## Accepted Manuscript

Design, synthesis, X-ray studies, and biological evaluation of novel macrocyclic HIV-1 protease inhibitors involving the P1'-P2' ligands

Arun K. Ghosh, W. Sean Fyvie, Margherita Brindisi, Melinda Steffey, Johnson Agniswamy, Yuan-Fang Wang, Manabu Aoki, Masayuki Amano, Irene T. Weber, Hiroaki Mitsuya

PII: S0960-894X(17)30892-2

DOI: http://dx.doi.org/10.1016/j.bmcl.2017.09.003

Reference: BMCL 25267

To appear in: Bioorganic & Medicinal Chemistry Letters

Received Date: 28 July 2017 Accepted Date: 3 September 2017



Please cite this article as: Ghosh, A.K., Sean Fyvie, W., Brindisi, M., Steffey, M., Agniswamy, J., Wang, Y-F., Aoki, M., Amano, M., Weber, I.T., Mitsuya, H., Design, synthesis, X-ray studies, and biological evaluation of novel macrocyclic HIV-1 protease inhibitors involving the P1'-P2' ligands, *Bioorganic & Medicinal Chemistry Letters* (2017), doi: http://dx.doi.org/10.1016/j.bmcl.2017.09.003

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

#### **Graphical Abstract**

To create your abstract, type over the instructions in the template box below. Fonts or abstract dimensions should not be changed or altered.

Design, synthesis, X-ray studies, and biological evaluation of novel macrocyclic HIV-1 protease inhibitors involving P1'-P2'-ligands

Leave this area blank for abstract info.

Arun K. Ghosh, <sup>a,b</sup> \* W. Sean Fyvie, <sup>a</sup> Margherita Brindisi, <sup>a</sup> Melinda Steffey, <sup>a</sup> Johnson Agniswamy, <sup>c</sup> Yuan-Fang Wang, <sup>c</sup> Manabu Aoki, <sup>d</sup> Masayuki Amano, <sup>d</sup> Irene T. Weber, <sup>c</sup> and Hiroaki Mitsuya <sup>d,e,f</sup>

<sup>a</sup>Department of Chemistry, Purdue University, West Lafayette, IN 47907; <sup>b</sup>Department of Medicinal Chemistry, Purdue University, West Lafayette, IN 47907 (USA); <sup>c</sup>Departments of Biology and Chemistry, Georgia State University, Atlanta, GA 30303 (USA); <sup>d</sup>Departments of Hematology and Infectious Diseases, Kumamoto University of Medicine, Kumamoto 860-8556 (Japan); <sup>c</sup>Experimental Retrovirology Section, HIV and AIDS Malignancy Branch National Cancer Institute, Bethesda, MD 20892 (USA); <sup>f</sup>Center for Clinical Sciences, National Center for Global Health and Medicine, Shinjuku, Tokyo 162-8655 (Japan).



## Bioorganic & Medicinal Chemistry Letters

journal homepage: www.elsevier.com

# Design, synthesis, X-ray studies, and biological evaluation of novel macrocyclic HIV-1 protease inhibitors involving the P1'-P2' ligands

Arun K. Ghosh, <sup>a,b \*</sup> W. Sean Fyvie, <sup>a</sup> Margherita Brindisi, <sup>a</sup> Melinda Steffey, <sup>a</sup> Johnson Agniswamy, <sup>c</sup> Yuan-Fang Wang, <sup>c</sup> Manabu Aoki, <sup>d</sup> Masayuki Amano, <sup>d</sup> Irene T. Weber, <sup>c</sup> and Hiroaki Mitsuya <sup>d,e,f</sup>

<sup>a</sup>Department of Chemistry, Purdue University, West Lafayette, IN 47907; <sup>b</sup>Department of Medicinal Chemistry, Purdue University, West Lafayette, IN 47907 (USA); <sup>c</sup>Departments of Biology and Chemistry, Georgia State University, Atlanta, GA 30303 (USA); <sup>d</sup>Departments of Hematology and Infectious Diseases, Kumamoto University of Medicine, Kumamoto 860-8556 (Japan); <sup>c</sup>Experimental Retrovirology Section, HIV and AIDS Malignancy Branch National Cancer Institute, Bethesda, MD 20892 (USA); <sup>f</sup>Center for Clinical Sciences, National Center for Global Health and Medicine, Shinjuku, Tokyo 162-8655 (Japan).

## ARTICLE INFO

#### Article history:

Received XX Month 2017 Revised XX Month 2017 Accepted XX Month 2017 Available online XX Month 2017

Keywords:
HIV protease
Drug resistance
P1'- P2' ligands
Macrocyclic inhibitors
Structure-based design

#### ABSTRACT

Design, synthesis, and evaluation of a new class of HIV-1 protease inhibitors containing diverse flexible macrocyclic P1'-P2' tethers are reported. Inhibitor  $\bf 5a$  with a pyrrolidinone-derived macrocycle exhibited favorable enzyme inhibitory and antiviral activity ( $K_i = 13.2$  nM, IC<sub>50</sub> = 22 nM). Further incorporation of heteroatoms in the macrocyclic skeleton provided macrocyclic inhibitors  $\bf 5m$  and  $\bf 5o$ . These compounds showed excellent HIV-1 protease inhibitory ( $K_i = 62$  pM and 14 pM, respectively) and antiviral activity (IC<sub>50</sub> = 5.3 nM and 2.0 nM, respectively). Inhibitor  $\bf 5o$  also remained highly potent against a DRV-resistant HIV-1 variant.

2009 Elsevier Ltd. All rights reserved.

 $<sup>*</sup> Corresponding \ author. \ Tel.: +1-765-494-5323; \ fax: +1-765-496-1612; \ e-mail: \\ \underline{akghosh@purdue.edu} \ (A.\ K.\ Ghosh).$ 

The introduction of combined active antiretroviral treatment (cART) in late nineteen-nineties marked the beginning of a breakthrough treatment for patients with HIV infection and AIDS. 1,2 cART treatment regimens with protease inhibitors and reverse transcriptase inhibitors dramatically improved HIVrelated disease progression and mortality.<sup>3,4</sup> The cART is not a cure, however, it has significantly improved quality of life and transformed the HIV/AIDS pandemic to a manageable chronic ailment with normal life expectancy.<sup>5,6</sup> The impact of cART is remarkable, however, current cART suffers from a number of major drawbacks. The most concerning is the rapid emergence of drug-resistant HIV-1 variants making cART ineffective for some HIV/AIDS patient groups.<sup>7-,9</sup> Current patients who achieve initial viral suppression may ultimately experience treatment failure. 10,11 Furthermore, it has been suggested that these drug-resistant variants can be transmitted to new individuals. The ability to provide long-term cART benefits remains a complex issue. HIV protease inhibitors (PIs) are critical components of cART regimens particularly for salvage treatments. Therefore, design and development of new, more potent, safer therapeutics with high genetic barrier against HIVs acquisition of drug resistance are very important.

Our laboratories have been involved in the design and synthesis of nonpeptide PIs that are active against HIV-1 variants resistant to the currently approved PIs. 12-14 One of the PIs is darunavir (DRV, 1), an FDA approved first-line therapeutic agent for the treatment of HIV/AIDS patients. 15,16 DRV contains a structure-based designed privileged template, (3*R*,3a*S*,6a*R-bis*-tetrahydrofuranyl urethane (*bis*-THF) as the P2 ligand imbedded in a hydroxyl ethylamine sulfonamide isostere (1, Figure 1). 14,17 DRV showed a high genetic barrier, to acquire drug-resistance associated mutations. 18-20 One of our key design strategies is to promote the extensive network of hydrogen bonding interactions

Figure 1. Structure of inhibitors 1-4 and general structures of macrocyclic inhibitors 5a-o.

with the active site backbone atoms of HIV-1 protease. <sup>13,17</sup> Based upon these strategies, we developed a range of PIs with broad-spectrum activity against multidrug-resistant HIV-1 variants. <sup>2,17</sup>

In an alternative approach to develop PIs with broad-spectrum activity, we have designed a number of macrocyclic PIs with exceptional antiviral activity and drug-resistance profiles.21-24 Among them, we have reported macrocyclic inhibitors typified by 13-membered unsaturated derivatives 3 and 4, modifying P1'-P2'-ligands of darunavir-like PIs (Figure 1). Both geometrical isomers displayed excellent inhibitory potency as well as antiviral activity. The corresponding saturated derivatives are significantly less potent. The rationale underlying the design of these inhibitors is based on crystallographic data and modeling studies indicating decreased van der Waals interactions and inhibitor side-chain repacking across representative mutant strains in the vicinity of the S1' subsite area. 25,26 In this context, the introduction of specific heterocyclic scaffolds or heteroatoms on the P1'-P2' tether of the macrocycle could allow effective inhibitor adaptation to a range of side chain mutations. With the aim of developing novel broad-spectrum HIV-1 PIs we explored the combination of a flexible macrocyclic P1'-P2' tether with the pyrrolidinone ring as the potential source for additional backbone interactions. Furthermore, we sought to investigate the outcome of a small set of flexible macrocycles incorporating suitably functionalized nitrogen and oxygen heteroatoms to interact with the backbone residues. Herein, we report design, synthesis and biological results of a series of potent macrocyclic HIV-1 protease inhibitors.

Based upon our previous studies, our current design plan was to further explore 13- and 14-membered macrocycles. 21 Early approaches of the work focused on the incorporation of 2pyrrolidinone heterocycles with (S)- and (R)-configuration to promote hydrogen bonding interaction with Gly27 backbone amide NH.<sup>27</sup> Furthermore, we planned to incorporate N-methyl sulfonamide and alkyl ether functionalities to promote interactions with backbone atoms in the active site. The synthesis of the pyrrolidinone-containing macrocyclic inhibitors are described 1. Commercially in Scheme available allylpyrrolidinone **6a** was converted to the corresponding tosylate derivative by treatment with tosyl chloride and triethylamine in the presence of a catalytic amount of DMAP in CH<sub>2</sub>Cl<sub>2</sub> at 23 °C. The resulting tosylate was reacted with NaH and di-tert-butyl iminodicarboxylate in DMF at 0 °C. The resulting mixture was then heated at 65 °C for 12 h to provide Boc-derivative 7a in excellent yield. Exposure of 7a to trifluoroacetic acid (TFA) at 23 °C provided the amine 8a in 82% yield. Reaction of amine 8a with commercially available optically active oxirane 9 in isopropanol at 60 °C for 24 h provided Boc-aminoalcohol derivative 10a in good yield. Amine 10a was reacted with the known<sup>21</sup> sulfonyl chloride **11a** in CH<sub>2</sub>Cl<sub>2</sub> in the presence of aqueous NaHCO<sub>3</sub> solution at 23 °C to provide diene derivative **12a.** Exposure of diene **12a** to ring closing metathesis (RCM) using Grubb's second-generation catalyst (Grubbs II) (5 mol%) in CH<sub>2</sub>Cl<sub>2</sub> at 23 °C for 14 h afforded macrocyclic derivative 13a as a E/Z mixture nearly 60:40 by HPLC analysis. Removal of the Boc group by treatment with TFA provided the corresponding amine. The reaction of the resulting amine with activated carbonate derivative 14 afforded a mixture of unsaturated derivatives. These E/Z isomers were separated by HPLC using a reverse phase C18 column to provide the pure E- and Z-isomers **5a** and **5b**, respectively. Catalytic hydrogenation of E/Z mixture in the presence of 10% Pd-C in ethyl acetate under a hydrogenfilled balloon afforded the saturated inhibitor 5c in good yield. The corresponding saturated derivative 5d containing (S)pyrrolidinone was synthesized following the same sequence of reactions using (S)-allylpyrrolidinone **6b** as the starting material.

**Scheme 1.** Reagents and conditions: (a) *p*-TsCl, DMAP, Et<sub>3</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, 16 h, (*R*) 63%, (*S*) 74%; (b) NHBoc<sub>2</sub>, NaH, dry DMF, 0 °C to 65 °C, 12 h, (*R*) 90%, (*S*) 93%; (c) TFA, CH<sub>2</sub>Cl<sub>2</sub>, 23 °C, 13 h, (*R*) 82%, (*S*) 75%; (d) **9**, *i*PrOH, 60 °C, 24 h, (*R*) 56%, (*S*) 57%; (e) **11**a, aqueous NaHCO<sub>3</sub>, CH<sub>2</sub>Cl<sub>2</sub>, 23 °C, 14 h, (*R*) 55%, (*S*) 80%; (f) Grubbs II, dry CH<sub>2</sub>Cl<sub>2</sub>, 23 °C, 12 h, (*R*) 64% (*E*/Z mixture 57:43 by HPLC), (*S*) 53%, (*E*/Z mixture); (g) TFA, CH<sub>2</sub>Cl<sub>2</sub>, 23 °C, 8 h; (h) **14**, *N*,*N*-DIPEA, MeCN, 23 °C, 5 days, (*R*) 64%, (*S*) 92%, (*E*/Z mixture); (i) H<sub>2</sub>, 10 % Pd/C, EtOAc, 23 °C, 12 h, (*R*) 73%, (*S*) 89%.

The synthesis of heteroatom-containing macrocyclic inhibitors is described in Scheme 2. Commercially available methyl allylaminopropionate 16 was reacted with MsCl in CH<sub>2</sub>Cl<sub>2</sub> in the presence of aqueous NaHCO<sub>3</sub> at 23 °C to provide the corresponding mesylate derivative. Saponification of the methyl ester with LiOH in aqueous THF at 23 °C furnished carboxylic acid 17. Curtius rearrangement of acid 17 with diphenylphosphoryl azide (DPPA) in the presence of triethylamine in toluene afforded amine 18 in good yield. For incorporation of ether functionality within the macrocycle, we utilized commercially available allyloxyethylamine 19. Both amines 18 and 19 were reacted with commercially available optically active oxirane 9 in isopropanol at 56 °C for 14 h to provide Boc-aminoalcohol derivatives 20 and 21, respectively. Amine 20 was then reacted with known<sup>21</sup> sulfonyl chlorides 11a and 11b in CH<sub>2</sub>Cl<sub>2</sub> in the presence of aqueous NaHCO<sub>3</sub> solution at 23 °C to afford diene sulfonamide derivatives 22 and 23, respectively. Similarly, reaction of amine 21 with sulfonyl chlorides 11a and 11b furnished diene sulfonamide derivatives 24 and 25 in excellent yields. Dienes 22-25 were converted macrocyclic inhibitors **5e-h** as follows. The acyclic dienes **22-25** were exposed to RCM using Grubbs' 2<sup>nd</sup> generation catalyst<sup>16</sup> to give the corresponding unsaturated macrocyclic derivatives. The Boc-group was deprotected using TFA and the resulting amines were reacted with the mixed carbonate of activated *bis*-THF derivative **14** providing the corresponding unsaturated macrocyclic derivatives. The resulting unsaturated compounds were hydrogenated over 10% Pd-C as catalyst to yield inhibitors **5e-h** 

**Scheme 2.** Reagents and conditions: (a) MsCl, aq. NaHCO<sub>3</sub>, CH<sub>2</sub>Cl<sub>2</sub>, 23 °C, 16 h, 86%; (d) LiOH·H<sub>2</sub>O, aq. THF, 23 °C, 18 h, 95%; (c) DPPA, Et<sub>3</sub>N, toluene, 23 °C to reflux, 5 h then aq. NaOH, 98 °C 12 h, 67%; (d) **9**, *i*-PrOH, 56 °C, 14 h, 56-63%; (e) **11a** or **11b**, aq. NaHCO<sub>3</sub>, CH<sub>2</sub>Cl<sub>2</sub>, 23 °C, 14 h, 90-98%; (f) Grubbs II, dry CH<sub>2</sub>Cl<sub>2</sub>, 40 °C, 3-6 h, 52-92% (*E/Z* mixture); (g) TFA, CH<sub>2</sub>Cl<sub>2</sub>, 23 °C, 48 h; (h) **14**, DIPEA, MeCN, 23 °C, 3-7 days, 23-60%; (i) H<sub>2</sub>, 10 % Pd/C, EtOAc, 23 °C, 12 h, 75-90%.

Based upon X-ray structure of our previous macrocyclic inhibitor-bound HIV-1 protease, we sought to investigate macrocycles with isomeric benzyl ether oxygen which would be within proximity to form hydrogen bonds with backbone atoms in the S2'-site. These benzyl ether derivatives may exhibit better stability than the phenyl ether derivatives. The synthesis of these benzyl ether-derived macrocyclic inhibitors is described in Scheme 3. Preparation of sulfonyl chloride 27 was carried out from commercially available 3-allyloxymethylanisole 26 as described by Blotny and co-workers<sup>28</sup> by treatment with chlorosulfuric acid at 0 °C followed by reaction of the resulting sulfonic acid with cyanuric chloride in dry acetone in the presence of triethylamine to provide sulfonyl chloride 27 in 37% yield. For the synthesis of benzyl ether derivative with 4-amino substitution, the corresponding sulfonyl chloride derivative 29 was prepared from commercially available 2-chloro-5nitrobenzyl alcohol 28. Reaction of alcohol with NaH in the

presence of TBSCl in dry THF provided the TBS ether. This was converted to the corresponding thiophenol derivative by reaction with sodium disulfide, freshly prepared from sodium sulfide and elemental sulfur in ethanolic solution in the presence of NaOH to provide a mixture of the corresponding thiol along with its oxidized disulfide derivative. <sup>29</sup> Oxidation of this mixture by a combination of *N*-chlorosuccinimide and dilute hydrochloric acid in MeCN afforded the corresponding sulfonyl chloride **29** in moderate yield. <sup>30</sup> Commercially available 5-hexenylamine and 6-heptenylamine **30** and **31** were reacted with chiral epoxide **9** 

Scheme 3. Reagents and conditions: (a)  $HSO_3Cl$ ,  $CH_2Cl_2$ , 0 °C, 20 min; (b) cyanuric chloride,  $Et_3N$ , dry acetone, 23 °C to 60 °C, 24 h, 37% over 2 steps; (c) TBSCl, NaH (60% susp. in mineral oil), TBAl, dry THF, 0 °C, 1 h, 90%; (d)  $Na_2S_2$  in EtOH, NaOH, EtOH, reflux, 2 h; (e) NCS, 2M HCl in MeCN, -10 °C to 20 °C, 30 min, 35% over 2 steps; (f) 9, iPrOH, 56 °C, 14 h, 82% for n=1, 63% for n=2; (g) 27 or 29, aqueous  $NaHCO_3$ ,  $CH_2Cl_2$ , 23 °C, 18 h, 85-92%; (h) TBAF, dry THF, 0 °C, 15 min, 78%; (o) ally1-tent-buty1carbonate,  $Pd(PPh_3)_4$ , dry THF, 60 °C, 3 h; (j) Grubbs II, dry  $CH_2Cl_2$ , 40 °C, 3-6 h, 85-93%; (k) TFA,  $CH_2Cl_2$ , 23 °C, 3 h; (l) 14, DIPEA, MeCN, 23 °C, 3 days, 34-60%; (m)  $H_2$ , 10% Pd/C, EtOAc, 23 °C, 12 h, 87-90%.

providing epoxide opening products 32 and 33, respectively in excellent yield. Reaction of these amines with sulfonyl chloride

derivative 27 afforded the corresponding diene sulfonamide derivatives 34 and 35. For the synthesis of diene 36, sulfonamide intermediate derived from sulfonyl chloride 29 was subjected to n-Bu<sub>4</sub>N<sup>+</sup>F<sup>-</sup> in THF. The resulting alcohol was subjected to Oallylation with allyl-tert-butylcarbonate in the presence of catalytic Pd(PPh<sub>3</sub>)<sub>4</sub> to provide 36. For the synthesis of the unsaturated inhibitors with p-OMe sulfonamides 5i-l, diene derivatives 34 and 35 were exposed to RCM to provide the corresponding unsaturated macrocycles with E/Z mixtures (approximately 1:2 E/Z ratio). Deprotection of Boc-group with TFA followed by reaction of the resulting amines with activated bis-THF derivative 14 furnished unsaturated inhibitors as a mixture of E/Z isomers. The mixture of isomers were separated by HPLC using a C18 column to furnish macrocyclic inhibitors 5i-5l (34-60% yield). Diene sulfonamide with a p-NO<sub>2</sub> group was converted to macrocyclic inhibitors 37 by similar sequence of reactions. Catalytic hydrogenation of unsaturated macrocycles over 10% Pd-C provided saturated inhibitors 5m and 5n. For the synthesis of the terminal p-NH<sub>2</sub>-derived inhibitor **50**, unsaturated macrocycle 37 was exposed to SnCl2-mediated reduction in EtOH to afford the corresponding aniline derivative in 82% yield. Hydrogenation of the resulting olefin mixture over 10% Pd-C furnished saturated macrocyclic inhibitor 50.

HIV-1 protease inhibitory potency of all synthesized inhibitors was evaluated using the assay protocol reported by Toth and Marshall.<sup>31</sup> These results are shown in Tables 1 and 2. A selected number of compounds was further evaluated in antiviral assay following a previously published assay protocol using MT-2 cells exposed to HIV-1<sub>LAI</sub>. 32 We first investigated a set of macrocyclic inhibitors containing both E- and Z-olefins along with R-pyrrolidinone on the macrocyclic tether to form backbone hydrogen bonding with Gly27 in the S1'-subsite. We specifically investigated (R)-pyrrolidinone in inhibitors 5a to 5c (entries 1-3, Table 1) as this stereochemistry showed enhanced potency over the (S)-isomer in acyclic inhibitors. As can be seen, inhibitor **5b** with an *E*-isomer is significantly potent in enzyme inhibitory assay. However, inhibitor 5a with Z-isomer showed better antiviral activity. Saturated inhibitor 5c displayed good enzyme activity however its antiviral activity was not improved over inhibitor 5a. We have also prepared inhibitor 5d incorporating a (S)-pyrrolidinone derivative. It showed significant reduction of enzyme  $K_i$  as well as antiviral activity. We previously observed that macrocyclic inhibitors with 13- and 14- membered rings are nicely accommodated by the S1-S2' subsites. In an effort to promote hydrogen bonding interactions in this region, we incorporated N-methylsulfonamide functionality. The corresponding 13- and 14-membered macrocyclic inhibitors 5e and 5f (entries 5 and 6, Table 1) showed reduced enzyme inhibitory activity compared to the corresponding inhibitors with carbon chains. The corresponding oxocyclic inhibitors 5g and 5h (entries 7 and 8, Table 1) showed improved enzyme inhibitory activity, however, inhibitor **5h** did not show appreciable antiviral activity (IC<sub>50</sub> > 1  $\mu$ M).

The X-ray crystal structure of the inhibitor 3-bound HIV-1 protease revealed that the phenolic oxygen on the macrocycle do not form any hydrogen bonds in the S1'-subsite. Based upon this X-ray structure, we envisioned that the corresponding positional isomer, particularly incorporation of oxygen at the benzylic position may lead to improved potency as this oxygen would be within proximity to interact with backbone atoms in the S2'-subsite. Inhibitors 5i and 5j incorporated the benzyl ether oxygen within the 13-membered ring cycle with both Z- and E-olefin (entries 1 and 2, Table 2). Both inhibitors showed excellent enzyme inhibitory activity. However, the antiviral activity of both compounds was significantly reduced compared to the corresponding phenolic ether derivatives 3 and 4. In contrast, the

Table 1. Structures and activity of inhibitors 5a-h

Entry Inhibitor structure		K <sub>i</sub>	IC <sub>50</sub> a,b
	0/=	(1	nM)
1. H	H OH NS 5a	13.2	22
2. H	O O O O O O O O O O O O O O O O O O O	0.47	176
3. H	O OME	e 0.46	122
4. H	Ph 5d	27.0	>1000
5. H	MŠN OME	33.3	nt
6. H	MsN OMe	13.3	nt
7. H	OH N S O 5g	4.21	nt
8. H	OH O OME	9.85	>1000

<sup>a</sup>Darunavir (1) exhibited  $K_1 = 16$  pM, antiviral  $IC_{50} = 3$  nM; <sup>b</sup>nt = not tested.

14-membered macrocyclic inhibitors with E- and Z-derivatives (compounds  $\bf 5k$  and  $\bf 5l$ , entries 3 and 4, Table 2) showed greater than 20-fold reduction of enzyme inhibitory activity over inhibitors  $\bf 5i$  and  $\bf 5j$ . Interestingly, both 13- and 14-membered saturated macrocyclic inhibitors  $\bf 5m$  and  $\bf 5n$  showed much improved enzyme inhibitory activity. Also, inhibitor  $\bf 5m$  with 13-membered macrocyclic e exhibited antiviral IC $_{\bf 50}$  of  $\bf 5.3$  nM.

Inhibitor **50** with a *para*-aminosulfonamide derivative showed excellent enzyme inhibitory potency as well as antiviral activity (entry 7, Table 2).

We selected inhibitors 5m and 5o for further evaluation against a DRV-resistant HIV-1 variants. These DRV-resistant HIV-1 $_{\rm DRV}^{\rm R}$  variants are highly resistant to all current clinically used PIs including DRV and nucleoside/nucleotide reverse transcriptase inhibitors such as tenofovir. In these assay, MT-4 cells  $(1\times10^4)$  were exposed to wild-type HIV-1 and a DRV-resistant variant (HIV-1 $_{\rm DRV}^{\rm R}_{\rm P20}$ ) and subjected to various concentrations of each PI. IC $_{50}$  values were determined using p24 assay.  $^{32,33}$  The results are shown in Table 3.

Inhibitor **50** potently blocked the replication of wild-type HIV- $1_{\rm NL4~3}$  showing improved antiviral activity compared to inhibitor **5m** and DRV. Furthermore, this inhibitor suppressed the replication of HIV- $1_{\rm DRV}^{\rm R}_{\rm P20}$ -resistent variant. As can be seen, the

Table 2. Structures and activity of inhibitors 5i-o

	Table 2. Structures and activity of himbitors 51-0					
Ent	try Inhibitor structure	K <sub>i</sub> (n	IC <sub>50</sub> a,b M)			
1.	OMe OH OH OS O O O O O O O O O O O O O	0.029	54			
2.	OMe H OPh Sj	0.015	46			
3.	H OPH OS 5k	0.64	nt			
4.	HO HO DO SI	0.55	nt			
5.	H OPh OS 5m	0.062	5.3			
6.	H OPH NS 5n	0.062	31			
7.	H O H N S So So	0.014	2.0			

<sup>a</sup>Darunavir (1) exhibited  $K_I = 16$  pM, antiviral IC<sub>50</sub> = 3 nM; <sup>b</sup>nt = not tested.

**Table 3.** Antiviral activity of inhibitors 5m and 5o against HIV-1<sub>DRV</sub> $^R_{P20}$  resistant HIV-1 variant.

Cpd –	mean IC <sub>50</sub> in nM (fold-change) <sup>a</sup>		
Сри	HIV-1 <sub>NL4 3</sub>	HIV-1 <sub>DRV</sub> P20	
5m	3.6	240 (67)	
<b>50</b>	1.5	20 (13)	
DRV (1)	1.8	87 (48)	

 $^aMT^{-4}$  cells (1 × 10<sup>4</sup>) were exposed to 50 TCID<sub>50</sub> of wild-type HIV-1<sub>NL4</sub>  $_3$  or HIV-1<sub>DRV</sub>R<sub>P20</sub>, and cultured in the presence of various concentrations of each PI, and the IC<sub>50</sub> values were determined using the p24 assay. The amino acid substitutions identified in protease of HIV-1<sub>DRV</sub>R<sub>P20</sub>, compared to HIV-1<sub>NL4</sub>  $_3$  include L10I/I15 V/K20R/L24I/V32I/M36I/M46L/L63P/V82A/L89M. All assays were conducted in triplicate, and the data shown represent mean values derived from the results of three independent experiments.

fold-difference in the  $IC_{50}$  values of **50** against HIV- $1_{DRV}{}^RP_{20}$  compared to wild-type HIV- $1_{NL4-3}$  was 13-fold, while the fold-differences for DRV (**1**) and inhibitor **5m** were 48- and 67-fold, respectively.

To gain molecular insight into the binding properties of macrocyclic inhibitors containing benzyl ether functionality on the macrocycle, we determined the X-ray crystal structure of inhibitor 5j and wild-type HIV-1 protease complex. The structure was refined at 1.27 Å resolution to give R-value of 15.9%. The X-ray structure contains a protease dimer and inhibitor 5j in two orientations related by a 180° rotation with relative occupancies of 65/35%. A stereoview of the active site interactions is shown in Figure 2. The X-ray showed similarity to the structure of darunavir-bound HIV-1 protease complex<sup>33</sup> with root mean square difference of 0.16Å for Cα atoms. Larger differences between the corresponding Cα atoms are less than 0.5Å. Inhibitor 5j shares identical P1 and P2 ligands like darunavir. However, the major difference is in the P1'-P2' regions where a 13membered macrocycle linking P1' and P2' ligands have been incorporated. Interestingly, this macrocycle differs from previously reported macrocyclic ligands with respect to a specific oxymethyl functionality on the macrocycle. 21 Most of the interaction of the bis-THF P2 ligand, phenylmethyl P1-ligand as well as the transition state hydroxyl group are comparable to those of the HIV-1 protease-darunavir complex.<sup>34</sup> The flexible P1'-P2' macrocyclic ring, containing an E-olefin, nicely packs between the S1'-S2' subsites in a zigzag crown-like shape. The

protease-inhibitor complex reveals interesting new water-mediated hydrogen bonding interactions of the oxymethyl oxygen with backbone atoms at the S2'-site. The ring oxygen makes water-mediated hydrogen bonds with backbone NH of Asp29' as well as with the carboxyl oxygen of Gly27', with distances ranging from 2.9-3.4 Å. The new backbone binding with the main chain atoms may be responsible for its ability to maintain high potency against multidrug-resistant HIV-1 variants. 12,17 Inhibitor maintains the water-mediated hydrogen bonding interactions with Ile50 and Ile50' amide NH<sub>8</sub> that are conserved in the majority of inhibitor-protease complexes. 53.36 Furthermore, inhibitor 5j makes weaker C—H···O interactions throughout the active site of HIV-1 protease. 37,38

In summary, we have designed novel HIV-1 protease inhibitors containing diverse flexible macrocyclic P1'-P2' tethers for the HIV-1 protease active site and investigated their biological activity. We based our rational design upon the premise that P1'-P2' tethers in the contrast of a flexible macrocycle enable effective inhibitor adaptation across a range of side chain mutations. With the aim of developing broad-spectrum inhibitors we also planned to establish additional contacts with key backbone residues. Accordingly, a series of pyrrolidinonefused macrocyclic inhibitors have been designed and synthesized, leading to the identification of inhibitor 5a endowed with a favorable enzyme inhibitory profile and relevant antiviral activity. We subsequently performed a systematic study involving the strategic placement of oxygen (and nitrogen) heteroatoms along the macrocycle skeleton which led to identification of derivative 5m and its aniline analogue 50 as potent inhibitors of HIV-1 protease in the picomolar range. Of particular importance, inhibitor 50 remained highly potent against a DRV-resistant HIV-1 variant. The flexible P1'-P2' macrocyclic nicely packs between the S1'-S2' subsites in a zigzag crown-like shape. To obtain molecular insight into the ligandbinding site interactions, we determined X-ray crystal structure of inhibitor **5i**-bound HIV-1 protease. The structure shows interesting new water-mediated hydrogen bonding interactions of the macrocyclic ring oxygen with backbone atoms at the S2'-site. This may be responsible for inhibitor's high affinity. Further design and improvement of inhibitor properties are currently in progress in our laboratory.

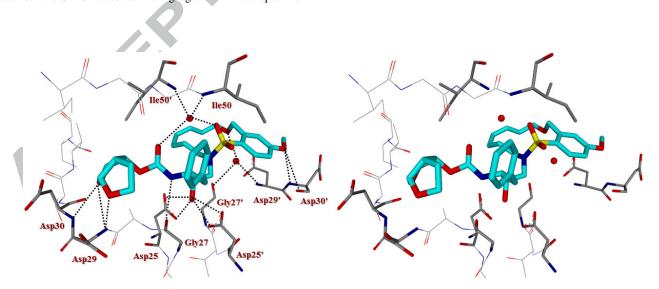


Figure 2. Stereoview of the X-ray structure of inhibitor 5j (turquoise)-bound HIV-1 protease (PDB code: 5WLO). All strong active site hydrogen bonding interactions of inhibitor 5j with HIV-1 protease are shown as dotted lines.

#### Acknowledgments

This research was supported by the National Institutes of Health (Grant GM53386, AKG and Grant GM62920, ITW). The X-ray data was collected at the Southeast Regional Collaborative Access Team (SER-CAT) beamline 22ID at the Advanced Photon Source, Argonne National Laboratory. This work was also supported by the Intramural Research Program of the Center for Cancer Research, National Cancer Institute, National Institutes of Health, and in part by a Grant-in-Aid for Scientific Research (Priority Areas) from the Ministry of Education, Culture, Sports, Science, and Technology of Japan (Monbu Kagakusho), a Grant for Promotion of AIDS Research from the Ministry of Health, Welfare, and Labor of Japan, and the Grant to the Cooperative Research Project on Clinical Epidemiological Studies of Emerging and Reemerging Infectious Diseases (Renkei Jigyo) of Monbu-Kagakusho. The authors would like to thank the Purdue University Center for Cancer Research, which supports the shared NMR and mass spectrometry facilities.

#### **Supplementary Material**

Supplementary data associated with this article can be found in the online version.

#### References and notes

- Edmonds, A.; Yotebieng, M.; Lusiama, J.; Matumona, Y.; Kitetele, F.; Napravnik, S.; Cole, S. R.; Van Rie, A.; Behets, F. PLoS Med. 2011. 8e1001044.
- Ghosh, A. K.; Osswald, H. L.; Prato, G. J. Med. Chem. 2016, 59, 5172-5208.
- Mitsuya, H.; Maeda, K.; Das, D.; Ghosh, A. K. Adv. Pharmacol. 2008, 56, 169-197.
- Hue, S.; Gifford, R. J.; Dunn, D.; Fernhill, E.; Pillay, D. J. Virol. 2009, 83, 2645-2654.
- Cohen, M. S.; Chen, Y.Q.; McCauley, M. N. Engl. J. Med. 2011, 365, 493-505.
- Diffenbach, C. W.; Fauci, A. S. Ann Intern Med. 2011, 154, 766-771.
- Patel, K.; Hernán, M. A.; Williams, P. L.; Seeger, J. D.; McIntosh, K.; Van Dyke, R. B.; Seage III, G. R. Clin. Infect. Dis. 2008, 46, 507-515.
- Gupta, R.; Hill, A.; Sawyer, A. W.; Pillay, D., Clin. Infect. Dis. 2008, 47, 712-722.
- 9. Cihlar, T.; Fordyce, M. Curr. Opin. Virol. 2016, 18, 50-56.
- 10. Pennigs, P. S. Infect. Dis. Rep. 2013, 5, 21-25.
- 11. Conway, B. Future Virol. 2009, 4, 39-41.
- Ghosh, A. K.; Chapsal, B.; Mitsuya, H. Wiley-VCH Verlag GmbH & Co. KGaA: Weinheim, 2010, 205-243.
- 13. Ghosh, A. K.; Chapsal, B. D.; Weber, I. T.; Mitsuya, H. Acc. Chem. Res. 2008, 41, 78-86.
- Ghosh, A. K.; Sridhar, P. R.; Kumaragurubaran, N.; Koh, Y.;
   Weber, I. T.; Mitsuya, H. ChemMedChem 2006, 1, 939-950.
- Ghosh, A. K.; Chapsal, B. D. Design of the anti-HIV protease inhibitor darunavir. In 'From Introduction to Biological and Small Molecule Drug Research and Development' Ed. Ganellin, C. R.; Roberts, S. M.; Jefferis, R. 2013, 355-384.
- Ghosh, A. K.; Dawson, Z. L.; Mitsuya, H. Bioorg. Med. Chem. 2007, 15, 756-7580.
- Ghosh, A. K.; Anderson, D. D.; Weber, I. T.; Mitsuya, H Angew. Chem. Int. Ed. 2012, 51, 1778-1802.
- Koh, Y.; Nakata, H.; Maeda, K.; Ogata, H.; Bilcer, G.; Devasamudram, T.; Kincaid, J. F.; Boross, P.; Wang, Y.-F.; Tie, Y.; Volarath, P.; Gaddis, L.; Harrison, R. W.; Weber, I. T.; Ghosh,

- A. K.; Mitsuya, H. Antimicrob. Agents Chemother 2003, 47, 3123-3129.
- de Béthune, M. P.; Sekar, V.; Spinosa-Guzman, S.; Vanstockem, M.; De Meyer, S.; Wigerinck, P.; Lefebvre, E. "Darunavir (Prezista, TMC114): From bench to clinic, improving treatment options for HIV-infected patients" Antiviral drugs: From basic discovery through clinical trials. John Wiley & Sons, Inc., 2011, 31-45.
- De Meyer, S.; Azijn, H.; Surleraux, D.; Jochmans, D.; Tahri, A.; Pauwels, R.; Wigerinck, P.; de Béthune, M. P. Antimicrob. Agents Chemother. 2005, 49, 2314-2321.
- Ghosh, A. K.; Kulkarni, S.; Anderson, D. D.; Hong, L.; Baldridge, A.; Wang, Y. F.; Chumanevich, A. A.; Kovalevsky, A. Y.; Tojo, Y.; Amano, M.; Koh, Y.; Tang, J.; Weber, I. T.; Mitsuya, H. J. Med. Chem. 2009, 52, 7689-7705.
- Ghosh, A. K.; Swanson, L.; Liu, C.; Cho, H.; Hussain, A.;
   Walters, D. E.; Holland, L. *Bioorg. Med. Chem. Lett.* 2002, 12, 1993-1996.
- Ghosh, A. K.; Swanson, L. M.; Cho, H.; Leshchenko, S.; Hussain, K. A.; Kay, S.; Walters, D. E.; Koh, Y.; Mitsuya, H. J. Med. Chem. 2005, 48, 3576-3585.
- Ghosh, A. K.; Schiltz, G. E.; Rusere, L. N.; Osswald, H. L.; Walters, D. E.; Amano, M.; Mitsuya, H. Org. Biomol. Chem. 2014, 12, 6842-6854.
- Erickson, J.; Neidhart, D. J.; VanDrie, J.; Kempf, D. J.; Wang, X. C.; Norbeck, D. W.; Plattner, J. J.; Rittenhouse, J. W.; Turon, M.; Wideburg, N.; Kohlbrenner, W. E.; Simmer, R.; Helfrich, R.; Paul, D. A.; Knigge, M.. Science 1990, 249, 527-533.
- Baldwin, E. T.; Bhat, T. N.; Liu, B.; Pattabriaman, N.; Erickson, J. W. *Nat. Struct. Biol.* 1995, 2, 244-249.
- Ghosh, A. K.; Leshchenko-Yashchuk, S.; Anderson, D. D.; Baldridge, A.; Noetzel, M.; Miller, H. B.; Tie, Y.; Wang, Y.-F.; Koh, Y.; Weber, I. T.; Mitsuya, H. J. Med. Chem. 2009, 52, 3902-3914.
- 28. Blotny, G. Tetrahedron Lett. 2003, 44, 1499-1501.
- 29. Price, C. C.; Stacy, G. W. J. Am. Chem. Soc. 1946, 68, 498-500.
- Nishiguchi, A.; Maeda, K.; Miki, S. Synthesis 2006, 24, 4131-4134.
- Toth, M. V.; Marshall, G. R. Int. J. Pept. Protein Res. 1990, 36, 544–550.
- Koh, Y.; Amano, M.; Towata, T.; Danish, M.; Leshchenko-Yashchuk, S.; Das, D.; Nakayama, M.; Tojo, Y.; Ghosh, A. K.; Mitsuya, H. J. Virol. 2010, 84, 11961–11969.
- Koh, Y.; Das, D.; Leschenko, S.; Nakata, H.; Ogata-Aoki, H.; Amano, M.; Nakayama, M.; Ghosh, A. K.; Mitsuya, H. Antimicrob. Agents Chemother. 2009, 53, 997–1006.
- Kovalevsky, A. Y.; Tie, Y.; Liu, F.; Boross, P. I.; Wang, Y. F.; Leshchenko, S.; Ghosh, A. K.; Harrison, R. W.; Weber, I. T. J. Med. Chem. 2006, 49, 1379-1387.
- Gustchina, A.; Sansom, C.; Prevost, M.; Richelle, J.; Wodak, S.; Wlodawer, A.; Weber, I. Protein Eng. 1994, 7, 309-317.
- Tie, Y.; Boross, P. I.; Wang, Y. F.; Gaddis, L.; Liu, F.; Chen, X.; Tozser, J.; Harrison, R. W.; Weber, I. T. FEBS J. 2005, 272, 5265-5277.
- 37. Panigrahi, T.; Desiraju, G. Proteins 2007, 67, 128-141.
- 38. PDB code: 5WLO. For details of X-ray studies, please see supplementary information.