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Glycosylation with 3,5-Dimethyl-4-(2'-phenylethynylphenyl)phenyl (EPP) Glycosides via a Dearomative Activation Mechanism

Zhifei Hu,^{+&} Yu Tang, ^{‡&} Biao Yu^{‡*}

 * School of Physical Science and Technology, ShanghaiTech University, 100 Haike Road, Shanghai 201210, China
* State Key Laboratory of Bioorganic and Natural Products Chemistry, Center for Excellence in Molecular Synthesis, Shanghai Institute of Organic Chemistry, University of Chinese Academy of Sciences, Chinese Academy of Sciences, 345 Lingling Road, Shanghai 200032, China

Supporting Information Placeholder

ABSTRACT: A highly effective and versatile glycosylation method is developed, which uses 3,5-dimethyl-4-(2²-phenylethynylphenyl)phenyl (EPP) glycosides as donors and NIS/TMSOTf as promoter and proceeds via an unprecedented dearomative activation mechanism.

The last decade witnesses the development of various new glycosylation methods in efforts to tackle the challenges occurring in the effective synthesis of glycans and glycoconjugates of great structural diversity.^{1,2} A category of these reactions capitalizes on selective activation of a C-C triple bond installed in the aglycone of the glycosylation donors by alkynophilic reagents (Figure 1).³ Upon activation, the ester type donors (e.g., A^{4a} and B^{4b}) readily undergo cleavage of the anomeric C-O bond, leading to the glycosyl oxacarbenium related species (e.g., I) for glycosylation.1d In comparison, the ether type donors (e.g., C^{5a}) require much stronger conditions for breaking of the glycosidic C-O bond, thus limiting the scope of the resultant glycosylation reactions. Nevertheless, the stability of the ether type donors would allow various protecting group manipulations, thus to facilitate the streamlined assembly of glycans and glycoconjugates (cf., the versatility of thioglycosides in the synthesis of glycan).^{1b,6} Although we have disclosed that the glycosyl orthoalkynylbenzoates (B) are the optimal ester type donors under the gold(I) catalysis,^{3a} our efforts toward development of an ether type donors basing on a similar activation mechanism met with no success. We found both glycosyl ortho-alkynylbenzyl and orthoalkynylphenyl glycosides were inert toward gold(I) catalyst (e.g., Ph₃PAuNTf₂ and Ph₃PAuOTf) under mild glycosylation conditions. Further engineering of the aglycone led to ortho-(methyltosylaminoethynyl)benzyl glycosides (**D**)^{5b} and ortho-(p-(E),^{5c} methoxyphenylethynyl)phenyl (MPEP) glycosides respectively. The former can undergo glycosylation under the catalysis of TMSOTf more effectively than under Ph₃PAuNTf₂ or Ph₃PAuOTf, and the latter, remaining inert toward the gold(I) catalyst, can be activated under NIS/TMSOTf to proceed glycosylation.

During the course of these studies, we once tried 2'-(phenylethynyl)biphenyl glycoside 1 as a potential glycosylation donor (Scheme 1). Glycoside 1 stayed inert in the presence of Ph₃PAuNTf₂ or Ph₃PAuOTf. Interestingly, it was converted under the action of NIS/TMSOTf into phenanthrene glycoside 3 (37%) and lactol **4** (43%). The phenanthrene derivative **3** was produced apparently via an intramolecular Friedel-Crafts type of reaction (Route 2). The presence of the hydrolyzed product **4** implied generation of the glycosyl oxacarbenium species (**I**) and aglycone derivative **III** in the reaction, that involved exactly the occurrence of the expected glycosylation pathway (Route 1). To our surprise, the aglycone derivative was determined to be spiroindene **5** rather than the expected benzo[c]chromene **III**.⁷ Thus, a dearomative pathway actually took place (Route 3).⁸ Such an activation mode resulting in the cleavage of a *O*-glycoside is unprecedented.⁹ Herein, we disclose a new glycosylation protocol which capitalizes on a relevant dearomative activation mechanism.



Figure 1. The glycosylation protocols based on activation of C-C triple bond and the representative glycosyl donors and promoters. Bu = butyl, Ts = p-toluenesulfonyl.

To facilitate the dearomative pathway and for the reason of easy preparation, we designed 3,5-dimethyl-4-(2'phenylethynylphenyl) phenyl (EPP) glycosides (9) as potential glycosylation donors (Scheme 2). Compared to the original biphenyl-2-yl glycoside 1, the new EPP glycosides are biphenyl-4yl glycosides, which are easier to synthesize, in addition, two methyl substituents are introduced to avoid the competitive Friedel-Crafts type of reaction. Indeed, an effective procedure was developed for the synthesis of EPP glycosides, which generally

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involved: (1) treatment of peracetyl monosaccharides (i.e., **6a-d**) and



Scheme 1. A proposed pathway for the glycosylation of 2'-(phenylethynyl)biphenyl glycoside 1 (Route 1) and the unexpected reactions (Routes 2 and 3). Bz = benzoyl, NIS = N-iodosuccinimide, TMSOTf = trimethylsilyl trifluoromethanesulfonate.

2'-iodo-2,6-dimethyl-[1,1'-biphenyl]-4-ol (7; CCDC1882371) with BF₃·OEt₂ or TMSOTf to give the corresponding phenyl glycosides (**8a-d**) (71-86%); (2) protecting group transformation or functional group conversion on the sugar unit; and (3) installation of the phenylethynyl moiety on the aglycone via Sonagashira coupling (86-97%).⁷ The resultant EPP glycosides (**9a-h**) are crystalline compounds and stay inert on shelf for at least three months. The structure of **9a** was confirmed by X-ray diffraction analysis (CCDC1881794).

Using perbenzoyl EPP glucopyranoside **9b** as donor and methyl 2,3,6-tri-*O*-benzyl- α -glucopyranoside (**2b**), a hindered sugar alcohol, as acceptor, we screened a series of promoters for the glycosylation in CH₂Cl₂ at RT, to provide the coupled (1 \rightarrow 4)-disaccharide **10a**.⁷ As expected, EPP glycoside **9b** stayed inert in the presence of Ph₃PAuNTf₂, Ph₃PAuOTf, AuCl₃, AgOTf, TMSOTf, and HOTf, nevertheless, it underwent glycosylation effectively under the action of NIS and TMSOTf.⁷ Where the glycosylation took place, the spiroindene derivative **12** was readily isolable, and its structure was confirmed unambiguously by X-ray diffraction analysis (CCDC1881796)(Scheme 3).



Scheme 2. A general procedure for the preparation of EPP glycosides and those studied in this report. Ac = acetyl, Bn = benzyl.

The model glycosylation reaction $(9b + 2b \rightarrow 10a)$, employing a disarmed donor and a hindered acceptor, could give the coupling product in 90% yield under the action of NIS (1.5 eq) and TMSOTF (0.3 eq) in CH₂Cl₂ at 0°C-RT. We were therefore confident that this new glycosylation protocol would possess a wide reaction scope. Indeed, under the present or slightly modified conditions, the glycosylation of EPP donors (**9a-9h**) with a wide array of alcoholic acceptors (**2a-j**)⁷ all led to the desired *O*-glycosides (**10a-x**) in satisfactory yields (Scheme 3). Thus, employing perbenzoyl EPP glucopyranoside **9b** as donor, β -(1 \rightarrow 6)-, (1 \rightarrow 2)-, (1 \rightarrow 3)-, besides the (1 \rightarrow 4)-disaccharide, were synthesized in excellent yields (**10af**; 85-98%); the glycosylation of aglycones, i.e., 1-adamantanol, menthol, and benzyl oleanolate, provided the coupled glycosides (**10g-i**) in ~90% yields. With perbenzyl EPP glucopyranoside (**9c**) as donor, the glycosylation proceeded smoothly and led expectedly

to an anomeric mixture of the glycosides (**10j-m**, 87-96%, $\alpha/\beta =$ 1:3 to 2.2:1). The peracetyl EPP glucopyranoside (**9a**), like the corresponding *ortho*-alkynylbenzoate,¹⁰ was a poor donor due to formation of orthoester derived products. The peracetyl EPP xylopyranosyl- and rhamnopyranosyl donors (**9d** and **9f**) reacted with the primary sugar alcohol **2b** to give the desired glycosides (**10n** and **10s**) in ~70% yields along with by-products derived from the donors;¹⁰ in comparison, their reactions with secondary alcohols and the reactions of their perbenzoyl counterparts (**9e** and **9g**) proceeded much cleanly, providing the corresponding coupled glycosides (**10o-q** and **10t-x**) in 84-98% yields. It is noteworthy that the corresponding MPEP α -rhamnopyranosides (**E**) did not undergo glycosylation under the activation of NIS/TMSOTf.^{5c}

The donor properties of the EPP glycosides were further tested in the *N*-glycosylation of pyrimidine and purine derivatives, which are known to be poor nucleophiles.¹¹ To our good fortune, purines (20 and 2p) were glycosylated with perbenzoyl EPP glucopyranoside (9b) and ribofuranoside (9h) to give the desired nucleosides (11d-f) in satisfactory yields (~70%). Upon activation and solvation with BSTFA in CH₃CN, pyrimidines (2m and 2n) were glycosylated to provide the corresponding pyrimidine nucleosides (11a-c) in excellent yields (>90%).



Scheme 3. Scope of the glycosylation reaction with EPP glycosides as donors under the action of NIS/TMSOTf. All = allyl, Boc = *tert*-butoxycarbonyl, BSTFA = *N*,*O*-bis(trimethylsilyl)trifluoroacetamide, Ph = phenyl, TBS = *tert*-butyldimethylsilyl. ^aThe amount of TMSOTf was increased to 0.5 equiv. ^bThe donor was used in excess (1.2 equiv) and the reaction performed at -20 °C-RT. ^cThe α/β ratio was determined by ¹H NMR analysis of the product mixture.

Given the fact that the EPP donors are derived from the corresponding 2'-iodo-2,6-dimethylbiphenyl-4-yl glycosides (8), which are inactive in the glycosylation conditions of the formers, these donors could be applied to the expeditious synthesis of glycans based on the 'latent-active' strategy.¹² To prove the concept, 2,3-di-O-benzoyl-4-O-benzyl-glucopyranoside 13 readily prepared from peracetyl 2'-iodo-2,6-dimethylbiphenyl-4-yl glycoside 8a via routine protecting group transformations (4 steps. 70% yield),⁷ was subjected to glycosylation with EPP xylosyl donor 9e. Under the present glycosylation conditions, disaccharide 14 was obtained in an excellent 98% yield, which was then converted readily into disaccharide EPP donor 15 via Sonagashira coupling (94%). Subjection of the nascent donor 15 to the next glycosylation with sugar alcohol 2b furnished trisaccharide 16 (85%).

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Scheme 4. A latent-active strategy for the synthesis of trisaccharide 16.

It is noted that the activation conditions (with NIS and TMSOTf as promoter) for the present EPP glycosides have been effectively applied for the glycosylation reactions with thioglycosides as donors.^{6,13} Thus, we compared the donor reactivity of two pairs of the EPP glycosides and thioglycosides (Scheme 5). In a mixture of EPP and *p*-tolylthic perbenzoyl glucopyranoside (9b and 17a, 1.0 eq each) and sugar alcohol **2b** (1.0 eq) in CH₂Cl₂ at -15 °C were added NIS (1.0 eq) and TMSOTf (0.3 eq). After 2 h, the EPP donor **9b** was completely consumed, leading to the coupled disaccharide 10a and spiroindene derivative 12 in 69% and 93% isolated yield, respectively; while the thioglycoside 17a was fully recovered (98%). Moreover, in a similar competition reaction of EPP donor 9b and perbenzyl thioglycoside 17b, which is ~2000 times more reactive than its perbenzoyl counterpart (17a),¹⁴ EPP donor 9b was still the one being fully activated, leading to coupled disaccharide 10a (67%) and spiroindene 12 (96%), while the thioglycoside 17b remained largely intact (94% recovery).15 This dramatic disparity in reactivity would enable a one-pot synthesis of glycan with combined use of an EPP donor and a thioglycoside acceptor without a concern on the protecting group pattern.^{1b,6} To prove the concept, EPP glycoside 9e was condensed with thioglycoside 18 under the action of NIS/TMSOTf to give the corresponding thiodisaccharide, upon addition of acceptor 13 and another portion of NIS/TMSOTf, the desired trisaccharide 19 was obtained in a high 89% yield. Trisaccharide 19, which bears a 2'-iodo-2,6dimethylbiphenyl aglycone, could be further applied in the latentactive synthesis (Scheme 5).



Scheme 5. Comparison of the donor reactivity of the EPP glycosides and thioglycosides under the action of NIS/TMSOTf and a one-pot synthesis of trisaccharide **19**.

In conclusion, we have developed a new glycosylation protocol with EPP glycosides (9) as donors, which proceeds via an unprecedented dearomative activation mechanism. This protocol, which can proceed under mild conditions, has demonstrated a broad reaction scope, including in the effective N-glycosylation of nucleobases. The stability of the EPP glycosides and their precursors allows various protecting group manipulations and enables application in the streamlined synthesis of glycans and glycoconjugates, as demonstrated here by the synthesis of trisaccharide 16 with the latent-active strategy. In comparison to the thioglycosides, which represent one of the most useful types of glycosylation donors, the present EPP glycosides can be activated much easily for glycosylation under the action of NIS/TMSOTf. In fact, the combined use of EPP glycosides and thioglycosides has demonstrated to be a new combination for one-pot synthesis of glycans. More importantly, the aglycone transfer side reaction of thioglycosides¹⁶ is completely avoided with the EPP donors, which release spiroindene derivative 12 as an innocuous side product. With these promising features, this new glycosylation method deserves further studies and shall find wide application.

ASSOCIATED CONTENT

Supporting Information

Supporting Information. Experimental procedures, analytical data (¹H and ¹³C NMR, MS) for all new compounds. This material is available free of charge via the Internet at http://pubs.acs.org.

AUTHOR INFORMATION

Corresponding Author

E-mail: byu@sioc.ac.cn

Author Contributions

& These authors contributed equally.

Notes

The authors declare no competing financial interests.

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