Contents lists available at ScienceDirect

European Journal of Medicinal Chemistry

journal homepage: http://www.elsevier.com/locate/ejmech

Original article

Design, synthesis and preliminary biological evaluation of *N*-hydroxy -4-(3-phenylpropanamido)benzamide (HPPB) derivatives as novel histone deacetylase inhibitors

Jie Jiao, Hao Fang, Xuejian Wang, Peng Guan, Yumei Yuan, Wenfang Xu*

Department of Medicinal Chemistry, School of Pharmaceutical Sciences, Shandong University, 44 West Wenhua Road, 250012 Ji'nan, Shandong, PR China

A R T I C L E I N F O

Article history: Received 26 February 2009 Received in revised form 29 May 2009 Accepted 8 June 2009 Available online 17 June 2009

Keywords: HPPB N-Hydroxybenzamide Histone deacetylase inhibitors Anticancer agents Cell-cycle arrest

ABSTRACT

A novel series of *N*-hydroxy-4-(3-phenylpropanamido)benzamide (HPPB) derivatives comprising *N*-hydroxybenzamide group as zinc-chelating moiety were designed, synthesized and evaluated for their ability to inhibit histone deacetylases. These compounds possessed inhibitory activity against the enzymes with IC₅₀ values as low as 4.0 μ M. Among them, the thiophene substituted derivative **5j** (IC₅₀ = 0.3 μ M) and benzo[d][1,3]dioxole derivative **5t** (IC₅₀ = 0.4 μ M) exhibited good antiproliferative activity against the growth of human colon carcinoma cell line HCT116 and non-small cell lung cancer cell (NSCLC) line A549. In addition, they were found to potently induce cell-cycle arrest at G2 phase.

1. Introduction

Histone deacetylases (HDACs) take charge of modification and remodeling of chromosomal histones by removing acetyl groups from ε -NH₂ of lysine residues in histones through zincdependent hydrolysis [1]. Acetylation results in the positive charge density on the N-termini of nucleosomal histone increases, which strengthens the interaction with the negative-charged DNA chain and blocks the access of RNA polymerase and transcription factor (TF) to DNA [2].

In many cancer cell lines, HDACs are over-expressed resulting in excessive deacetylated histones being tightly packed with DNA to form an abnormal "compact structure" of chromatin. In this process, expression of the cell-cycle inhibitor p21^{WAF1} is inhibited and activity of the onco-suppressor p53 is down-regulated, whereas tumor activators HIF-1 and VEGF are up-regulated. Therefore, the inhibition of HDAC activity is considered as a potential strategy for cancer therapy [3]. HDAC inhibitors (HDACi) become epigenetic anticancer agents due to their capacity to achieve significant biological effects in preclinical models of cancer. They can induce cell growth arrest, differentiation and

apoptotic cell death of transformed cells in vitro and vivo [4,5]. Some HDACi have been developed into clinical trials for therapy of various solid tumors and hematological malignancy [6-8]. One inhibitor, Vorinostat (Zolinza[®], SAHA), has been approved by FDA for treatment of cutaneous T-cell lymphoma (CTCL) in 2006 and became the first HDACi therapeutic agent [9]. Vorinostat is a representative linear fatty chain HDACi, and the common features of this class of HDACi have been summarized by Manfred and Soon (Fig. 1) [10,11]. There are two rules to the structureactivity relationship (SAR) of Vorinostat-like derivatives: 1) introduction of hydrophobic groups to the *para* position of the benzene ring results in a higher level of activity, and 2) the optimal chain length between the benzene ring and zinc-binding group (ZBG) is 7-8 atoms. As reported in the literature [8,12], three structural units are essential for an HDACi, an aromatic group (Ar), a zincbinding group (ZBG) and a spacer linking Ar and ZBG. N-Hydroxy-4-(3-phenylpropanamido)benzamide (HPPB) [13], another HDACi which contains N-hydroxybenzamide as zinc-chelating moiety could provide a molecular framework for the design of novel HPPB-like HDACi. However, so far the structure-activity relationship (SAR) information for HPPB-like derivatives was not reported. In this paper, the design, synthesis and biological evaluation of a novel series of HPPB derivatives as potent HDACi has been described.





^{*} Corresponding author. Tel./fax: +86 531 88382264. *E-mail address:* xuwenf@sdu.edu.cn (W. Xu).

^{0223-5234/\$ –} see front matter @ 2009 Elsevier Masson SAS. All rights reserved. doi:10.1016/j.ejmech.2009.06.010



Fig. 1. Structural features of HPPB derivatives.

2. Chemistry

Twenty compounds selected for biological evaluation were prepared as described in Scheme 1. The synthesis of compound **5a** has been described in our previous paper [14], and the synthetic route has been modified based on chemical experimental details of this paper. 3-(4-(Benzyloxy)-3-methoxyphenyl)propanoic acid (1) was coupled with methyl 4-aminobenzoate in the presence of dimethylpropyl ethyl carbodiimide hydrochloride (EDCI) to obtain the coupled product (2a). Deprotection of 2a using 10% Pd/C in MeOH yielded the corresponding phenol product (3). Etherification of compound **3** with various alkyl bromides and benzyl chloride generated phenolic ether derivatives 2b-s. Deprotection of methyl group in compound 3 with boron tribromide produced diphenol derivative (4), which was closed to form a benzo d [1,3] dioxole ring containing derivative (2t) with diiodomethane. Compounds 2a-t were hydrolyzed with sodium hydroxide to give carboxylic acids, followed by condensation with isobutylchloroformate (ClCOOBu-i) and then reaction with hydroxylamine in THF yields the final hydroximic acid compounds 5a-t.

3. Results and discussion

In vitro bioactivity evaluation of compounds 5a-t was performed by HDAC activity assays using an HDAC colorimetric activity assay kit (AK501, Biomol Research Laboratories). The source of HDACs was HeLa nuclear extracts comprising HDAC1 and HDAC2 (the major contributors to HDAC activity in HeLa nuclear extracts), and the substrate was a type of $[{}^{3}H]$ acetylated histone peptide. HDAC1 and HDAC2 are known to be the key nucleus enzymes in charge of deacetylation of histones [15]. Assays were performed according to kit instructions. The test results of compounds 5a-t for inhibitory activity against HDACs in vitro have been presented in Table 1. Vorinostat was used as positive control and IC₅₀ values for both HPPB and its derivatives have been determined. These compounds showed potent inhibitory activities against HDACs with IC₅₀ values as low as 4.0 µM. The alkyl substituted derivatives (**5b**-**h**) with IC₅₀ values ranging from 1.6 μ M to 4.0 μ M displayed the order of increasing activity of R as: n-butyl > n-propyl > sbutyl > isopentyl \approx ethyl > methyl > cyclopentyl. The most potent one is *n*-butyl substituted derivative **5e** which is still less potent than the benzylether derivative **5a**. Compound **5j**, the thiophene analogue of **5a**, was 1-fold more potent than **5a**, whereas compound **5i**, the pyridine analogue of **5a**, showed very poor inhibitory potency. Compounds 5k-s are various substituted derivatives of the lead compound 5a. Among them, ortho bromine substituted benzylether derivative 5n was the most potent one and nearly as potent as Vorinostat. Shifting the Br from ortho position into the para (5p) or meta (5o) position resulted in a little loss of inhibitory potency, while the chlorine substituted benzylether derivatives (5q-s) and methyl substituted benzylether derivatives (**5k-m**) were all less potent than **5n**. In addition, benzo[d][1,3]dioxole derivative **5t** exhibited higher inhibitory potency than Vorinostat. In general, this series of compounds showed potent inhibitory activities against HDACs. Among them, compounds 5a, 5j, 5n, 5o, 5p and 5t were found to be more potent than the paradigmatic hydroxamate HDACi-HPPB, furthermore 5j and 5t exhibited higher inhibitory activity than Vorinostat. Therefore, compounds 5a, 5j, 5n and 5t were chosen as the representative active ones for evaluation of their antiproliferative activities against the growth of cancer cells. Human colon carcinoma cell line HCT116 was the primary antiproliferative model in vitro because HDACs express very highly in this cell line. Besides it, non-small cell lung cancer cell line A549 is a minor model in vitro due to its low expressed level of HDACs. The difference of the expressed level of HDACs between the two cancer cell lines was utilized for better elucidation of anticancer activities of the compounds. The antiproliferative activities were determined by MTT assays and the results have been summarized in Table 2. These compounds were found to have potent antiproliferative activities compared with Vorinostat. Compound 5j greatly inhibits the carcinoma cells' growth with IC_{50} values of 0.03 mM (HCT116) and 0.08 mM (A549), **5t** was a little less potent than **5j** with IC₅₀ values of 0.06 mM (HCT116) and 0.5 mM (A549). They are both more potent than Vorinostat with IC₅₀ values of 0.08 mM (HCT116) and 0.6 mM (A549). Compound **5a** and its bromine analogue **5n** were nearly as potent as Vorinostat. Further study was made on the effects of 5j and 5t upon the cell-cycle by flow cytometric analysis. It has been known that many HDACi including Vorinostat can potently induce cell-cycle arrest at G1 phase or G2 phase which is associated with



Scheme 1. Reagents and conditions: a) EDCI, Et₃N, THF, methyl 4-aminobenzoate, r.t.; b) H₂, 10% Pd–C, CH₃OH, r.t. c) BBr₃, CH₂Cl₂, -40 °C; d) K₂CO₃, CH₂l₂, acetone, reflux; e) KOH, RBr, DMF, 50 °C; f) 2 mol/L NaOH, EtOH, 75 °C (for two steps); CICOOBu-s, Et₃N, THF, NH₂OH·HCl, r.t.

activation of cyclin-dependent kinase (CDK) inhibitor p21 expression or apoptotic cell death pathway [14]. The effects of **5j** and **5t** on the cell-cycle were measured against human HCT116 cells at the concentration of 1 μ mol/L. The analytic results have been shown in Table 3. The population proportion of HCT116 cells in each phase of cell-cycle has been calculated. The two compounds were found to strongly induce cell-cycle arrest at G₂ phase which is associated with p21-independent apoptotic cell death pathway and the anticancer mechanism of these two compounds is worthy of further scrutiny in the future.

4. Conclusion

In conclusion, we have synthesized a novel series of HPPB derivatives with *N*-hydroxybenzamide as zinc-chelating moiety to inhibit HDACs. Several HPPB derivatives exhibited subnanomolar inhibitory activities against HDACs with IC₅₀ values below 1 μ M. In this series, the thiophene substituted derivative **5j** (IC₅₀ = 0.3 μ M) and benzo[*d*][1,3]dioxole derivative **5t** (IC₅₀ = 0.4 μ M) were the most active and notably inhibited the growth of human carcinoma cells HCT116 and A549. In addition, they can potently induce cell-cycle arrest in G2 phase which exhibits their good anticancer bioactivities. They can be good lead compounds for the development of novel anticancer agents in future.

5. Experimental procedure

5.1. Chemistry

5.1.1. General

All materials and reagents used in this work are analytical reagents. All reactions were monitored by thin-layer chromatography (TLC) on 0.25 mm silica gel plates (60GF₂₅₄) and visualized with UV light. ¹H NMR spectra were obtained on a Brucker Avance 300 spectrometer using TMS as an internal standard in DMSO- d_6 solutions. Chemical shifts were reported in ppm (δ) downfield from TMS. All the coupling constants (*J*) are in hertz. IR spectra were recorded on a Perkin–Elmer FTIR 1600 spectrometer. ESI-MS were determined on an API 4000 spectrometer. Melting points were determined on an electrothermal melting point apparatus and without correction. All reported yields are for purified products.

5.1.2. Methyl 4-(3-(4-(benzyloxy)-3-methoxyphenyl) propanamido)benzoate (**2a**)

To a solution of 3-(4-(benzyloxy)-3-methoxyphenyl)propanoic acid **1** (9.0 g, 32 mmol) in dry THF (200 mL) were added EDCI (7.4 g, 38.4 mmol) and Et₃N (9.0 mL, 64 mmol) at room temperature. The mixture was stirred for 2 h and then a solution of methyl 4-aminobenzoate (4.8 g, 32 mmol) in dry THF (100 mL) was added into this mixture. After stirring for 6 h, the reacted mixture was concentrated under reduced pressure. To the residue was added 1 mol/L HCI (100 mL), and then extracted by ethyl acetate (3×100 mL), washed with water (3×100 mL) and dried over Na₂SO₄. After filtration and evaporation of the solvent *in vacuo*, methyl 4-(3-(4-(benzyloxy)-3-methoxyphenyl)propanamido)benzoate **2a** was obtained as white crystal. (9.6 g, 72.5%). ¹H NMR (DMSO-*d*₆, 300 MHz) δ 10.25 (s, 1H), 7.92–7.71 (dd, 4H), 7.43–7.31 (m, 5H), 6.94–6.71 (m, 3H), 5.02 (s, 2H), 3.82 (s, 3H), 3.73 (s, 3H), 2.85 (t, 2H, *J* = 7.5 Hz), 2.64 (t, 2H, *J* = 7.5 Hz). ESI-MS *m/z* 420.4 (MH⁺).

5.1.3. Methyl 4-(3-(4-hydroxy-3-methoxyphenyl) propanamido)benzoate (**3**)

To a solution of methyl 4-(3-(4-(benzyloxy)-3-methoxyphenyl) propanamido)benzoate **2a** (5.0 g, 12 mmol) in MeOH (100 mL) was added 10% wt. Pd–C (0.5 g). The reaction mixture was stirred for 10 h under H₂ pressure at room temperature, filtered with Celite, the solvent was removed *in vacuo* to give methyl 4-(3-(4-hydroxy-3-methoxyphenyl)propanamido)benzoate **3** as a white solid (3.5 g, 88.7%). ¹H NMR (DMSO-*d*₆, 300 MHz) δ 10.24 (s, 1H), 8.70 (s, 1H), 7.92–7.70 (dd, 4H), 6.80–6.60 (m, 3H), 3.82 (s, 3H), 3.72 (s, 3H), 2.81 (t, 2H, *J* = 7.5 Hz), 2.62 (t, 2H, *J* = 7.5 Hz). ESI-MS *m*/*z* 330.3 (MH⁺).

5.1.4. Methyl 4-(3-(3,4-dihydroxyphenyl)propanamido) benzoate (**4**)

To a solution of methyl 4-(3-(4-hydroxy-3-methoxyphenyl) propanamido)benzoate **3** (3.0 g, 9 mmol) in dry CH₂Cl₂ (150 mL) was added added KOH powder (0.34 g, 6.0 mmol). at -40 °C. The mixture was stirred for 1 h at -40 °C and then the solvent was removed *in vacuo*. To the residue was added 1 mol/L Na₂CO₃ (100 mL), then extracted by ethyl acetate (3 × 100 mL), washed with water (3 × 50 mL) and dried over MgSO₄. After filtration and evaporation of the solvent *in vacuo*, methyl 4-(3-(3,4-dihydroxyphenyl)propana mido)benzoate **4** was obtained as a yellow solid (2.4 g, 83.6%). ¹H NMR (DMSO-*d*₆, 300 MHz) δ 10.23 (s, 2H), 8.71 (s, 1H), 7.90–7.72

Table 1

The inhibitory activity results of HPPB derivatives against HDACs in vitro.



Compd	R	HDACs	Compd	R	HDACs		
		IC ₅₀ (μWI)			iC ₅₀ (μινι)		
5a	C Set	0.7	5k	provide the second seco	1.8		
5b	CH ₃ -	3.3	51		2.6		
5c	C ₂ H ₅ -	2.9	5m	A Contraction of the second se	1.5		
5d	CH ₃ (CH ₂) ₂ -	1.7	5n	Br	0.6		
5e	CH ₃ (CH ₂) ₃ -	1.6	50	Br	0.9		
5f	CH ₃ CH ₂ (CH ₃)CH-	2.2	5p	Br	0.8		
5g	(CH ₃) ₂ CH(CH ₂) ₂ -	2.9	5q	CI p ²	1.2		
5h		4.0	5r	Cl provide the second s	1.6		
5i	N Str	3.9	5s	CI	1.4		
5j	S S S S S S S S S S S S S S S S S S S	0.3	5t		0.4		
HPPB Vorinostat		1.1 0.5					

(dd, 4H), 6.79-6.58 (m, 3H), 3.83 (s, 3H), 2.80 (t, 2H, J = 7.5 Hz), 2.61 (t, 2H, J = 7.5 Hz). ESI-MS m/z 316.3 (MH⁺).

5.1.5. Methyl 4-(3-(3,4-dimethoxyphenyl)propanamido) benzoate (**2b**)

To a solution of methyl 4-(3-(4-hydroxy-3-methoxyphenyl) propanamido)benzoate **3** (1.0 g, 3 mmol) in DMF (10 mL) was added KOH powder (0.34 g, 6.0 mmol). The reaction mixture was

stirred for 1 h and then CH₃I (1.3 g, 9.0 mmol) was added at room temperature. The mixture was heated to 50 °C and stirred for 5 h, then was added H₂O (50 mL) and then extracted by ether (3 × 50 mL). The organic layer was merged and washed with water (3 × 50 mL) and dried over MgSO₄. After filtration and evaporation of the solvent *in vacuo*, methyl 4-(3-(3,4-dimethoxyphenyl) propanamido)benzoate **2b** was obtained as a yellow solid (0.9 g, 87.4%). ¹H NMR (DMSO-*d*₆, 300 MHz) δ 8.70 (s, 1H), 7.90–7.68

4474 Table 2

Growth inhibitory results of HDACi against HCT116 and A549 cell lines (IC₅₀, mM).

Compd	HCT116	A549
5a	0.12	0.6
5j	0.03	0.08
5n	0.10	0.6
5t	0.06	0.5
Vorinostat	0.08	0.6

(dd, 4H), 6.79–6.63 (m, 3H), 3.83 (s, 3H), 3.73 (s, 6H), 2.80 (t, 2H, J = 7.5 Hz), 2.65 (t, 2H, J = 7.5 Hz). ESI-MS m/z 344.4 (MH⁺).

5.1.6. General procedure for the synthesis of ethers 2c-t

To a solution of methyl 4-(3-(4-hydroxy-3-methoxyphenyl) propanamido)benzoate **3** (1.0 g, 3 mmol) in DMF (10 mL) was added KOH powder (0.34 g, 6.0 mmol). The reaction mixture was stirred for 1 h and then corresponding alkyl halide (9.0 mmol) was added at room temperature. The mixture was heated to 50 °C and stirred for 5 h, then H₂O was added (50 mL) and extracted by ether (3 × 50 mL). The organic layer was merged and washed with water (3 × 50 mL) and dried over MgSO₄. After filtration and evaporation of the solvent *in vacuo*, the ethers **2c–s** were obtained.

5.1.7. General procedure for the synthesis of hydroxamic acids **5a**-**t** from methyl esters **2a**-**t**

The corresponding methyl esters 2a-t (5.0 mmol) were dissolved in EtOH (50 mL), then 2 mol/L NaOH (10 mL) was added. The solution was stirred at 75 °C for 5–6 h. and then concentrated under reduced pressure. To the residue was added 1 mol/L HCl (50 mL), then the solution was filtered to get the sediment which was washed with water for several times. The sediment was dried and dissolved in dry THF (50 mL), then Et₃N (1.0 g, 10 mmol) was added. The solution was stirred under room temperature and ClCOOBu-*i* (0.7 g, 5.3 mmol) was dropped into the solution. After 1–3 min, NH₂OH/MeOH (5 mL, 2 mmol/mL) was dropped into the reaction mixture. After stirring for 5 h, the reaction mixture was filtered and the filtrate was evaporated under reduced pressure. To the residue was added 1 mol/L HCl (30 mL) and extracted by EtOAc $(3 \times 20 \text{ mL})$ for 3 times. The organic layer was merged and washed with water $(3 \times 20 \text{ mL})$ and then dried with MgSO₄. The solution was filtered and evaporated to give a crude product which was purified by column chromatography.

5.1.7.1. 4-(3-(4-(Benzyloxy)-3-methoxyphenyl)propanamido)-N-hydroxybenzamide (**5a**). Yield: 28.6%. M.p. 193–195 °C. IR (KBr): ν (cm⁻¹) 3266, 1674, 1513, 1256. ¹H NMR (DMSO- d_6 , 300 MHz) δ 10.09 (s, 1H), 10.12 (s, 1H), 8.94 (s, 1H), 7.71–7.62 (m, 4H), 7.44–7.29 (m, 5H), 6.92 (d, 1H, *J* = 8.1 Hz), 6.88 (s, 1H, *J* = 8.1 Hz), 6.73 (d, 1H, *J* = 8.1 Hz), 5.02 (s, 2H), 3.73 (s, 3H), 2.85 (t, 2H, *J* = 7.5 Hz), 2.62 (t, 2H, *J* = 7.5 Hz). ESI-MS *m*/*z* 421.4 (MH⁺).

5.1.7.2. 4-(3-(3,4-Dimethoxyphenyl)propanamido)-N-hydroxybenzamide (**5b**). Yield: 19.0%. M.p. 178–179 °C. IR (KBr): ν (cm⁻¹) 3302, 1662, 1608, 1592, 1515, 1255. ¹H NMR (DMSO- d_6 , 300 MHz) δ 11.10 (s, 1H), 10.13 (s, 1H), 8.95 (s, 1H), 7.75–7.62 (dd, 4H), 6.86–6.73 (m, 3H), 3.71 (s, 3H), 3.70 (s, 3H), 2.85 (t, 2H, J = 7.5 Hz), 2.62 (t, 2H, J = 7.5 Hz). ESI-MS m/z 345.3 (MH⁺).

5.1.7.3. 4-(3-(4-Ethoxy-3-methoxyphenyl)propanamido)-N-hydroxybenzamide (**5c**). Yield: 57.6%. M.p. 179–180 °C. IR (KBr): ν (cm⁻¹) 3299, 1660, 1609, 1515, 1258. ¹H NMR (DMSO- d_6 , 300 MHz) δ 11.10 (s, 1H), 10.13 (s, 1H), 8.95 (s, 1H), 7.75–7.62 (dd, 4H), 6.85–6.70 (m, 3H), 3.94 (d, 2H, J = 6.9 Hz), 3.71 (s, 3H), 2.84 (s, 2H), 2.63 (d, 2H, J = 7.2 Hz), 1.29 (t, 3H, J = 6.9 Hz). ESI-MS m/z 359.5 (MH⁺).

Table 3

The effects of **5j** and **5t** on the cell-cycle were measured against human HCT116 cells at the concentration of 1 μ mol/L, and the population proportion of HCT116 cells in each phase of cell-cycle has been calculated.

Compd concentration (1 µmol/L)	Proportion of HCT116 cells (%)		
	G1 phase	S phase	G ₂ phase
5j	28.51	24.33	47.16
5t	17.09	34.11	48.80
Control	34.60	46.55	18.85

5.1.7.4. *N*-Hydroxy-4-(3-(3-*methoxy*-4-*propoxyphenyl*)*propanamido*)*benzamide* (**5d**). Yield: 19.2%. M.p. 182–183 °C. IR (KBr): ν (cm⁻¹) 3306, 1671, 1639, 1610, 1592, 1514, 1258. ¹H NMR (DMSO- d_6 , 300 MHz) δ 11.10 (s, 1H), 10.13 (s, 1H), 8.94 (s, 1H), 7.72–7.62 (dd, 4H), 6.85–6.71 (m, 3H), 3.84 (t, 2H, *J* = 6.6 Hz), 3.72 (s, 3H), 2.85 (t, 2H, *J* = 7.5 Hz), 2.62 (t, 2H, *J* = 7.5 Hz), 1.75–1.63 (m, 2H), 0.95 (t, 3H, *J* = 7.5 Hz). ESI-MS *m*/*z* 373.2 (MH⁺).

5.1.7.5. 4-(3-(4-Butoxy-3-methoxyphenyl)propanamido)-N-hydroxybenzamide (**5e**). Yield: 28.2%. M.p. 163–165 °C. IR (KBr): ν (cm⁻¹) 3302, 1668, 1514, 1255. ¹H NMR (DMSO- d_6 , 300 MHz) δ 11.10 (s, 1H), 10.13 (s, 1H), 8.95 (s, 1H), 7.71–7.62 (m, 4H), 6.85–6.82 (m, 2H), 6.72 (d, 1H, J=8.1), 3.88 (t, 2H, J=6.6), 3.71 (s, 3H), 2.84 (t, 2H, J=7.5 Hz), 2.62 (t, 2H, J=7.5 Hz), 1.70–1.61 (m, 2H), 1.47–1.35 (m, 2H), 0.91 (t, 3H, J=7.5 Hz). ESI-MS m/z 387.4 (MH⁺).

5.1.7.6. 4-(3-(4-sec-Butoxy-3-methoxyphenyl)propanamido)-N-hydroxybenzamide (**5f**). Yield: 38.4%. M.p. 156–158 °C. IR (KBr): ν (cm⁻¹) 3261, 1664, 1626, 1608, 1512, 1259. ¹H NMR (DMSO- d_6 , 300 MHz) δ 11.09 (s, 1H), 10.12 (s, 1H), 8.94 (s, 1H), 7.71–7.62 (dd, 4H), 6.85–6.70 (m, 3H), 4.25–4.19 (m, 1H, J = 6.0 Hz), 3.71 (s, 3H), 2.84 (t, 2H, J = 7.5 Hz), 2.62 (t, 2H, J = 7.5 Hz), 1.67–1.46 (m, 2H), 1.16 (d, 3H, J = 6.0 Hz), 0.90 (t, 3H, J = 7.5 Hz). ESI-MS m/z 387.4 (MH⁺).

5.1.7.7. N-Hydroxy-4-(3-(4-(isopentyloxy)-3-methoxyphenyl)propanamido)benzamide (**5g**). Yield: 38.4%. M.p. 171–172 °C. IR (KBr): ν (cm⁻¹) 3301, 1670, 1643, 1609, 1590, 1515, 1255. ¹H NMR (DMSO-d₆, 300 MHz) δ 11.08 (s, 1H), 10.12 (s, 1H), 8.94 (s, 1H), 7.71–7.62 (dd, 4H), 6.87–6.71 (m, 3H), 3.91 (t, 2H, *J* = 6.6 Hz), 3.71 (s, 3H), 2.84 (t, 2H, *J* = 7.5 Hz), 2.62 (t, 2H, *J* = 7.5 Hz), 1.81–1.72 (m, 1H), 1.61–1.54 (q, 2H, *J* = 6.6 Hz), 0.92 (s, 3H), 0.90 (s, 3H). ESI-MS *m*/z 401.5 (MH⁺).

5.1.7.8. 4-(3-(4-(Cyclopentyloxy)-3-methoxyphenyl)propanamido)-*N*-hydroxybenzamide (**5h**). Yield: 77.0%. M.p. 188–190 °C. IR (KBr): $v \,(\text{cm}^{-1})$ 3309, 1670, 1641, 1609, 1593, 1511, 1256. ¹H NMR (DMSO- d_6 , 300 MHz) δ 11.11 (s, 1H), 10.13 (s, 1H), 8.99 (s, 1H), 7.72–7.63 (dd, 4H), 6.84–6.71 (m, 3H), 4.70 (s, 1H), 3.70 (s, 3H), 2.83 (d, 2H, *J* = 7.5 Hz), 2.63 (d, 2H, *J* = 7.5 Hz), 1.81–1.54 (m, 8H). ESI-MS *m*/*z* 399.4 (MH⁺).

5.1.7.9. *N*-Hydroxy-4-(3-(3-*methoxy*-4-(*pyridin*-2-*ylmethoxy*)*phenyl*)propanamido)benzamide (**5i**). Yield: 57.9%. M.p. 205–206 °C. IR (KBr): ν (cm⁻¹) 3290, 1655, 1641, 1603, 1592, 1518, 1257. ¹H NMR (DMSO- d_6 , 300 MHz) δ 11.09 (s, 1H), 10.12 (s, 1H), 8.92 (s, 1H), 8.55 (s, 1H), 7.83 (t, 1H, J = 7.5 Hz), 7.71–7.62 (dd, 4H), 7.50 (d, 1H, J = 7.8 Hz), 7.33 (s, 1H), 6.93–6.71 (m, 3H), 5.10 (s, 2H), 3.76 (s, 3H), 2.83 (d, 2H, J = 7.5 Hz), 2.63 (d, 2H, J = 7.5 Hz). ESI-MS *m*/*z* 422.4 (MH⁺).

5.1.7.10. N-Hydroxy-4-(3-(3-methoxy-4-(thiophene-2-ylmethoxy)phenyl)propanamido)benzamide (**5***j*). Yield: 19.4%. M.p. 169–170 °C. IR (KBr): ν (cm⁻¹) 3305, 1674, 1638, 1609, 1589, 1512, 1255. ¹H NMR (DMSO- d_6 , 300 MHz) δ 11.09 (s, 1H), 10.12 (s, 1H), 8.94 (s, 1H), 7.71–7.62 (dd, 4H), 7.53 (d, 1H, J = 4.8 Hz), 7.15 (s, 1H), 7.01 (t, 1H, J = 4.2 Hz), 6.97–6.72 (m, 3H), 5.20 (s, 2H), 3.73 (s, 3H), 2.85 (t, 2H, J = 7.5 Hz), 2.63 (t, 2H, J = 7.5 Hz). ESI-MS m/z 427.5 (MH⁺).

5.1.7.11. 4-(3-(4-(2-Methylbenzyloxy)-3-methoxyphenyl)propanamido)-N-hydroxybenzamide (**5**k). Yield: 48.3%. M.p. 173–175 °C. IR (KBr): ν (cm⁻¹) 3215, 1667, 1608, 1515, 1257. ¹H NMR (DMSO- d_6 , 300 MHz) δ 11.09 (s, 1H), 10.12 (s, 1H), 8.96 (s, 1H), 7.71–7.62 (dd, 4H), 7.38– 7.22 (m, 4H), 6.98–6.76 (m, 3H), 5.00 (s, 2H), 3.73 (s, 3H), 2.86 (t, 2H, J = 7.5 Hz), 2.63 (t, 2H, J = 7.5 Hz), 2.31 (s, 3H). ESI-MS m/z 435.5 (MH⁺).

5.1.7.12. 4-(3-(4-(3-Methylbenzyloxy)-3-methoxyphenyl)propanamido)-N-hydroxybenzamide (**51**). Yield: 19.4%. M.p. 178–179 °C. IR (KBr): ν (cm⁻¹) 3308, 1673, 1643, 1610, 1590, 1513, 1257. ¹H NMR (DMSO d_6 , 300 MHz) δ 10.99 (s, 1H), 10.12 (s, 1H), 8.94 (s, 1H), 7.71–7.62 (dd, 4H), 7.29–7.13 (m, 4H), 6.93–6.71 (m, 3H), 4.97 (s, 2H), 3.71 (s, 3H), 2.85 (t, 2H, *J* = 7.5 Hz), 2.63 (t, 2H, *J* = 7.5 Hz), 2.31 (s, 3H). ESI-MS *m*/ *z* 435.4 (MH⁺).

5.1.7.13. 4-(3-(4-(4-Methylbenzyloxy)-3-methoxyphenyl)propanamido)-N-hydroxybenzamide (**5m**). Yield: 58.0%. M.p. 216–218 °C. IR (KBr): ν (cm⁻¹) 3314, 1675, 1592, 1513, 1254. ¹H NMR (DMSO- d_6 , 300 MHz) δ 11.09 (s, 1H), 10.12 (s, 1H), 8.94 (s, 1H), 7.87–7.62 (m, 4H), 7.31–7.16 (dd, 4H), 6.92–6.70 (m, 3H), 4.97 (s, 2H), 3.73 (s, 3H), 2.85 (t, 2H, J = 7.5 Hz), 2.64 (t, 2H, J = 7.5 Hz), 2.30 (s, 3H). ESI-MS m/z 435.5 (MH⁺).

5.1.7.14. 4-(3-(4-(2-Bromobenzyloxy)-3-methoxyphenyl)propanamido)-N-hydroxybenzamide (**5n**). Yield: 16.2%. M.p. 162–164 °C. IR (KBr): ν (cm⁻¹) 3246, 1661, 1595, 1514, 1558. ¹H NMR (DMSO- d_6 , 300 MHz) δ 11.07 (s, 1H), 10.11 (s, 1H), 8.93 (s, 1H), 7.72–7.29 (m, 8H), 6.94–6.74 (m, 3H), 5.05 (s, 2H), 3.75 (s, 3H), 2.87 (s, 2H), 2.64 (s, 2H). ESI-MS m/z 499.3/501.3 (MH⁺).

5.1.7.15. 4-(3-(4-(3-Bromobenzyloxy)-3-methoxyphenyl)propanamido)-N-hydroxybenzamide (**50**). Yield: 16.2%. M.p. 200–202 °C. IR (KBr): ν (cm⁻¹) 3285, 1671, 1611, 1512, 1256. ¹H NMR (DMSO- d_6 , 300 MHz) δ 11.09 (s, 1H), 10.12 (s, 1H), 8.93 (s, 1H), 7.71–7.62 (m, 5H), 7.53–7.32 (m, 3H), 6.91 (d, 2H, J = 8.7 Hz), 6.73 (d, 1H, J = 8.1 Hz), 5.04 (s, 2H), 3.75 (s, 3H), 2.85 (t, 2H, J = 7.5 Hz), 2.62 (t, 2H, J = 7.5 Hz). ESI-MS 499.3/501.3 (MH⁺).

5.1.7.16. 4-(3-(4-(4-Bromobenzyloxy)-3-methoxyphenyl)propanamido)-N-hydroxybenzamide (**5p**). Yield: 16.2%. M.p. 208–210 °C. IR (KBr): ν (cm⁻¹) 3313, 1674, 1637, 1607, 1591, 1514, 1256. ¹H NMR (DMSO-d₆, 300 MHz) δ 11.09 (s, 1H), 10.12 (s, 1H), 8.93 (s, 1H), 7.71–7.56 (m, 6H), 7.38 (d, 2H, *J* = 7.8 Hz), 6.90 (d, 1H, *J* = 8.4 Hz), 6.74 (s, 1H), 5.01 (s, 2H), 3.74 (s, 3H), 2.85 (t, 2H, *J* = 7.5 Hz), 2.62 (t, 2H, *J* = 7.5 Hz). ESI-MS *m*/*z* 499.2/501.2 (MH⁺).

5.1.7.17. 4-(3-(4-(2-Chlorobenzyloxy)-3-methoxyphenyl)propanamido)-N-hydroxybenzamide (**5q**). Yield: 35.0%. M.p. 206–208 °C. IR (KBr): ν (cm⁻¹) 3253, 1663, 1609, 1594, 1514, 1259. ¹H NMR (DMSO-d₆, 300 MHz) δ 11.09 (s, 1H), 10.12 (s, 1H), 8.93 (s, 1H), 7.71–7.56 (m, 4H), 7.52–7.35 (m, 4H), 6.95–6.70 (m, 3H), 5.08 (s, 2H), 3.74 (s, 3H), 2.86 (t, 2H, *J* = 7.5 Hz), 2.61 (t, 2H, *J* = 7.5 Hz). ESI-MS *m*/*z* 455.3 (MH⁺).

5.1.7.18. 4-(3-(4-(3-Chlorobenzyloxy)-3-methoxyphenyl)propanamido)-N-hydroxybenzamide (**5r**). Yield: 35.0%. M.p. 238–240 °C. IR (KBr): ν (cm⁻¹) 3260, 1671, 1644, 1610, 1591, 1513, 1256. ¹H NMR (DMSO-d₆, 300 MHz) δ 11.09 (s, 1H), 10.12 (s, 1H), 8.94 (s, 1H), 7.71–7.62 (m, 4H), 7.49–7.39 (m, 3H), 6.93–6.72 (m, 3H), 5.05 (s, 2H), 3.74 (s, 3H), 2.86 (t, 2H, *J* = 7.5 Hz), 2.63 (t, 2H, *J* = 7.5 Hz). ESI-MS *m*/*z* 455.3 (MH⁺). 5.1.7.19. 4-(3-(4-(4-Chlorobenzyloxy)-3-methoxyphenyl)propanamido)-N-hydroxybenzamide (**5s**). Yield: 17.6%. M.p. 240–242 °C. IR (KBr): ν (cm⁻¹) 3307, 1671, 1641, 1607, 1592, 1514, 1256. ¹H NMR (DMSO-d₆, 300 MHz) δ 11.09 (s, 1H), 10.12 (s, 1H), 8.94 (s, 1H), 7.71–7.65 (m, 4H), 7.44 (s, 4H), 6.91 (d, 2H, *J* = 8.7 Hz), 6.73 (d, 1H, *J* = 7.8 Hz), 5.03 (s, 2H), 3.74 (s, 3H), 2.86 (t, 2H, *J* = 7.5 Hz), 2.62 (t, 2H, *J* = 7.5 Hz). ESI-MS *m*/*z* 455.3 (MH⁺).

5.1.7.20. 4-(3-(Benzo[d][1,3]dioxol-5-yl)propanamido)-N-hydroxybenzamide (**5***t*). Yield: 19.0%. M.p. 230–232 °C. IR (KBr): ν (cm⁻¹) 3280, 1658, 1608, 1534, 1501, 1255. ¹H NMR (DMSO-*d*₆, 300 MHz) δ 11.09 (s, 1H), 10.10 (s, 1H), 8.93 (s, 1H), 7.71–7.61 (dd, 4H), 6.83–6.69 (m, 3H), 5.95 (s, 2H), 2.83 (t, 2H, *J* = 7.5 Hz), 2.60 (t, 2H, *J* = 7.5 Hz). ESI-MS *m*/*z* 329.4 (MH⁺).

5.2. HDAC inhibitory activity assays

We performed assays according to kit instructions. The source of HDACs was HeLa nuclear extracts including HDAC1 and HDAC2 (the major contributors to HDAC activity in HeLa nuclear extracts), and the substrate was a type of [³H]acetylated histone peptide. Both HDAC1 and HDAC2 are known to be nucleus proteins in charge of the deacetylation of histones. The compound samples and the control drug were diluted to various concentrations. On the 96-well plate, HDACs (5 μ L/well) were incubated at 37 °C with 10 μ L of various concentrations of samples and 25 μ L of substrate. After reacting for 30 min, Color de Lys Developer (50 μ L/well) was added. Then, after 15 min the ultraviolet absorption of the wells was measured on a microtiter-plate reader at 405 nm. The % inhibition was calculated from the ultraviolet absorption readings of inhibited wells relative to those of control wells. Finally, the IC₅₀ values were determined using a regression analysis of the concentration/inhibition data.

5.3. Antiproliferative activity evaluation of HDAC inhibitors by MTT assays

HCT116 cells were maintained in McCoy's 5a medium with 10% fetal bovine serum (FBS) while A549 cells were cultured in Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% FBS. Appropriate numbers of cells $(2.0 \times 10^5/mL)$ were inoculated into 96-well plates (50 µL/well). After 4 h, compounds of various concentrations were dosed, and the cells were cultured for 2 days. Then 0.5% MTT (10 µL/well) was added into each well. After additional 4 h incubation, OD570 and OD630 as references were measured, and the IC₅₀ values were calculated according to a regression analysis of the concentration/inhibition data.

Acknowledgments

This work was supported by National "863" Foundation (No. 2007AA02Z314) of PR China and the National Natural Foundation Research Grant (Grant Nos. 30772654; 36072541).

References

- [1] S. Minucci, P.G. Pelicci, Nat. Rev. Cancer 6 (2006) 38-51.
- [2] G. Legube, D. Trouche, EMBO Rep. 4 (2003) 944–947.
- [3] D.H. Kim, M. Kim, H.J. Kwon, J. Biochem. Mol. Biol. 36 (2003) 110-119.
- [4] J.S. Carew, F.J. Giles, S.T. Nawrocki, Cancer Lett. 269 (2008) 7-17.
- [5] P.A. Marks, V.M. Richon, R.A. Rifkind, J. Natl. Cancer Inst. 92 (2000) 1210–1216.
- [6] M.R. Acharya, A. Sparreboom, J. Venitz, W.D. Figg, Mol. Pharmacol. 68 (2005) 917–932.
- 7] C. Monneret, Eur. J. Med. Chem. 40 (2005) 1–13.
- [8] M. Paris, M. Porcelloni, M. Binaschi, D. Fattori, J. Med. Chem. 51 (2008) 1505–1529.

- [12] T.A. Miller, D.J. Witter, S. Belvedere, J. Med. Chem. 46 (2003) 5097–5116.
 [13] Q. Lu, D.S. Wang, C.S. Chen, Y.D. Hu, C.S. Chen, J. Med. Chem. 48 (2005)
 - 5530-5535. [14] J. Jiao, Q. Wang, H.W. Zhu, F. Hao, W.F. Xu, Chin. Chem. Lett. 19 (2008) 673-675.
 - [15] L. Pan, J. Lu, B. Huang, Cell. Mol. Immunol. 4 (2007) 337–343.
- [9] D. Madeleine, V. Jenny, Expert Opin. Investig. Drugs 16 (2007) 1111–1120.
 [10] M. Jung, G. Brosch, D. Kolle, H. Scherf, C. Gerhauser, P. Loidl, J. Med. Chem. 42 (1999) 4669–4679.
- S.H. Woo, S. Frechette, E.A. Khalil, G. Bouchain, A. Vaisburg, N. Bernstein,
 O. Moradei, S. Leit, M. Allan, M. Fournel, M. Trachy-Bourget, Z. Li,
 J.M. Besterman, D. Delorme, J. Med. Chem. 45 (2002) 2877–2885.