

0040-4039(95)02238-4

Practical, Convergent Total Synthesis of Polyamine Amide Spider Toxin NSTX-3

Ian S. Blagbrough,* Eduardo Moya, and Steven P. Walford

Department of Medicinal Chemistry, School of Pharmacy and Pharmacology, University of Bath, Claverton Down, Bath BA2 7AY, U.K.

Abstract: A practical, total synthesis of polyamine amide spider toxin NSTX-3, a potent glutamate receptor antagonist with potential as a neuroprotective agent, is reported. The unsymmetrical polyamine moiety was built by a conjugate addition to afford putreanine and regioselective acylation of L-asparaginyl-cadaverine.

Nephila clavata is an orb-weaver spider which envenomates with essentially millimolar glutamate and a complex mixture of unsymmetrical polyamine amides toxins e.g. NSTX-3 (1) ¹ and peptide based toxins, NPTXs.^{2,3} The polyamine amide components are open-channel glutamate receptor blockers.¹ The unusual structure of NSTX-3 (1) was solved and published by Nakajima, Kawai, and their co-workers.^{1,4} This spider toxin contains an unsymmetrical polyamine (5. β -Ala.4.Arg), regioselectively acylated on the primary amino functional group of the cadaverine (1,5-diaminopentane) moiety with 2,4-dihydroxyphenylacetyl-L-asparagine. The terminal amine of the putreanine (β -Ala.4) moiety is acylated with L-arginine. Polyamine amide β -Ala.4.Arg carries up to three positive charges, at physiological pH. Confirmation of the structure of this regioselectively diacylated unsymmetrical polyamine came with the total syntheses of NSTX-3 (1) ^{5,6} and the closely related tripeptide (Ala.Gly.Arg) containing spider toxin clavamine (2).⁷⁻⁹ There is continuing interest in polyamine amides as channel blockers for glutamic acid and/or nicotinic acetylcholine-gated cation channels, and certain voltage-sensitive calcium channels^{1,10-16} and as novel natural products containing polyamine amides.¹⁷ In this *Letter*, we present a convergent total synthesis of NSTX-3 (1) ¹⁸ based upon a strategy which allows the putreanine moiety (β -Ala.4) to be incorporated first. This practical route ensures sufficient material is available for pharmacological evaluation.



NSTX-3 (1) synthesis: 2,4-Dibenzyloxyphenylacetic acid activated as its *N*-hydroxysuccinimide ester (3) was prepared by an Arndt-Eistert chain homologation strategy from 2,4-dibenzyloxybenzoic acid (4). After conversion of acid (4) into the corresponding acid chloride (5) (oxalyl chloride, 1.2 eq., pyr., 1.1 eq., PhMe, 0 to 20°C, 30 mins), reaction with an ethereal (EtOH free) solution of diazomethane (10 eq., 0 to 20°C, 2 h) gave diazoketone (6) as a yellow solid mp 98-99°C dec., in 81 % yield from acid (4). Arndt-Eistert reaction (anhydrous DMF, 20°C, PhCOOAg 0.2 eq., 75 mins) gave the desired activated ester (3) as a white solid (80 %) mp 145-146°C (lit. ⁵ mp 143-143.5°C), after silica gel chromatography, *via* trapping of the presumed ketene intermediate (7) *in situ* with *N*-hydroxysuccinimide (10 eq.).

The cadaverine moiety of NSTX-3 (1) was incorporated in (2,4-dibenzyloxy)phenylacetyl-L-Asnmono-BOC cadaverine (8) which was designed for selective deprotection to afford free primary amine (9) for coupling with a putreanine containing polyamine amide moiety.⁶ Thus, mono-BOC cadaverine was prepared by reacting cadaverine (1,5-diaminopentane) (3.0 eq.) with BOC₂O (1.0 eq., THF, 0°C, 16 h, 62 %).¹⁹ *N*-BOC-1, 5-Diaminopentane was then acylated with Z-L-AsnOpNP (10) (1.1 eq., DCM, 20°C, 16 h) which efficiently gave orthogonally protected Asn-cadaverine (11) (69 %). Hydrogenolysis (H₂, 1 atm, 10 % Pd/C, MeOH, 15°C, 16 h) of Z-protected Asn (11) gave free amine (12) (86 %) which was then *N*-acylated with activated chromophore (3) (DCM, NEt₃ 1.1 eq., 20°C, 16 h) affording BOC protected amine (8) (79 %). Free amine (9),²⁰ incorporating the required protected chromophore-L-Asn-cadaverine moiety, was obtained by brief treatment (1 h) of BOC protected amine (8) with TFA in DCM (1:1) at 0°C (79 % as the free base after silica gel chromatography DCM:MeOH:conc. NH₄OH 75:10:1 v/v/v).¹⁹

The β -Ala.4.Arg molecular of NSTX-3 (1) was designed to be incorporated by acylation of primary amine (9). Therefore, polyamine-Arg (13) was prepared from putrescine (1,4-diaminobutane) (14). Mono-Z protection of putrescine (14) to afford carbamate $(15)^{21}$ was not found to be a practical strategy, yields were typically <3% using Z-Cl in aq. NaOH/THF at 0°C where the di-Z protected diamine predominated.²¹ A convenient way around this problem was via mono-BOC-mono-Z-putrescine. Mono-BOC protection of putrescine (14) (3.0 eq.) (BOC₂O 1.0 eq., THF, 0°C, 16 h) afforded carbamate (16) (76 %) which was then reacted with Z-Cl (1.1 eq.) under Schotten-Baumann conditions (1 M aq. NaOH, 1.1 eq., 0 to 20°C, 16 h) to give the required unsymmetrical dicarbamate (82 %) which was selectively deprotected with TFA in DCM (1:1) (0°C, 1 h) to yield mono-Z-putrescine (15)²¹ (76 %). Amine (15) underwent 1,4-Michael addition with t-butyl acrylate (1.1 eq.) (MeOH, 20°C, 16 h) to afford the desired conjugate (17) (39%). The protected terminal amine of conjugate (17) was hydrogenolysed (H2, 1 atm, 10 % Pd/C, MeOH, 15°C, 16 h) to yield primary amine t-butyl ester (18) (96 %) which was acylated with Z₃ArgOH (1.1 eq.) (DCM, DCC 1.5 eq., HOBt 0.05 eq., 20°C, 16 h) to afford the desired amide (19) (75%). The secondary amine functional group of the amide ester (19) was protected by carbamoylation with Z-Cl (1.1 eq.), in DCM, using NaOEt (1.1 eq. in EtOH) to yield the fully protected polyamine amide (20) (73 %) whose t-butyl ester was deprotected with TFA in DCM (1:1) (20°C, 16 h) to afford the desired protected β -Ala.4.Arg as the free acid (13) (91 %).

Putreanine moiety (13) was acylated with primary amine (9) (1.0 eq., DMF, 7 d, 20°C) after the acid had been activated as its pentafluorophenyl ester [free acid (13) activated with pentafluorophenol (1.1 eq.) (THF, DCC 1.5 eq., 16 h, 20°C)]. The activated ester was not isolated, but the solution was used directly in the next step, to afford protected NSTX-3 (21), in 65 % overall yield from primary amine (9).





Acknowledgements: We acknowledge the generous financial support of BBSRC/AFRC (AG86/521), The Wellcome Trust (Project Grant 039750), and MRC (G9219146N). ISB is the recipient of a Nuffield Foundation Science Lecturer's award (SCI/180/91/15/G).

REFERENCES

- 1. Aramaki, Y.; Yasuhara, T.; Higashijima, T.; Yoshioka, M.; Miwa, A.; Kawai, N.; Nakajima, T. Proc. Japan Acad. Sci. 1986, 62B, 359-362.
- 2. Toki, T.; Yasuhara, T.; Aramaki, Y; Kawai, N.; Nakajima, T. Biomedical Res. 1988, 9, 75-79.
- 3. Toki, T.; Yasuhara, T.; Aramaki, Y; Osowa, K.; Miwa, A.; Kawai, N.; Nakajima, T. *Biomedical Res.* 1988, 9, 421-428.
- 4. Teshima, T.; Wakamiya, T.; Aramaki, Y.; Nakajima, T.; Kawai, N.; Shiba, T. *Tetrahedron Letters* 1987, 28, 3509-3510.
- 5. Teshima, T.; Matsumoto, T.; Wakamiya, T.; Shiba, T.; Aramaki, Y.; Nakajima, T.; Kawai, N. Tetrahedron 1991, 47, 3305-3312.
- 6. Nason, D. M.; Jasys, V. J.; Kelbaugh, P. R.; Phillips, D.; Saccomano, N. A.; Volkmann, R. A. *Tetrahedron Letters* 1989, *30*, 2337-2340.
- Teshima, T.; Matsumoto, T.; Wakamiya, T.; Shiba, T.; Nakajima, T.; Kawai, N. *Tetrahedron* 1990, 46, 3813-3818.
- 8. Teshima, T.; Matsumoto, T.; Miyagawa, M.; Wakamiya, T.; Shiba, T.; Narai, N.; Yoshioka, M. *Tetrahedron* 1990, 46, 3819-3822.
- 9. Yoshioka, M.; Narai, N.; Teshima, T.; Matsumoto, T.; Wakamiya, T.; Shiba, T.; Tokoro, N.; Okauchi, T.; Kono, Y. Chem. Pharm. Bull. 1992, 40, 3005-3008.
- Ino, H.; Nakade, S.; Niinobe, M.; Ikenaka, K.; Teshima, T.; Wakamiya, T.; Matsumoto, T.; Shiba, T.; Kawai, N.; Mikoshiba, K. *Neuroscience Research* 1990, 8, 29-39.
- 11. Draguhn, A.; Jahn, W.; Witzemann, V. Neuroscience Letters 1991, 132, 187-190.
- 12. Usherwood, P. N. R.; Blagbrough, I. S. Pharmacol. Therap. 1991, 52, 245-268.
- 13. Blagbrough, I. S.; Usherwood, P. N. R. Proc. Roy. Soc. Edin. 1992, 99B, 67-81.
- 14. Raditsch, M.; Ruppersberg, J. P.; Kuner, T.; Gunther, W.; Schoepfer, R.; Seeburg, P. H.; Jahn, W; Witzemann, V. FEBS Letters 1993, 324, 63-66.
- Herlitze, S.; Raditsch, M.; Ruppersberg, J. P.; Jahn, W.; Monyer, H.; Schoepfer, R.; Witzemann, V. Neuron 1993, 10, 1131-1140.
- 16. Carter, C. (ed.) The Neuropharmacology of Polyamines, Academic Press, London, 1994, 1-318.
- 17. Chiba, T.; Akizawa, T.; Matsukawa, M.; Pan-Hou, H.; Yoshioka, M. Chem. Pharm. Bull. 1994, 42, 1864-1869.
- For recent reviews, in the general area of polyamine amide syntheses, see: McCormick, K. D.; Meinwald, J. J. Chem. Ecol. 1993, 19, 2411-2451; Schäfer, A.; Benz, H.; Fiedler, W.; Guggisberg, A.; Bienz, S.; Hesse, M. The Alkaloids 1994, 45, 1-125; Moya, E.; Blagbrough, I. S. in: The Neuropharmacology of Polyamines, (ed. Carter, C.), Academic Press, London 1994, 167-184.
- 19. Moya, E.; Blagbrough, I. S. Tetrahedron Letters 1994, 35, 2061-2062.
- Hashimoto, Y.; Endo, Y.; Shudo, K.; Aramaki, Y.; Kawai, N.; Nakajima, T. Tetrahedron Letters 1987, 28, 3511-3514.
- 21. Atwell, G. J.; Denny, W. A. Synthesis 1984, 1032-1033.

(Received in UK 3 October 1995; accepted 24 November 1995)