ORIGINAL RESEARCH

New aminoadamantane derivatives with antiproliferative activity

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Abstract 1-Benzyl-2-aminoadamantanes 2a–c, conformationally constrained aminoadamantanes 3a, b and 4 and diaminoadamantanes derivatives 5a–c, 6a–c were synthesized and tested as antiproliferative agents. The in vitro biological evaluation showed a significant difference in activity between 1-phenyl and 1-benzyl derivatives.

Keywords Aminoaromatic adamantanes · Synthesis · Antiproliferative activity

Introduction

Adamantane derivatives have a broad pharmacological profile, involving antibacterial (Aigami *et al.*, 1975), antifungal (Kadi *et al.*, 2007), antiviral (Kolocouris *et al.*, 1994, 1996), trypanocidal (Papanastasiou *et al.*, 2008) and

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A. Vamvakides Anavex Life Sciences, 27 Marathonos Avenue, 153 51 Pallini, Athens, Greece anti-Parkinson (Chakrabarti et al., 1976) activity. Moreover, it is well documented that the incorporation of the adamantyl moiety into the skeleton of various molecules increases their antiproliferative potency, possibly due to the lipophilic character of adamantane or its unique structure. Such examples include aromatic carboxylic acids (Cincinelli et al., 2003; Dawson et al., 2008), pyrimidines, pyridines (Kazimierczuk et al., 2001) and cyclic imides (Shibata et al., 1995; Tsuji et al., 2000; Wang et al., 1997, 1998, 2002) with anticancer activity. Recently, aminoaromatic adamantane (Wang et al., 2004) and bisubstituted adamantane derivatives, with marked antiproliferative activity on different human cell lines, were reported (Zefirova et al., 2002; Smith et al., 2006; Wang et al., 2003). In our effort to develop new antiproliferative agents, we have synthesized 1-benzyl-2-aminoadamantanes 2a-c and the conformationally constrained fused three-membered ring systems 3 and 4. We have also introduced into the skeleton of derivatives 2 a second nitrogen atom, 2 or 3 carbon atoms away from the first nitrogen (diamines 5 and 6, respectively) to probe the influence on anticancer potency of two nitrogens in these scaffolds (Fig. 1).

Materials and methods

Cell cultures and cell lines

All human cancer cell lines were obtained from the National Cancer Institute, NIH (Bethesda, MD, USA). All cell lines were adapted to propagate in RPMI 1640 medium supplemented with 5 % heat-inactivated fetal calf serum, 2 mM L-glutamine and antibiotics. The cultures were grown in a humidified 37 °C incubator in 5 % CO_2 atmosphere.

Fig. 1 New adamantane derivatives with antiproliferative activity



In vitro cytotoxic activity

Cell viability was assessed at the beginning of each experiment by the trypan blue dye exclusion method and was always greater than 95 %. Cells were seeded into 96-well microtiter plates in 100 µL of medium at the corresponding density (3,500–30,000 cells per well) and, subsequently, the plates were incubated at standard conditions for 24 h to allow the cells to resume exponential growth prior to addition of the agents to be tested. Then to measure the cell population, cells in one plate were fixed in situ with TCA followed by SRB staining, as described elsewhere (Skehan et al., 1990; Keepers et al., 1991). To determine the activity, each compound was dissolved in DMSO and then added at tenfold dilutions (from 100 to 0.01 μ M) and incubation continued for an additional period of 48 h. The assay was terminated by addition of cold TCA followed by SRB staining and absorbance measurement at 540 nm, in a DAS plate reader, to determine GI₅₀, that is, the concentration required in the cell culture to inhibit cell growth by 50 %, and TGI, the concentration that is required to completely inhibit cell growth (Skehan et al., 1990; Keepers et al., 1991).

Synthesis

Melting points were determined using a Büchi capillary apparatus and are uncorrected. IR spectra were recorded on a Perkin-Elmer 833 spectrometer. ¹H NMR spectra were recorded on a Bruker MSL 400 and ¹³C NMR spectra were taken on a Bruker AC 200 spectrometer using CDCl₃ as solvent and TMS as internal standard. Carbon multiplicities were established by DEPT experiments. 2D NMR experiments (HMQC, COSY and NOESY) were performed for the elucidation of the structures of the new compounds. Microanalyses were carried out by the Service Central de Microanalyse (CNRS), France, and the results obtained had a maximum deviation of ± 0.4 % from the theoretical values.

General procedure for the synthesis of 1benzyltricyclo[$3.3.1.1^{3,7}$]decan-2-ketoxime (**8a**), 1-(4fluorobenzyl)-tricyclo[$3.3.1.1^{3,7}$]decan-2-ketoxime (**8b**) and 1-(4-methoxybenzyltricyclo[$3.3.1.1^{3,7}$]decan-2ketoxime (**8c**)

Hydroxylamine hydrochloride (1.4 g, 20.2 mmol) and sodium acetate trihydrate (4.26 g, 31.3 mmol) were added to a 90° ethanolic solution (50 mL) of ketones **7a** (Papanastasiou *et al.*, 2008), **7b** (Papanastasiou *et al.*, 2009) and **7c** (Papanastasiou *et al.*, 2009) (10.1 mmol), respectively, and the resulting mixture was refluxed for 5 h. Upon completion of the reaction, ethanol was evaporated, chilled water (20 mL) was added and the precipitate formed was filtered off, washed with water and dried to give **8a**, **8b** and **8c**, respectively, as solids in quantitative yield; **8a**: mp 210 °C (ether), **8b**: mp 216 °C (ether–pentane), **8c**: mp 182 °C (ether–pentane).

General procedure for the synthesis of 1benzyltricyclo[3.3.1.1^{3,7}]*decan-2-amine* (**2a**), 1-(4-fluoro*benzyl)tricyclo*[3.3.1.1^{3,7}]*decan-2-amine* (**2b**) and 1-(4*methoxybenzyltricyclo*[3.3.1.1^{3,7}]*decan-2-amine* (**2c**)

Oximes 8a, 8b and 8c (13.7 mmol) in absolute ethanol (40 mL) were hydrogenated under pressure (55 psi) over Raney-Ni (200 mg) for 5 h, at 70 °C. The catalyst was filtered off through Celite and the solvent was evaporated under vacuum to give the crude title compounds as viscous oils; amines 2a and 2b were crystallized upon standing. 2a mp 38 °C (Papanastasiou *et al.*, 2008); **2b** mp 40 °C. ¹H NMR (CDCl₃) δ 1.26–1.97 (13H, m, H_{3–10}), 1.98 (2H, brs, NH₂, D₂O exchangeable), 2.36–2.49 (2H, AB, J = 13.2, $\Delta v = 0.05$, CH₂Ph), 2.61 (1H, s, H₂), 6.84–7.03 (4H, m, H_{2,3,5,6}-arom.); ¹³C NMR (CDCl₃) δ 28.0 (C₅), 28.1 (C₇), 30.4 (C₆), 35.64 (C₄), 35.8 (C₃), 36.8 (C₁), 37.0 (C₁₀), 37.4 (C₉), 40.9 (C₈), 45.0 (CH₂Ph), 57.8 (C₂), 114.2-114.4 $(C_{3,5}\text{-}arom.), 131.8\text{-}133.9 (C_{2,6}\text{-}arom.), 133.4 (C_1\text{-}arom.),$ 162.6 (C₄-arom.); **2c**, ¹H NMR (CDCl₃): δ 1.25–1.83 (13H, m, H₃₋₁₀), 1.85 (2H, brs, NH₂, D₂O exchangeable), 2.39–2.50 (2H, AB, J = 13.6, $\Delta v = 0.04$, CH_2Ph), 2.66 (1H, s, H₃), 3.78 (3H, s, CH₃O), 6.78–7.05 (4H, m, H_{2.3.5.6}arom.); ¹³C NMR (CDCl₃) δ 28.1 (C₅), 28.2 (C₇), 30.5 (C₆), 35.7 (C₄), 35.8 (C₃), 37.1 (C₁₀), 37.5 (C₉), 42.6 (C₁), 40.9 (C₈), 44.9 (CH₂Ph), 55.1 (CH₃O) 55.7 (C₂), 112.9-113.1 (C_{3.5}-arom.), 129.8 (C₁-arom.), 131.4 (C_{2.6}arom.), 157.7 ppm (C₄-arom.). Elemental analyses measurements were performed on the hydrochloride salts of amines **2b** and **2c**. **2b** (hydrochloride, mp >250 °C); Anal calcd for C17H23ClFN: C 69.02, H 7.84, found C 68.86, H 7.86. 2c (hydrochloride, mp >250 °C); Anal calcd for C₁₈H₂₆ClNO: C 70.22, H 8.51, found C 70.31, H 8.43.

1-Phenyltricyclo[3.3.1.1^{3,7}]decan-2-carboxylic acid (12)

Dry dimethylsulfoxide (105 mL) was added dropwise to a mixture of trimethylsulfoxonium iodide (7.92 g, 36 mmol) and sodium hydride 60 % (1.44 g, 36 mmol), under an argon atmosphere, and stirred for 30 min before ketone **9** (Lenoir, 1973; Tseng *et al.*, 1998) (6 g, 26.5 mmol) was added. The resulting mixture was stirred at 20 °C for 30 min and then heated at 55–58 °C for 2 h. The mixture was then poured into chilled water and extracted with *n*-hexane 3×20 mL without shaking the separatory funnel. The combined organic layers were dried over Na₂SO₄ and evaporated at 40 °C to afford 6.21 g of a liquid, which was diluted with dry benzene (120 mL) and shaken well with BF₃·Et₂O complex (3.6 g, recently distilled) in the separatory funnel. After 3 min at rest,

the organic phase was washed with H₂O (3×15 mL) and dried over Na₂SO₄. The organic phase was partially evaporated (40-50 mL) at 30 °C, acetone was added, followed by evaporation; this procedure was repeated twice. Then, acetone was added to give a volume of 150-200 mL, and the solution was oxidized with Jones reagent (~33 mL, 1 M) at 15 °C. The reaction mixture was stirred overnight at 20 °C, then quenched by the addition of isopropyl alcohol (3 mL) and filtered through cotton wool. H₂O (20 mL) was added to the filtrate and acetone was removed under reduced pressure. The residue was extracted with ether and the organic extracts were washed with an aqueous solution of NaOH (5 %). The combined aqueous phases were washed with ether and acidified with conc. HCl at 0 °C. The crystals formed were filtered off, washed with H₂O and dried to afford carboxylic acid 12 (4.73 g, 70 %); mp: 127 °C (ether); ¹H NMR (CDCl₃) δ 1.51-2.75 (13H, m, H₃₋₁₀), 3.04 (1H, s, H₂), 7.06-7.24 (5H, m, H-arom.), 10.5 (1H, brs, COOH); ¹³C NMR (CDCl₃) δ 27.9 (C₅), 28.8 (C₇), 31.6 (C₆), 31.8 (C₃), 35.1 (C₄), 36.8 (C₁₀), 37.5 (C₁), 38.0 (C₉), 47.1 (C₈), 53.8 (C₂), 124.9 (C_{2.6}-arom.), 125.8 (C₄-arom.), 128.1 (C_{3.5}-arom.), 148.5 (C₁-arom.), 179.0 (CO); IR (Nujol): $v = 1,689-1,634 \text{ cm}^{-1}$ (CO).

Indeno[1,2-a]tricyclo[3.3.1.1^{3,7}]decan-5(5aH)-one (13)

Carboxylic acid 12 (3 g, 11.7 mmol) was treated with freshly distilled thionyl chloride (8 mL), at 65 °C, during 15-20 min. After removal of residual thionyl chloride by co-evaporation with benzene under reduced pressure, carbon disulfide (45 mL) and anhydrous aluminum chloride (1.8 g, 13.4 mmol) were added to the reaction mixture, which was stirred at RT for 30 min and then refluxed for 30 min. The reaction was cooled, quenched by the addition of ice and extracted with diethyl ether $(3 \times 20 \text{ mL})$. The combined organic phases were washed with H₂O, dried over Na₂SO₄ and evaporated to afford a residue, which crystallized upon standing. Further purification by flash column chromatography over silica gel at gradient eluent (pentane to pentane:ether, 6:1) gave compound 13 (2.81 g, almost quantitative) mp: 56 °C; 1H NMR (CDCl₃) δ 1.93-2.57 (13H, m, H₆₋₁₃), 2.57 (1H, s, H_{5a}), 7.19-7.66 (4H, m, H-arom.); ¹³C NMR (CDCl₃) δ 26.9 (C₈), 27.9 (C₁₀), 28.6 (C₆), 32.10 (C₉), 36.0 (C₇), 38.2 (C₁₃), 38.7 (C12), 41.5 (C11a), 46.9 (C11), 62.8 (C5a), 121.6 (C1), 124.0 (C₃), 127.0 (C₂), 133.3 (C₄), 135.7 (C_{11b}), 164.5 (C_{4a}), 204.6 (CO); IR (Nujol): $v = 1,724 \text{ cm}^{-1}(\text{CO})$.

Indeno[1,2-a]tricyclo[3.3.1.1^{3,7}]decan-5-ketoxime (14)

A mixture of ketone **13** (0.31 g, 1.3 mmol), hydroxylamine hydrochloride (0.27 g, 3.9 mmol) and sodium acetate trihydrate (0.81 g, 6 mmol) in ethanol (90°) (15 mL) was

refluxed for 7 h. The reaction mixture was then cooled to room temperature, ethanol was evaporated and chilled water was added. The precipitate formed was filtered off, washed with water and dried to give ketoxime **14** as a solid (0.32 g, yield quantitative) mp: 164 °C (ethanol–water, then dried with hexane).

5,5*a*-Dihydroindeno[1,2-*a*]tricyclo[3.3.1.1^{3,7}]decan-5amines (**3a**) and (**3b**)

Acetic anhydride (4.32 g, 42.4 mmol) was added to a chilled solution of oxime 14 (0.59 g, 2.34 mmol) in pyridine (10 mL). The reaction mixture was stirred at 0 °C for 7 h and 18 h at 3 °C and then diluted with diethyl ether prior to addition of ice. The aqueous phase was discarded and the organic laver sequentially washed with H₂O, saturated aqueous solution of NaHCO₃ and H₂O. The organic phase was dried over Na₂SO₄ and concentrated in vacuo to give oxime-ester 15 (0.72 g, yield almost quantitative); IR (net): $v = 1,764 \text{ cm}^{-1}$ (CO), 1,644 cm⁻¹ (CN). To a stirred solution of oxime-ester 15 (0.7 g, 2.17 mmol) in anhydrous tetrahydrofuran (10 mL) was added sodium borohydride (0.65 g, 17.4 mmol) and a solution of iodine (1.67 g, 6.61 mmol) in dry tetrahydrofuran (10 mL) at 0 °C. The reaction mixture was refluxed for 8 h and then quenched at 0 °C by the addition of an aqueous solution of HCl (3N). The mixture was then made alkaline with aqueous conc. KOH, extracted with diethyl ether, washed with H₂O and dried over anhydrous Na₂CO₃. The ethereal phase was treated with a saturated ethanolic solution of gaseous HCl under ice cooling and the precipitate formed was filtered off, washed with cold diethyl ether and dried to afford the respective hydrochloride salts of the desired compounds, which were then converted to the free bases by the addition of a solution of Na₂CO₃. These were further purified by flash column chromatography, eluting with a 1:1 MeOH-Et₂O solution to give viscous oils, which were then separated to the two diastereomers, respectively, eluting with a 13:1 Et₂O-MeOH mixture. The less polar compounds correspond to the cis-diastereomers, which are solids, while the more polar to the trans-diastereomers, which are liquid (80 mg each diastereomer, total yield 31.5 %). 3a: ¹H NMR (CDCl₃) δ 1.35–2.22 (16H, m, H_{5a-13}, NH₂), 4.12 $(1H, d, J = 10.6, H_5), 6.99-7.14 (4H, m, H-arom.); {}^{13}C NMR$ (CDCl₃) δ 27.7 (C₈), 28.1 (C₁₀), 29.1 (C₆), 31.10 (C₉), 37.4 (C₇), 39.1 (C₁₃), 40.6 (C₁₂), 40.8 (C₁₁), 42.5 (C_{11a}), 55.7 (C_{5a}), $64.0 \hspace{0.2cm} (C_5), \hspace{0.2cm} 120.6 \hspace{-0.2cm} -123.7 \hspace{-0.2cm} -126.4 \hspace{-0.2cm} -126.9 \hspace{0.2cm} (C_{1-4}), \hspace{0.2cm} 146.7 \hspace{-0.2cm} -151.5$ (C_{4a-11b}) . **3b**: ¹H NMR (CDCl₃) δ 1.66–2.29 (16H, m, H_{5a-13}, NH_2), 4.25 (1H, d, J = 7.16, H_5), 7.05–7.15 (4H, m, H-arom); ¹³C NMR (CDCl₃) δ 28.2 (C₈), 29.7 (C₁₀), 30.0 (C₆), 33.0 (C₉), 37.5 (C7), 39.1 (C11a), 40.9 (C13), 42.0 (C12), 44.3 (C11), 54.0 58.6 (C_5), 121.6–125.3–126.7–127.5 (C_{1-4}), $(C_{5a}),$ 146.9–152.6 (C_{4a-11b}). The precipitation of the hydrochloride salts from the ethereal solution of the amine was effected by the addition of *n*-pentane. Hydrochloride (*trans*-amine) **3a**; mp >255 °C. Anal. calcd for $C_{17}H_{22}CIN$: C 74.03, H 8.04, found C 73.91, H 8.12. Hydrochloride (*cis*-amine) **3b**; mp >255 °C. Anal. calcd for $C_{17}H_{22}CIN$: C 74.03, H 8.04, found C 73.96, H 8.05.

5H,6H-tricyclodecan[1,2-a]isoquinoline (4)

A mixture of oxime 14 (0.9 g, 3.55 mmol) and phosphorus pentachloride (1 g, 4.79 mmol) in benzene (20 mL) was refluxed for 1 h. After cooling, ice was added and the mixture extracted with benzene. The organic phase was washed sequentially with a saturated aqueous solution of NaHCO₃ and H₂O, dried over Na₂SO₄ and evaporated to give a residue, which was treated with a mixture of ether: n-pentane (1:1) and chilled to give a precipitate, which was filtered off and washed with *n*-pentane. Recrystallization from methanol gave 16 >250 °C; (260 mg, 29 %); mp IR (Nujol): $v = 3,435-3,175 \text{ cm}^{-1}(\text{NH}), 1,667 \text{ cm}^{-1}(\text{CO}).$ Compound 16 (238 mg, 0.94 mmol) was then added dropwise into a suspension of lithium aluminum hydride (400 mg, 10.5 mmol) in anhydrous tetrahydrofuran (30 mL) and the reaction mixture was refluxed for 12 h. Quenching by the sequential addition of H2O and an aqueous solution of NaOH (5 %) gave a suspension, which was filtered off, the filtrate dried over Na₂CO₃ and the solvent evaporated to give a residue, which was transformed into the hydrochloride salt of 4. The salt was recrystallized from ethanol-ether and then converted to the free base 4 upon treatment with Na₂CO₃. Base 4 was further purified by flash column chromatography using a 1:9 MeOH-Et₂O mixture as eluent to give 4 as a viscous oil, which was crystallized on standing (0.13 g, 58 %); mp: 79 °C; ¹H NMR (CDCl₃) δ 1.50–2.31 (13H, m, H_{6a-14} , NH), 2.83 (1H, s, H_{6a}), 4.01–4.18 (2H, AB, J = 16, H₅), 6.91–7.22 (4H, m, H-arom.); 13 C NMR (CDCl₃) δ 28.4 (C₉), 28.9 (C₁₁), 30.68 (C₁₀), 33.9 (C₇), 35.5 (C_{12a}), 37.2 (C₈), 37.6 (C₁₄), 40.6 (C₁₃), 41.2 (C₁₂), 49.4 (C₅), 62.2 (C_{6a}), 124.9–125.5–126.0–126.2 (C_{1-4}), 134.6–144.5 (C_{4a-12b}); hydrochloride salt: mp >250 °C. Anal calcd for $C_{17}H_{22}CIN$: C 74.03, H 8.04, found C 74.15, H 8.08.

General procedure of synthesis of 1-benzyltricycle [3.3.1.1^{3,7}] decan-2-(2-dimethylaminoethyl)amine (**5a**), 1-benzyltricycle [3.3.1.1^{3,7}]decan-2-(N-piperidinylethyl)amine (**5b**), 1-benzyltricyclo[3.3.1.1^{3,7}]decan-2-(N-piperazinylethyl) amine (**5c**), 1-benzyltricyclo[3.3.1.1^{3,7}]decan-2-(3-dimethyl aminopropyl)amine (**6a**), 1-benzyltricyclo[3.3.1.1^{3,7}]decan-2-(N-morpholinopropyl)amine (**6b**) and 1-benzyltricyclo [3.3.1.1^{3,7}]decan-2-(1-imidazolylpropyl)amine (**6c**)

A mixture of ketone **7a** (8.66 mmol) and the requisite diamine (10 mmol) in anhydrous toluene (20 mL) was refluxed for 6 h, while removing H_2O azeotropically. The solvent was then evaporated to give a viscous oil, which was reduced with

lithium aluminum hydride (50 mmol) in anhydrous diethyl ether (30 mL), upon refluxing for 10 h. The reaction was then quenched by the sequential addition of H₂O and an aqueous solution of NaOH (5 %) and the solid filtered off. The filtrate was dried over Na₂CO₃ and evaporated to give a residue, which was further purified by flash column chromatography over silica gel using a mixture of Et₂O:MeOH as eluent to give the desired amine, as an oil, which was transformed to the respective fumarate salt. Compound 5a: purification by column chromatography-eluent system Et₂O:MeOH (2:1), yield 67 %; ¹H NMR (CDCl₃) δ 1.32–1.90 (13H, m, H_{3–10}), 2.18 (7H, s, $(CH_3)_2N$, CHNH), 2.31–2.63 (2H, AB, J = 12.8, CH₂Ph), 2.40–2.43 (3H, m, NHCHH + $CH_2N(CH_3)_2$), 2.74 (1H, m, NHCHHCH₂N(CH₃)₂), 7.07–7.17 (5H, m, H-arom.); ¹³C NMR (CDCl₃) δ 28.1 (C₅), 28.2 (C₇), 30.3 (C₃), 30.7 (C₆), 37.2 $(C_4),$ 37.3 $(C_{10}),$ 37.4 $(C_9),$ 41.6 $(C_8),$ 45.0 (NHCH₂CH₂N(CH₃)₂), 45.5 ((CH₃)₂N), 45.8 (CH₂Ph), 59.7 (NHCH₂CH₂N(CH₃)₂), 63.7 (C₂), 71.8 (C₁), 125.5 (C₄-arom.), 127.4 (C_{2.6}-arom), 130.8 (C_{3.5}-arom.), 138.5 (C₁-arom.); fumarate, mp: 171 °C. Anal calcd for C₂₅H₃₆N₂O₄: C 70.06, H 8.47, found C 69.97, H 8.41. Compound 5b: purification by column chromatography-eluent system Et₂O:MeOH (1:1), yield 92 %;¹H NMR (CDCl₃) δ 1.13–1.78 (19H, m, H_{3–10}, H₃₄₅-piper.), 2.20 (1H, ~d, CHNH), 2.33–2.45 (8H, AB, J = 12.8, CHHPh, NHCHH + CH₂N, H_{2,6}-piper.), 2.64 (1H, AB, J = 12.8, CH*H*Ph), 2.60–2.64 (1H, m, NHC*H*HCH₂N), 7.08-7.18 (5H, m, H-arom.); ¹³C NMR (CDCl₃) δ 24.4 (C₄pip), 26.0 (C_{3.5}-piper.), 28.1 (C₅), 28.2 (C₇), 30.3 (C₃), 30.7 $(C_6),$ 37.1-37.2-37.3 $(C_{4.9,10}), 41.5$ $(C_8),$ 44.36 (NHCH₂CH₂N), 45.8 (CH₂Ph), 54.7 (C_{1.6}-piper.), 59.0 (NHCH₂CH₂N), 64.0 (C₂), 71.8 (C₁), 125.5(C₄-arom.), 127.4 (C2.6-arom.), 130.7 (C3.5-arom.), 138.5 (C1-arom.); fumarate, mp: 187 °C. Anal calcd for C₂₈H₄₀N₂O₄: C 71.76, H 8.60, found C 71.82, H 8.63. Compound 5c: purification by column chromatography-eluent system Et₂O:MeOH (3:2), yield 40 %; ¹H NMR (CDCl₃) δ 1.43–1.97 (13H, m, H_{3–10}), 2.27 (1H, ~s, J = 0.2, CHNH), 2.40–2.53 (8H, AB, J = 12.8, CHHPh, NHCHH + CH₂N, H_{3.5}-piperaz.), 2.64 (1H, AB, J = 12.8, CHHPh), 2.81 (1H, m, NHCHHCH₂N), 2.90 (4H, m, H₂₆piperaz.), 7.14–7.26 (5H, m, H-arom.); 13 C NMR (CDCl₃) δ 28.2 (C₅), 28.3 (C₇), 30.4 (C₃), 30.8 (C₆), 37.2-37.3-37.4 (C₄₉₁₀), 41.6 (C₈), 44.1 (NHCH₂CH₂N), 45.9 (CH₂Ph), 46.3 (C₄-piperaz.), 54.7 (C₂-piperaz.), 59.0 (NHCH₂CH₂N), 64.0 (C₂), 125.5 (C₄-arom.), 127.4 (C_{2.6}-arom.), 130.7 (C_{3.5}-arom.), 138.5 (C1-arom.); fumarate, mp: 168 °C. Anal calcd for C₂₇H₃₉N₃O₄: C 69.05, H 8.37, N 8.95, found C 69.12, H 8.32. Compound 6a: purification by column chromatography-eluent system Et₂O:MeOH (3:2), yield 61 %; ¹H NMR (CDCl₃) δ 1.25-2.00 (15H, m, H₃₋₁₀, CH₂CH₂CH₂), 2.24 (6H, s, $(CH_3)_2N$), 2.28 (1H, ~s, CHNH), 2.36–2.43 (4H, m, AB, J = 12.8, CHHPh, NHCHHCH₂CH₂N(CH₃)₂), 2.68–2.78 (2H, m, AB, J = 12.8, CH*H*Ph, NHCH*H*CH₂CH₂N(CH₃)₂), 7.14-7.26 (5H, m, H-arom.); ¹³C NMR (CDCl₃) δ 28.1 (C₅),

28.2 (C₇), 28.9 (CH₂CH₂CH₂), 30.1 (C₃), 30.8 (C₆), 37.16 (C₄), 37.3 (C₁₀), 37.4 (C₉), 41.7 (C₈), 45.6 ((CH₃)₂N), 45.8 (NHCH₂CH₂CH₂N(CH₃)₂), 45.8 (CH₂Ph), 58.3 (NHCH₂CH₂ CH₂N(CH₃)₂), 63.7 (C₂), 71.8 (C₁), 125.5 (C₄-arom.), 127.4 (C_{2.6}-arom.), 130.7 (C_{3.5}-arom.), 138.6 (C₁-arom.); fumarate, mp: 170 °C. Anal calcd for C₂₆H₃₈N₂O₄: C 70.56, H 8.65, found C 70.45, H 8.71. Compound 6b:purification by column chromatography-eluent system Et₂O:MeOH (3:2), yield 52 %; ¹H NMR (CDCl₃) δ 1.24–2.00 (15H, m, H_{3–10}, CH₂CH₂CH₂), 2.27 (1H, \sim s, CHNH), 2.37–2.70 (8H, m, AB, J = 14, CHHPh, NHCHHCH₂CH₂N, H_{3 5}-morph.), 2.73–2.85 (2H, m, AB, J = 14, CH*H*Ph, NHCH*H*CH₂CH₂N), 3.72–3.77 (4H, m, H_{2.6}-morph.), 7.13–7.26 (5H, m, H-arom.); ¹³C NMR (CDCl₃) δ 27.5 (CH₂CH₂CH₂), 28.2 (C₅), 28.3 (C₇), 30.1 (C₃), 30.9 (C_6) , 37.2 (C_4) , 37.3 (C_{10}) , 37.6 (C_9) , 41.7 (C_8) , 45.9 (NHCH₂CH₂CH₂N), 46.0 (CH₂Ph), 53.9 (C_{3.5}-morph.), 57.9 (NHCH₂CH₂CH₂N), 63.6 (C₂), 67.1 (C_{2.6}-morph.), 71.8 (C₁), 125.5 (C₄-arom.), 127.4 (C_{2.6}-arom), 130.7 (C_{3.5}-arom.), 138.6 (C₁-arom.); fumarate, mp: 178 °C. Anal calcd for C₂₈H₄₀N₂O₅: C 69.39, H 8.32, found C 69.43, H 8.47. Compound 6c: purification by column chromatography-eluent system Et₂O:MeOH (2:1), yield 60 %; ¹H NMR (CDCl₃) δ 1.26–1.97 (15H, m, H_{3-10} , $CH_2CH_2CH_2$), 2.27 (1H, ~s, CHNH), 2.41 (1H, quint, NHCHHCH2CH2N), 2.58 (8H, AB, J = 13.2, CHHPh), 2.72 (1H, quint, NHCHHCH₂CH₂N), 4.12 (2H, t, NHCH₂CH₂CH₂N), 6.94–7.50 (8H, m, H-phen.+imid.); ${}^{13}C$ NMR (CDCl₃) δ 27.9 (C₅), 28.1 (C₇), 30.1 (C₃), 30.7 (C₆), 32.07 (NHCH₂CH₂CH₂N) 36.9 (C₄), 37.0 (C₁₀), 37.3 $(C_9),$ 41.7 $(C_8),$ 43.6 (NHCH₂CH₂CH₂N), 44.9 (NHCH₂CH₂CH₂N), 46.0 (CH₂Ph), 64.0 (C₂), 118.8 (C-imid.), 125.6 (C₄-phen.), 127.5 (C_{2.6}-phen.), 129.3 (C-imid.),130.6 (C_{3 5}-arom.), 137.2 (C-imid.), 138.3 (C₁-phen.); fumarate, mp: 171 °C. Anal calcd for C₂₇H₃₅N₃O₄: C 69.65, H 7.58, found C 69.54, H 7.66.

Results and discussion

Chemical synthesis

For the preparation of 1-benzyl-2-aminoadamantanes $2\mathbf{a}-\mathbf{c}$, the previously reported ketones $7\mathbf{a}-\mathbf{c}$ (Papanastasiou *et al.*, 2009) were converted to their respective oximes $8\mathbf{a}-\mathbf{c}$, which were then catalytically reduced to the desired amines $2\mathbf{a}-\mathbf{c}$ (Scheme 1).

Amine 1 was prepared from 1-phenyl-2-adamantanone (Lenoir, 1973; Tseng *et al.*, 1998; Zoidis *et al.*, 2010), in a similar way to amines 2. For the preparation of the diastereomeric amines **3a** and **3b**, phenylketone 9 (Lenoir, 1973; Tseng *et al.*, 1998) was treated with the appropriate sulfur ylide to give epoxide 10. This was treated with $BF_3 \cdot Et_2O$ complex to give aldehyde 11, which was immediately oxidized, under Jones conditions, to the



Scheme 1 Reagents and conditions: $a \text{ NH}_2\text{OH}$ HCl, CH₃CO-ONa·3H₂O, EtOH, reflux for 5 h, (quant.); $b \text{ H}_2/\text{Ni-Raney}$, EtOH, 55 psi, 70 °C for 5 h, (91–95 %)

carboxylic acid **12** in good yield. This acid was then converted to its chloride, which, in the presence of AlCl₃, was cyclized to ketone **13**. Treatment of **13** with hydroxylamine gave oxime **14**, which was transformed to the oxime-ester **15**. This was reduced by diborane, prepared in situ from NaBH₄ and iodine, to the diastereomeric amines **3a** and **3b**. The two diastereomers were separated by flash column chromatography and characterized by NMR spectral analysis. $J_{6H-6aH} \approx 7.16$ Hz coupling is attributed to the *cis*-**3b** amine and $J_{6H-6aH} \approx 10.6$ Hz to the *trans*-**3a** amine (Scheme 2).

Beckmann rearrangement of oxime 14 on treatment with PCl₅ led to the formation of lactam 16, which was selectively reduced by $LiAlH_4$ to provide amine 4 (Scheme 3).

The selectivity of this reaction can be attributed to the stability of the produced cation **16a**, which leads to lactam **16**. Conversely, cation **17a** is less stable and thus the corresponding lactam **17** is not formed (Fig. 2).



Scheme 3 Reagents and conditions: *a* PCl₅, anhydrous C_6H_6 , reflux for 1 h. *b* LiAlH₄, anhydrous THF, reflux 12 h

The elucidation of the structure of amine **4** was based on the NMR spectra. Thus, the 5-CH₂ peak appears as an AB system with J = 16.1 Hz, which justifies its linkage to the phenolic ring and not with the C2 carbon atom of the adamantane skeleton.

The synthesis of the ethanodiamines **5** and propanodiamines **6** was effected by heating the appropriate diamines with ketone **7a**, followed by azeotropic removal of water. The intermediate imines **18** were not isolated, but reduced directly to the corresponding amines **5a**, **5b**, **5c** and **6a**, **6b**, **6c** (Scheme 4).

Pharmacology

The results from the in vitro drug screening of the new compounds on cancer cell lines (colon, prostate, breast, ovarian, central nervous system, leukemia, pancreas, liver) and on normal cell lines are shown in Table 1.

Considering the antiproliferative activity of the active analogs on various cancer cell lines, it appears that aminoadamantanes **2a–c** are the most potent compounds, derivative **2b** being the most active across all examined cancer cell lines. Moreover, the OMe-substituted analog **2c** proved to be more efficient than the parent compound **2a**. The C2-diaminosubstituted adamantanes (compounds **5b** and **6a**) show a

Scheme 2 Reagents and conditions: a trimethylsulfoxonium iodide, NaH, anhydrous DMSO, 20 °C for 30 min, 55-58 °C for 2 h. b BF₃·Et₂O, anh. C₆H₆. c CrO₃, H₂SO₄ (8N), 15-20 °C. d SOCl₂, 65 °C for 15 min. e AlCl₃, CS₂, rt for 30 min, reflux for 30 min. f NH₂OH.HCl, CH₃COONa·3H₂O, EtOH, reflux for 7 h, (quant.). g (CH₃CO)₂O, Py, 0 °C for 7 h and 3 °C for 18 h, (quant.). h NaBH₄/I₂, anhydrous. THF, reflux for 8 h





Scheme 4 Reagents and conditions: a n = 2, i, 2-dimethylaminoethylamine, toluene, reflux for 6 h. ii, 2-(piperidin-1-yl)ethylamine, toluene, reflux for 6 h. iii, 2-(piperazin-1-yl)ethylamine, reflux for 6 h; n = 3, i, 3-dimethylaminopropylamine, toluene, reflux for 6 h. ii,

3-morpholinopropylamine, toluene, reflux for 6 h. iii, 3-(1*H*-imidazol-1-yl)propylamine, toluene, reflux for 6 h. *b* LiAlH₄, anhydrous Et₂O, reflux 10 h, (40–92 %)

Compound	Values ^a	Cancer cell lines											
		Colon			Renal	Prostate		Breast					
		Colo205	HCT-116	HCT-15	Caki	DU145	PC3	LNCap	MDA MB231	MCF7			
1 ^b	TGI	100				100	89.3			100			
	GI50	84.53				100	82.92			69.3			
2a ^b	TGI	33.23	35.84		26.84	29.77			46.6	9.5			
	GI50	7.12	6.59		6.54	6.68			6.68	5.5			
2 b ^b	TGI		17.46	53.82						8.7			
	GI50		5.6	19.39						4.6			
2c ^b	TGI		52.17	75.05						42.3			
	GI50		18.49	38.81						7.4			
3a ^b	TGI	69.89				37.62		61.2		62.47			
	GI50	34.95				26.63		28.53		32.62			
3b ^b	TGI					66.13	56.74						
	GI50					31.45	21.34						
4	TGI		100.0							100.0			
	GI50		63.92							52.0			
5a ^c	TGI	59.94	55.87					54.7		54.09			
	GI50	25.44	24.02					24.36		25.86			

Table 1 Results from the in vitro drug screening

Table 1 continued

		Cancer cell lines											
Compound		Colon			Renal	Prostate			Breast				
	Values ^a	Colo205	HCT-116	HCT-15	Caki	DU145	PC3	LNCap	MDA M	IB23 1	MCF7		
5b ^c	TGI		51.69								33.2		
	GI50		9.94								6.75		
5c ^c	TGI		50.0								52.72		
	GI50		7.02								21.1		
6a ^c	TGI		34.8								42.85		
	GI50		6.76								9.11		
6b ^c	TGI		100.0								58.54		
	GI50		43.98								28.94		
6c ^c	TGI		64.7								56.15		
	GI50		26.82								26.96		
5-FU	TGI		>100								>100		
	GI50		9.51								8.40		
ETO	TGI												
	GI ₅₀												
	Cano	er cell lines											
	Ovar	rian		CNS	Non	small lung	S	Small lung M	Ielanoma	Normal	cells		

Compound	Values ^a	IGROV-1	OVCAR-5	ADR-res NCI	SF268	U251	NCI-H460	EKVX	A549	DMS 114	SK-MEL-28	hMSC	NHDF
1 ^b	TGI				100	100	100						
	GI ₅₀				100	100	100						
2a ^b	TGI	36.5	37.8	30.00	56.82			29.25	43.7			45.56	
	GI ₅₀	7.6	8.35	6.7	20.98			6.94	8.67			9.55	
2 b ^b	TGI			24.4	37.44	50.87	51.95			50.39	44.98		52.8
	GI ₅₀			7.4	7.83	21.81	13.72			9.38	13.24		17.9
2c ^b	TGI			55.3	50.7	57.98	62.38			73.94	60.71	56.05	58.8
	GI ₅₀			29.3	21.17	31.23	31.18			40.05	31.79	24.83	27.7
3a ^b	TGI					56.23							
	GI ₅₀					31.15							
3b ^b	TGI					56.56							
	GI ₅₀					33.05							
4	TGI				100.0		100.0						
	GI50				100.0		100.0						
5a ^c	TGI				57.05					58.07			
	GI50				29.34					26.48			
5b ^c	TGI				45.85		32.18						
	GI50				13.09		6.32						
5c ^c	TGI				53.61		46.78						
	GI50				22.19		7.49						
6a ^c	TGI				40.93		25.83						
	GI50				9.09		5.78						
6b ^c	TGI				100.0		100.0						
	GI ₅₀				56.39		50.73						
6c ^c	TGI				62.77		67.52						
	GI ₅₀				28.78		31.48						

Table 1 continued

		Cancer cell lines											
		Ovarian		CNS		Non small lung		Small lung	Melanoma	Normal cells			
Compound	Values ^a	IGROV-1	OVCAR-5	ADR-res NCI	SF268	U251	NCI-H460	EKVX	A549	DMS 114	SK-MEL-28	hMSC	NHDF
5 -FU	TGI				>100		>100						
	GI50				49.21		5.95						
ETO	TGI				92.67								
	GI ₅₀				4.21								

5-FU 5-fluorouracil, ETO etoposide

 a TGI is the concentration (μ M) resulting in total growth inhibition, GI₅₀ is the concentration (μ M) resulting in growth inhibition of 50 %

^b Hydrochloride

° Monofumarate

similar profile, while the tricyclic derivative 4 is inactive and **3a–c** are marginally active. The presence of morpholine in analog 6b does not contribute to antiproliferative activity. On the other hand, the presence of piperidine, piperazine and imidazole moieties on aminoadamantanes 5b, 5c and 6c, respectively, has similar contribution to potency. The size of $(CH_2)_n$ linker, 2 or 3 methylene units, does not affect the antiproliferative potency. Interestingly, 1-phenyl-2-aminoadamantane 1 proved to be inactive, while its 1-benzyl congener 2a is sufficiently potent. It seems that the increase of the distance between the aromatic ring and the adamantane moiety enhances the antiproliferative potency. The activity shown by the benzylic compounds is consistent with previous reported findings on diaryl-substituted adamantanes (Riganas et al., 2012a, b, c). These results should aid in the future design of more effective antiproliferative aminoadamantanes.

References

- Aigami K, Inamoto Y, Takaishi N, Hattori K (1975) Biologically active polycycloalkanes. 1. Antiviral adamantane derivatives. J Med Chem 18:713–721
- Chakrabarti JK, Hotten TM, Sutton S, Tupper DE (1976) Adamantane and protoadamantanealkanamines as potential anti-Parkinson agents. J Med Chem 19:967–969
- Cincinelli R, Dallavalle S, Merlini L, Penco S, Pisano C, Carminati P, Giannini G, Vesci L, Gaetano C, Illy B, Zuco V, Supino R, Zunino F (2003) A novel atypical retinoid endowed with proapoptotic and antitumor activity. J Med Chem 46:909–912
- Dawson MI, Xia Z, Jiang T, Ye M, Fontana JA, Farhana L, Patel B, Xue LP, Bhuiyan M, Pellicciari R, Macchiarulo A, Nuti R, Zhang XK, Han YH, Tautz L, Hobbs PD, Jong L, Waleh N, Chao WR, Feng GS, Pang Y, Su Y (2008) Adamantylsubstituted retinoid-derived molecules that interact with the orphan nuclear receptor small heterodimer partner: effects of replacing the 1-adamantyl or hydroxyl group on inhibition of cancer cell growth, induction of cancer cell apoptosis, and inhibition of SRC homology 2 domain-containing protein tyrosine phosphatase-2 activity. J Med Chem 51:5650–5662

- Kadi AA, El-Brollosy NR, Al-Deeb OA, Habib EE, Ibrahim TM, El-Emam AA (2007) Synthesis, antimicrobial, and anti-inflammatory activities of novel 2-(1-adamantyl)-5-substituted-1,3,4-oxadiazoles and 2-(1-adamantylamino)-5-substituted-1,3,4thiadiazoles. Eur J Med Chem 42:235–242
- Kazimierczuk Z, Gorska A, Switaj T, Lasek W (2001) Adamantylaminopyrimidines and -pyridines are potent inducers of tumor necrosis factor-alpha. Bioorg Med Chem Lett 11:1197–1200
- Keepers YP, Pizao PE, Peters GJ, Van Ark-Otte J, Winograd B, Pinedo HM (1991) Comparison of the sulforhodamine B protein and tetrazolium (MMT) assays for in vitro chemosensitivity testing. Eur J Cancer 27:897–900
- Kolocouris N, Foscolos GB, Kolocouris A, Marakos P, Pouli N, Fytas G, Ikeda S, De Clercq E (1994) Synthesis and antiviral activity evaluation of some aminoadamantane derivatives. J Med Chem 37:2896–2902
- Kolocouris N, Kolocouris A, Foscolos GB, Fytas G, Neyts J, Padalko E, Balzarini J, Snoeck R, Andrei G, De Clercq E (1996) Synthesis and antiviral activity evaluation of some new aminoadamantane derivatives. 2. J Med Chem 39:3307–3318
- Lenoir D (1973) Solvolyse von 1-Aryl-2-adamantyl-tosylaten, Einfluss von β -Aryl-Substitution auf verbrückte und unverbrückte Carbonium-Ionen. Chem Ber 106:78–90
- Papanastasiou I, Tsotinis A, Kolocouris N, Prathalingam SR, Kelly JM (2008) Design, synthesis, and trypanocidal activity of new aminoadamantane derivatives. J Med Chem 51:1496–1500
- Papanastasiou I, Tsotinis A, Zoidis G, Kolocouris N, Prathalingam SR, Kelly JM (2009) Design and synthesis of Trypanosoma brucei active 1-alkyloxy and 1-benzyloxyadamantano 2-guanylhydrazones. Chem Med Chem 4:1059–1062
- Riganas S, Papanastasiou I, Foscolos GB, Tsotinis A, Bourguignon JJ, Serin G, Mirjolet JF, Dimas K, Kourafalos VN, Eleutheriades A, Moutsos VI, Khan H, Georgakopoulou S, Zaniou A, Prassa M, Theodoropoulou M, Pondiki S, Vamvakides A (2012a) Synthesis, σ1, σ2-receptors binding affinity and antiproliferative action of new C1-substituted adamantanes. Bioorg Med Chem 20:3323–3331
- Riganas S, Papanastasiou I, Foscolos GB, Tsotinis A, Dimas K, Kourafalos VN, Eleutheriades A, Moutsos VI, Khan H, Prassa M, Georgakopoulou S, Zaniou A, Theodoropoulou M, Mantelas A, Pondiki A, Vamvakides A (2012b) New adamantane derivatives with sigma affinity and antiproliferative activity. Med Chem 8:569–586
- Riganas S, Papanastasiou I, Foscolos GB, Tsotinis A, Serin G, Mirjolet JF, Dimas K, Kourafalos VN, Eleutheriades A, Moutsos VI, Khan H, Georgakopoulou S, Zaniou A, Prassa M,

Theodoropoulou M, Mantelas A, Pondiki S, Vamvakides A (2012c) New adamantane phenylalkylamines with σ -receptor binding affinity and anticancer activity, associated with putative antagonism of neuropathic pain. J Med Chem 55:10241–10261

- Shibata Y, Shiehita M, Sasaki K, Nishimura IL, Hashimoto Y, Iwasaki S (1995) Synthesis and study of some new N-substituted imide derivatives as potential antibacterial agents. Chem Pharm Bull 43:177–179
- Skehan P, Storeng R, Scudiero D, Monks A, McMahon J, Vistica D, Warren JT, Bokesch H, Kenney S, Boyd MR (1990) New colorimetric cytotoxicity assay for anticancer-drug screening. J Natl Cancer Inst 82:1107–1112
- Smith CD, French KJ, Zhuang Y (2006) Sphingosine kinase inhibitors. US 2006/0287317
- Tseng CC, Handa I, Abdel-Sayed AN, Bauer L (1998) N-[(aryl substitute adamantane)alkyl] 2-mercaptoacetamidines, their corresponding disulfides and 5-phosphorothioates. Tetrahedron 44:1893–1904
- Tsuji M, Koiso Y, Takahashi H, Hashimoto Y, Endo Y (2000) Modulators of tumor necrosis factor alpha production bearing dicarba-closo-dodecaborane as a hydrophobic pharmacophore. Biol Pharm Bull 23:513–516
- Wang JJ, Wang SS, Leeb CF, Chung MA, Chern YT (1997) In vitro antitumor and antimicrobial activities of N-substituents of maleimide by adamantane and diamantane. Chemotherapy 43:182–189

- Wang JJ, Chern YT, Liu TY, Chi CW (1998) In vitro and in vivo growth inhibition of cancer cells by adamantylmaleimide derivatives. Anticancer Drug Des 13:779–796
- Wang JJ, Chern YT, Chang YF, Liu TY, Chi CW (2002) Dimethyladamantylmaleimide-induced in vitro and in vivo growth inhibition of human colon cancer Colo205 cells. Anticancer Drugs 13:533–543
- Wang JJ, Chang YF, Chern YT, Chi CW (2003) Study of in vitro and in vivo effects of 1,6-Bis[4-(4-amino-3-hydroxyphenoxy)phenyl]diamantane (DPD), a novel cytostatic and differentiation inducing agent, on human colon cancer cells. Br J Cancer 89:1995–2003
- Wang JJ, Chen YC, Chi CW, Huang KT, Chern YT (2004) In vitro and in vivo growth inhibition and G1 arrest in human cancer cell lines by diaminophenyladamantane derivatives. Anticancer Drugs 15:697–705
- Zefirova ON, Selyunina EV, Averina NV, Zyk NV, Zefirov NS (2002) Synthetic approach to preparation of polycyclic compounds possessing physiological activity: I. Synthesis of 1,4disubstituted adamantanes with amino acid fragment. Rus J Org Chem 38:1125–1129
- Zoidis G, Kolocouris N, Kelly JM, Prathalingam SR, Naesens L, De Clercq E (2010) Design and synthesis of bioactive adamantanaminoalcohols and adamantanamines. Eur J Med Chem 45:5022–5030