue symbols), from their constituent components. Both reaction profiles are sigmoidal, demonstrating the striking effect of recognition-mediated replication processes on product formation. At $t = 4$ h, replicator **T^p** was formed at a concentration of 0.53 mM, with complete diastereoselectivity for the *trans* cycloadduct (*i.e.*, *cis* < limit of detection by $^{19}\text{F}\{^1\text{H}\}$ NMR spectroscopy). This concentration increased further to 3.34 mM after 16 h. The reaction that forms **T^m** was shown to be less diastereoselective, affording *trans*-**T^m** and *cis*-**T^m** at 0.30 mM and 0.13 mM after 4 h, respectively. After 16 h, the concentration of these cycloadducts increased to 2.10 mM and 0.40 mM. The ratio of the two diastereoisomer products ([*trans*]/[*cis*]) increased from 2.3 at 4 h to 5.2 after 16 h.

In the next stage of our analysis of this system, the formation of each replicator was examined in the presence of 20 mol% (1 mM) of the respective *trans* template (*e.g.*, **T^p** added to **M^p** and **N**) in order to test whether **T^p** and **T^m** possess the capacity to template their own formation. The reaction profile for the formation of **T^p** in the presence of **T^p** (**Figure 2c**, filled red symbols) and **T^m** in the presence of **T^m** (**Figure 2d**, filled blue symbols) revealed disappearance of the lag period in each case, thereby confirming the abilities of **T^p** and **T^m** to self-replicate. In the presence of the autocatalytic template **T^p**, formation of **T^p** proceeded with significantly increased efficiency, reaching a

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2 concentration of 1.26 mM after 4 h, and 3.69 mM after 16 h. As expected, the addition of preformed
3 template induces the most striking enhancement in the concentration of replicator at early reaction
4 times. The enhancement in the rate of formation of *trans*-**T^m** was, in contrast, less pronounced. The
5 concentration of *trans*-**T^m** reached 0.86 mM and 2.82 mM after 4 and 16 h, respectively. Catalytically-
6 inert *cis*-**T^m** reached a concentration of 0.09 mM and 0.32 mM after 4 and 16 h, giving [*trans*]/[*cis*]
7 ratios of 10.1 and 8.1, respectively.
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10 Thus far, the kinetic analyses confirmed that both **T^P** and **T^m** are capable of self-replication.
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12 Next, we examined the two crosscatalytic pathways—**T^m** acting as a template for the formation of **T^P**
13 (**Figure 2c**, filled blue symbols) and *vice versa* (**Figure 2d**, filled red symbols). To this end, each set of
14 reactants was instructed with 20 mol% of the corresponding crosscatalytic template. Interestingly, the
15 addition of **T^m** to the reactants required for the formation of **T^P** resulted in no observable enhancement
16 in the concentration of **T^P**. In fact, no change in the lag period for **T^P** formation was observed and the
17 reaction profile determined in the presence of **T^m** was essentially identical to that obtained in the
18 absence of instruction. In terms of concentration, cycloadduct *trans*-**T^P** reached 0.43 mM after 4 h—
19 similar to that (0.53 mM) reached in the absence of any instructing template. After 16 h, this
20 concentration increased to 3.15 mM. Strikingly, however, instructing the building blocks required for
21 the formation of **T^m** with template **T^P** resulted in the unambiguous disappearance of the lag period,
22 demonstrating that **T^P** *can* template the formation of *trans*-**T^m**. After 4 h, *trans*-**T^m** and *cis*-**T^m** were
23 formed at concentrations of 0.87 mM and 0.08 mM, respectively. These concentrations increased
24 further after 16 h, reaching 2.52 mM and 0.22 mM, respectively ([*trans*]/[*cis*] ratios of 10.3 and 11.4).
25 The slight increase in diastereoselectivity for the *trans* product in the presence of preformed **T^P**
26 compared to **T^m** suggests that template **T^P**, which locates the two recognition sites at a slightly different
27 separation and geometry with respect to each other than **T^m**, can promote the formation of the *trans*
28 diastereoisomer more effectively than **T^m**.
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45 In order to quantify the changes in replication efficiency observed in the template-instructed
46 experiments relative to those lacking any instruction, we calculated the percentage enhancement
47 factor¹⁸ (%EF) for each *trans* product determined after both 4 h and 16 h (**Figure 2**). Examination of
48 these results for **T^P** (**Figure 2e**) and **T^m** (**Figure 2f**) reveals a number of trends. Firstly, the %EFs are
49 significantly higher after 4 h than after 16 h, reflecting the fact that the presence of preformed template
50 exerts a stronger effect at earlier reaction times when the overall concentration of catalytically active
51 template within the uninstructed reaction mixture is typically low. Further, **Figure 2e** shows that the
52 addition of **T^P** to the building blocks required for its own formation enhanced its formation
53 significantly: 138 ± 7% at 4 h, and 10 ± 1% at 16 h. In contrast, the addition of **T^m** to the same mixture
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of building blocks resulted in a suppression of \mathbf{T}^{P} formation (4 h = $-19 \pm 5\%$, 16 h = $-6 \pm 1\%$) relative to the uninstructed reaction. This observation suggests that \mathbf{T}^{P} might be sequestered within the $[\mathbf{T}^{\text{P}}\cdot\mathbf{T}^{\text{m}}]$ heteroduplex, which reduces the quantity of free template available for reaction. The same analysis of the enhancements determined for \mathbf{T}^{m} reveals that the addition of autocatalytic template \mathbf{T}^{m} (4 h = $185 \pm 15\%$, 16 h = $36 \pm 2\%$) affords a marginally higher enhancement than the addition of crosscatalytic template \mathbf{T}^{P} (4 h = $187 \pm 16\%$, 16 h = $22 \pm 1\%$). Overall, a comparison of the EFs showed that the addition of preformed template produces a more significant change in the formation of \mathbf{T}^{m} than in the formation of \mathbf{T}^{P} . Most likely, this difference can be attributed to the fact that replicator \mathbf{T}^{m} is less efficient at making itself than \mathbf{T}^{P} in the absence of template and is therefore affected less by the decreasing availability of the reagents required for its formation at longer reaction times. In summary, the comprehensive evaluation of all auto- and crosscatalytic pathways available to \mathbf{T}^{P} and \mathbf{T}^{m} confirmed that only three catalytic channels out of the four possible operate efficiently.

Kinetic fitting and simulations

In order to develop a better understanding of the differences in the kinetic behaviors of \mathbf{T}^{P} and \mathbf{T}^{m} in terms of the key kinetic and thermodynamic parameters that characterize the auto- and crosscatalytic cycles, we fitted the reaction time courses shown in **Figure 3** to appropriate kinetic models (see Supporting Information). These kinetic models describe the interactions and reactions leading to the formation of \mathbf{T}^{P} or \mathbf{T}^{m} and using established protocols,^{8a,13b,14a} we were able to determine the bimolecular reaction rate constants (k_{bi}) for the formation of both *trans* and *cis* cycloadducts, the unimolecular rate constants (k_{auto} , k_{cross}), and duplex association constants ($K_{\text{a}}^{\text{Duplex}}$) pertaining to all auto- and crosscatalytic pathways (**Table 1**).

Table 1. Overview of rate and duplex association constants determined using SimFit^{13b} for the bimolecular and recognition-mediated auto- and crosscatalytic pathways of replicators *trans*- \mathbf{T}^{P} and *trans*- \mathbf{T}^{m} . The rate constants for the formation of catalytically inactive *cis* diastereoisomers of \mathbf{T}^{P} and \mathbf{T}^{m} through the corresponding bimolecular template-independent pathways were determined to be 0.431 and $0.704 \cdot 10^{-4} \text{ M}^{-1} \text{ s}^{-1}$, respectively.

	Reactions	k	$\text{EM}_{\text{kinetic}} / \text{M}$
Bimolecular $k_{\text{bi}} / 10^{-4} \text{ M}^{-1} \text{ s}^{-1}$	$\text{N} + \text{M}^{\text{P}} \rightarrow \text{trans-}\mathbf{T}^{\text{P}}$	1.12	—
	$\text{N} + \text{M}^{\text{m}} \rightarrow \text{trans-}\mathbf{T}^{\text{m}}$	1.79	—
Autocatalytic $k_{\text{auto}} / 10^{-4} \text{ s}^{-1}$	$[\text{N}\cdot\text{M}^{\text{P}}\cdot\text{trans-}\mathbf{T}^{\text{P}}] \rightarrow [\text{trans-}\mathbf{T}^{\text{P}}\cdot\text{trans-}\mathbf{T}^{\text{P}}]$	71.6	64.0
	$[\text{N}\cdot\text{M}^{\text{m}}\cdot\text{trans-}\mathbf{T}^{\text{m}}] \rightarrow [\text{trans-}\mathbf{T}^{\text{m}}\cdot\text{trans-}\mathbf{T}^{\text{m}}]$	17.0	9.47
Crosscatalytic $k_{\text{cross}} / 10^{-4} \text{ s}^{-1}$	$[\text{N}\cdot\text{M}^{\text{P}}\cdot\text{trans-}\mathbf{T}^{\text{m}}] \rightarrow [\text{trans-}\mathbf{T}^{\text{P}}\cdot\text{trans-}\mathbf{T}^{\text{m}}]$	4.67	4.21
	$[\text{N}\cdot\text{M}^{\text{m}}\cdot\text{trans-}\mathbf{T}^{\text{P}}] \rightarrow [\text{trans-}\mathbf{T}^{\text{P}}\cdot\text{trans-}\mathbf{T}^{\text{m}}]$	20.3	18.3

	Equilibria	K_a	EM_{thermo} / M
Binary $K_a^{\text{Ind}} / M^{-1}$	$N + M^p \leftrightarrow [N \cdot M^p]$	830	—
	$N + M^m \leftrightarrow [N \cdot M^m]$	3320	—
Template $K_a^{\text{Duplex}} / 10^6 M^{-1}$	$trans\text{-}T^p + trans\text{-}T^p \leftrightarrow [trans\text{-}T^p \cdot trans\text{-}T^p]$	17.9	26.0
	$trans\text{-}T^m + trans\text{-}T^m \leftrightarrow [trans\text{-}T^m \cdot trans\text{-}T^m]$	68.6	6.22
	$trans\text{-}T^p + trans\text{-}T^m \leftrightarrow [trans\text{-}T^p \cdot trans\text{-}T^m]$	35.0	12.7

These parameters, together with association values for the formation of $[M \cdot N]$ complexes (K_a^{Ind}) were subsequently used to calculate the kinetic effective molarity¹⁹ (EM_{kinetic}) and thermodynamic effective molarity²⁰ (EM_{thermo}) for each replication pathway. Examination of the parameters shown in **Table 1** reveals that both *trans* and *cis* diastereoisomeric products of T^m are formed with a higher bimolecular rate constant their T^p counterparts. The initial advantage provided to T^m replicator as a result of the higher bimolecular rate (T^m should reach the concentration required for initiating autocatalysis earlier than T^p) is countered by the lower k_{auto} for the formation of T^m . Comparison of the kinetic profiles and values of EM_{kinetic} determined for T^p and T^m confirmed that, in isolation, T^p is significantly more efficient at directing its own formation than T^m . The fitting procedure allowed the determination of the association constants²¹ for replicator homoduplexes for *trans*- T^p ($17.9 \times 10^6 M^{-1}$) and *trans*- T^m ($68.6 \times 10^6 M^{-1}$), respectively. The higher stability of the $[T^m \cdot T^m]$ duplex relative to the $[T^p \cdot T^p]$ duplex indicates that T^m is hindered more significantly by product inhibition than T^p . Interestingly, while T^m is associated with a higher duplex stability, it possesses lower EM_{thermo} than T^p . This observation suggests a lower degree of complementarity and, hence, cooperativity within the T^m duplex.

The fitting procedure employed in the determination of kinetic parameters pertaining to the two crosscatalytic pathways required the introduction of an additional parameter—*i.e.*, the association constant for the formation of heteroduplex $[T^p \cdot T^m]$. This heteroduplex contains one more rotatable bond than $[T^m \cdot T^m]$ and one rotatable bond less than $[T^p \cdot T^p]$ and therefore, its stability should be bracketed by the stabilities of the two homoduplexes. Applying rotor increments, as described^{18a} by Page and Jencks, the stability of $[T^p \cdot T^m]$ was estimated as $35.0 \times 10^6 M^{-1}$, and this value was employed in the determination of k_{cross} for the formation of each *trans* product on the corresponding crosscatalytic template (**Table 1**).

The parameters obtained through the kinetic fitting of crosscatalytic kinetic data allowed us to assess the differences observed in the corresponding time course profiles of both replicators. Specifically, T^m was found to be significantly worse at catalyzing the formation of T^p than T^p was at catalyzing its own formation. In fact, the EM_{kinetic} for this crosscatalytic pathway was *ca.* $15 \times$ smaller than that determined for the autocatalytic pathway leading to T^p . By contrast, the EM_{kinetic} for the

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2 crosscatalytic pathway that involves \mathbf{T}^m being made on \mathbf{T}^p was actually somewhat higher (18.3 M) than
3 the corresponding autocatalytic process (9.47 M). These differences in the relative efficiencies of the
4 auto- and crosscatalytic pathways directed by \mathbf{T}^p and \mathbf{T}^m can explain the absence of a template effect
5 for \mathbf{T}^p being synthesized on a \mathbf{T}^m template.
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9 An important consequence of the order of stabilities determined for the duplexes in the full
10 network ($[\mathbf{T}^m \cdot \mathbf{T}^m] > [\mathbf{T}^p \cdot \mathbf{T}^m] > [\mathbf{T}^p \cdot \mathbf{T}^p]$), is the preferential incorporation of \mathbf{T}^p within the more stable
11 heteroduplex $[\mathbf{T}^p \cdot \mathbf{T}^m]$ in situations where \mathbf{T}^m is present at significant concentrations. The kinetic data
12 derived from NMR spectroscopy reveals that the exchange between the various template species
13 present in solution is fast on the NMR chemical shift timescale (*i.e.*, only a single resonance arising
14 from the *trans* isoxazolidine proton is observed for each replicator). For this reason, we exploited
15 kinetic simulations and the relevant reaction parameters obtained through fitting to demonstrate how
16 the presence of \mathbf{T}^m serves to decrease the amount of catalytically active \mathbf{T}^p in solution. Specifically,
17 early in the reaction where \mathbf{T}^p is being synthesized in the presence of preformed \mathbf{T}^m (**Figure 3a**), the
18 higher stability of the $[\mathbf{T}^p \cdot \mathbf{T}^m]$ heteroduplex relative to that of the $[\mathbf{T}^p \cdot \mathbf{T}^p]$ homoduplex leads to the
19 template \mathbf{T}^p formed within the system being sequestered into the more stable heteroduplex. The higher
20 proportion of \mathbf{T}^p in the $[\mathbf{T}^p \cdot \mathbf{T}^m]$ heteroduplex than in the $[\mathbf{T}^p \cdot \mathbf{T}^p]$ homoduplex reduces the catalytic
21 availability of this template and, also, the overall rate of production of \mathbf{T}^p because the $[\mathbf{T}^p \cdot \mathbf{T}^m]$
22 heteroduplex has a lower K_d . At a certain point in the reaction, however, the concentration of \mathbf{T}^p
23 reaches a critical threshold and the fraction of \mathbf{T}^p bound in the $[\mathbf{T}^p \cdot \mathbf{T}^p]$ homoduplex becomes higher
24 than the fraction present within the $[\mathbf{T}^p \cdot \mathbf{T}^m]$ heteroduplex.
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37 By contrast, this order of stabilities exerts the opposite effect on the crosscatalytic pathway
38 where the formation of \mathbf{T}^m is simulated in the presence of \mathbf{T}^p (**Figure 3b**). In this situation, *trans*- \mathbf{T}^p is
39 the dominant product early in the reaction and therefore, the molecules of \mathbf{T}^m formed through the
40 crosscatalytic pathway are present predominantly in the form of the heteroduplex $[\mathbf{T}^p \cdot \mathbf{T}^m]$. This
41 heteroduplex is less stable than the $[\mathbf{T}^m \cdot \mathbf{T}^m]$ duplex that is present exclusively in the absence of \mathbf{T}^p ,
42 resulting in higher concentrations of monomeric of \mathbf{T}^m at early time points. Nevertheless, the affinity of
43 \mathbf{T}^p for \mathbf{M}^m and \mathbf{N} is lower than the affinity of \mathbf{T}^m for these components, and as the concentration of \mathbf{T}^m
44 continues to increase, homoduplex $[\mathbf{T}^m \cdot \mathbf{T}^m]$ becomes the dominant species in solution, and the
45 efficiency of the crosscatalytic pathway ($\mathbf{T}^p \rightarrow \mathbf{T}^m$) decreases.
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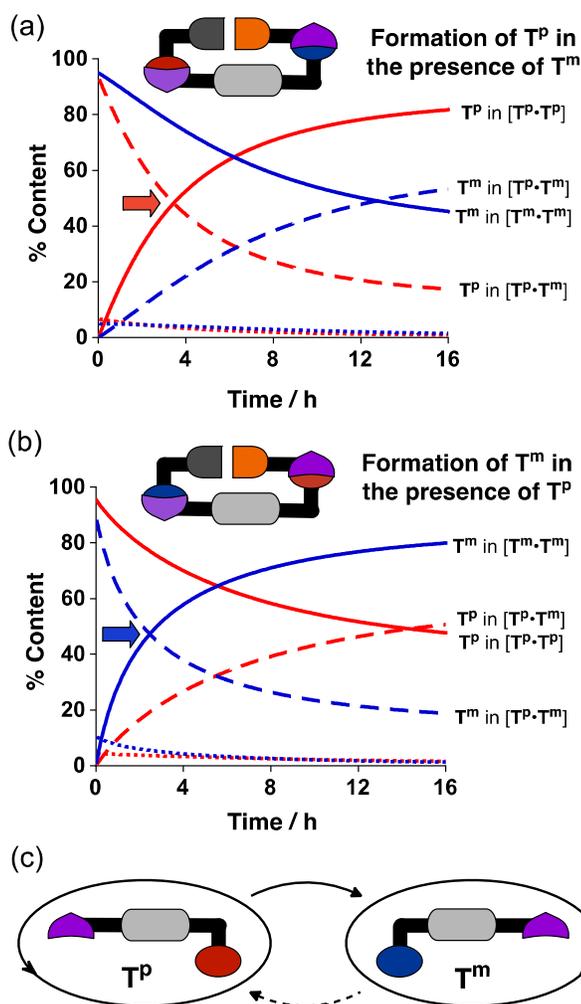


Figure 3. Simulations predict the distribution of templates T^m and T^P in homo- (full lines) and heteroduplexes (dashed lines) in experiments where the reagents required for the formation of (a) T^P are instructed with 20 mol% of *trans*- T^m (1 mM), and (b) the reagents for the formation of T^m are instructed with 20 mol% of *trans*- T^P (1 mM). Templates T^P and T^m present within the various categories are represented in red and blue, respectively. Dotted lines represent the % content of T^P and T^m in their monomeric form or in other binary complexes. Red and blue arrows highlight the time point in the reaction at which the % content of the homoduplex of the template being formed is higher than its content in the corresponding heteroduplex. (c) Schematic overview of the catalytic relationship between replicators T^P and T^m . Dashed line signifies absence of effective catalysis.

Kinetic analysis of competition between replicators

Kinetic analyses of our two replicators in isolation established clearly that T^P possesses the capacity to template both its own formation and to crosscatalyze the formation of T^m , while T^m is capable of autocatalytic activity only. Building on these results, we examined these replicators under conditions where M^P and M^m compete for the nitrene N building block. In order to explore the possibility of directing the reaction network towards selective formation of either T^P or T^m , we employed a series of template-instructed experiments. In each of these experiments, an equimolar solution (5 mM) containing all three reaction components, N , M^P , and M^m , was prepared in $CDCl_3$ and the formation of the replicators was monitored at 5 °C in the absence of any added template using 470.3 MHz $^{19}F\{^1H\}$

NMR spectroscopy (see Supporting Information, **Figure S8**). The instructed experiments examined the formation of T^P and T^m in the presence of 20 mol% of one or both preformed replicators. A number of scenarios were examined — (i) only *trans*- T^P added, (ii) only *trans*- T^m added and (iii) both *trans*- T^P and *trans*- T^m added simultaneously (see Supporting Information, **Figure S8**). The results of these experiments are summarized in **Figure 4**. Interestingly, the uninstructed competition experiment revealed that, as a result of the interplay between the recognition and reaction processes taking place simultaneously in the network, the less efficient template *trans*- T^m reached a higher concentration than *trans*- T^P , the replicator found to be more efficient in isolation ($[trans-T^m]/[trans-T^P] = 2.0$ after 4 h, **Figure 4a**).

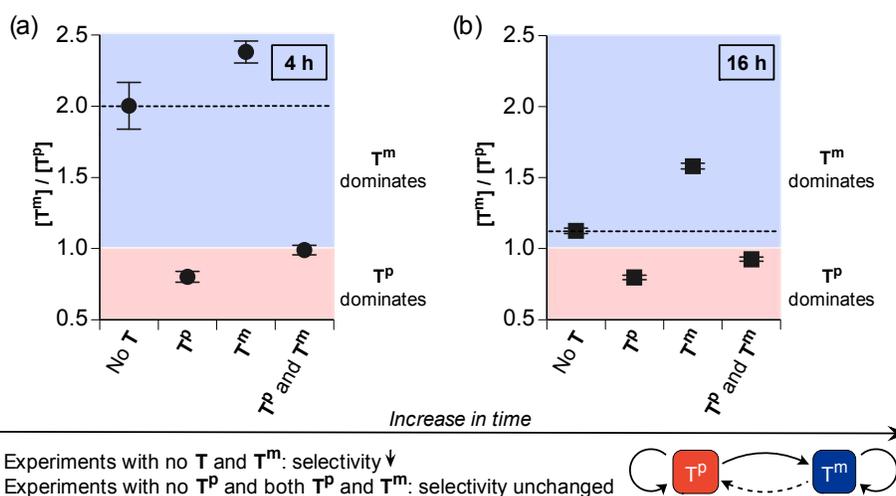


Figure 4. The effect of instructing the T^P - T^m network with preformed template (added at $t = 0$), on the ratio of $[T^m]/[T^P]$ determined after (a) 4 h (black circles) and (b) 16 h (black squares). Dashed lines represent the $[T^m]/[T^P]$ ratio determined in the uninstructed kinetic experiments, for comparison. Concentrations were determined using 470.3 MHz $^{19}\text{F}\{^1\text{H}\}$ NMR spectroscopy ($[\text{M}^P] = [\text{M}^m] = [\text{N}] = 5 \text{ mM}$, if present, $[\text{template}] = 1 \text{ mM}$, CDCl_3 , $5 \text{ }^\circ\text{C}$).

In the presence of preformed *trans*- T^P , added to the reaction mixture at the start of the reaction, the observed outcome was altered significantly—the instructing replicator, *trans*- T^P , outperformed *trans*- T^m in the competition for N and the two products were formed in a ratio of 0.80 ($[trans-T^m]/[trans-T^P]$ after 4 h, **Figure 5a**). When preformed *trans*- T^m was added at the start of the reaction, the network displayed the opposite behavior—*i.e.*, formation of replicator *trans*- T^m was enhanced further ($[trans-T^m]/[trans-T^P] = 2.38$ after 4 h, **Figure 5a**). Interestingly, when both *trans*- T^P and *trans*- T^m were used to instruct the replication processes simultaneously, the two replicators were formed at a ratio of 0.99 ($[trans-T^m]/[trans-T^P]$) after 4 h (**Figure 5a**). The simultaneous addition of both T^P and T^m enhances the formation of *trans*- T^P relative to the uninstructed scenario, nevertheless *trans*- T^m remained the dominant species in solution in this experiment after 4 h. This observation can be attributed to the fact

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2 that the formation of replicator *trans*- \mathbf{T}^m proceeds through two template-mediated channels, whilst
3 replicator *trans*- \mathbf{T}^p is formed through a single autocatalytic pathway only.

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5 The ratios of the two replicators, $[\textit{trans}\text{-}\mathbf{T}^m]/[\textit{trans}\text{-}\mathbf{T}^p]$, were re-examined after 16 h, revealing
6 a trend similar to that observed for the %EFs discussed earlier for the auto- and crosscatalytic pathways
7 in isolation. Specifically, the values of $[\textit{trans}\text{-}\mathbf{T}^m]/[\textit{trans}\text{-}\mathbf{T}^p]$ at $t = 16$ h were determined to be 1.13,
8 0.80, 1.58, and 0.93 in the presence of no template, *trans*- \mathbf{T}^p , *trans*- \mathbf{T}^m , and both *trans*- \mathbf{T}^p and *trans*-
9 \mathbf{T}^m , respectively (**Figure 5b**). Comparison of the ratios determined after 4 and 16 h showed that the
10 selectivity in experiments instructed with \mathbf{T}^p only changed less over time (4 h \rightarrow 16 h) than the
11 selectivity in experiments employing no template or \mathbf{T}^m . These differences in product ratio as a
12 function of time can be attributed to the fact that only a single crosscatalytic pathway is active in this
13 network and thus, while the presence of \mathbf{T}^p affects the formation of both templates, addition of \mathbf{T}^m
14 enhances the production of itself exclusively. Overall, these results demonstrate that the $[\textit{trans}\text{-}\mathbf{T}^m]/[\textit{trans}\text{-}\mathbf{T}^p]$
15 ratio is strongly dependent on *both* the reaction time and the identity of the instructing
16 template or templates.
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28 **Examining the role of the $[\mathbf{T}^p\cdot\mathbf{T}^m]$ heteroduplex**

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30 The kinetic analyses revealed that the two replicators examined in this study exhibit markedly different
31 replication efficiencies as a result of small structural changes engineered in their molecular
32 frameworks. Although the kinetics of the competition experiments utilized data from ^{19}F NMR
33 spectroscopy, ^1H NMR spectroscopic data was also acquired at the same time during each of the
34 competition experiments. Examination of these ^1H NMR spectra revealed interesting changes in the
35 chemical shifts of the resonances arising from the *trans* protons (**Figure 5a**) on the isoxazolidine rings
36 present in \mathbf{T}^p and \mathbf{T}^m . These changes are visible most clearly in the competition kinetic experiment
37 where the reaction mixture is instructed with 20 mol% of \mathbf{T}^p (**Figure 5a**).
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45 At the beginning of this reaction, replicator \mathbf{T}^p , added at a concentration of 1 mM, is the
46 dominant template in solution and no \mathbf{T}^m is present at the start of the reaction. In the presence of
47 preformed \mathbf{T}^p template, \mathbf{M}^p , \mathbf{M}^m , and \mathbf{N} , are transformed quickly into both \mathbf{T}^p and \mathbf{T}^m . As the
48 concentration of \mathbf{T}^m increases in the reaction mixture over time, a significant change in the chemical
49 shift of the resonance associated with the *trans* proton present in \mathbf{T}^p is observed. However, the
50 resonance arising from replicator \mathbf{T}^m itself is apparently unperturbed. These observations are
51 instructive, since they demonstrate strikingly the ability of replicators \mathbf{T}^p and \mathbf{T}^m to assemble into both
52 homoduplexes ($[\mathbf{T}^p\cdot\mathbf{T}^p]$ and $[\mathbf{T}^m\cdot\mathbf{T}^m]$) and the corresponding heteroduplex ($[\mathbf{T}^p\cdot\mathbf{T}^m]$). The presence of
53 a single resonance for each template indicates that the exchange between these species is fast on the ^1H
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NMR chemical shift timescale. For each replicator, the observed chemical shift depends on the mole fraction of the replicator bound within its homoduplex and the heteroduplex.

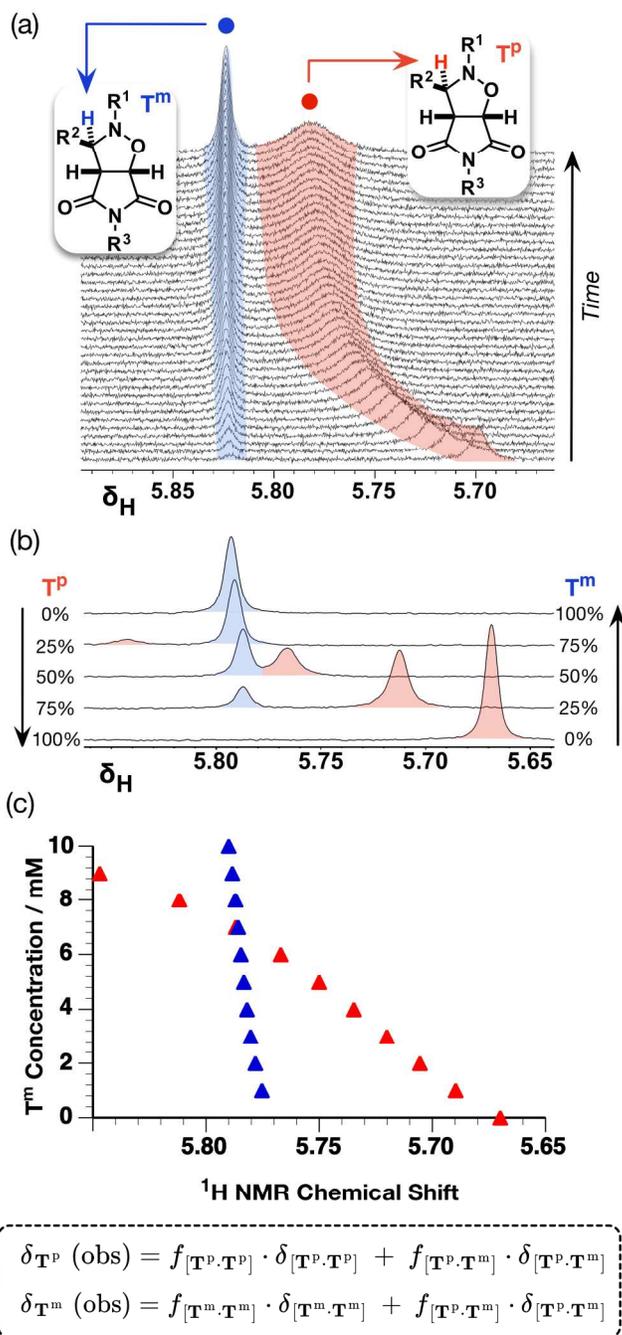


Figure 5. (a) Arrayed partial 499.9 MHz 1H NMR spectra showing the changes in the chemical shifts of the resonances associated with the *trans* protons on the isoxazolidine ring of T^P (red) and T^m (blue) replicators formed within a competition experiment instructed with T^P template ($[M^P] = [M^m] = [N] = 5$ mM, $[trans-T^P] = 1$ mM, $CDCl_3$, 5 °C) over time. (b) Overlay of partial 400.1 MHz 1H NMR spectra (RT) showing the *trans* resonances belonging to the T^P and T^m cycloadducts. The chemical shift changes for these *trans* products were examined at five different conditions: starting with T^P only and progressively increasing the content of T^m to 100% (combined concentration of T^P and $T^m = 10$ mM). (c) Simulated changes in the chemical shifts of the isoxazolidine *trans* proton (see (a)) in *trans*- T^P (red triangles) and in *trans*- T^m (blue triangles), arising as a result of the distribution of these products in homo and heteroduplexes. Data are derived from simulations run using Gepasi 3 (Ref. 22), using the duplex association constants determined through fitting of kinetic data and estimated/determined chemical shifts for each duplex in the system.

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2 In order to develop a better understanding of these chemical shift changes, we examined the ^1H
3 NMR spectra (**Figure 5b**) recorded on five solutions of T^{P} and T^{m} at a combined concentration of 10
4 mM in CDCl_3 , starting with a composition of 100% T^{P} and increasing the fraction of T^{m} progressively
5 from 0% to 25%, 50%, 75%, and, finally, 100%. Each mixture was analyzed by 500.1 MHz ^1H NMR
6 spectroscopy (**Figure 5b**, for the corresponding $^{19}\text{F}\{^1\text{H}\}$ NMR spectra, see **Figure S9**). These five
7 experiments allowed the changes in the chemical shifts arising from the resonances of both replicators
8 to be examined under conditions where only homoduplex $[\text{T}^{\text{P}}\cdot\text{T}^{\text{P}}]$ or homoduplex $[\text{T}^{\text{m}}\cdot\text{T}^{\text{m}}]$ are present,
9 as well as the three intermediate conditions where both homo- and heteroduplexes are present together.
10 Analysis of the chemical shift changes observed in the isoxazolidine ring protons colored red and blue
11 (**Figure 5**) as a function of the T^{P} and T^{m} composition revealed a markedly different behavior for
12 replicator T^{P} relative to T^{m} . While the chemical shift of the resonance arising from T^{m} varied very little
13 across the four different compositions examined (25% T^{m} \rightarrow 100% T^{m}), the chemical shift of the
14 resonance arising from T^{P} exhibited a dramatic downfield shift (>0.2 ppm, **Figure 5b**) as the fraction
15 of T^{m} in the solution increases.
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18 We modelled the steady state distributions of T^{P} and T^{m} within the various product duplexes
19 present in solution and, consequently, the expected chemical shift for the resonances arising from the
20 isoxazolidine protons present in T^{P} and T^{m} (colored red and blue in **Figure 5c**) by combining the data
21 in **Figure 5b** with the corresponding association constants for the homo- and heteroduplexes
22 determined earlier through kinetic fitting. **Figure 5c** shows the calculated chemical shifts for the
23 appropriate proton resonances in T^{P} and T^{m} at nine different concentration ratios. Comparison of these
24 calculated chemical shifts to the experimentally determined ^1H NMR data shows a clear qualitative
25 agreement. The high stability of $[\text{T}^{\text{m}}\cdot\text{T}^{\text{m}}]$ is reflected by the fact that the chemical shift of the
26 isoxazolidine proton associated with this product remains virtually unchanged throughout the
27 experiments. Since $[\text{T}^{\text{P}}\cdot\text{T}^{\text{P}}]$ is the least stable duplex, as the concentration of T^{m} is increased, the
28 distribution of T^{P} is shifted towards the more stable $[\text{T}^{\text{P}}\cdot\text{T}^{\text{m}}]$ duplex, evidenced by the 0.2 ppm
29 downfield shift in the resonance arising from *trans*- T^{P} . The analysis of the behavior of the $[\text{T}^{\text{P}}\cdot\text{T}^{\text{m}}]$
30 heteroduplex demonstrated that the differences in duplex stabilities have a dramatic influence over not
31 only the auto- and crosscatalytic efficiencies of the two replicators, but also the chemical shifts of the
32 resonances associated with these templates. In this case, we were able to use the thermodynamic
33 parameters derived through kinetic fitting to successfully relate the variation in duplex and heteroduplex
34 stabilities to the variation in the chemical shifts observed for templates T^{P} and T^{m} within the highly
35 interconnected network. Additionally, these differences in the strengths of recognition processes that
36 underlie the formation of replicators T^{P} and T^{m} and the associated complexes/duplexes also help to
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2 determine the outcome of competition in this network of replicators— T^m is not only able to exploit T^p
3 for its formation, it is also capable of sequestering T^p in a duplex that decreases its availability in
4 solution and, as a result, the efficiency of the autocatalytic pathway leading to the formation of T^p .
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9 **What governs the outcome of competition in T^p - T^m network?**

10 The comprehensive set of kinetic experiments described here allowed us to examine each catalytic
11 pathway in isolation and within a number of competition scenarios. The observed results demonstrate
12 clearly that the absence of one crosscatalytic relationship (T^m is not a catalyst for the formation of T^p)
13 has a significant influence on the behavior of the replicator network in a situation where T^m and T^p
14 have to compete for a shared building block. The kinetic fitting demonstrates that there are significant
15 differences between the two replicators in terms of the duplex association constants and unimolecular
16 rate constants for the reactions that take place on templates. The origin of the disparity in the
17 crosscatalytic efficiencies of T^m and T^p , identified by our kinetic simulations and fitting, must lie in
18 subtle differences in the stabilities and structures of the ternary complexes, $[N \cdot M^p \cdot T^m]$ and
19 $[N \cdot M^m \cdot T^p]$, and the transition states accessed from these complexes. In order to gain a better
20 understanding of these processes, we undertook a series of density functional theory calculations that
21 examined the structures of the ternary complexes $[N \cdot M^p \cdot T^m]$ and $[N \cdot M^m \cdot T^p]$, the heteroduplex
22 $[T^p \cdot T^m]$, and the transition states (TSs) that connect these complexes. All of these calculations were
23 performed at the ω B97X/def2-SVP level of theory using a continuum solvation model (PCM) for
24 chloroform (see Supporting Information for details). In addition, we compared the transition state
25 structures calculated for the template-directed reactions with that computed for the prototypical 1,3-
26 dipolar cycloaddition of diphenylnitrone to *N*-phenyl maleimide. Although the relative stereochemistry
27 of the forming cycloadduct is controlled very well in favor of the *trans* diastereoisomer, the absolute
28 stereochemical relationships between the templates are more complex. The template-directed reactions
29 can proceed by one of two pathways. The stereochemistry of the newly formed replicator can match
30 that of the template, for example, $RSR-T^p \rightarrow RSR-T^m$ — we will refer to this pathway as “matched”.
31 Alternatively, the stereochemistry of the newly formed replicator can be mismatched compared to that
32 of the template, for example, $RSR-T^p \rightarrow SRS-T^m$ — we will refer to this pathway as “mismatched”.
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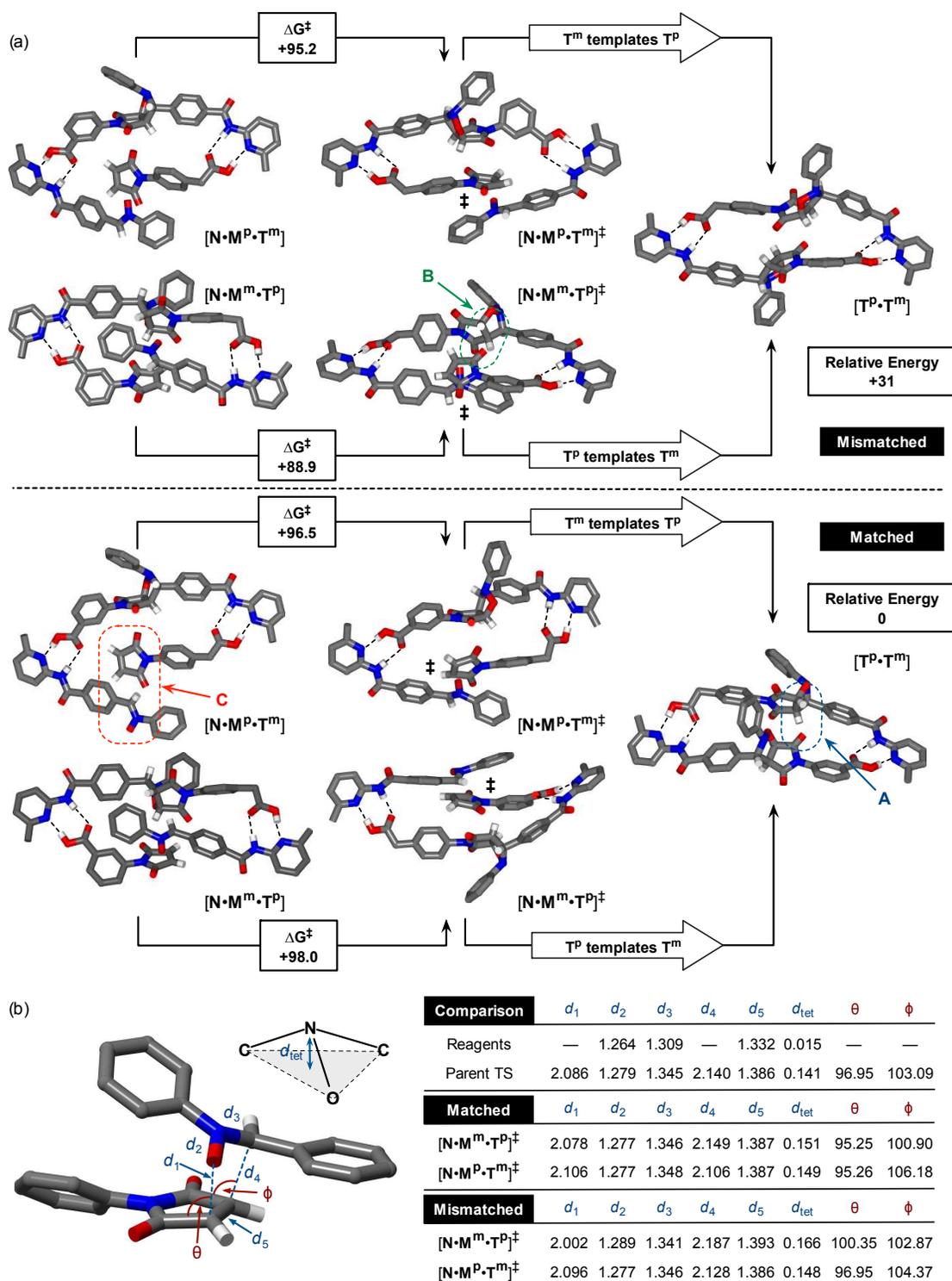


Figure 6. (a) Calculations reveal that the ternary complexes $[N\cdot M^m\cdot T^P]$ and $[N\cdot M^P\cdot T^m]$ can react to form the template duplex $[TP\cdot T^m]$ through two sets of transition states—in which, the stereochemistries of the forming cycloadducts can be either matched or mismatched with respect to the replicator template—giving rise to four possible reaction pathways. Calculations were performed at the ω B97X/def2-SVP level of theory using a continuum solvation model for chloroform. Energies are in kJ mol^{-1} . Hydrogen bonds are represented by dashed lines. The colored shaded areas indicated by letters are discussed in the main text. (b) Structural parameters describing the calculated structures (ω B97X/def2-SVP) for the four transition states accessed from either $[N\cdot M^m\cdot T^P]$ or $[N\cdot M^P\cdot T^m]$. Data for the corresponding parameters for the transition state located for the reaction of diphenylnitrene and *N*-phenyl maleimide are provided for comparison. All distances are in Å and angles are in degrees.

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2 The consequence of these relationships is that heteroduplex $[\mathbf{T}^p \cdot \mathbf{T}^m]$, and the transition states leading to
3 it, can exist either as a structure in which the stereochemistries of the templates are matched or as a
4 structure in which the stereochemistries of the templates are mismatched. We computed the four
5 possible pathways—matched and mismatched for both $\mathbf{T}^p \rightarrow \mathbf{T}^m$ and $\mathbf{T}^m \rightarrow \mathbf{T}^p$. The results of these
6 calculations are summarized in **Figure 6**.
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10 In the case of the template duplexes, the matched form of $[\mathbf{T}^p \cdot \mathbf{T}^m]$ is predicted to be around
11 30 kJ mol^{-1} more stable than the corresponding mismatched duplex. In part, this observation may be
12 explained by the presence of stabilizing C–H \cdots O interactions along the spine of the template (marked
13 **A** and highlighted in blue in **Figure 6a**) that are present in the matched duplex, but not in the
14 mismatched one. Additionally, there are some distortions in the conformations of the fused bicyclic
15 cycloadducts present in the mismatched $[\mathbf{T}^p \cdot \mathbf{T}^m]$ duplex that are required in order to maintain the four
16 hydrogen bonds connecting the two templates.
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23 Comparison of the transition states that were located for the four available pathways reveals that
24 one structure—mismatched $\mathbf{T}^p \rightarrow \mathbf{T}^m$ (**Figure 6a**)—is significantly lower in energy than the other
25 three. This observation is consistent with the experimental results indicating that \mathbf{T}^p can act as a
26 catalyst for the formation of \mathbf{T}^m , but \mathbf{T}^m is not a particularly efficient catalyst for the formation of \mathbf{T}^p .
27 Comparison of the transition state structures with that of the parent cycloaddition between
28 diphenylnitron and *N*-phenyl maleimide (**Figure 6b**) reveals that the mismatched transition state
29 $[\mathbf{N} \cdot \mathbf{M}^m \cdot \mathbf{T}^p]^\ddagger$ has a geometry that differs considerably from the other three and from the parent. In this
30 transition state, C–O bond formation is more advanced (C \cdots O = 2.002 \AA vs. 2.086 \AA in parent TS) and
31 C–C bond formation is much less advanced (C \cdots C = 2.187 \AA vs. 2.146 \AA in parent TS) than in any of
32 the other three transition states accessible from the ternary complexes. This distortion suggests a
33 transition state that may be slightly more polar²³ than the other three and may be stabilized by a series
34 of C–H \cdots O interactions present along the spine of the template (marked **B** and highlighted in green in
35 **Figure 6a**).
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46 Whilst the structures of the transition states and the duplexes have limited conformational
47 freedom as they adopt co-conformations that maintain four hydrogen bonds, the ternary complexes are
48 much less restricted in a conformational sense. Bruce and Lightstone have shown²⁴ that the rates of
49 intramolecular reactions can be related to the fraction of low energy conformations that are present as
50 near-attack conformations (NACs). Previously, we have applied²⁵ this type of analysis successfully to a
51 recognition-mediated pseudointramolecular cycloaddition reaction. Therefore, when considering these
52 structures, we restricted our examination to structures that could be categorized loosely as NACs, *i.e.*,
53 we examined the structures in which the termini of the 4π and 2π components of the cycloaddition were
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2 in closest proximity. Interestingly, in the lowest energy structure located for each ternary complex,
3 there is a striking difference between the distances that separate the reactive termini of the 4π and 2π
4 components ($C\cdots O$ and $C\cdots C$) depending on the identity of the replicator that is acting as the template.
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7 In the ternary complex $[N\cdot M^P\cdot T^m]$, where T^m is templating the formation of T^P , all of the
8 $C\cdots O$ and $C\cdots C$ distances are above 4 Å and, in the case of matched $[N\cdot M^P\cdot T^m]$, over 5 Å (marked C
9 and highlighted in red in **Figure 6a**). These large distances suggest that the low energy co-
10 conformations of $[N\cdot M^P\cdot T^m]$ are unlikely to contain a high fraction of NACs. This observation is
11 consistent with lower efficiency of this ternary complex determined experimentally. By contrast, in
12 $[N\cdot M^m\cdot T^P]$, where T^P is templating the formation of T^m , all of the $C\cdots O$ and $C\cdots C$ distances are
13 below 4 Å and in the case of the mismatched ternary complex, these distances are both below 3.40 Å.
14 These very short distances suggest that the low energy co-conformations of mismatched $[N\cdot M^P\cdot T^m]$
15 are likely to contain a high fraction of NACs and this observation is consistent with this ternary
16 complex accessing the lowest energy transition state.
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25 These calculations also offer a window into a more complex world that is currently beyond the
26 reach of our experiments, in which the stereochemical relationships between replicators can play an
27 important role in determining the outcome of competition^{8b,8e,26} between the replicators. Going
28 forward, these results demand that, in order to maximize the efficiency of a template-directed process,
29 it will be necessary to consider the detailed relationship between individual molecular structures and
30 their co-conformations within complexes.
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37 Conclusions

38 Small networks of minimal replicators may serve²⁷ as simple models for primitive metabolic pathways
39 in that the population of an individual replicator within these networks can be manipulated by
40 exploiting the autocatalytic properties of the replicating template in order to specifically upregulate the
41 rate of formation of that template. In this work, we describe the experimental implementation of such a
42 network constructed from three simple building blocks—two maleimides (M^m and M^P) and a single
43 nitron (N)—that react to create two synthetic replicators (T^m and T^P) that can template their own
44 formation. Within this network, the two replicating templates compete for a common reagent, namely
45 nitron N . In order to understand the system-level behavior of this network, it is important to
46 characterize each replicator in isolation. In this case, our kinetic studies on T^m and T^P in isolation
47 reveal that T^P is the more efficient replicator of the pair. In isolation, T^P benefits from a significantly
48 higher effective molarity for the key cycloaddition step than T^m . Additionally, the $[T^P\cdot T^P]$ duplex is
49 somewhat less stable than the $[T^m\cdot T^m]$ duplex, thereby ensuring that the concentration of catalytically
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1 active, monomeric T^P is higher than in the corresponding reaction involving T^m . The abilities of both
2 T^P and T^m to replicate are confirmed by their capacity to template their own formation as evidenced by
3 experiments in which pre-formed template is added to a mixture of the nitron and the appropriate
4 maleimide at the start of the reaction. The structural similarity between T^P and T^m opens the possibility
5 that a mutual crosscatalytic relationship may exist between these two templates. By exploring the
6 response of reactions that form either T^P or T^m to the addition of the other template as an instruction,
7 we established that, although T^P can template the formation of T^m , T^m does not catalyze the formation
8 of T^P effectively. The absence of reciprocity in the crosscatalytic relationship between T^P and T^m is
9 critical to understanding the outcome of experiments in which T^P and T^m are placed in a competitive
10 situation. Analysis of experiments in which T^P and T^m replicate in the same flask reveal the apparently
11 anomalous observation that the replicator that is less efficient in isolation (T^m) becomes the dominant
12 species in the complete network. This observation is a direct result of the presence of the non-
13 reciprocal crosscatalytic interaction between T^P and T^m , which allows T^m to exploit T^P for its
14 formation but not *vice versa*. The system-level outcome is also driven by the subtle interplay of the
15 recognition and reaction processes. Replicator T^m forms more stable complexes and duplexes, which
16 allows it to reduce the replication efficiency of T^P by sequestering it within the $[T^P \cdot T^m]$ heteroduplex
17 that is more stable than the $[T^P \cdot T^P]$ homoduplex. However, our attempts to direct the competition
18 between the replicators using preformed templates under the well-stirred batch reactor conditions
19 employed here always result in the erosion of the imbalance between the two replicators generated at
20 early time points. This effect is a direct result of the exhaustion of building blocks that limits the
21 efficiency of replication processes, thereby preventing the initially dominant replicator from enhancing
22 its advantage. This observation illustrates clearly that the ability of one replicator to dominate within
23 this network depends not only on its catalytic relationship with the other replicator in the network but
24 also their reaction environment. The current work represents a firm foundation for the expansion of the
25 study of replicator networks under conditions where dynamic processes or diffusion contribute to the
26 outcome of replicator competition—*i.e.*, reaction conditions where replication processes can operate
27 with increased efficiency. These studies are currently underway in our laboratory.
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Acknowledgements

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Supporting information:

Supporting information is available free of charge at:

General experimental procedures; synthetic procedures and compound characterization; details of kinetic analyses, fitting (including fitted kinetic profiles), and simulations, and details of DFT calculations.

Notes and References

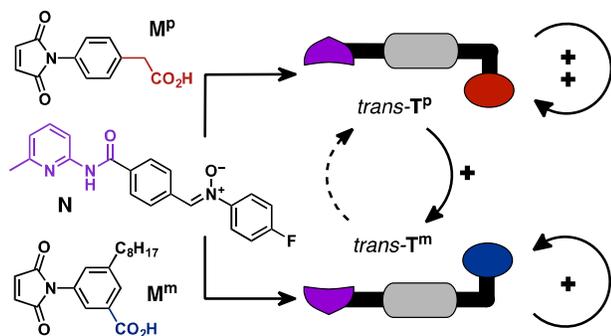
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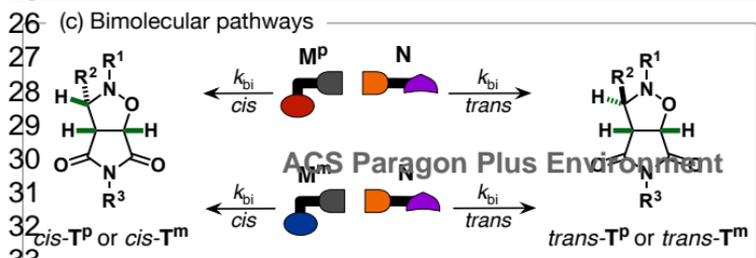
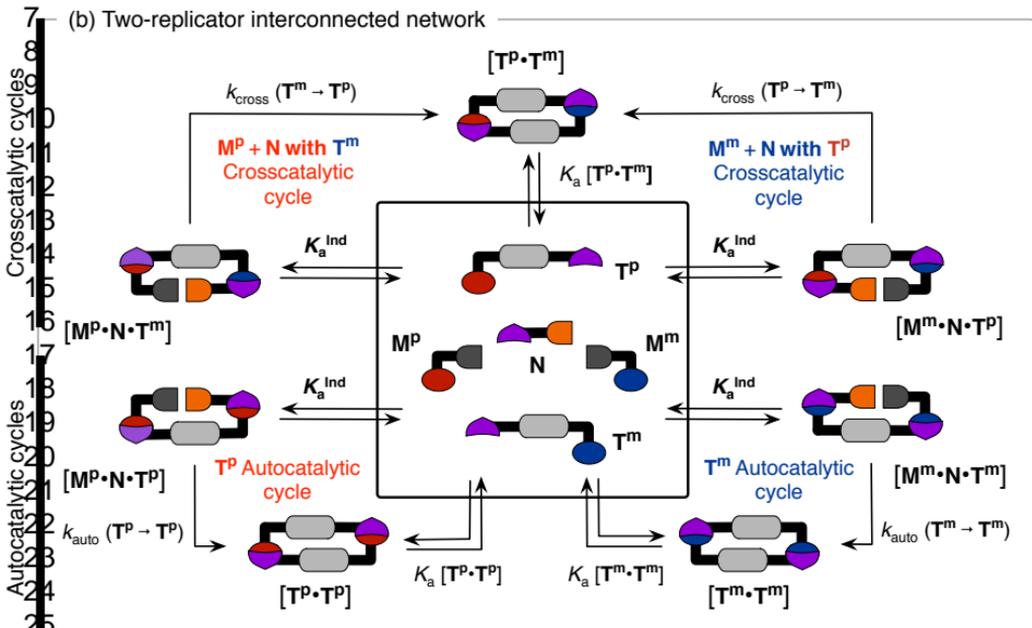
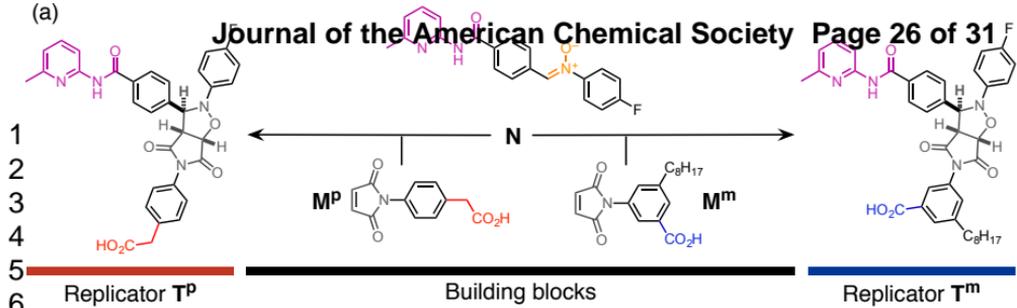
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42 (16) The 1,3-dipolar cycloaddition reaction between a nitron and a maleimide can give rise to two
43 diastereoisomers, *trans* and *cis*. Only the *trans* diastereoisomers are capable of taking part in
44 template-directed replication processes in the network of T^P and T^M replicators. For this reason,
45 the notation for the *trans* cycloadducts capable of replication will generally omit the *trans*
46 throughout this work (for example, *trans*- T^P will be generally referred to as T^P , unless emphasis
47 on the identity of the diastereoisomer is necessary). All preformed templates used in instructed
48 experiments consisted exclusively of the *trans* diastereoisomer of the relevant replicator.
- 49
50 (17) This relationship presents a slight generalization, as whilst generally true, the increase in the
51 strength of the recognition between two network components might also result in the formation
52 of $[M \cdot N]$ type of complexes, which can reduce the efficiency of the autocatalytic pathway.
- 53
54 (18) The error values for the calculated %EFs were determined using standard methods for
55 calculation of normally distributed errors. These calculations employed the estimated value of
56 concentration error of ± 0.02 mM determined for all of the kinetic experiments described in this
57 work (for details, see Supporting Information).
- 58
59 (19) Kinetic effective molarity provides a measure of the enhancement in the template-mediated
60 pathway relative to the bimolecular pathway. $EM_{kinetic}$ also tells us about the concentration at
which the reaction would have to be performed in order for the bimolecular pathway to perform

1
2 at the efficiency of the recognition-mediated pathway. For this reason, rate acceleration takes
3 place even if $EM_{\text{kinetic}} < 1$, as long as the concentration at which the reaction is conducted is
4 below the value of EM_{kinetic} .

- 5
6 (20) Thermodynamic effective molarity provides information about the stability of the template
7 duplex relative to the strength of the ternary complex.
- 8 (21) The values of template duplex association constants determined through kinetic fitting in this
9 work were in the range of ~ 17.9 to $68.6 \times 10^6 \text{ M}^{-1}$, *i.e.*, a situation where the concentration of
10 the catalytically active ternary complex will be in the μM range (see Ref. 8c for detailed
11 analysis) under the conditions employed experimentally. Ideally, self-replicating systems would
12 be examined at a concentration (C) that promotes the dissociation of the template duplex
13 ($[C]_{\text{initial}} = 1/K_{\text{a}}^{\text{Duplex}}$). At such a concentration, however, the formation of the catalytically active
14 ternary complex, which is driven by the considerably smaller $K_{\text{a}}^{\text{Ind}}$ would be strongly
15 disfavoured. The optimum conditions, therefore, need to balance these two competing
16 influences.
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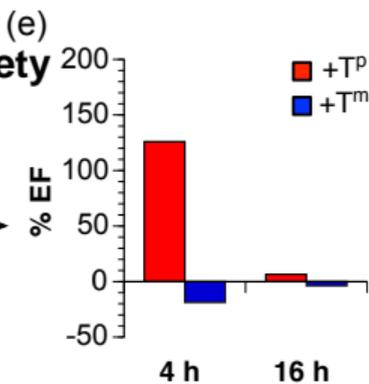
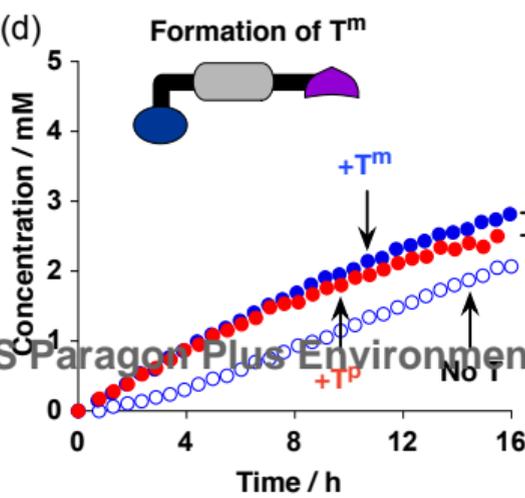
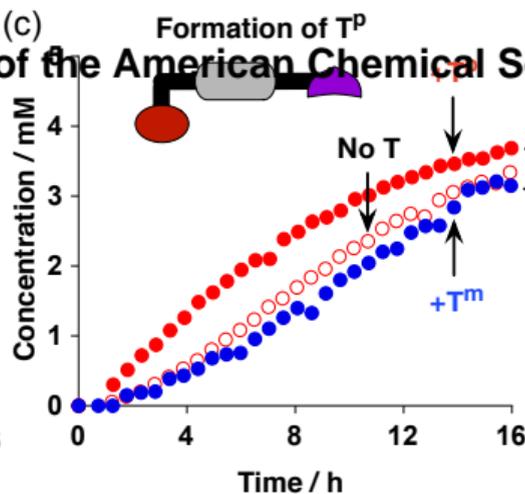
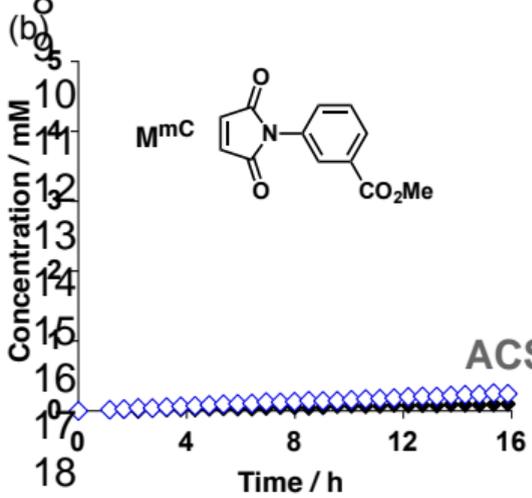
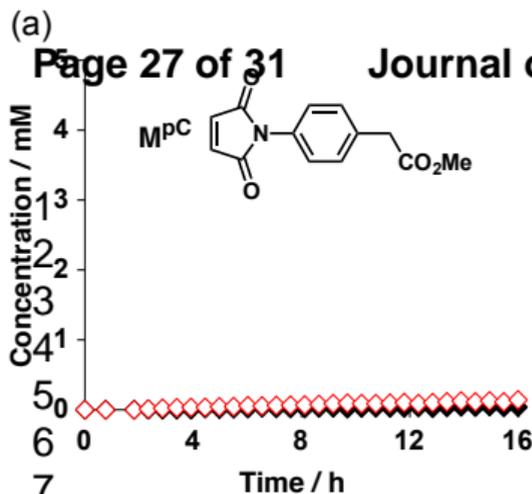
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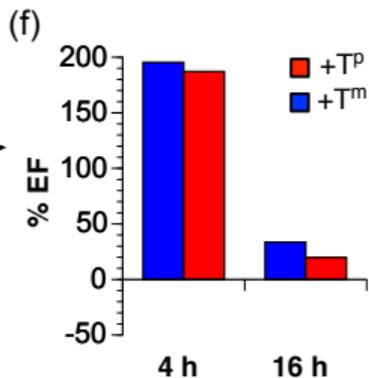


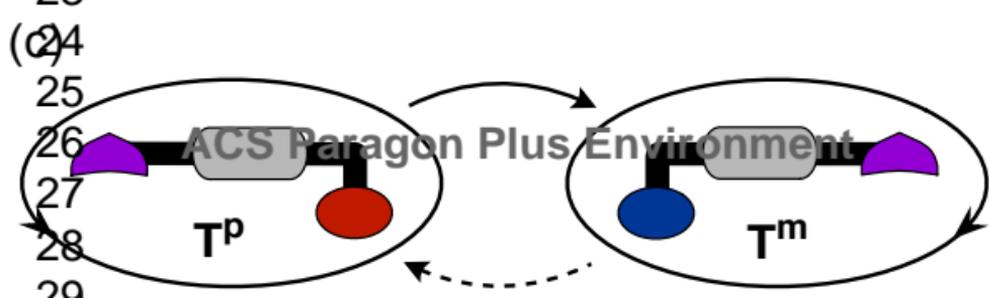
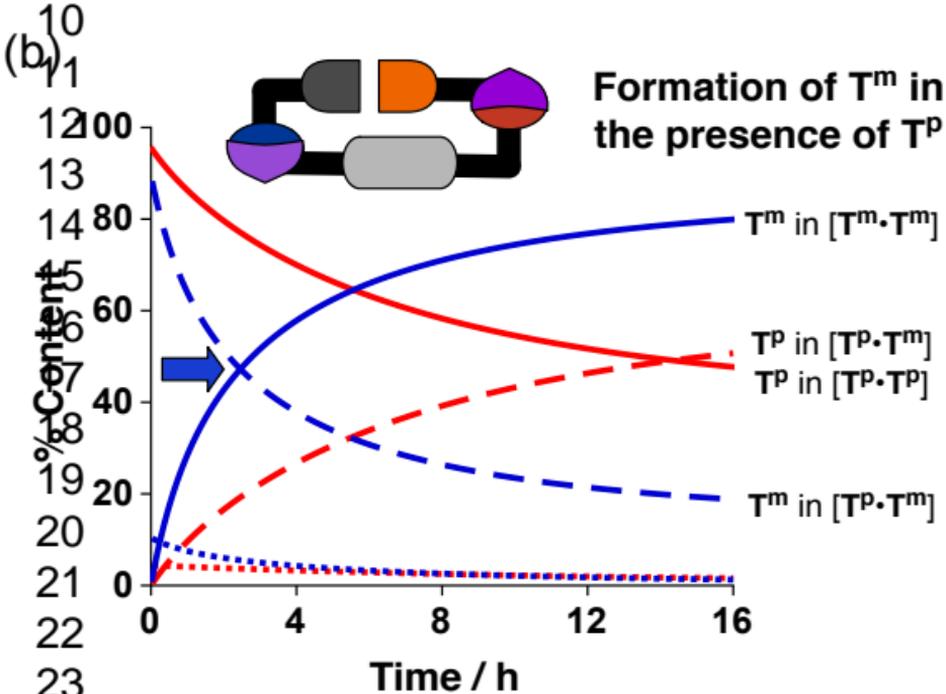
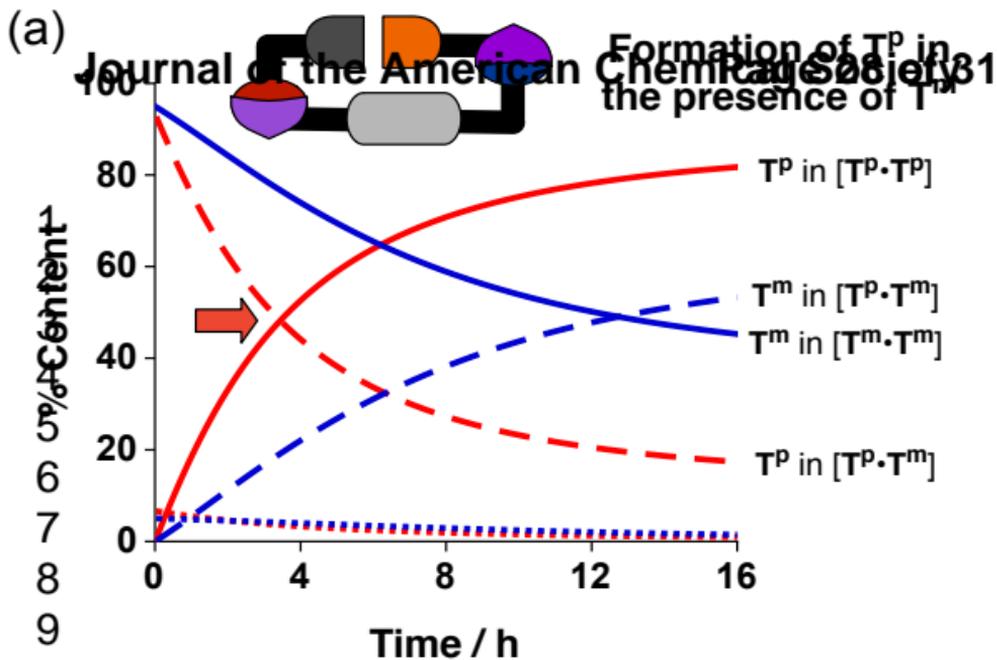
(d) K_a values

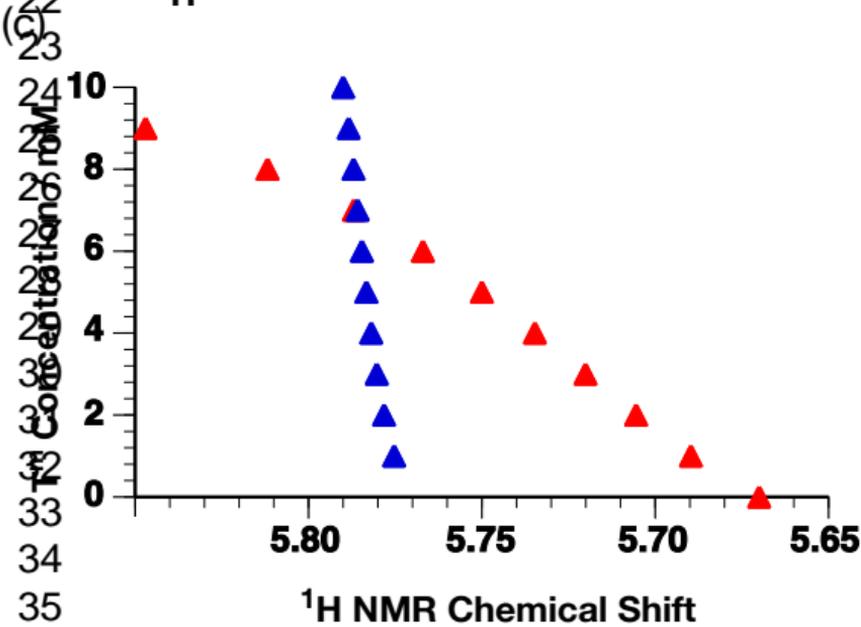
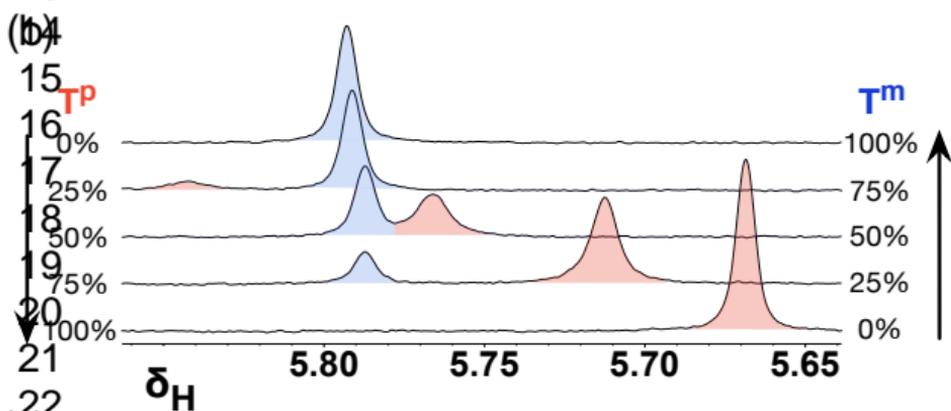
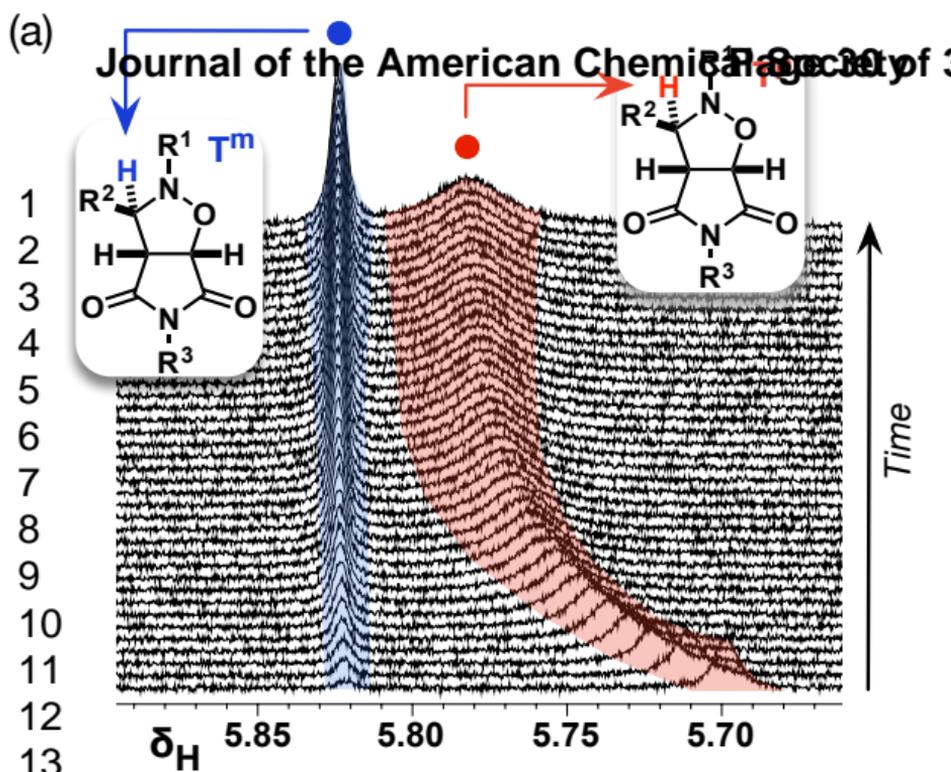
$K_a^{\text{Ind}} / \text{M}^{-1}$
830
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$$\%EF = 100 \cdot \frac{[T]_i - [T]_u}{[T]_u}$$



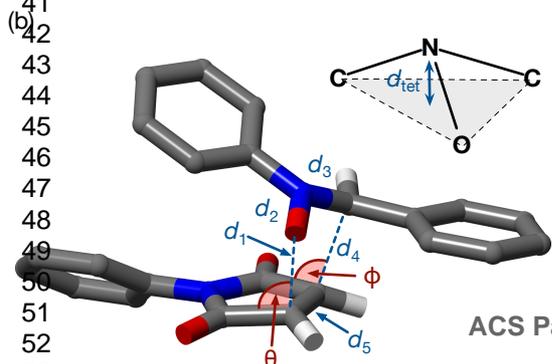
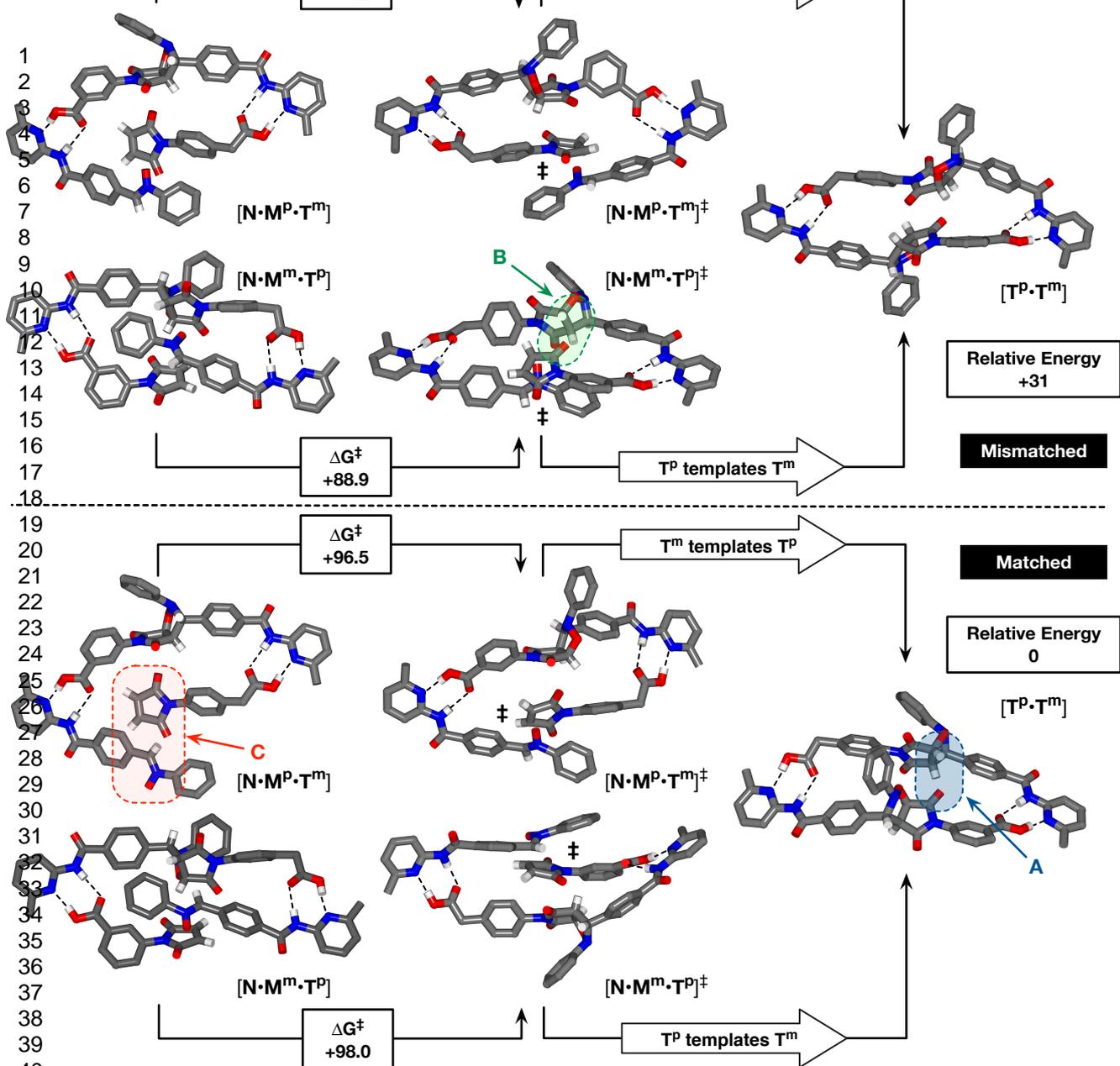




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$$\delta_{\text{TP}}^{\text{(obs)}} = f_{[\text{TP} \cdot \text{TP}]} \cdot \delta_{[\text{TP} \cdot \text{TP}]} + f_{[\text{TP} \cdot \text{T}^m]} \cdot \delta_{[\text{TP} \cdot \text{T}^m]}$$

$$\delta_{\text{T}^m}^{\text{(obs)}} = f_{[\text{T}^m \cdot \text{T}^m]} \cdot \delta_{[\text{T}^m \cdot \text{T}^m]} + f_{[\text{TP} \cdot \text{T}^m]} \cdot \delta_{[\text{TP} \cdot \text{T}^m]}$$



Comparison	d_1	d_2	d_3	d_4	d_5	d_{tet}	θ	ϕ
Reagents	—	1.264	1.309	—	1.332	0.015	—	—
Parent TS	2.086	1.279	1.345	2.140	1.386	0.141	96.95	103.09
Matched	d_1	d_2	d_3	d_4	d_5	d_{tet}	θ	ϕ
$[N \cdot M^m \cdot T^P]^\ddagger$	2.078	1.277	1.346	2.149	1.387	0.151	95.25	100.90
$[N \cdot M^P \cdot T^m]^\ddagger$	2.106	1.277	1.348	2.106	1.387	0.149	95.26	106.18
Mismatched	d_1	d_2	d_3	d_4	d_5	d_{tet}	θ	ϕ
$[N \cdot M^m \cdot T^P]^\ddagger$	2.002	1.289	1.341	2.187	1.393	0.166	100.35	102.87
$[N \cdot M^P \cdot T^m]^\ddagger$	2.096	1.277	1.346	2.128	1.386	0.148	96.95	104.37