Bioorganic & Medicinal Chemistry Letters 21 (2011) 164-167

Contents lists available at ScienceDirect



Bioorganic & Medicinal Chemistry Letters

journal homepage: www.elsevier.com/locate/bmcl



Fused bicyclic derivatives of 2,4-diaminopyrimidine as c-Met inhibitors

Linda R. Weinberg^{*}, Mark S. Albom, Thelma S. Angeles, Jean Husten, Joseph G. Lisko, Robert J. McHugh, Karen L. Milkiewicz, Seetha Murthy, Gregory R. Ott, Jay P. Theroff, Rabindranath Tripathy, Ted L. Underiner, Craig A. Zificsak, Bruce D. Dorsey

Worldwide Discovery Research, Cephalon, Inc., 145 Brandywine Parkway, West Chester, PA 19380-4245, USA

ARTICLE INFO

Article history: Received 30 September 2010 Revised 4 November 2010 Accepted 8 November 2010 Available online 11 November 2010

Keywords: c-Met Kinase inhibitors Mesenchymal epithelial transition factor Diaminopyrimidine

ABSTRACT

The HGF-c-Met signaling axis is an important paracrine mediator of epithelial-mesenchymal cell interactions involving the regulation of multiple cellular activities including cell motility, mitogenesis, morphogenesis, and angiogenesis. Dysregulation of c-Met signaling (e.g., overexpression or increased activation) is associated with the development of a wide range of tumor types; thus, inhibiting the HGF-c-Met pathway is predicted to lead to anti-tumor effects in many cancers. Elaboration of a 2-aryla-minopyrimidine scaffold led to a series of potent c-Met inhibitors bearing a C4-2-amino-*N*-methylbenza-mide group. Specifically, a series of C2-benzazepinone analogs demonstrated potent inhibition of c-Met in enzymatic and cellular assays. Kinase selectivity could be tuned by varying the nature of the alkyl group on the benzazepinone nitrogen.

© 2010 Elsevier Ltd. All rights reserved.

Mesenchymal epithelial transition factor (c-Met) kinase and its natural ligand, hepatocyte growth factor (HGF) are involved in signaling pathways for embryological development, wound healing, tissue regeneration, angiogenesis, proliferation, survival, scattering, and morphogenic differentiation. The pathway regulates a series of intracellular downstream events in many different cell types that lead to cell growth, motility, and invasion.

Due to the c-Met/HGF role of regulation of cellular motility, cells expressing high levels of HGF and/or c-Met are likely to be metastatic.^{1,2} The cells can become solid tumors in the liver, breast, pancreas, lung, kidney, bladder, ovary, brain, prostate, and gallbladder.^{1,2}

A number of Type I (DFG-in) and Type II (DFG-out) c-Met inhibitors are currently being evaluated clinically and several Type I inhibitors which are currently in clinical trials are shown in Figure 1.³

In connection with other work focused on optimizing a 2,4-diaminopyrimidine scaffold to inhibit ALK, it was discovered that compounds with an aminobenzamide appended at C4 consistently demonstrated potent c-Met inhibition (Fig. 2).^{4,5} This putative Type I kinase inhibitor platform served as a starting point in a medicinal chemistry effort to optimize c-Met cellular activity while improving overall kinase selectivity. This Letter describes the synthesis and c-Met SAR around a series of diaminopyrimidine analogs bearing a C4-aminobenzamide and a C2-benzazepinone substituent and how kinase selectivity can be tuned by varying the nature of these appendages (Fig. 2).

* Corresponding author. Tel.: +1 610 738 6595.

E-mail address: lweinber@cephalon.com (L.R. Weinberg).

In general, these diaminopyrimidine targets were prepared by first coupling 2-aminobenzamide analogs **2**, derived from the corresponding isatoic anhydrides **1**, with trichloropyrimidine to afford aminopyrimidines **3**; these intermediates were then coupled to aminobenzazepin-2-ones **4** or **5** (Scheme 1).

Aminobenzazepin-2-one isomers (**4** and **5**) were prepared as described in Schemes 2 and 3. Beckmann rearrangement of the oxime derived from tetralone **6** produced benzazepinone **8**.⁵ Nitration of **8a** or **8b** followed by either reduction or alkylation and reduction afforded intermediates **4a–d**.



Figure 1. Type I c-Met inhibitors in clinical trials.

⁰⁹⁶⁰⁻⁸⁹⁴X/ $\$ - see front matter \odot 2010 Elsevier Ltd. All rights reserved. doi:10.1016/j.bmcl.2010.11.045



for c-Met inhibition, R' = CONHMe

X = H, F, CI or MeY = H or Me





Scheme 1. Target synthesis. Reagents and conditions: (a) R^1R^2NH , THF; (b) 2,4,5-trichloropyrimidine, DIPEA, NMP, 100 °C; (c) DL-10-camphorsulfonic acid, IPA, μ W at 120 °C.



Scheme 2. Synthesis of 7-amino-1,3,4,5-tetrahydro-benzo[*b*]azepin-2-one (**4**). Reagents and conditions: (a) NaOAc, NH₂OH·HCl, EtOH, water, reflux; (b) PPA, 125 °C; (c) H₂SO₄, HNO₃; (d) R³X, Cs₂CO₃, DMF; (e) 10% Pd/C, N₂H₄·H₂O, EtOH, 60 °C.

A similar synthesis was carried out to make the isomeric 8-amino-1,3,4,5-tetrahydro-benzo[b]azepin-2-ones **5**, and is shown in Scheme 3. Nitration of tetralone **6** afforded **10**. A Beckmann rearrangement of the corresponding oxime **11** produced the benzazepinone **12a**. Both **12a** and **12b**⁶ were either reduced or alkylated and then reduced to give anilines **5**.⁵



Scheme 3. Synthesis of 8-amino-1,3,4,5-tetrahydro-benzo[*b*]azepin-2-one (5). Reagents and conditions: (a) H_2SO_4 , HNO_3 ; (b) NaOAc, NH_2OH -HCl, EtOH, water, reflux; (c) PPA, 125 °C; (d) R^3X , Cs_2CO_3 , DMF; (e) 10% Pd/C, N_2H_4 ·H₂O, EtOH, 60 °C.

In addition to optimizing for c-Met inhibition, medicinal chemistry efforts were directed at enhancing selectivity against the insulin receptor kinase (IR)⁷ in particular, and against a broad panel of kinases in general. Kinase selectivity was determined using Ambit Bioscience KINOMEscanTM technology and is expressed as an *S*(90) value, which indicates the fraction of kinases inhibited >90% when screened at 1 μ M across a panel of 256 kinases.⁸

The benzamide SAR is summarized in Table 1. Increasing size of the secondary amide group (**13** vs **16**) led to reduced c-Met inhibitory activity. In this series, analog **14** was the most potent cellular c-Met inhibitor and demonstrated fourfold selectivity for c-Met over IR. Nonetheless, this analog demonstrated >90% inhibition of 151 out of 256 kinases when screened at 1 μ M. Interestingly, the propargyl benzamide analog **17** was also quite active while the corresponding *N*-Me tertiary amide **18** was >20-fold less active. Optimization thus focused on analogs of methylbenzamide **14**.

Kinase selectivity was dramatically improved upon incorporation of a C3 substituent on the benzamide ring (e.g., a halogen or methyl group, Table 2). For example, the S(90) value fell from 0.57 for analog **19** (X = H) to 0.02 for analog **21** (X = Cl). Although analog **21** had improved selectivity, it had an unacceptably high GTL-16 cell IC₅₀. Analog **20** (X = F) represented a better compromise; although the IR/c-Met IC₅₀ ratio remained at 4, the S(90) value fell to 0.31 with no significant impact on c-Met potency or cell translation. The C4-substituent was fixed as 2-amino-3-fluoro-*N*methylbenzamide, and medicinal chemistry focused on optimizing the C2-benzazepin-2-one.

Table 1

c-Met SAR and kinase selectivity of benzamide analogs 13-18



Compd	R ¹	R ¹	IC ₅₀ ^a (nM)				
			c-Met enzyme	c-Met cell ^b	IR enzyme	IR/c-Met enzyme	<i>S</i> (90) ^c
13	Н	Н	7	85	4	0.6	0.69
14	Me	Н	6	10	23	4	0.59
15	<i>n</i> -Pr	Н	73	202	11	0.2	NT ^d
16	<i>n</i> -Bu	Н	161	NT ^d	34	0.2	NT ^d
17	CH ₂ CCH	Н	17	38	8	0.5	0.42
18	CH ₂ CCH	Me	484	NT ^d	NT ^d	NT ^d	0.34

^a Average of at least two separate determinations; see Supplementary data for assay conditions.

^b GTL-16 cells.

 $^{\rm c}$ Fraction of 256 kinases inhibited >90% when screened at 1 $\mu M.^8$

^d Not tested.

Table 2

c-Met SAR and kinase selectivity of benzamide analogs 19-23



Compd	Х	R ³	IC_{50}^{a} (nM)					
			c-Met enzyme	c-Met cell ^b	IR enzyme	IR/c-Met enzyme	<i>S</i> (90) ^c	
19	Н	Н	54	80	30	0.6	0.57	
20	F	Н	45	102	200	4	0.31	
21	Cl	Н	159	1652	>6500	>40	0.02	
22	Me	Н	203	NT ^d	NT ^d	NT ^d	0.04	
23	F	Et	56	64	334	6	0.23	

^a Average of at least two separate determinations; see Supplementary data for assay conditions.

^b GTL-16 cells.

^c Kinase selectivity was determined using the Ambit Bioscience KINOMEscan™ technology.

^d Not tested.

Incorporation of an ethyl group on the 7-aminobenzazepinone nitrogen **23** maintained c-Met cell potency (cell $IC_{50} = 64 \text{ nM}$) and improved kinase selectivity (S(90) = 0.23, Table 2). However, the IR IC_{50} was modestly affected and the IR/c-Met IC_{50} ratio remained <10.

Initial results indicated that the IR/c-Met ratio was improved in the 8-aminobenzazepinone series (Table 3). For example, benzazepinone **25**, R^3 = methoxyethyl, maintained cellular c-Met inhibition and overall kinase selectivity but had increased separation from IR activity (IR/c-Met IC₅₀ ratio = 115). This SAR was more fully explored in the 8-amino-5,5-dimethylbenzazepin-2-one series (Table 4); c-Met inhibition fell while kinase selectivity (i.e., *S*(90) values) improved with increasing α -branching of the lactam substituent (R³). In this series, analog **27** (R³ = Et) possessed the best balance of c-Met cellular potency (cell IC₅₀ = 44 nM), selectivity against IR (IR/c-Met IC₅₀ ratio = 73), and kinase selectivity [*S*(90) = 0.17] see Supplementary data for a list of kinases inhibited by this compound greater than 90% at1 µM). Analog **24** demonstrated modest IV and oral exposure in rat (1 mg/kg IV, 5 mg/kg PO: IV $t_{1/2} = 0.7$ h; CL = 15 mL/min/kg; IV AUC_{0-∞} = 1261 ng h/mL; PO AUC_{0-∞} = 592 ng h/mL; %*F* = 8%). Analog **26**, with its putatively metabolic benzylic sites protected with methyl groups demonstrated lower oral exposure (1 mg/kg IV, 5 mg/kg PO: IV $t_{1/2} = 0.3$ h; CL = 16 mL/min/kg; IV AUC_{0-∞} = 1057 ng h/mL; PO AUC_{0-∞} = 46 ng h/mL; %*F* <1%), and analog **27** had even lower IV and oral coverage. Current efforts are directed at improving the PK properties of these analogs.

In summary, a series of 2,4-diaminopyrimidine analogs bearing a C4-*N*-methylbenzamide and a C2-benzazepin-2-one was prepared and representative compounds were shown to be potent inhibitors of c-Met kinase. Kinase selectivity could be increased without compromising c-Met cellular activity by incorporating a fluorine atom at the C3 position of the pendant aminobenzamide. Further improvement in kinase selectivity in general, and over IR kinase in particular, could be realized by substitution of the benzazepinone nitrogen. Thus, the *S*(90) values dropped from 0.57

Table 3

c-Met SAR and kinase selectivity of 8-aminobenzazepinone analogs 24-25



Compd	R ³	IC_{50}^{a} (nM)					
		c-Met enzyme	c-Met cell ^b	IR enzyme	IR/c-Met enzyme	<i>S</i> (90) ^c	
24 25	H (CH ₂) ₂ OMe	30 10	99 61	369 1150	12 115	0.24 0.25	
25	$(CH_2)_2OMe$	10	61	1150	115	0.25	

^a Average of at least two separate determinations; see Supplementary data for assay conditions.

^b GTL-16 cells.

^c Kinase selectivity was determined using the Ambit Bioscience KINOMEscan™ technology.

Table 4

c-Met SAR and kinase selectivity of 8-amino-5,5-dimethylbenzazepinone analogs 26-30



Compd	R ³	IC_{50}^{a} (nM)					
		c-Met enzyme	c-Met cell ^b	IR enzyme	IR/c-Met enzyme	<i>S</i> (90) ^c	
26	Н	54	10	153	3	0.29	
27	Et	10	44	730	73	0.17	
28	i-Pr	376	NT ^d	404	1	NT ^d	
29	i-Bu	84	214	840	10	0.07	
30	(CH ₂) ₂ OMe	18	85	231	13	0.23	

^a Average of at least two separate determinations; see Supplementary data for assay conditions.

^b GTL-16 cells.

^c Kinase selectivity was determined using the Ambit Bioscience KINOMEscan™ technology.

^d Not tested.

for our initial lead compound **19** to 0.17 for substituted analog **27** and the IR/c-Met IC₅₀ ratio increased from 0.6 to 73 without impacting c-Met cellular IC₅₀ values (80 nM vs 44 nM, respectively).

Supplementary data

Supplementary data (biological assay conditions, the kinase profile for compound **27**, experimental procedures and spectral characterization) associated with this article can be found, in the online version, at doi:10.1016/j.bmcl.2010.11.045.

References and notes

- 1. Cui, J. J. Expert Opin. Ther. Patents 2007, 17, 1035.
- 2. Peruzzi, B.; Bottaro, D. P. Clin. Cancer Res. 2006, 12, 3657.
- (a) Underiner, T. L.; Herbertz, T.; Miknyoczki, S. J. Anti-Cancer Agents Med. Chem. 2010, 10, 7; (b) Dussault, I.; Bellon, S. Anti-Cancer Agents Med. Chem. 2009, 9, 221.

- (a) Ott, G. R.; Tripathy, R.; Cheng, M.; McHugh, R.; Anzalone, A. V.; Underiner, T. L.; Curry, M. A.; Quail, M. R.; Lu, L.; Wan, W.; Angeles, T. S.; Albom, M. S.; Aimone, L. D.; Ator, M. A.; Ruggeri, B. A.; Dorsey, B. D. ACS Med. Chem. Lett. 2010. doi:10.1021/ml100158; (b) Zificsak, C. A.; Theroff, J. P.; Aimone, L. D.; Albom, M. S.; Angeles, T. S.; Brown, R.; Galinis, D.; Grobelny, J. V.; Herbertz, T.; Husten, J.; Kocsis, L. S.; LoSardo, C.; Miknyoczki, S. J.; Murthy, S.; Rolon-Steele, D.; Underiner, T. L.; Worrell, C. S.; Zeigler, K.; Dorsey, B. D. Bioorg. Med. Chem. Lett., submitted for publication.
- Ahmed, G.; Bohnstedt, A.; Breslin, H. J.; Burke, J.; Curry, M. A.; Diebold, J. L.; Dorsey, B.; Dugan, B. J.; Feng, D.; Gingrich, D. E.; Guo, T.; Ho, K.; Learn, K. S.; Lisko, J. G.; Liu, R.; Mesaros, E. F.; Milkiewicz, K.; Ott, G. R.; Parrish, J.; Theroff, J. P.; Thieu, T. V.; Tripathy, R.; Underiner, T. L.; Wagner, J. C.; Weinberg, L.; Wells, G. J.; You, M.; Zificsak, C. A. International Patent WO 2008051547A1, 2008.
- 6. Smith, P. A. S.; Berry, W. L. J. Org. Chem. 1961, 26, 27.
- 7. As a key regulator of glucose metabolism, selectivity for c-Met over IR was preferred to be >10-fold in enzyme assays.
- Fabian, M. A.; Biggs, W. H.; Treiber, D. K.; Atteridge, C. E.; Azimioara, M. D.; Benedetti, M. G.; Carter, T. A.; Ciceri, P.; Edeen, P. T.; Floyd, M.; Ford, J. M.; Galvin, M.; Gerlach, J. L.; Grotzfeld, R. M.; Herrgard, S.; Insko, D. E.; Insko, M. A.; Lai, A. G.; Lelias, J.; Mehta, S. A.; Milanov, Z. V.; Velasco, A. M.; Wodicka, L. M.; Patel, H. K.; Zarrinkar, P. P.; Lockhart, D. J. Nat. Biotechnol. 2005, 23, 329.