Journal of Medicinal Chemistry



Article

Development of protein degradation inducers of androgen receptor by conjugation of androgen receptor ligands and inhibitor of apoptosis protein (IAP) ligands

Norihito Shibata, Katsunori Nagai, Yoko Morita, Osamu Ujikawa, Nobumichi Ohoka, Takayuki Hattori, Ryokichi Koyama, Osamu sano, Yasuhiro Imaeda, Hiroshi Nara, Nobuo Cho, and Mikihiko Naito

J. Med. Chem., Just Accepted Manuscript • Publication Date (Web): 08 Jun 2017 Downloaded from http://pubs.acs.org on June 9, 2017

Just Accepted

"Just Accepted" manuscripts have been peer-reviewed and accepted for publication. They are posted online prior to technical editing, formatting for publication and author proofing. The American Chemical Society provides "Just Accepted" as a free service to the research community to expedite the dissemination of scientific material as soon as possible after acceptance. "Just Accepted" manuscripts appear in full in PDF format accompanied by an HTML abstract. "Just Accepted" manuscripts have been fully peer reviewed, but should not be considered the official version of record. They are accessible to all readers and citable by the Digital Object Identifier (DOI®). "Just Accepted" is an optional service offered to authors. Therefore, the "Just Accepted" Web site may not include all articles that will be published in the journal. After a manuscript is technically edited and formatted, it will be removed from the "Just Accepted" Web site and published as an ASAP article. Note that technical editing may introduce minor changes to the manuscript text and/or graphics which could affect content, and all legal disclaimers and ethical guidelines that apply to the journal pertain. ACS cannot be held responsible for errors or consequences arising from the use of information contained in these "Just Accepted" manuscripts.



Journal of Medicinal Chemistry is published by the American Chemical Society. 1155 Sixteenth Street N.W., Washington, DC 20036

Published by American Chemical Society. Copyright © American Chemical Society. However, no copyright claim is made to original U.S. Government works, or works produced by employees of any Commonwealth realm Crown government in the course of their duties.

Development of protein degradation inducers of androgen receptor by conjugation of androgen receptor ligands and inhibitor of apoptosis protein (IAP) ligands

Norihito Shibata[†], Katsunori Nagai[‡], Yoko Morita[‡], Osamu Ujikawa[‡], Nobumichi Ohoka[†], Takayuki Hattori[†], Ryokichi Koyama[‡], Osamu Sano[‡], Yasuhiro Imaeda[‡], Hiroshi Nara[‡], Nobuo Cho[‡], Mikihiko Naito^{*,†}

†Divisions of Molecular Target and Gene Therapy Products, National Institute of Health Sciences, 1-18-1 Kamiyoga, Setagaya-ku, Tokyo 158-8501, Japan.

‡Pharmaceutical Research Division, Takeda Pharmaceutical Co. Ltd., 26-1,Muraoka-Higashi 2-chome, Fujisawa, Kanagawa 251-0012, Japan

Targeted protein degradation using small molecules is a novel strategy for drug development. We have developed hybrid molecules named Specific and Non-genetic inhibitor of apoptosis protein [IAP]-dependent Protein Erasers (SNIPERs) that recruit IAP ubiquitin ligases to degrade target proteins. Here, we show novel SNIPERs capable of inducing proteasomal degradation of the androgen receptor (AR). Through derivatization of the SNIPER(AR) molecule at AR ligand, IAP ligand and linker, we developed 42a (SNIPER(AR)-51), which shows effective protein knockdown activity against AR. Consistent with the degradation of the AR protein, 42a inhibits AR-mediated gene expression and proliferation of androgen-dependent prostate cancer cells. In addition, 42a efficiently induces caspase activation and apoptosis in prostate cancer cells, which was not observed in the cells treated with AR antagonists. These results suggest that SNIPER(AR)s could be leads for an anti-cancer drug against prostate cancers that exhibit AR-dependent proliferation.

INTRODUCTION

Prostate cancer (PC) is the second-leading cause of cancer deaths, after lung cancer, in American males.¹ Since the progression of PC is primarily stimulated by androgens, inhibition of the secretion and/or action of androgens is highly effective in controlling the growth of PC.²⁻⁴ Besides surgical castration (orchiectomy) or androgen synthesis inhibitors, androgen receptor (AR) antagonists are widely used to treat PC. These antagonists suppress androgen signaling by inhibiting the binding of androgens to AR. Currently, AR antagonists such as flutamide, bicalutamide and enzalutamide are used clinically.⁵ In combination, AR antagonists and androgen synthesis inhibitors show significant synergistic effects by blocking androgen signaling as well as by suppressing transient increase in testosterone levels. Although such therapy is initially effective, a considerable population of patients develops castration-resistant prostate cancer (CRPC) after the prolonged use of an AR antagonist, primarily because of an increased level of AR expression in clinical situations.⁶⁻⁸ Therefore, down-regulation of the AR level is regarded as a promising strategy to treat

Down-regulation of the AR protein can be achieved by genetic knockdown technologies, such as antisense oligonucleotides, RNA interference and genome editing. Despite its great therapeutic potential, genetic knockdown remains clinically challenging.9, ¹⁰ because oligonucleotides are not efficiently taken up into cells and delivery to the desired target tissues is technically difficult. An alternative approach is protein knockdown, which induces the degradation of target proteins via the ubiquitin proteasome system (UPS). Based on this approach, molecules named Specific and Non-genetic inhibitor of apoptosis protein [IAP]-dependent Protein Erasers (SNIPERs)¹¹⁻²² and Proteolysis Targeting Chimeras (PROTACs)²³⁻³² have been developed. Both classes represent hybrid molecules composed of two different ligands connected by a linker; one ligand is for the protein of interest (POI) and the other is for E3 ubiquitin ligases. Accordingly, these molecules are expected to crosslink the POIs and E3 ubiquitin ligases in cells, resulting in the ubiquitylation and subsequent degradation of the POIs via the UPS. To date, we have developed several SNIPERs by connecting POI ligands to 79 (bestatin)³³ a ligand for

cellular IAP 1 (cIAP1), which induces the degradation of the POIs comprising cellular retinoic acid binding protein-II,^{13, 21} estrogen receptor,¹⁶ transforming acidic coiled-coil-3¹⁷ and BCR-ABL.¹⁹

In this study, we designed and synthesized SNIPER(AR)s that induce proteasomal degradation of the AR protein by conjugating an AR antagonist to an IAP ligand, **24** (LCL-161³⁴ derivative). We also showed that SNIPER(AR)s not only inhibit the growth but also induce the cell death of PC cells, which exhibit AR-dependent proliferation.

CHEMISTRY

The syntheses of **4a-d** (SNIPER(AR)-2, 8, 23 and 26, respectively) were performed according to Scheme 1. Removal of the Boc group of bestatin analog **1**, followed by condensation with the AR ligand part **2a-d** afforded the corresponding coupling products **3a-d**, which were treated with dimethylamine to give the bestatin-based SNIPER(AR)s

4a-d.

Scheme 1. Synthesis of 4a-d^a



^{*a*}Reagents and conditions: (a) (1) aq. HCl, THF, rt; (2) **2a**, **2b**, **2c**, or **2d**, EDC, HOBt·H₂O, DIPEA, THF, rt, 41–76%; (b) Me₂NH, MeOH, rt, 68–98%.

Compound **4e** (SNIPER(AR)-27) having a different pyrazole-based AR ligand was prepared by the route shown in Scheme 2. Protection of the pyrazole nitrogen atom of **5** with 2-(trimethylsilyl)ethoxymethyl (SEM) group and subsequent alkaline hydrolysis of the ester group yielded the benzoic acid derivative **7**, which was condensed with the Boc-protected diamine **8** to give **9**. Treatment of **9** with hydrochloride–cyclopropyl methyl ether (CPME) provided the fully-deprotected intermediate, which was reacted with excess di-*tert*-butyl dicarbonate under basic conditions, and subsequently treated with sodium methoxide to furnish the pyrazole analog **10**. Nucleophilic substitution of the fluorine atom of **11** with **10** gave **12**. Boc-deprotection of **12** followed by condensation with **13** produced **3e**, which was converted to the target compound **4e** by Fmoc-deprotection.

Scheme 2. Synthesis of 4e^{*a*}



^{*a*}Reagents and conditions: (a) SEMCl, NaH, DMF, 0 °C to rt, 89%; (b) aq. NaOH, THF, MeOH, rt, 84%; (c) **8**, EDC, HOBt, DMF, rt, 97%; (d) (1) 4 M HCl in CPME, MeOH, THF, reflux; (2) Boc₂O, aq. NaHCO₃, THF, rt, then NaOMe, MeOH, rt, 77%; (e) **11**, NaH, DMF, 0 °C to rt, 97%; (f) (1) 4M HCl in CPME, THF, dioxane, rt; (2) **13**, EDC, HOBt, DIPEA, DMF, rt, 73%; (g) Me₂NH, MeOH, rt, 81%.

Compound 18 (SNIPER(AR)-12) was synthesized by the route depicted in Scheme 3. The carboxylate 2a was condensed with the Boc-protected diamine 14 to produce 15. Removal of the Boc group of 15 and subsequent condensation of the obtained amine with 16 yielded the coupling product 17. Final Boc-deprotection of 17 afforded the

desired compound 18.

Scheme 3. Synthesis of 18^a



^{*a*} Reagents and conditions: (a) **14**, EDC, HOBt, THF, rt, 98%; (b) (1) 2 M HCl in CPME, THF, rt; (2) **16**, HATU, DIPEA, DMF, 0 °C, 95%; (c) 4 M HCl in CPME, THF, rt, 84%.

The synthetic route to **23** (SNIPER(AR)-31) is presented in Scheme 4. Mitsunobu reaction of the compound **19** with azido-alcohol **20** gave the corresponding coupling product, which was subjected to Staudinger reduction to provide the primary amine **21**. Coupling of **21** with the carboxylic acid **2a** afforded **22**, which was converted to the target

compound **23** by Boc-deprotection. The reference compound **24** was obtained from **19** under Mitsunobu reaction conditions.

Scheme 4. Synthesis of 23 and 24^a



^{*a*} Reagents and conditions: (a) (1) **20**, DEAD, Ph₃P, THF, 0 °C to rt; (2) Ph₃P, H₂O, quant.; (b) **2a**, EDC, HOBt, THF, DMF, rt, 37%; (c) 4 M HCl in CPME, THF, rt, 74%; (d) (1) MeOH, DEAD, Ph₃P, THF, 0 °C to rt; (2) 4 M HCl in CPME, rt, 42%.

Compounds 32a (SNIPER(AR)-60) and 32b (SNIPER(AR)-70) having different linker

moieties were synthesized according to Scheme 5. Sonogashira coupling of the

bromobenzene analog 25 with the acetylene 26 produced 27, which was sulfonylated with *p*-toluenesulfonyl chloride to give 28. Reaction of 28 with 19 under basic conditions, followed by removal of the Boc group of 31a furnished the desired product 32a. Palladium catalyzed hydrogenation of the alkyne derivative 27 produced 29, which was converted to the target compound 32b via the three-step procedure, i.e. tosylation, substitution with 19, and final deprotection.

Scheme 5. Synthesis of 32a and 32b^a





^{*a*}Reagents and conditions: (a) **26**, Pd(PPh₃)₄, CuI, Et₃N, DMF, 60 °C, 49%; (b) TsCl, pyridine, 0 °C to rt, 72–83%; (c) H₂, 10% Pd–C, EtOAc, rt, 84%; (d) **28** or **30**, K₂CO₃, DMF, 50 °C, 67–72%; (e) TFA, rt, 78–86%.

The syntheses of 42a-e (SNIPER(AR)-51, -53, -52, -171, and -65, respectively) having

different substitution pattern and linker length were carried out by the procedure depicted in Scheme 6. Various tosylated precursors (**36a–e**) were prepared by the three protocols: (i) reaction of the phenol analogs **33a,b** with the diol **34** under Mitsunobu reaction conditions, followed by tosylation; (ii) coupling of **33a** with the di-tosylated diol **37a** or **37b**; (iii) coupling of **38** with **37c**, mesylation of the benzyl alcohol **39**, and subsequent reaction with the pyrrole **40**. The tosylates **36a–e** were converted to the desired products **42a–e** in a similar procedure described for the preparation of **32** (Scheme 5).

Scheme 6. Synthesis of $42a-e^{a}$



^{*a*}Reagents and conditions: (a) **34**, DEAD, Ph₃P, THF, 0 °C to rt, 73–94%; (b) TsCl, pyridine, 0 °C to rt, 68–75%; (c) **37a** or **37b**, K₂CO₃, DMF, 50 °C, 46–51%; (d) **37c**, K₂CO₃, DMF, 40 °C, 46%; (e) (1) MsCl, Et₃N, THF, 0 °C; (2) **40**, NaH, DMF, 0 °C, 83%; (f) **36a**, **36b**, **36c**, **36d**, or **36e**, K₂CO₃, DMF, rt to 60 °C, 67–81%; (g) TFA, rt, 70–90%.



Scheme 7. Synthesis of 48a–c^a



^{*a*}Reagents and conditions: (a) TsCl, pyridine, 0 °C, 52–56%; (b) **33a**, K₂CO₃, DMF, 60 °C, 82–99%; (c) TsCl, pyridine, 0 °C to rt, 83–87%; (d) **19**, K₂CO₃, DMF, rt to 50 °C, 74–82%; (e) TFA, rt, 85–89%.

Induction of AR protein degradation by SNIPER(AR)

We designed a hybrid molecule **4a**, composed of **79** and **76** (AR antagonist-1), which possesses strong binding affinity to AR.³⁵ Based on the co-crystal structure of AR ligand-binding domain (W741L mutant) complexed with bicalutamide (Protein Data Bank (PDB) ID: 1Z95),³⁶ the fluorophenyl moiety of bicalutamide is located at the AR surface. Therefore, the PEG (polyethylene glycol) linker was ligated to the corresponding benzyl moiety of **76** at one end, and **79** at the other end, as shown in Figure 1A.

To examine whether **4a** reduces the AR protein level, human PC 22Rv1 cells were treated with graded concentrations of **4a**, and the whole-cell lysates were analyzed by western blot. The AR protein level was significantly decreased by treatment with **4a** at 30 μ M for 6 h (Figure 1B). Combination treatment with 30 μ M **76** and 30 μ M **80** (methyl bestatin)³³ did not decrease the AR protein level, indicating that linking the two ligands into a single molecule is essential for the reduction of the AR protein (Figure 1B).

We next conjugated **79** to various AR ligands,^{35, 37} whose binding affinities to AR ranged from 10^{-9} to 10^{-6} M. Compounds **4a**, **4c** and **4d**, which contain AR ligands with the binding affinity of 0.0014, 0.0014 and 0.053 µM, showed IC₅₀ values of 0.027, 0.19, and 0.19 µM in binding affinity to AR, and DC₅₀ values of 21.6, 11.7 and 20.5 µM in the reduction of AR protein expression, respectively (Figure 1C, D). However, **4b** and **4e**, which contain AR ligands with IC₅₀ values of 0.054 and 1.5 µM, had IC₅₀ values of 1.1 and >10 µM, and DC₅₀ values of >30 and >30 µM, respectively. These results suggest that incorporation of an AR ligand with a higher binding affinity is preferable to develop a SNIPER(AR) with effective protein knockdown activity.

57 58 59

60



Figure 1 Reduction of AR protein levels by bestatin-conjugated SNIPER(AR)s.

(A) Chemical structures of the SNIPER(AR)s. (B) Reduction of AR protein by 4a

(SNIPER(AR)-2). (C) Reduction of AR protein by SNIPER(AR)s containing various AR ligands. 22Rv1 cells were incubated with the indicated concentrations of SNIPER(AR)s or a mixture of the two chemically separate ligands (**76** and **80**, 30 μ M each) for 6 h. Numbers below the AR panel represent AR/GAPDH ratio normalized by vehicle control as 100. Data in the graphs are means \pm standard deviation (n = 3). *, *P*<0.05 compared with vehicle control (B, C). (D) Binding affinity and protein knockdown activity of the bestatin-conjugated SNIPER(AR)s. IC₅₀ values and DC₅₀ values (concentrations of the SNIPER(AR)s required to reduce the protein expression of AR by 50%) of SNIPER(AR)s were obtained from three separate experiments. The binding affinities of the AR ligands were cited from References 34 and 36.

Optimization of SNIPER(AR)

We selected **4a** as a lead, and derivatized it at several locations in turn – the E3 ligase ligand, the connection point (joint) between the AR ligand and linker, and finally the linker itself – to improve the protein knockdown efficacy against AR. First, we replaced bestatin-moiety

with two other specific IAP antagonists, methyl (2S)-2-(((2S)-1-((2S)-2-cyclohexyl-2-(((2S)-2-(methylamino)propanoyl)amino)acetyl)pyrro lidine-2-carbonyl)amino)-3,3-diphenylpropanoate (MV-1)³⁸ and 24, to generate 18 and 23, respectively (Figure 2A), since 79 is not a specific ligand for cIAP1.^{39, 40} For 4a and 18, the minimum concentrations necessary to reduce the AR protein levels were 30 and 20 μ M, respectively, while 23 was effective at 3 μ M and above. Thus, among the three tested IAP ligands, incorporation of 24 conferred the greatest AR protein degradation activity on the corresponding SNIPER(AR).

Compound 23 was further derivatized by modification of the joint (the functional group connecting 76 and the linker), i.e., amide (23), ether (42a), acetylene (32a) or alkyl groups (32b). These compounds all showed comparable abilities to reduce the AR protein level (Figure 2B). From 42a, we further derivatized the molecule by varying the position at which the linker was connected to 76: either the *o*-, *m*- or *p*-position on the benzyl moiety. The *m*-analog (42b) showed the same ability to reduce the AR protein level as did the *p*-analog (42a) (Figure 2C). However, the *o*-analog (42e) showed no activity against the AR

protein at concentrations of 1 or 3 μM (Figure 2C). Since the benzyl moiety of 76 is likely

to be located on the surface of the AR protein, conjugation of the linker at the o-position

may have hindered the binding of **42e** to the AR protein.



ACS Paragon Plus Environment

Figure 2 Derivatization of SNIPER(AR) by changing the cIAP1 ligand (A), joint (functional group between **76** and linker) (B), and position of joint (C). 22Rv1 cells were incubated with the indicated concentrations of SNIPER(AR)s for 6 h. Numbers below the AR panel represent AR/GAPDH ratio normalized by vehicle control as 100. Data in the graphs are means \pm standard deviation (n = 3). *, *P*<0.05 compared with vehicle control.

From **42a**, we further synthesized analogs with different linker lengths. SNIPER(AR)s with different lengths of PEG linker chain (n = 3, 4, 5) substantially reduced the AR protein levels at 3 μ M, while combination treatment with 3 μ M **76** and 3 μ M **24** did not affect the AR protein level (Figure 3A). Next, we examined the effect of linker flexibility on the protein knockdown activity. In contrast to **42a**, which contains a flexible PEG linker, the series **48a–c**, with rigid phenyl linkers, showed no activity against AR protein (Figure 3B), suggesting that flexibility of the linker is important for the reduction of AR protein levels by SNIPER(AR)s. On the basis of these results, we chose **42a** for further biological study.



linker molecules (B). 22Rv1 cells were incubated with the indicated concentrations of SNIPER(AR)s or 3 μ M of the two chemically separate ligands (**76** and **24**) for 6 h. Numbers below the AR panel represent AR/GAPDH ratio normalized by vehicle control as

100. Data in the graphs are means \pm standard deviation (n = 3). *, *P*<0.05 compared with vehicle control.

Biological Evaluation

SNIPER(AR)s are presumed to induce the ubiquitylation and subsequent degradation of AR via the UPS. Treatment of 22Rv1 cells with **42a** at 3 μ M reduced the AR protein level in a time-dependent manner, while this reduction of AR by **42a** was abrogated by a proteasome inhibitor, benzyl

N-((2S)-4-methyl-1-(((2S)-4-methyl-1-(((2S)-4-methyl-1-oxopentan-2-yl)amino)-1-oxopent an-2-yl)amino)-1-oxopentan-2-yl)carbamate **77** (MG132)⁴¹ (Figure 4A). These results indicate that **42a** induces the proteasomal degradation of AR. The proteasome-dependent reduction of AR protein by **42a** was similarly observed in another PC cell line, VCaP, which has an endogenous AR gene amplification and exhibits androgen-dependent proliferation⁴² (Figure 4B).



Figure 4 SNIPER(AR) induces the proteasomal degradation of AR protein. (A,

B) Effect of proteasome inhibitor, 77, on protein knockdown activity of 42a in 22Rv1 (A)

and VCaP (B) cells. Cells were incubated with 3 μM 42a with or without 10 μM 77 for the

indicated time. Numbers below the AR panel represent AR/GAPDH ratio normalized by

vehicle control as 100. Data in the graphs are means \pm standard deviation (n = 3). *, *P*<0.05 compared with vehicle control.

Next, we tested the effect of 42a on AR-mediated gene expression and proliferation in androgen-dependent PC VCaP cells. When cells were treated with 1 nM dihydrotestosterone (DHT), the expressions of AR-regulated genes (PSA, TMPRSS2, KLK2 and NKX3.1)⁴³⁻⁴⁵ were up-regulated. Compound **42a** at 3 μ M suppressed the expression of these AR-regulated genes (Figure 5A), which is consistent with the reduction of AR protein. In line with this, 42a at 3 µM dramatically inhibited the growth of VCaP cells (Figure 5B). A similar growth-inhibitory effect of 42a was also observed in another PC cell line, LNCaP (Figure 5C), which expresses the AR protein and exhibits androgen-dependent proliferation. However, 42a did not inhibit cell growth in the androgen-independent cell lines PC-3, A549 or HT1080 under the same conditions (Figure 5C). Treating the cells with 42a at 100 µM for 6 h seriously reduced the viability of all the cell lines, suggesting the toxicity of this compound. These data indicate that AR Page 29 of 140

degradation by 42a results in the inhibition of (i) AR-mediated gene expression and (ii)

androgen-dependent PC cell proliferation.





of and rogen-dependent PC cells. (A) Expression of and rogen-responsive genes in VCaP cells. Cells were incubated with 1 nM DHT in the absence or presence of 3 μ M 42a or 76

for 24 h. Expression levels are relative to vehicle treatment, which was arbitrarily set to 1. *, P < 0.05. (B, C) Effect of **42a** on the growth of various cells. VCaP cells were incubated with the indicated concentrations of **42a** (red line), bicalutamide (blue line) or combination of **76** and **24** (green line) for 48 h (B). The indicated cells were incubated with 3 μ M **42a** for 48 h (C). Data in the graphs are means \pm standard deviation (n = 3). *, P < 0.05compared with vehicle control.

By microscopy, we observed that **42a** exhibited a pronounced effect on the cell morphology of VCaP cells, appearing to induce apoptosis (Figure 6A). Therefore, we examined whether **42a** indeed induced apoptosis in the VCaP cells. Figure 6B shows that **42a** at 3 μ M, but not **76** or bicalutamide, induced caspase-3 activation and poly (ADP-ribose) polymerase (PARP) cleavage in VCaP cells. These effects of **42a** were abrogated by a pan-caspase inhibitor, **78** (zVAD-FMK)⁴⁶ (Figure 6C). In line with this, **42a** treatment increased the count of Annexin V-positive apoptotic cells, which was suppressed by **78** (Figure 6D). The induction of apoptosis by **42a** was prevented by **77** (Figure 6E).

These results suggest that the degradation of the AR protein by SNIPER(AR)s hampers androgen signaling, resulting in the inhibition of cell proliferation and induction of apoptosis in the AR-dependent PC cells.



Figure 6 SNIPER(AR) induces apoptosis. (A) VCaP cells were treated with 3 μ M **42a** or **76** for 24 h. Bar: 50 μ m. (B, C) Effect of **42a** on caspase activation in VCaP cells. Cells were incubated with the indicated concentrations of compounds (B), or 3 μ M **42a** with or without the indicated concentrations of **78** (C) for 24 h. (D) Cell death by **42a** was determined using Annexin V and PI (propidium iodide) staining. VCaP cells were incubated with 3 μ M **42a** with or without 50 μ M **78** for 24 h. Double negative cells represent living cells (lower left), whereas Annexin V-positive cells represent apoptotic cells (lower right). PI-positive (upper left) and double positive (upper right) cells represent necrotic cells. (E) Proteasome inhibitor prevents apoptosis induced by **42a** in VCaP cells. Cells were incubated with 3 μ M **42a** and/or 10 μ M **77** for 9 h.

CONCLUSION

Here, we reported the development of SNIPER(AR)s, which induce proteasomal degradation of the AR protein. Compound 42a inhibits androgen signaling and cell proliferation, and induces apoptosis in androgen-dependent PC cells. Previously, Crews et al. reported PROTACs against AR that induce AR protein degradation by connecting an AR ligand to ligands for von Hippel-Lindau (VHL) and Mdm2 E3 ubiquitin ligases.^{24, 26, 27} Thus, ligands for three E3 ubiquitin ligases, VHL, Mdm2 and cIAP1, can be used to produce small-molecule-based AR degradation inducers. We have shown that the ligand for the target protein, ligand for E3 ligase, linker composition and linker length are all important factors that affect the efficacy of target-protein degradation by hybrid molecules such as SNIPERs and PROTACs. Since protein knockdown is a promising strategy for the treatment of CRPC, further optimization is necessary to develop more efficient AR degradation inducers for clinical use in future.

Compared with PROTACs, SNIPERs induce the degradation of both POIs and cIAP1, which may limit the full potential and duration of knockdown activity by SNIPERs.

However, cIAP1 is an anti-apoptotic protein frequently overexpressed in human tumor cells,

which is believed to be involved in the resistance to tumor therapy.^{47, 48} Therefore,

degradation of cIAP1 simultaneously with POIs by SNIPERs could be beneficial to

effectively kill cancer cells.

EXPERIMENTAL SECTION

1. Chemistry.

1-1. General Procedures, Materials, and Information.

All solvents and chemicals were used as purchased without further purification. The progress of all reactions was monitored by thin-layer chromatography (TLC) analysis on silica gel 60 F₂₅₄ plates (Merck) or NH TLC plates (Fuji Silysia Chemical Ltd.). Column chromatography was performed with a silica gel column [(Merck Kieselgel 60, 70-230 mesh, Merck) or (Chromatorex[®] NH-DM1020, 100-200 mesh, Fuji Silysia Chemical Ltd.)], or with prepacked Purif-Pack columns [silica gel or NH (3-aminopropyl-functionalized) silica gel, particle size: 60 µm, Fuji Silysia Chemical Ltd.]. The proton nuclear magnetic resonance (¹H NMR) spectra were recorded on a Bruker AVANCE II (300 or 400 MHz) spectrometer. Chemical shifts for ¹H NMR were reported in parts per million (ppm) downfield from tetramethylsilane (δ) as the internal standard in deuterated solvent and coupling constants (J) are in Hertz (Hz). The following abbreviations are used for spin
multiplicity: s = singlet, d = doublet, t = triplet, q = quartet, quin = quintet, dd = doublet of doublets, dt = doublet of triplets, qd = quartet of doublets, dquin = doublet of quintets, m = multiplet, and br s = broad singlet. Low-resolution mass spectra (MS) were acquired using Agilent LC/MS (Agilent1200SL/Agilent6130MS, system an Agilent1200SL/Agilent1956MS or Agilent1200SL/Agilent6110MS), Shimadzu UFLC/MS (Shimadzu LC-20AD/LCMS-2020) operating in electron spray ionization mode (ESI+). The column used was an L-column 2 ODS (3.0×50 mm I.D., 3 µm, CERI, Japan) with a temperature of 40 °C and a flow rate of 1.2 or 1.5 mL/min. Condition 1: Mobile phases A and B under acidic conditions were 0.05% TFA in water and 0.05% TFA in MeCN, respectively. The ratio of mobile phase B was increased linearly from 5 to 90% over 0.9 min, and held at 90% over the next 1.1 min. Condition 2: Mobile phases A and B under neutral conditions were a mixture of 5 mmol/L AcONH₄ and MeCN (9:1, v/v) and a mixture of 5 mmol/L AcONH₄ and MeCN (1:9, v/v), respectively. The ratio of mobile phase B was increased linearly from 5 to 90% over 0.9 min, and held at 90% over the next 1.1 min. High-resolution mass spectrometry (HRMS) were carried out by Takeda

Analytical Research Laboratories, Sumika Chemical Analysis Service, Ltd., or measured by Bruker Autoflex Speed (MALDI-TOF Mass). Elemental analyses were carried out by Takeda Analytical Laboratories Ltd. The purities of all compounds tested in biological systems were assessed as being >95% using analytical HPLC. Analytical HPLC was performed with NQAD (Nano Quantity Analyte Detector) or Corona CAD (Charged Aerosol Detector). The column was an L-column 2 ODS (30 × 2.1 mm I.D., CERI, Japan) or a Capcell Pak C18AQ (50 mm × 3.0 mm I.D., Shiseido, Japan) with a temperature of 50 °C and a flow rate of 0.5 mL/min. Mobile phases A and B under neutral conditions were a mixture of 50 mmol/L ammonium acetate, water and acetonitrile (1:8:1, v/v/v) and a mixture of 50 mmol/L ammonium acetate and MeCN (1:9, v/v), respectively. The ratio of mobile phase B was increased linearly from 5 to 95% over 3 min, and held at 95% over the next 1 min. Optical rotation analyses were carried out by Sumika Chemical Analysis Service, Ltd. Specific rotations $\left[\alpha\right]_{D}^{25}$ are given in units of deg cm³ g⁻¹ dm⁻¹.

tert-Butyl

((5R,6S,9S)-5-benzyl-1-(9H-fluoren-9-yl)-6-hydroxy-9-isobutyl-3,7,10-trioxo-2,14,17,20-te

ACS Paragon Plus Environment

traoxa-4,8,11-triazadocosan-22-yl)carbamate 1 was synthesized according to the procedures of Ohoka.¹⁷ Methyl 4-((3,5-dimethyl-1*H*-pyrazol-4-yl)methyl)benzoate 5, 4-(4-cyanophenyl)-2,5-dimethyl-1H-pyrrole-3-carbonitrile 49. 2-chloro-4-(4-iodo-3,5-dimethyl-1H-pyrazol-1-yl)benzonitrile 56. and the tested 1-benzyl-4-(4-cyanophenyl)-2,5-dimethyl-1H-pyrrole-3-carbonitrile 76 were synthesized 37 Yamamoto.^{35,} according procedures of to the N-(4-Cyano-3-(trifluoromethyl)phenyl)-2-methyloxirane-2-carboxamide 52 and .49 bicalutamide synthesized according procedures Tucker were to the of 4-((1-(4-Cyano-3-(trifluoromethyl)phenyl)-3,5-dimethyl-1H-pyrazol-4-yl)methyl)benzoic acid tert-butyl and (2S)-2-(4-(methoxy(methyl)carbamoyl)-1,3-thiazol-2-yl)pyrrolidine-1-carboxylate 72 were synthesized according to the procedures described in patent applications.^{34, 50} Compound **79** ((2S)-2-(((2S,3R)-3-amino-2-hydroxy-4-phenylbutanoyl)amino)-4-methylpentanoate) was OChem purchased from Inc. 80 (methyl and (2S)-2-(((2S,3R)-3-amino-2-hydroxy-4-phenylbutanoyl)amino)-4-methylpentanoate) was

ACS Paragon Plus Environment

synthesized as described previously.³³ Schemes and experimental procedures for synthesis of AR and IAP ligand parts are provided in the Supporting Information.

1-2.

N-(13-(4-((3-Cyano-4-(4-cyanophenyl)-2,5-dimethyl-1*H*-pyrrol-1-yl)methyl)phenyl)-13 -oxo-3,6,9-trioxa-12-azatridec-1-yl)-*N*²-((2*S*,3*R*)-3-(((9*H*-fluoren-9-ylmethoxy)carbonyl)amino)-2-hydroxy-4-phenylbutanoyl)-L-leucinamide (3a).

To a solution of **1** (300 mg, 0.373 mmol) in THF (5 mL) was added 4 M aq. HCl (3.7 mL, 15 mmol) at room temperature. The mixture was stirred at room temperature for 5 h. The mixture was concentrated to give a residual oil (276 mg); MS m/z 705 (M+H)⁺.

To a solution of the obtained oil (276 mg), HOBt·H₂O (72 mg, 0.47 mmol), DIPEA (0.34 mL, 2.0 mmol), and **2a** (140 mg, 0.394 mmol) in THF (5 mL) was added EDC (82 μ L, 0.47 mmol) at room temperature. The mixture was stirred at room temperature overnight. The mixture was poured into water at room temperature and extracted with EtOAc. The organic layer was separated, washed with water and brine, dried over MgSO₄, and concentrated *in*

vacuo. The residue was purified by column chromatography on silica gel (eluted with 0– 15% MeOH in EtOAc) to give the title compound (274 mg, 67%) as a colorless amorphous solid. ¹H NMR (300 MHz, CDCl₃) δ 0.73–0.90 (6H, m), 1.47–1.71 (3H, m), 2.14 (3H, s), 2.32 (3H, s), 2.83–3.06 (2H, m), 3.18–3.32 (1H, m), 3.32–3.72 (16H, m), 4.04–4.34 (5H, m), 4.40–4.56 (1H, m), 5.09 (2H, s), 5.45 (1H, d, *J* = 8.4 Hz), 6.88–7.00 (3H, m), 7.14–7.33 (8H, m), 7.36–7.43 (2H, m), 7.45–7.55 (4H, m), 7.69 (2H, d, *J* = 8.4 Hz), 7.75 (2H, d, *J* = 7.5 Hz), 7.82 (2H, d, *J* = 8.4 Hz); MS *m/z* 1042.9 (M+H)⁺.

1-3.

9H-Fluoren-9-ylmethyl

((2*R*,3*S*,6*S*)-21-(4-((3-((4-cyano-3-(trifluoromethyl)phenyl)amino)-2-hydroxy-2-methyl -3-oxopropyl)sulfonyl)phenyl)-3-hydroxy-6-isobutyl-4,7,21-trioxo-1-phenyl-11,14,17-tr ioxa-5,8,20-triazahenicosan-2-yl)carbamate (3b).

Compound **3b** was prepared from **1** and **2b** in a similar manner to that described for compound **3a** and obtained as a colorless amorphous solid (76%). ¹H NMR (300 MHz, CD₃OD) δ 0.72–0.88 (6H, m), 1.41–1.67 (6H, m), 2.79–3.01 (2H, m), 3.17–3.41 (2H, m),

3.42–3.72 (15H, m), 3.93–4.14 (4H, m), 4.21–4.37 (2H, m), 4.38–4.59 (1H, m), 7.07–7.43 (9H, m), 7.48–7.64 (2H, m), 7.70–7.82 (2H, m), 7.83–8.06 (6H, m), 8.26 (1H, d, *J* = 1.6 Hz); MS *m/z* 1143 (M+H)⁺.

1-4.

N-(13-(4-((1-(4-Cyano-3-(trifluoromethyl)phenyl)-3,5-dimethyl-1H-pyrazol-4-yl)methy l)phenyl)-13-oxo-3,6,9-trioxa-12-azatridec-1-yl)- N^2 -((2S,3R)-3-(((9H-fluoren-9-ylmetho xy)carbonyl)amino)-2-hydroxy-4-phenylbutanoyl)-L-leucinamide (3c).

Compound **3c** was prepared from **1** and **2c** in a similar manner to that described for compound **3a** and obtained as a colorless amorphous solid (49%); MS m/z 1086.8 (M+H)⁺.

1-5.

N-(13-(4-(2-(1-(3-Chloro-4-cyanophenyl)-3,5-dimethyl-1H-pyrazol-4-yl)ethyl)phenyl)-13-oxo-3,6,9-trioxa-12-azatridec-1-yl)- $N^2-((2S,3R)-3-(((9H-fluoren-9-ylmethoxy)carbonyl))$ nyl)amino)-2-hydroxy-4-phenylbutanoyl)-L-leucinamide (3d). Compound **3d** was prepared from **1** and **2d** in a similar manner to that described for compound **3a** and obtained as a colorless amorphous solid (41%); MS m/z 1066.8 (M+H)⁺.

1-6.

N-(13-(4-((1-(2-Chloro-4-cyanophenyl)-3,5-dimethyl-1H-pyrazol-4-yl)methyl)phenyl)-13-oxo-3,6,9-trioxa-12-azatridec-1-yl)- $N^2-((2S,3R)-3-(((9H-fluoren-9-ylmethoxy)carbonyl))$ nyl)amino)-2-hydroxy-4-phenylbutanoyl)-L-leucinamide (3e).

To a solution of **12** (261 mg, 0.408 mmol) in THF (1 mL) was added 4 M HCl in CPME (2.0 mL, 8.2 mmol). Dioxane (5 mL) was added to the mixture, and the whole mixture was stirred at room temperature for 2 h, then concentrated. The residue was diluted with toluene, and concentrated *in vacuo*, then dissolved in DMF (4 mL). To the solution were added DIPEA (0.356 mL, 2.04 mmol), **13** (260 mg, 0.490 mmol), HOBt (83 mg, 0.61 mmol), and EDC (0.11 mL, 0.61 mmol). The mixture was stirred at room temperature for 4 h. The mixture was poured into sat. aq. NaHCO₃, and extracted with EtOAc. The organic layer was separated, washed with brine, dried over MgSO₄, and concentrated *in vacuo*. The

residue was purified by column chromatography on silica gel (eluted with 0-5% MeOH in EtOAc) and preparative HPLC. The desired fraction was neutralized with sat. aq. NaHCO₃, and extracted with EtOAc. The organic layer was separated, dried over MgSO₄, and concentrated *in vacuo* to give the title compound (315 mg, 73%) as a colorless amorphous solid. MS *m/z* 1052.8 (M+H)⁺.

1-7. Purification conditions of preparative HPLC: Instrument: Waters 2525 separations module; Column: L-column 2 ODS ($20 \times 150 \text{ mm I.D.}, 5 \mu \text{m}$); Mobile phase: A: H₂O/TFA (1000:1, v/v); B: MeCN/TFA (1000:1, v/v); Gradient: B from 60% to 95% in 7 min; Flow

rate: 20 mL/min; Column temperature: room temperature; Wavelength: 220 nm.

1-8.

*N*²-((2*S*,3*R*)-3-Amino-2-hydroxy-4-phenylbutanoyl)-*N*-(13-(4-((3-cyano-4-(4-cyanophe nyl)-2,5-dimethyl-1*H*-pyrrol-1-yl)methyl)phenyl)-13-oxo-3,6,9-trioxa-12-azatridec-1-yl)-L-leucinamide (4a).

To a solution of **3a** (100 mg, 0.0959 mmol) in MeOH (1 mL) was added 2 M Me₂NH in MeOH (4.8 mL, 9.6 mmol) at room temperature. The mixture was stirred at room temperature overnight. The mixture was concentrated and purified by column chromatography on NH silica gel (eluted with 0-15% MeOH in EtOAc) to give the title compound (77 mg, 98%) as a colorless amorphous solid. ¹H NMR (300 MHz, CD₃OD) δ 0.89-1.01 (6H, m), 1.51-1.73 (3H, m), 2.22 (3H, s), 2.38 (3H, s), 2.66 (1H, dd, J = 13.4, 7.8 Hz), 2.90 (1H, dd, J = 13.4, 6.9 Hz), 3.24–3.40 (3H, m), 3.44–3.69 (14H, m), 3.94 (1H, d, J = 3.0 Hz), 4.38–4.48 (1H, m), 5.33 (2H, s), 7.10 (2H, d, J = 8.3 Hz), 7.17–7.34 (5H, m), 7.61 (2H, d, J = 8.5 Hz), 7.76–7.89 (4H, m); MS m/z 820.5 (M+H)⁺; HRMS m/z $(M+H)^+$ Calcd for C₄₆H₅₈N₇O₇: 820.4398, Found: 820.4373; Purity: 100% (HPLC); $[\alpha]_{D}^{25}$ -16.9 (c 0.200, methanol).

1-9.

*N*²-((2*S*,3*R*)-3-Amino-2-hydroxy-4-phenylbutanoyl)-*N*-(13-(4-((3-((4-cyano-3-(trifluoro methyl)phenyl)amino)-2-hydroxy-2-methyl-3-oxopropyl)sulfonyl)phenyl)-13-oxo-3,6,9 -trioxa-12-azatridec-1-yl)-L-leucinamide (4b).

ACS Paragon Plus Environment

Compound 4b was prepared from 3b in a similar manner to that described for compound 4a
and obtained as a colorless amorphous solid (89%). $^1\mathrm{H}$ NMR (300 MHz, CD ₃ OD) δ 0.84–
1.02 (6H, m), 1.49 (3H, s), 1.56–1.74 (3H, m), 2.65 (1H, dd, <i>J</i> = 13.4, 7.8 Hz), 2.89 (1H,
dd, J = 13.4, 6.9 Hz), 3.19–3.40 (3H, m), 3.44–3.72 (15H, m), 3.92 (1H, d, J = 3.0 Hz),
4.08 (1H, d, <i>J</i> = 14.7 Hz), 4.36–4.47 (1H, m), 7.13–7.34 (5H, m), 7.83–8.08 (6H, m), 8.17–
8.37 (1H, m); MS m/z 921.3 (M+H) ⁺ ; HRMS m/z (M+H) ⁺ Calcd for C ₄₃ H ₅₆ F ₃ N ₆ O ₁₁ S:
921.3680, Found: 921.3629; Purity: 98.1% (HPLC); $[\alpha]_{D}^{25}$ -16.7 (<i>c</i> 0.200, methanol).

1-10.

*N*²-((2*S*,3*R*)-3-Amino-2-hydroxy-4-phenylbutanoyl)-*N*-(13-(4-((1-(4-cyano-3-(trifluoro methyl)phenyl)-3,5-dimethyl-1*H*-pyrazol-4-yl)methyl)phenyl)-13-oxo-3,6,9-trioxa-12-a zatridec-1-yl)-L-leucinamide (4c).

Compound **4c** was prepared from **3c** in a similar manner to that described for compound **4a** and obtained as a colorless solid (68%). ¹H NMR (300 MHz, CD₃OD) δ 0.88–0.97 (6H, m), 1.54–1.72 (3H, m), 2.15 (3H, s), 2.38 (3H, s), 2.61–2.72 (1H, m), 2.86–2.95 (1H, m), 3.25–

3.39 (3H, m), 3.42–3.68 (14H, m), 3.85–3.96 (3H, m), 4.37–4.46 (1H, m), 7.15–7.32 (7H, m), 7.76 (2H, d, J = 8.3 Hz), 7.91–7.98 (1H, m), 8.06–8.13 (2H, m); MS *m/z* 864.5 (M+H)⁺; HRMS *m/z* (M+H)⁺ Calcd for C₄₅H₅₇F₃N₇O₇: 864.4272, Found: 864.4228; Purity: 97.6% (HPLC); $[\alpha]_{\rm p}^{25}$ –13.6 (*c* 0.278, methanol).

1-11.

*N*²-((2*S*,3*R*)-3-Amino-2-hydroxy-4-phenylbutanoyl)-*N*-(13-(4-(2-(1-(3-chloro-4-cyanop henyl)-3,5-dimethyl-1*H*-pyrazol-4-yl)ethyl)phenyl)-13-oxo-3,6,9-trioxa-12-azatridec-1-yl)-L-leucinamide (4d).

Compound **4d** was prepared from **3d** in a similar manner to that described for compound **4a** and obtained as a colorless amorphous solid (86%). ¹H NMR (300 MHz, CD₃OD) δ 0.93 (6H, t, *J* = 6.0 Hz), 1.54–1.71 (3H, m), 2.08 (6H, s), 2.60–2.93 (6H, m), 3.26–3.35 (3H, m), 3.43–3.68 (14H, m), 3.93 (1H, d, *J* = 3.0 Hz), 4.37–4.47 (1H, m), 7.14–7.32 (7H, m), 7.54 (1H, d, *J* = 8.5 Hz), 7.69–7.77 (3H, m), 7.88 (1H, dd, *J* = 8.5, 2.0 Hz); MS *m/z* 844.5

 $(M+H)^+$; HRMS *m/z* $(M+H)^+$ Calcd for C₄₅H₅₉ClN₇O₇: 844.4165, Found: 844.4133; Purity: 96.4% (HPLC); $[\alpha]_{D}^{25}$ -13.7 (*c* 0.503, methanol).

1-12.

 N^2 -((2*S*,3*R*)-3-amino-2-hydroxy-4-phenylbutanoyl)-*N*-(13-(4-((1-(2-chloro-4-cyanophe nyl)-3,5-dimethyl-1*H*-pyrazol-4-yl)methyl)phenyl)-13-oxo-3,6,9-trioxa-12-azatridec-1-yl)-L-leucinamide (4e).

A mixture of **3e** (0.34 g, 0.32 mmol) and 2 M Me₂NH in MeOH (4.0 mL, 8.0 mmol) was stirred at room temperature for 3.5 h, then concentrated. The residue was purified by column chromatography on NH silica gel (eluted with 0–5% EtOAc in hexane) to give the title compound (216 mg, 81%) as a colorless amorphous solid. ¹H NMR (300 MHz, CD₃OD) δ 0.93 (6H, t, *J* = 5.9 Hz), 1.56–1.71 (3H, m), 2.05 (3H, s), 2.13 (3H, s), 2.60–2.69 (1H, m), 2.83–2.94 (1H, m), 3.27–3.35 (3H, m), 3.43–3.68 (14H, m), 3.87–3.95 (3H, m), 4.38–4.46 (1H, m), 7.14–7.33 (7H, m), 7.65 (1H, d, *J* = 8.2 Hz), 7.77 (2H, d, *J* = 8.2 Hz), 7.87 (1H, dd, *J* = 8.2, 1.5 Hz), 8.08 (1H, d, *J* = 1.5 Hz); MS *m/z* 830.5 (M+H)⁺; HRMS *m/z*

 $(M+H)^+$ Calcd for C₄₄H₅₇ClN₇O₇: 830.4008, Found: 830.3980; Purity: 99.4% (HPLC); $[\alpha]_D^{25}$ -13.3 (*c* 0.514, methanol).

1-13.

Methyl

4-((3,5-dimethyl-1-((2-(trimethylsilyl)ethoxy)methyl)-1*H*-pyrazol-4-yl)methyl)benzoat e (6).

To a solution of **5** (305 mg, 1.25 mmol) in DMF (12 mL) was added NaH (60% w/w, 54.9 mg, 1.37 mmol) at 0 °C. The mixture was stirred at 0 °C for 30 min. SEMCl (0.26 mL, 1.5 mmol) was added to the mixture, and the whole mixture was stirred at room temperature for 1.5 h. The mixture was poured into iced water and extracted with EtOAc. The organic layer was separated, washed with water and brine, dried over MgSO₄, and concentrated *in vacuo*. The residue was purified by column chromatography on silica gel (eluted with 0–25% EtOAc in hexane) to give the title compound (415 mg, 89%) as a colorless oil. ¹H NMR (300 MHz, CDCl₃) δ –0.03 (9H, s), 0.84–0.92 (2H, m), 2.09 (3H, s), 2.20 (3H, s), 3.52–

3.60 (2H, m), 3.78 (2H, s), 3.90 (3H, s), 5.34 (2H, s), 7.16 (2H, d, J = 8.2 Hz), 7.92 (2H, d, J = 8.2 Hz); MS m/z 375.3 (M+H)⁺.

1-14.

4-((3,5-Dimethyl-1-((2-(trimethylsilyl)ethoxy)methyl)-1*H*-pyrazol-4-yl)methyl)benzoic acid (7).

To a solution of **6** (252 mg, 0.673 mmol) in THF (2 mL) and MeOH (2 mL) was added 2 M aq. NaOH (2.0 mL, 4.0 mmol). The mixture was stirred at room temperature overnight. The mixture was acidified with 2 M aq. HCl at 0 °C, and extracted with EtOAc. The organic layer was separated, dried over MgSO₄, and concentrated in *vacuo* to give the title compound (203 mg, 84%) as a colorless oil. ¹H NMR (300 MHz, DMSO-*d*₆) δ –0.07 (9H, s), 0.78 (2H, t, *J* = 7.9 Hz), 1.98 (3H, s), 2.19 (3H, s), 3.48 (2H, t, *J* = 7.9 Hz), 3.76 (2H, s), 5.28 (2H, s), 7.21 (2H, d, *J* = 8.2 Hz), 7.83 (2H, d, *J* = 8.2 Hz), 12.79 (1H, br s); MS *m/z* 361.3 (M+H)⁺.

1-15.

(13-(4-((3,5-dimethyl-1-((2-(trimethylsilyl)ethoxy)methyl)-1*H*-pyrazol-4-yl)methyl)phe nyl)-13-oxo-3,6,9-trioxa-12-azatridec-1-yl)carbamate (9).

A mixture of 7 (200 mg, 0.555 mmol), 8 (197 mg, 0.674 mmol), EDC (0.15 mL, 0.84 mmol), and HOBt (114 mg, 0.844 mmol) in DMF (6 mL) was stirred at room temperature for 2 days. The mixture was poured into sat. aq. NaHCO₃, and extracted with EtOAc. The organic layer was separated, washed with brine, dried over MgSO₄, and concentrated *in vacuo*. The residue was purified by column chromatography on silica gel (eluted with 30–100% EtOAc in hexane) to give the title compound (345 mg, 97%) as a colorless oil. ¹H NMR (300 MHz, CDCl₃) δ –0.04––0.01 (9H, m), 0.82–0.93 (2H, m), 1.43 (9H, s), 2.09 (3H, s), 2.20 (3H, s), 3.22–3.32 (2H, m), 3.45–3.70 (16H, m), 3.76 (2H, s), 4.97–5.08 (1H, m), 5.34 (2H, s), 6.70 (1H, br s), 7.14 (2H, d, *J* = 8.1 Hz), 7.69 (2H, d, *J* = 8.1 Hz); MS *m/z* 635.6 (M+H)⁺.

1-16.

(13-(4-((3,5-dimethyl-1*H*-pyrazol-4-yl)methyl)phenyl)-13-oxo-3,6,9-trioxa-12-azatridec

To a solution of 9 (345 mg, 0.543 mmol) in MeOH (1 mL) was added 4 M HCl in CPME (1.5 mL, 6.0 mmol) at room temperature. The mixture was refluxed for 1 h. To the mixture was added THF (1 mL), and the whole mixture was refluxed for 7 h, then concentrated. The residue was suspended with THF (4 mL) and neutralized with sat. aq. NaHCO₃ at 0 °C. To the mixture was added Boc₂O (0.14 mL, 0.60 mmol), and the whole mixture was stirred at room temperature overnight. Additional Boc₂O (0.139 mL, 0.60 mmol) was added to the mixture, and the whole mixture was stirred at room temperature for 5 h. The mixture was poured into water and extracted with EtOAc. The organic layer was separated, dried over MgSO₄, and concentrated *in vacuo*. The residue was purified by column chromatography on silica gel (eluted with 0-10% MeOH in EtOAc). The residue was dissolved in MeOH (3.5 mL). To the solution was added sodium methoxide in MeOH (28% w/w, 21 mg, 0.11 mmol). The mixture was stirred at room temperature for 1 h. The mixture was poured into

water and extracted with EtOAc. The organic layer was separated, dried over MgSO₄, and concentrated *in vacuo*. The residue was purified by column chromatography on silica gel (eluted with 0–10% EtOAc in hexane) to give the title compound (213 mg, 77%) as a colorless oil. ¹H NMR (300 MHz, DMSO-*d*₆) δ 1.36 (9H, s), 2.04 (6H, br s), 3.04 (2H, q, *J* = 5.7 Hz), 3.34–3.56 (14H, m), 3.70 (2H, s), 6.74 (1H, t, *J* = 5.5 Hz), 7.17 (2H, d, *J* = 8.1 Hz), 7.73 (2H, d, *J* = 8.1 Hz), 8.39 (1H, t, *J* = 5.3 Hz), 11.99 (1H, br s); MS *m/z* 505.4 (M+H)⁺.

1-17.

tert-Butyl

(13-(4-((1-(2-chloro-4-cyanophenyl)-3,5-dimethyl-1*H*-pyrazol-4-yl)methyl)phenyl)-13oxo-3,6,9-trioxa-12-azatridec-1-yl)carbamate (12).

To a solution of **10** (213 mg, 0.422 mmol) in DMF (4 mL) was added NaH (60% w/w, 20 mg, 0.51 mmol) at 0 °C. The mixture was stirred at 0 °C for 30 min. To the mixture was added **11** (131 mg, 0.842 mmol), and the whole mixture was stirred at room temperature overnight. The mixture was poured into sat. aq. NaHCO₃, and extracted with EtOAc. The

Journal of Medicinal Chemistry

organic layer was separated, washed with brine, dried over MgSO₄, and concentrated *in vacuo*. The residue was purified by column chromatography on silica gel (eluted with 0– 100% EtOAc in hexane) to give the title compound (261 mg, 97%) as a colorless oil. ¹H NMR (300 MHz, CDCl₃) δ 1.43 (9H, s), 2.03 (3H, s), 2.18 (3H, s), 3.22–3.32 (2H, m), 3.48–3.53 (2H, m), 3.59–3.71 (12H, m), 3.84 (2H, s), 5.02 (1H, br s), 6.74 (1H, br s), 7.20 (2H, d, *J* = 8.1 Hz), 7.55 (1H, d, *J* = 8.1 Hz), 7.66–7.78 (3H, m), 7.84 (1H, d, *J* = 1.7 Hz); MS *m/z* 640.5 (M+H)⁺.

1-18.

tert-Butyl

(13-(4-((3-cyano-4-(4-cyanophenyl)-2,5-dimethyl-1*H*-pyrrol-1-yl)methyl)phenyl)-13-ox o-3,6,9-trioxa-12-azatridec-1-yl)carbamate (15).

A mixture of **2a** (110 mg, 0.310 mmol), **14** (110 mg, 0.376 mmol), HOBt (57 mg, 0.42 mmol), and EDC (74 μ l, 0.42 mmol) in THF (3 mL) was stirred at room temperature overnight. The mixture was diluted with EtOAc, washed with 5% aq. NaHCO₃ and brine, dried over Na₂SO₄, and concentrated *in vacuo*. The residue was purified by column

chromatography on silica gel (eluted with 0–20% MeOH in EtOAc) to give the title compound (185 mg, 98%) as a colorless amorphous powder. ¹H NMR (300 MHz, CDCl₃) δ 1.42 (9H, s), 2.19 (3H, s), 2.37 (3H, s), 3.26 (2H, q, *J* = 5.0 Hz), 3.50 (2H, t, *J* = 5.2 Hz), 3.56–3.74 (12H, m), 5.04 (1H, br s), 5.15 (2H, s), 6.92 (1H, br s), 6.99 (2H, d, *J* = 8.0 Hz), 7.48–7.59 (2H, m), 7.66–7.77 (2H, m), 7.83 (2H, d, *J* = 7.4 Hz); MS *m/z* 530.5 (M+H– Boc)⁺.

1-19.

1-((2*S*)-2-((*N*-(*tert*-Butoxycarbonyl)-*N*-methyl-L-alanyl)amino)-2-cyclohexylacetyl)-L-p rolyl-*N*-(13-(4-((3-cyano-4-(4-cyanophenyl)-2,5-dimethyl-1*H*-pyrrol-1-yl)methyl)pheny l)-13-oxo-3,6,9-trioxa-12-azatridec-1-yl)-β-phenyl-L-phenylalaninamide (17).

To a mixture of **15** (185 mg, 0.290 mmol) in THF (0.75 mL) was added 2 M HCl in CPME (4.5 mL, 9.0 mmol). The mixture was stirred at room temperature for 1.5 h. The mixture was diluted with toluene (5 mL), and concentrated *in vacuo* to give a colorless amorphous powder (181 mg). To a mixture of the obtained amorphous powder (164 mg, 0.290 mmol),

16 (192 mg, 0.290 mmol), and DIPEA (0.15 mL, 0.87 mmol) in DMF (4.5 mL) was added
HATU (165 mg, 0.430 mmol) at 0 °C. The mixture was stirred at 0 °C for 1 h. The mixture
was diluted with EtOAc, washed with 5% aq. NaHCO3 and brine, dried over Na2SO4, and
concentrated in vacuo. The residue was purified by column chromatography on silica gel
(eluted with 0–30% MeOH in EtOAc), then on NH silica gel (eluted with 0–20% MeOH in
EtOAc) to give the title compound (322 mg, 95%) as a pale yellow solid. ¹ H NMR (300
MHz, CD ₃ OD) δ 0.95–1.42 (9H, m), 1.42–1.55 (9H, m), 1.56–1.88 (9H, m), 2.17 (3H, s),
2.34 (3H, s), 2.76–3.22 (7H, m), 3.32–3.47 (2H, m), 3.49–3.83 (12H, m), 4.23–4.64 (4H,
m), 5.08–5.21 (1H, m), 5.29 (2H, s), 7.00–7.47 (12H, m), 7.57 (2H, d, <i>J</i> = 8.5 Hz), 7.71–
7.94 (4H, m); MS m/z 1175.0 (M+H) ⁺ .

1-20.

1-((2*S*)-2-Cyclohexyl-2-((*N*-methyl-L-alanyl)amino)acetyl)-L-prolyl-*N*-(13-(4-((3-cyano -4-(4-cyanophenyl)-2,5-dimethyl-1*H*-pyrrol-1-yl)methyl)phenyl)-13-oxo-3,6,9-trioxa-1 2-azatridec-1-yl)-beta-phenyl-L-phenylalaninamide (18).

3	
4	
5	
6	
7	
8	
ă	
10	
10	
11	
12	
13	
14	
15	
16	
17	
18	
19	
20	
21	
22	
23	
24	
25	
20	
20	
21	
28	
29	
30	
31	
32	
33	
34	
35	
36	
37	
38	
39	
40	
11	
41	
42	
43	
44	
45	
46	
47	
48	
49	
50	
51	
52	
53	
54	
55	
56	
57	
51	
50	
59	
60	

A mixture of 17 (308 mg, 0.260 mmol) and 4 M HCl in CPME (3.0 mL, 12 mmol) in THF
(1.5 mL) was stirred at room temperature for 3 h. The mixture was diluted with toluene (5
mL) and concentrated in vacuo. The residue was diluted with sat. aq. NaHCO ₃ , and
extracted with EtOAc-THF (3:1). The organic layer was separated, washed with brine,
dried over Na ₂ SO ₄ , and concentrated <i>in vacuo</i> . The residue was purified by column
chromatography on NH silica gel (eluted with 0-10% MeOH in EtOAc) to give the title
compound (237 mg, 84%) as a colorless solid. $^1\mathrm{H}$ NMR (300 MHz, CD_3OD) δ 0.95–1.48
(9H, m), 1.56–2.06 (9H, m), 2.17 (3H, s), 2.24–2.43 (6H, m), 2.79–3.04 (2H, m), 3.04–3.27
(3H, m), 3.33–3.45 (2H, m), 3.47–3.93 (12H, m), 4.19–4.52 (3H, m), 5.07–5.22 (1H, m),
5.25–5.41 (2H, m), 7.01–7.48 (12H, m), 7.52–7.62 (2H, m), 7.72–7.87 (4H, m); MS m/z
1074.7 $(M+H)^+$; HRMS $m/z (M+H)^+$ Calcd for C ₆₂ H ₇₆ N ₉ O ₈ : 1074.5817, Found: 1074.5761;
Purity: 99.8% (HPLC); $[\alpha]_{D}^{25}$ -13.8 (<i>c</i> 0.202, methanol).

1-21.

tert-Butyl

2-yl)pyrrolidin-1-yl)-1-cyclohexyl-2-oxoethyl)amino)-1-oxopropan-2-yl)methylcarbam ate (21).

To a mixture of Ph₃P (425 mg, 1.62 mmol) in THF (1.5 mL) was added 40% DEAD in toluene (0.71 g, 1.6 mmol) at 0 °C. After the mixture was stirred at 0 °C for 10 min, a solution of 19 (194 mg, 0.320 mmol) and 20 (0.5 M MTBE solution: 3.3 mL, 1.7 mmol) in THF (3 mL) was added to the mixture. After the mixture was stirred at room temperature for 5 h, water (0.4 mL) and Ph₃P (850 mg, 3.24 mmol) were added to the mixture. The mixture was stirred at room temperature overnight. The mixture was diluted with EtOAc, washed with brine, dried over Na₂SO₄, and concentrated in vacuo. The residue was purified by column chromatography on NH silica gel (eluted with 0-20% MeOH in EtOAc) to give the title compound (308 mg, quant.) as a colorless oil. ¹H NMR (300 MHz, CD₃OD) δ 0.97-1.39 (8H, m), 1.41-1.52 (9H, m), 1.52-1.94 (6H, m), 2.07-2.53 (4H, m), 2.67-2.95 (5H, m), 3.45-4.30 (12H, m), 4.42-4.70 (2H, m), 5.38-5.75 (1H, m), 7.24 (1H, dd, J = 7.8, 100)1.9 Hz), 7.44 (1H, t, J = 8.2 Hz), 7.67–7.84 (2H, m), 8.32 (1H, s); MS m/z 730.6 (M+H)⁺.

1-22. tert-Butyl ((2S)-1-(((1S)-2-((2S)-2-(4-(3-(2-(2-((4-((3-cyano-4-(4-cyanophenyl)-2,5-dimethyl-1H -pyrrol-1-yl)methyl)benzoyl)amino)ethoxy)ethoxy)ethoxy)benzoyl)-1,3-thiazol-2-yl)py rrolidin-1-yl)-1-cyclohexyl-2-oxoethyl)amino)-1-oxopropan-2-yl)methylcarbamate (22).

A mixture of 21 (234 mg, 0.321 mmol), 2a (341 mg, 0.960 mmol), HOBt (130 mg, 0.962 mmol), and EDC (0.17 mL, 0.97 mmol) in THF (3 mL) and DMF (1 mL) was stirred at room temperature for 3 h. The mixture was diluted with EtOAc, washed with 5% aq. NaHCO₃ and brine, dried over Na₂SO₄, and concentrated in vacuo. The residue was purified by column chromatography on NH silica gel (eluted with 50-100% EtOAc in hexane then 0-10% MeOH in EtOAc) and by column chromatography on silica gel (eluted with 0-10% MeOH in EtOAc) to give the title compound (125 mg, 37%) as a colorless solid. ¹H NMR (300 MHz, CD₃OD) δ 1.04–1.39 (8H, m), 1.40–1.49 (9H, m), 1.49–1.85 (6H, m), 2.06–2.37 (10H, m), 2.66–2.90 (3H, m), 3.50–4.02 (13H, m), 4.11–4.18 (2H, m), 4.43–4.66 (2H, m), 5.27 (2H, s), 5.41–5.67 (1H, m), 7.03 (2H, d, *J* = 8.3 Hz), 7.17 (1H, dd,

J = 7.9, 2.1 Hz), 7.39 (1H, t, *J* = 8.0 Hz), 7.56 (2H, d, *J* = 8.6 Hz), 7.63–7.86 (6H, m), 8.26– 8.33 (1H, m); MS *m*/*z* 1067.9 (M+H)⁺.

1-23.

4-((3-Cyano-4-(4-cyanophenyl)-2,5-dimethyl-1*H*-pyrrol-1-yl)methyl)-*N*-(2-(2-(2-(3-((2-(3-((2-(3-((2S)-1-((2S)-2-cyclohexyl-2-((*N*-methyl-L-alanyl)amino)acetyl)pyrrolidin-2-yl)-1,3-thi azol-4-yl)carbonyl)phenoxy)ethoxy)ethoxy)ethyl)benzamide (23).

A mixture of **21** (120 mg, 0.112 mmol) in THF (3 mL) and 4 M HCl in CPME (3.0 mL, 12 mmol) was stirred at room temperature for 4 h. The mixture was diluted with toluene (5 mL), and concentrated *in vacuo*. The residue was dissolved in EtOAc–THF(1:1), washed with sat. aq. NaHCO₃ and brine, dried over Na₂SO₄, and concentrated *in vacuo*. The residue was purified by column chromatography on NH silica gel (eluted with 0–10% MeOH in EtOAc) to give the title compound (81 mg, 74%) as a colorless solid. ¹H NMR (300 MHz, CD₃OD) δ 0.95–1.35 (8H, m), 1.52–1.83 (6H, m), 2.07–2.42 (13H, m), 2.86–3.19 (1H, m), 3.50–4.04 (12H, m), 4.13 (2H, t, *J* = 4.5 Hz), 4.47–4.62 (1H, m), 5.26 (2H, s), 5.41–5.70

(1H, m), 7.03 (2H, d, J = 8.4 Hz), 7.16 (1H, dd, J = 8.3, 1.7 Hz), 7.38 (1H, t, J = 8.0 Hz), 7.50–7.86 (8H, m), 8.25–8.32 (1H, m); MS m/z 967.6 (M+H)⁺; HRMS m/z (M+H)⁺ Calcd for C₅₄H₆₃N₈O₇S: 967.4540, Found: 967.4543; Purity: 96.5% (HPLC); $[\alpha]_{D}^{25}$ –71.0 (c 0.202, methanol).

1-24.

N-((1*S*)-1-Cyclohexyl-2-((2*S*)-2-(4-(3-methoxybenzoyl)-1,3-thiazol-2-yl)pyrrolidin-1-yl) -2-oxoethyl)-*N*²-methyl-L-alaninamide (24).

To a mixture of **19** (120 mg, 0.200 mmol), Ph₃P (158 mg, 0.602 mmol), and MeOH (24 μ L, 0.59 mmol) in THF (3 mL) was added 40% DEAD in toluene (0.28 g, 0.64 mmol) at 0 °C. The mixture was stirred at room temperature overnight, and concentrated *in vacuo*. The residue was purified by column chromatography on silica gel (eluted with 50–100% EtOAc in hexane) to give the title compound as a colorless gum. A mixture of the obtained gum and 4 M HCl in CPME (3.0 mL, 12 mmol) was stirred at room temperature for 1 h. The resultant solid precipitate was collected by filtration and washed with EtOAc. The solid was

neutralized with saturated aq. NaHCO3 and extracted with EtOAc. The organic layer was
washed with brine, dried over Na ₂ SO ₄ , and concentrated <i>in vacuo</i> . The residue was purified
by column chromatography on NH silica gel (eluted with 50-100% EtOAc in hexane), and
crystallized from EtOAc-hexane to give the title compound (43 mg, 42%) as colorless
crystals. ¹ H NMR (300 MHz, CD ₃ OD) δ 0.96–1.30 (8H, m), 1.54–1.86 (6H, m), 2.06–2.50
(7H, m), 2.89–3.22 (1H, m), 3.83–4.09 (5H, m), 4.49–4.61 (1H, m), 5.42–5.71 (1H, m),
7.21 (1H, ddd, <i>J</i> = 8.3, 2.6, 0.9 Hz), 7.43 (1H, t, <i>J</i> = 7.8 Hz), 7.65–7.77 (2H, m), 8.27–8.36
(1H, m); MS m/z 513.4 (M+H) ⁺ ; HRMS m/z (M+H) ⁺ Calcd for C ₂₇ H ₃₇ N ₄ O ₄ S: 513.2536,
Found: 513.2508; Purity: 97.8% (HPLC); $[\alpha]_{D}^{25}$ -146.9 (<i>c</i> 0.103, methanol).

1-25.

4-(4-Cyanophenyl)-1-(4-(3-(2-(2-(2-hydroxyethoxy)ethoxy)prop-1-yn-1-yl)benz yl)-2,5-dimethyl-1*H*-pyrrole-3-carbonitrile (27).

A mixture of **25** (585 mg, 1.50 mmol), **26** (565 mg, 3.00 mmol), Et₃N (0.84 mL, 6.0 mmol), copper(I) iodide (29 mg, 0.15 mmol), and Pd(Ph₃P)₄ (173 mg, 0.150 mmol) in DMF (4.5

mL) was stirred at 60 °C under Ar overnight. After cooling, the mixture was diluted with EtOAc, washed with sat. aq. NH₄Cl and brine, dried over Na₂SO₄, and concentrated *in vacuo*. The residue was purified by column chromatography on NH silica gel (eluted with 0–20% MeOH in EtOAc) to give the title compound (365 mg, 49%) as a pale yellow oil. ¹H NMR (300 MHz, CDCl₃) δ 2.18 (3H, s), 2.36 (3H, s), 2.48 (1H, t, *J* = 6.2 Hz), 3.56–3.84 (12H, m), 4.42 (2H, s), 5.10 (2H, s), 6.87 (2H, d, *J* = 8.4 Hz), 7.43 (2H, d, *J* = 8.3 Hz), 7.49–7.58 (2H, m), 7.66–7.76 (2H, m); MS *m/z* 498.4 (M+H)⁺.

1-26.

2-(2-((3-((3-((3-Cyano-4-(4-cyanophenyl)-2,5-dimethyl-1*H*-pyrrol-1-yl)methyl)pheny l)prop-2-yn-1-yl)oxy)ethoxy)ethoxy)ethyl 4-methylbenzenesulfonate (28).

To a mixture of **27** (320 mg, 0.643 mmol) in pyridine (4.5 mL) was added TsCl (0.18 g, 0.96 mmol) at 0 °C. The mixture was stirred at room temperature for 4 h. To the mixture was then added TsCl (0.18 g, 0.96 mmol), and the mixture was stirred at room temperature for 1 h. The mixture was diluted with EtOAc, washed with 1 M HCl, sat. aq. NaHCO₃, and

brine, dried over Na₂SO₄, and concentrated *in vacuo*. The residue was purified by column chromatography on silica gel (eluted with 50–100% EtOAc in hexane) to give the title compound (302 mg, 72%) as a pale yellow oil. ¹H NMR (300 MHz, CDCl₃) δ 2.19 (3H, s), 2.36 (3H, s), 2.44 (3H, s), 3.61 (4H, s), 3.64–3.78 (6H, m), 4.12–4.19 (2H, m), 4.41 (2H, s), 5.10 (2H, s), 6.88 (2H, d, *J* = 8.3 Hz), 7.34 (2H, d, *J* = 8.1 Hz), 7.43 (2H, d, *J* = 8.2 Hz), 7.53 (2H, d, *J* = 8.4 Hz), 7.71 (2H, d, *J* = 8.4 Hz), 7.79 (2H, d, *J* = 8.3 Hz); MS *m/z* 652.5 (M+H)⁺.

1-27.

4-(4-Cyanophenyl)-1-(4-(3-(2-(2-(2-hydroxyethoxy)ethoxy)propyl)benzyl)-2,5-d imethyl-1*H*-pyrrole-3-carbonitrile (29).

A mixture of **27** (365 mg, 0.730 mmol) and 10% Pd–C (73 mg) in EtOAc (3 mL) was stirred under a hydrogen atmosphere at room temperature for 2 h. The mixture was filtered, and the filtrate was concentrated *in vacuo*. The residue was purified by column chromatography on silica gel (eluted with EtOAc) to give the title compound (310 mg,

84%) as a pale yellow oil. ¹H NMR (300 MHz, CDCl₃) δ 1.81–1.96 (2H, m), 2.20 (3H, s),
2.37 (3H, s), 2.52–2.73 (3H, m), 3.47 (2H, t, J = 6.4 Hz), 3.55–3.77 (12H, m), 5.07 (2H, s),
6.84 (2H, d, J = 8.2 Hz), 7.18 (2H, d, J = 8.1 Hz), 7.49–7.58 (2H, m), 7.67–7.75 (2H, m);
MS *m/z* 502.4 (M+H)⁺.

1-28.

2-(2-(3-(4-((3-Cyano-4-(4-cyanophenyl)-2,5-dimethyl-1*H*-pyrrol-1-yl)methyl)phenyl)propoxy)ethoxy)ethoxy)ethyl 4-methylbenzenesulfonate (30).

To a mixture of **29** (310 mg, 0.618 mmol) in pyridine (4.5 mL) was added TsCl (353 mg, 1.85 mmol) at 0 °C. The mixture was stirred at room temperature for 3 h. The mixture was diluted with EtOAc, washed with 1 M HCl, sat. aq. NaHCO₃ and brine, dried over Na₂SO₄, and concentrated *in vacuo*. The residue was purified by column chromatography on silica gel (eluted with 40–90% EtOAc in hexane) to give the title compound (338 mg, 83%) as a colorless oil. ¹H NMR (300 MHz, CDCl₃) δ 1.81–1.96 (2H, m), 2.20 (3H, s), 2.37 (3H, s), 2.44 (3H, s), 2.61–2.74 (2H, m), 3.46 (2H, t, *J* = 6.4 Hz), 3.53–3.73 (10H, m), 4.12–4.19

(2H, m), 5.07 (2H, s), 6.84 (2H, d, *J* = 8.1 Hz), 7.17 (2H, d, *J* = 8.1 Hz), 7.33 (2H, d, *J* = 8.0 Hz), 7.49–7.57 (2H, m), 7.65–7.74 (2H, m), 7.75–7.84 (2H, m); MS *m*/*z* 656.5 (M+H)⁺.

1-29.

tert-Butyl

((2*S*)-1-(((1*S*)-2-((2*S*)-2-(4-(3-(2-(2-((3-((4-((3-cyano-4-(4-cyanophenyl)-2,5-dimethyl-1*H*-pyrrol-1-yl)methyl)phenyl)prop-2-yn-1-yl)oxy)ethoxy)ethoxy)ethoxy)benzoyl)-1,3thiazol-2-yl)pyrrolidin-1-yl)-1-cyclohexyl-2-oxoethyl)amino)-1-oxopropan-2-yl)methyl carbamate (31a).

A mixture of **28** (302 mg, 0.460 mmol), **19** (277 mg, 0.460 mmol), and K₂CO₃ (77 mg, 0.56 mmol) in DMF (2 mL) was stirred at room temperature overnight, and then heated at 50 °C for 24 h. After cooling, the mixture was diluted with EtOAc, washed with 5% aq. NaHCO₃ and brine, dried over Na₂SO₄, and concentrated *in vacuo*. The residue was purified by column chromatography on silica gel (eluted with 50–100% EtOAc in hexane) and then NH silica gel (eluted with 50–100% EtOAc in hexane) to give the title compound (334 mg, 67%) as a colorless amorphous solid. ¹H NMR (300 MHz, CD₃OD) δ 0.94–1.38 (8H, m),

1.40–1.49 (9H, m), 1.50–1.83 (6H, m), 2.04–2.42 (10H, m), 2.63–2.90 (3H, m), 3.62–3.77 (8H, m), 3.80–4.03 (4H, m), 4.14–4.21 (2H, m), 4.38 (2H, s), 4.45–4.68 (2H, m), 5.23 (2H, s), 5.41–5.67 (1H, m), 6.94 (2H, d, *J* = 8.3 Hz), 7.20 (1H, dd, *J* = 8.4, 1.7 Hz), 7.34–7.45 (3H, m), 7.57 (2H, d, *J* = 8.5 Hz), 7.65–7.73 (2H, m), 7.77 (2H, d, *J* = 8.5 Hz), 8.26–8.33 (1H, m); MS *m*/*z* 1078.8 (M+H)⁺.

1-30.

tert-Butyl

((2*S*)-1-(((1*S*)-2-((2*S*)-2-(4-(3-(2-(2-(2-(2-(3-(4-((3-cyano-4-(4-cyanophenyl)-2,5-dimethyl-1 *H*-pyrrol-1-yl)methyl)phenyl)propoxy)ethoxy)ethoxy)ethoxy)benzoyl)-1,3-thiazol-2-yl) pyrrolidin-1-yl)-1-cyclohexyl-2-oxoethyl)amino)-1-oxopropan-2-yl)methylcarbamate (31b).

Compound **31b** was prepared from **19** and **30** in a similar manner to that described for compound **31a** and obtained as a colorless amorphous powder (72%). ¹H NMR (300 MHz, CD₃OD) δ 0.90–1.37 (8H, m), 1.39–1.50 (9H, m), 1.51–1.88 (8H, m), 2.06–2.43 (10H, m), 2.64 (2H, t, *J* = 7.6 Hz), 2.68–2.90 (3H, m), 3.37–3.47 (2H, m), 3.50–3.75 (8H, m), 3.79–

4.02 (4H, m), 4.17 (2H, dd, *J* = 5.4, 3.9 Hz), 4.44–4.69 (2H, m), 5.17 (2H, s), 5.42–5.66 (1H, m), 6.87 (2H, d, *J* = 8.2 Hz), 7.11–7.23 (3H, m), 7.39 (1H, t, *J* = 8.1 Hz), 7.52–7.61 (2H, m), 7.66–7.80 (4H, m), 8.23–8.33 (1H, m); MS *m/z* 1082.9 (M+H)⁺.

1-31.

N-((1S)-2-((2S)-2-(4-(3-(2-(2-(2-((3-(4-((3-Cyano-4-(4-cyanophenyl)-2,5-dimethyl-1H-p yrrol-1-yl)methyl)phenyl)prop-2-yn-1-yl)oxy)ethoxy)ethoxy)ethoxy)benzoyl)-1,3-thiaz ol-2-yl)pyrrolidin-1-yl)-1-cyclohexyl-2-oxoethyl)-N²-methyl-L-alaninamide (32a).

A mixture of **31a** (334 mg, 0.310 mmol) in TFA (3 mL) was stirred at room temperature for 10 min. The mixture was diluted with toluene (5 mL), and concentrated *in vacuo*. The residue was dissolved in EtOAc, washed with sat. aq. NaHCO₃ and brine, dried over Na₂SO₄, and concentrated *in vacuo*. The residue was purified by column chromatography on NH silica gel (eluted with 0–10% MeOH in EtOAc) to give the title compound (237 mg, 78%) as a colorless solid. ¹H NMR (300 MHz, CD₃OD) δ 0.96–1.35 (8H, m), 1.49–1.84 (6H, m), 2.04–2.45 (13H, m), 2.87–3.20 (1H, m), 3.63–3.76 (8H, m), 3.83–4.04 (4H, m),

4.14–4.21 (2H, m), 4.38 (2H, s), 4.49–4.62 (1H, m), 5.23 (2H, s), 5.43–5.67 (1H, m), 6.94 (2H, d, J = 8.2 Hz), 7.16–7.24 (1H, m), 7.34–7.45 (3H, m), 7.53–7.61 (2H, m), 7.65–7.73 (2H, m), 7.77 (2H, d, J = 8.5 Hz), 8.25–8.31 (1H, m); MS *m/z* 978.4 (M+H)⁺; HRMS *m/z* (M+H)⁺ Calcd for C₅₆H₆₄N₇O₇S: 978.4588, Found: 978.4528; Purity: 99.2% (HPLC); $[\alpha]_{D}^{25}$ –77.6 (*c* 0.201, methanol).

1-32.

N-((1*S*)-2-((2*S*)-2-(4-(3-(2-(2-(2-(2-(3-(4-((3-Cyano-4-(4-cyanophenyl)-2,5-dimethyl-1*H*-py rrol-1-yl)methyl)phenyl)propoxy)ethoxy)ethoxy)ethoxy)benzoyl)-1,3-thiazol-2-yl)pyrr olidin-1-yl)-1-cyclohexyl-2-oxoethyl)- N^2 -methyl-L-alaninamide (32b).

Compound **32b** was prepared from **31b** in a similar manner to that described for compound **32b** and obtained as a colorless solid (86%). ¹H NMR (300 MHz, CD₃OD) δ 0.96–1.39 (8H, m), 1.50–1.93 (8H, m), 2.02–2.42 (13H, m), 2.64 (2H, t, *J* = 7.6 Hz), 3.14 (1H, q, *J* = 6.9 Hz), 3.42 (2H, t, *J* = 6.3 Hz), 3.50–3.76 (8H, m), 3.81–4.03 (4H, m), 4.17 (2H, dd, *J* = 5.5, 3.9 Hz), 4.47–4.64 (1H, m), 5.17 (2H, s), 5.42–5.68 (1H, m), 6.87 (2H, d, *J* = 8.2 Hz),

7.10–7.23 (3H, m), 7.33–7.44 (1H, m), 7.52–7.60 (2H, m), 7.66–7.81 (4H, m), 8.24–8.31 (1H, m); MS m/z 982.4 (M+H)⁺; HRMS m/z (M+H)⁺ Calcd for C₅₆H₆₈N₇O₇S: 982.4901, Found: 982.4825; Purity: 99.8% (HPLC); $[\alpha]_{D}^{25}$ –77.6 (*c* 0.200, methanol).

1-33.

4-(4-Cyanophenyl)-1-(4-(2-(2-(2-(2-(2-hydroxyethoxy)ethoxy)ethoxy)ethoxy)benzyl)-2,5-d imethyl-1*H*-pyrrole-3-carbonitrile (35a).

To a mixture of **33a** (392 mg, 1.20 mmol), **34** (1.28 mL, 7.41 mmol), and Ph₃P (960 mg, 3.66 mmol) in THF (12 mL) was dropwise added 40% DEAD in toluene (3.20 g, 7.35 mmol) at 0 °C. The mixture was stirred at room temperature overnight. The mixture was diluted with EtOAc, washed with water and brine, dried over Na₂SO₄, and concentrated *in vacuo*. The residue was purified by column chromatography on silica gel (eluted with 0– 20% MeOH in EtOAc) to give the title compound (440 mg, 73%) as a colorless oil. ¹H NMR (300 MHz, CDCl₃) δ 2.20 (3H, s), 2.37 (3H, s), 2.64 (1H, br s), 3.56–3.78 (12H, m),

3.81–3.90 (2H, m), 4.06–4.24 (2H, m), 5.04 (2H, s), 6.79–6.95 (4H, m), 7.48–7.58 (2H, m), 7.65–7.76 (2H, m); MS *m/z* 504.4 (M+H)⁺.

1-34.

4-(4-Cyanophenyl)-1-(3-(2-(2-(2-(2-hydroxyethoxy)ethoxy)ethoxy)ethoxy)benzyl)-2,5-d imethyl-1*H*-pyrrole-3-carbonitrile (35b).

Compound **35b** was prepared from **33b** in a similar manner to that described for compound **35a** and obtained as a colorless oil (94%). ¹H NMR (300 MHz, CDCl₃) δ 2.20 (3H, s), 2.37 (3H, s), 2.63 (1H, br s), 3.52–3.79 (12H, m), 3.80–3.90 (2H, m), 4.05–4.13 (2H, m), 5.07 (2H, s), 6.44–6.57 (2H, m), 6.84 (1H, dd, *J* = 8.1, 1.7 Hz), 7.21–7.31 (1H, m), 7.53 (2H, d, *J* = 8.3 Hz), 7.71 (2H, d, *J* = 8.3 Hz); MS *m/z* 504.4 (M+H)⁺.

1-35.

2-(2-(2-(2-(4-((3-Cyano-4-(4-cyanophenyl)-2,5-dimethyl-1*H*-pyrrol-1-yl)methyl)pheno xy)ethoxy)ethoxy)ethyl 4-methylbenzenesulfonate (36a).

To a mixture of 35a (220 mg, 0.437 mmol) in pyridine (3 mL) was added TsCl (125 mg,
0.656 mmol) at 0 °C. The mixture was stirred at room temperature for 3 h. The mixture was
diluted with EtOAc, washed with 1 M HCl, sat. aq. NaHCO ₃ , and brine, dried over Na ₂ SO ₄ ,
and concentrated in vacuo. The residue was purified by column chromatography on silica
gel (eluted with 50–100% EtOAc in hexane) to give the title compound (216 mg, 75%) as a
colorless oil. ¹ H NMR (300 MHz, CDCl ₃) & 2.20 (3H, s), 2.37 (3H, s), 2.44 (3H, s), 3.54-
3.76 (10H, m), 3.81–3.89 (2H, m), 4.07–4.22 (4H, m), 5.04 (2H, s), 6.79–6.95 (4H, m),
7.33 (2H, d, <i>J</i> = 8.0 Hz), 7.52 (2H, d, <i>J</i> = 8.5 Hz), 7.70 (2H, d, <i>J</i> = 8.4 Hz), 7.79 (2H, d, <i>J</i> =
8.3 Hz); MS m/z 658.5 (M+H) ⁺ .

1-36.

2-(2-(2-(3-((3-Cyano-4-(4-cyanophenyl)-2,5-dimethyl-1*H*-pyrrol-1-yl)methyl)pheno xy)ethoxy)ethoxy)ethoxy)ethyl 4-methylbenzenesulfonate (36b).

Compound **36b** was prepared from **35b** in a similar manner to that described for compound **36a** and obtained as a colorless oil (68%). ¹H NMR (300 MHz, CDCl₃) δ 2.19 (3H, s), 2.37
(3H, s), 2.44 (3H, s), 3.59 (4H, s), 3.62–3.75 (6H, m), 3.80–3.90 (2H, m), 4.06–4.19 (4H, m), 5.06 (2H, s), 6.45–6.56 (2H, m), 6.78–6.88 (1H, m), 7.21–7.30 (1H, m), 7.33 (2H, d, *J* = 8.1 Hz), 7.53 (2H, d, *J* = 8.4 Hz), 7.65–7.74 (2H, m), 7.79 (2H, d, *J* = 8.3 Hz); MS *m/z* 658.5 (M+H)⁺.

1-37.

2-(2-(4-((3-Cyano-4-(4-cyanophenyl)-2,5-dimethyl-1*H*-pyrrol-1-yl)methyl)phenoxy) ethoxy)ethoxy)ethyl 4-methylbenzenesulfonate (36c).

A mixture of **70a** (197 mg, 0.602 mmol), **74** (0.33 g, 0.73 mmol), and K₂CO₃ (0.13 g, 0.92 mmol) in DMF (5 mL) was stirred at 50 °C for 10 h. The mixture was poured into water and extracted with EtOAc. The organic layer was separated, washed with water and brine, dried over MgSO₄, and concentrated *in vacuo*. The residue was purified by column chromatography on silica gel (eluted with 0–75% EtOAc in hexane) to give the title compound (190 mg, 51%) as a colorless gum. ¹H NMR (300 MHz, CDCl₃) δ 2.17–2.22 (3H, m), 2.35–2.39 (3H, m), 2.41–2.46 (3H, m), 3.58–3.73 (6H, m), 3.83 (2H, dd, *J* = 5.4,

4.1 Hz), 4.06–4.18 (4H, m), 5.04 (2H, s), 6.82–6.94 (4H, m), 7.33 (2H, d, *J* = 7.9 Hz), 7.49–7.56 (2H, m), 7.66–7.73 (2H, m), 7.76–7.82 (2H, m); MS *m/z* 614.4 (M+H)⁺.

1-38.

14-(4-((3-Cyano-4-(4-cyanophenyl)-2,5-dimethyl-1*H*-pyrrol-1-yl)methyl)phenoxy)-3,6, 9,12-tetraoxatetradec-1-yl 4-methylbenzenesulfonate (36d).

Compound **36d** was prepared from **33a** and **37b** in a similar manner to that described for compound **36c** and obtained as a colorless amorphous powder (46%). ¹H NMR (300 MHz, CDCl₃) δ 2.20 (3H, s), 2.37 (3H, s), 2.44 (3H, s), 3.55–3.75 (14H, m), 3.81–3.88 (2H, m), 4.07–4.18 (4H, m), 5.04 (2H, s), 6.81–6.92 (4H, m), 7.33 (2H, d, *J* = 8.0 Hz), 7.49–7.56 (2H, m), 7.67–7.73 (2H, m), 7.79 (2H, d, *J* = 8.3 Hz); MS *m/z* 702.4 (M+H)⁺.

1-39.

2-(2-(2-(2-((3-Cyano-4-(4-cyanophenyl)-2,5-dimethyl-1*H*-pyrrol-1-yl)methyl)pheno xy)ethoxy)ethoxy)ethoxy)ethyl 4-methylbenzenesulfonate (36e).

To a mixture of **39** (1.02 g, 2.24 mmol) and Et_3N (0.63 mL, 4.5 mmol) in THF (6 mL) was added MsCl (0.21 mL, 2.7 mmol) at 0 °C. The mixture was stirred at 0 °C for 30 min. The mixture was diluted with EtOAc, washed with water and brine, dried over Na₂SO₄, and concentrated *in vacuo* to give a colorless oil. A solution of the obtained oil in DMF (3 mL) was added to a mixture of 40 (332 mg, 1.50 mmol) and NaH (60% w/w, 72 mg, 1.8 mmol) in DMF (3 mL) at 0 °C. The mixture was stirred at 0 °C for 1.5 h and was poured into 5% aq. NaHCO₃.The mixture was extracted with EtOAc, washed with 5% aq. NaHCO₃ and brine, dried over Na₂SO₄, and concentrated *in vacuo*. The residue was purified by column chromatography on silica gel (eluted with 40-90% EtOAc in hexane) to give the title compound (0.82 g, 83%) as a pale yellow oil. ¹H NMR (300 MHz, CDCl₃) δ 2.17 (3H, s), 2.33 (3H, s), 2.44 (3H, s), 3.53-3.78 (10H, m), 3.85-3.96 (2H, m), 4.10-4.29 (4H, m), 5.10 (2H, s), 6.34 (1H, dd, J = 7.4, 0.8 Hz), 6.83-6.96 (2H, m), 7.22-7.30 (1H, m), 7.33 (2H, d, J)J = 8.1 Hz), 7.54 (2H, d, J = 8.3 Hz), 7.66–7.74 (2H, m), 7.78 (2H, d, J = 8.3 Hz); MS m/z $658.5 (M+H)^+$.

1-40. 2-(2-(2-(2-(Hydroxymethyl)phenoxy)ethoxy)ethoxy)ethoxy)ethyl 4-methylbenzenesulfonate (39).

A mixture of **38** (620 mg, 4.99 mmol), **37c** (5.03 g, 10.0 mmol), and K₂CO₃ (0.83 g, 6.0 mmol) in DMF (10 mL) was stirred at 40 °C overnight. After cooling, the mixture was diluted with EtOAc, washed with 5% aq. NaHCO₃ and brine, dried over Na₂SO₄, and concentrated *in vacuo*. The residue was purified by column chromatography on silica gel (eluted with 50–100% EtOAc in hexane) to give the title compound (1.05 g, 46%) as a colorless oil. ¹H NMR (300 MHz, CDCl₃) δ 2.43 (3H, s), 3.19 (1H, t, *J* = 6.8 Hz), 3.57 (4H, s), 3.60–3.74 (6H, m), 3.85 (2H, dd, *J* = 5.5, 3.7 Hz), 4.14 (2H, dd, *J* = 5.4, 4.2 Hz), 4.20 (2H, dd, *J* = 5.4, 3.7 Hz), 4.65 (2H, d, *J* = 6.4 Hz), 6.84–6.98 (2H, m), 7.19–7.28 (2H, m), 7.33 (2H, d, *J* = 8.1 Hz), 7.79 (2H, d, *J* = 8.3 Hz).

1-41.

tert-Butyl

((2S)-1-(((1S)-2-((2S)-2-(4-(3-(2-(2-(2-(2-(4-((3-cyano-4-(4-cyanophenyl)-2,5-dimethyl-1

H-pyrrol-1-yl)methyl)phenoxy)ethoxy)ethoxy)ethoxy)benzoyl)-1,3-thiazol-2-yl)

ACS Paragon Plus Environment

pyrrolidin-1-yl)-1-cyclohexyl-2-oxoethyl)amino)-1-oxopropan-2-yl)methylcarbamate (41a).

A mixture of 36a (210 mg, 0.319 mmol), 19 (190 mg, 0.317 mmol), and K₂CO₃ (49 mg, 0.35 mmol) in DMF (1.5 mL) was stirred at room temperature overnight, and then heated at 60 °C for 4 h. After cooling, the mixture was diluted with EtOAc, washed with 5% aq. NaHCO₃ and brine, dried over Na₂SO₄, and concentrated in vacuo. The residue was purified by column chromatography on silica gel (eluted with 50–100% EtOAc in hexane, then 0-10% MeOH in EtOAc), and then on NH silica gel (eluted with 50-100% EtOAc in hexane) to give the title compound (231 mg, 67%) as a colorless amorphous powder. ¹H NMR (300 MHz, CD₃OD) δ 0.94–1.37 (8H, m), 1.38–1.49 (9H, m), 1.49–1.88 (6H, m), 2.02-2.44 (10H, m), 2.65-2.91 (3H, m), 3.50-4.26 (18H, m), 4.43-4.69 (2H, m), 5.13 (2H, s), 5.41–5.67 (1H, m), 6.85–6.93 (4H, m), 7.15–7.24 (1H, m), 7.40 (1H, t, *J* = 8.2 Hz), 7.56 (2H, d, J = 8.5 Hz), 7.66–7.73 (2H, m), 7.76 (2H, d, J = 8.5 Hz), 8.26–8.31 (1H, m); MS m/z 1084.9 (M+H)⁺.

 1-42.
 tert-Butyl

 ((2S)-1-(((1S)-2-((2S)-2-(4-(3-(2-(2-(2-(2-(2-(3-((3-cyano-4-(4-cyanophenyl)-2,5-dimethyl-1

 H-pyrrol-1-yl)methyl)phenoxy)ethoxy)ethoxy)ethoxy)benzoyl)-1,3-thiazol-2-yl)

 pyrrolidin-1-yl)-1-cyclohexyl-2-oxoethyl)amino)-1-oxopropan-2-yl)methylcarbamate

 (41b).

Compound **41b** was prepared from **36b** and **19** in a similar manner to that described for compound **41a** as a colorless amorphous powder (81%). ¹H NMR (300 MHz, CD₃OD) δ 0.94–1.39 (8H, m), 1.40–1.49 (9H, m), 1.52–1.85 (6H, m), 2.03–2.41 (10H, m), 2.66–2.91 (3H, m), 3.56–4.21 (18H, m), 4.42–4.68 (2H, m), 5.17 (2H, s), 5.41–5.66 (1H, m), 6.46– 6.58 (2H, m), 6.83 (1H, dd, *J* = 8.2, 2.3 Hz), 7.13–7.29 (2H, m), 7.39 (1H, t, *J* = 8.2 Hz), 7.56 (2H, d, *J* = 8.4 Hz), 7.65–7.72 (2H, m), 7.76 (2H, d, *J* = 8.4 Hz), 8.23–8.33 (1H, m); MS *m/z* 1084.9 (M+H)⁺.

1-43.

tert-Butyl

((2S)-1-(((1S)-2-((2S)-2-(4-(3-(2-(2-(2-(4-((3-cyano-4-(4-cyanophenyl)-2,5-dimethyl-1H-

ACS Paragon Plus Environment

pyrrol-1-yl)methyl)phenoxy)ethoxy)ethoxy)benzoyl)-1,3-thiazol-2-yl)pyrrolidin -1-yl)-1-cyclohexyl-2-oxoethyl)amino)-1-oxopropan-2-yl)methylcarbamate (41c).

Compound **41c** was prepared from **36c** and **19** in a similar manner to that described for compound **41a** and obtained as a colorless amorphous powder (75%). ¹H NMR (300 MHz, CD₃OD) δ 0.94–1.37 (8H, m), 1.39–1.49 (9H, m), 1.51–1.84 (6H, m), 2.03–2.38 (10H, m), 2.65–2.90 (3H, m), 3.66–4.24 (14H, m), 4.43–4.71 (2H, m), 5.13 (2H, s), 5.40–5.65 (1H, m), 6.85–6.93 (4H, m), 7.19 (1H, dd, *J* = 8.3, 1.7 Hz), 7.39 (1H, t, *J* = 7.9 Hz), 7.56 (2H, d, *J* = 8.4 Hz), 7.66–7.73 (2H, m), 7.76 (2H, d, *J* = 8.4 Hz), 8.26–8.33 (1H, m); MS *m/z* 1040.8 (M+H)⁺.

1-44.

tert-Butyl

((2*S*)-1-(((1*S*)-2-((2*S*)-2-(4-(3-((14-(4-((3-cyano-4-(4-cyanophenyl)-2,5-dimethyl-1*H*-pyr rol-1-yl)methyl)phenoxy)-3,6,9,12-tetraoxatetradec-1-yl)oxy)benzoyl)-1,3-thiazol-2-yl) pyrrolidin-1-yl)-1-cyclohexyl-2-oxoethyl)amino)-1-oxopropan-2-yl)methylcarbamate (41d).

Compound **41d** was prepared from **36d** and **19** in a similar manner to that described for compound **41a** and obtained as a colorless amorphous powder (67%). ¹H NMR (300 MHz, CD₃OD) δ 0.95–1.20 (5H, m), 1.27–1.49 (12H, m), 1.52–1.83 (6H, m), 2.07–2.42 (10H, m), 2.67–2.89 (3H, m), 3.58–3.71 (12H, m), 3.75–4.01 (6H, m), 4.04–4.10 (2H, m), 4.13– 4.21 (2H, m), 4.45–4.66 (2H, m), 5.14 (2H, s), 5.42–5.65 (1H, m), 6.85–6.96 (4H, m), 7.20 (1H, dt, *J* = 8.2, 1.3 Hz), 7.35–7.45 (1H, m), 7.52–7.60 (2H, m), 7.66–7.80 (4H, m), 8.26– 8.31 (1H, m); MS *m/z* 1128.7 (M+H)⁺.

1-45.

tert-Butyl

((2*S*)-1-(((1*S*)-2-((2*S*)-2-(4-(3-(2-(2-(2-(2-(2-((3-cyano-4-(4-cyanophenyl)-2,5-dimethyl-1 *H*-pyrrol-1-yl)methyl)phenoxy)ethoxy)ethoxy)ethoxy)benzoyl)-1,3-thiazol-2-yl) pyrrolidin-1-yl)-1-cyclohexyl-2-oxoethyl)amino)-1-oxopropan-2-yl)methylcarbamate (41e).

Compound **41e** was prepared from **36e** and **19** in a similar manner to that described for compound **41a** and obtained as a colorless amorphous powder (76%). ¹H NMR (300 MHz,

> CD₃OD) δ 0.94–1.38 (8H, m), 1.40–1.49 (9H, m), 1.51–1.84 (6H, m), 2.04–2.43 (10H, m), 2.65–2.90 (3H, m), 3.60–3.74 (8H, m), 3.78–4.01 (6H, m), 4.05–4.24 (5H, m), 4.42–4.68 (2H, m), 5.17 (2H, s), 5.42–5.66 (1H, m), 6.42 (1H, dd, *J* = 7.5, 0.9 Hz), 6.87 (1H, t, *J* = 7.4 Hz), 7.00 (1H, d, *J* = 8.2 Hz), 7.17 (1H, dd, *J* = 7.8, 2.1 Hz), 7.20–7.30 (1H, m), 7.38 (1H, t, *J* = 7.9 Hz), 7.50–7.59 (2H, m), 7.64–7.79 (4H, m); MS *m/z* 1084.9 (M+H)⁺.

1-46.

N-((1*S*)-2-((2*S*)-2-(4-(3-(2-(2-(2-(2-(2-(4-((3-Cyano-4-(4-cyanophenyl)-2,5-dimethyl-1*H*-py rrol-1-yl)methyl)phenoxy)ethoxy)ethoxy)ethoxy)ethoxy)benzoyl)-1,3-thiazol-2-yl)pyrr olidin-1-yl)-1-cyclohexyl-2-oxoethyl)- N^2 -methyl-L-alaninamide (42a).

A mixture of **41a** (225 mg, 0.208 mmol) in TFA (3 mL) was stirred at room temperature for 10 min. The mixture was diluted with toluene (5 mL), and concentrated *in vacuo*. The residue was dissolved in EtOAc washed with sat. aq. NaHCO₃ and brine, dried over Na₂SO₄, and concentrated *in vacuo*. The residue was purified by column chromatography on NH silica gel (eluted with 0–10% MeOH in EtOAc) to give the title compound (143 mg,

70%) as a colorless solid. ¹ H NMR (300 MHz, CD ₃ OD) δ 0.98–1.38 (8H, m), 1.49–1.84
(6H, m), 2.03–2.42 (13H, m), 3.14 (1H, q, <i>J</i> = 6.8 Hz), 3.59–3.73 (8H, m), 3.75–4.02 (6H,
m), 4.02–4.08 (2H, m), 4.15 (2H, dd, J = 5.4, 3.8 Hz), 4.45–4.61 (1H, m), 5.12 (2H, s),
5.41–5.67 (1H, m), 6.83–6.92 (4H, m), 7.19 (1H, ddd, <i>J</i> = 8.2, 2.5, 0.9 Hz), 7.34–7.44 (1H,
m), 7.51–7.59 (2H, m), 7.65–7.72 (2H, m), 7.73–7.82 (2H, m), 8.25–8.30 (1H, m); MS <i>m/z</i>
984.6 $(M+H)^+$; HRMS $m/z (M+H)^+$ Calcd for C ₅₅ H ₆₆ N ₇ O ₈ S: 984.4694, Found: 984.4657;
Purity: 98.3% (HPLC); $[\alpha]_{D}^{25}$ -77.5 (<i>c</i> 0.115, methanol).

1-47.

N-((1S)-2-((2S)-2-(4-(3-(2-(2-(2-(2-(2-(3-((3-Cyano-4-(4-cyanophenyl)-2,5-dimethyl-1H-py rrol-1-yl)methyl)phenoxy)ethoxy)ethoxy)ethoxy)ethoxy)benzoyl)-1,3-thiazol-2-yl)pyrr olidin-1-yl)-1-cyclohexyl-2-oxoethyl)-N²-methyl-L-alaninamide (42b).

Compound **42b** was prepared from **41b** in a similar manner to that described for compound **42a** and obtained as a colorless amorphous powder (85%). ¹H NMR (300 MHz, CD₃OD) δ 0.96–1.40 (8H, m), 1.50–1.86 (6H, m), 2.03–2.43 (13H, m), 2.88–3.21 (1H, m), 3.57–4.21

(18H, m), 4.46–4.64 (1H, m), 5.17 (2H, s), 5.41–5.67 (1H, m), 6.46–6.58 (2H, m), 6.83 (1H, dd, J = 8.1, 2.1 Hz), 7.14–7.31 (2H, m), 7.34–7.45 (1H, m), 7.56 (2H, d, J = 8.5 Hz), 7.65–7.73 (2H, m), 7.76 (2H, d, J = 8.5 Hz), 8.23–8.35 (1H, m); MS m/z 984.4 (M+H)⁺; HRMS m/z (M+H)⁺ Calcd for C₅₅H₆₆N₇O₈S: 984.4694, Found: 984.4625; Purity: 100% (HPLC); $[\alpha]_{D}^{25}$ –75.3 (c 0.202, methanol).

1-48.

N-((1*S*)-2-((2*S*)-2-(4-(3-(2-(2-(2-(4-((3-Cyano-4-(4-cyanophenyl)-2,5-dimethyl-1*H*-pyrr ol-1-yl)methyl)phenoxy)ethoxy)ethoxy)ethoxy)benzoyl)-1,3-thiazol-2-yl)pyrrolidin-1-yl)-1-cyclohexyl-2-oxoethyl)-*N*²-methyl-L-alaninamide (42c).

Compound **42c** was prepared from **41c** in a similar manner to that described for compound **42a** and obtained as a colorless amorphous powder (85%). ¹H NMR (300 MHz, CD₃OD) δ 0.97–1.39 (8H, m), 1.46–1.86 (6H, m), 2.02–2.41 (13H, m), 3.14 (1H, q, *J* = 6.8 Hz), 3.71 (4H, s), 3.76–4.22 (10H, m), 4.47–4.61 (1H, m), 5.12 (2H, s), 5.41–5.67 (1H, m), 6.84–6.93 (4H, m), 7.15–7.23 (1H, m), 7.39 (1H, t, *J* = 8.0 Hz), 7.55 (2H, d, *J* = 8.4 Hz), 7.66–7.73

(2H, m), 7.76 (2H, d, J = 8.5 Hz), 8.23–8.32 (1H, m); MS m/z 940.6 (M+H)⁺; HRMS m/z(M+H)⁺ Calcd for C₅₃H₆₂N₇O₇S: 940.4431, Found: 940.4383; Purity: 100% (HPLC); $[\alpha]_{D}^{25}$ -73.8 (*c* 0.200, methanol).

1-49.

N-((1*S*)-2-((2*S*)-2-(4-(3-((14-((3-Cyano-4-(4-cyanophenyl)-2,5-dimethyl-1*H*-pyrrol-1 -yl)methyl)phenoxy)-3,6,9,12-tetraoxatetradec-1-yl)oxy)benzoyl)-1,3-thiazol-2-yl)pyrr olidin-1-yl)-1-cyclohexyl-2-oxoethyl)-*N*²-methyl-L-alaninamide (42d).

Compound **42d** was prepared from **41d** in a similar manner to that described for compound **42a** and obtained as a colorless amorphous powder (90%). ¹H NMR (300 MHz, CD₃OD) δ 0.99–1.26 (8H, m), 1.54–1.82 (6H, m), 2.06–2.41 (13H, m), 3.14 (1H, q, *J* = 6.8 Hz), 3.59–3.71 (12H, m), 3.76–4.03 (6H, m), 4.07 (2H, dd, *J* = 5.5, 3.7 Hz), 4.17 (2H, dd, *J* = 5.4, 3.8 Hz), 4.49–4.59 (1H, m), 5.14 (2H, s), 5.43–5.65 (1H, m), 6.88–6.93 (4H, m), 7.17–7.23 (1H, m), 7.37–7.44 (1H, m), 7.53–7.59 (2H, m), 7.67–7.73 (2H, m), 7.73–7.80 (2H, m), 8.27–8.31 (1H, m). NH protons were omitted. MS *m/z* 1028.3 (M+H)⁺; HRMS *m/z* (M+H)⁺

Calcd for C₅₇H₇₀N₇O₉S: 1028.4956, Found: 1028.4945; Purity: 98.2% (HPLC); $[\alpha]_{D}^{25}$ -73.7 (*c* 0.200, methanol).

1-50.

N-((1*S*)-2-((2*S*)-2-(4-(3-(2-(2-(2-(2-((3-Cyano-4-(4-cyanophenyl))-2,5-dimethyl-1*H*-py rrol-1-yl)methyl)phenoxy)ethoxy)ethoxy)ethoxy)ethoxy)benzoyl)-1,3-thiazol-2-yl)pyrr olidin-1-yl)-1-cyclohexyl-2-oxoethyl)- N^2 -methyl-L-alaninamide (42e).

Compound **42e** was prepared from **41e** in a similar manner to that described for compound **42a** and obtained as a colorless amorphous powder (87%). ¹H NMR (300 MHz, CD₃OD) δ 0.96–1.38 (8H, m), 1.51–1.84 (6H, m), 2.02–2.40 (13H, m), 2.85–3.20 (1H, m), 3.60–4.03 (14H, m), 4.09–4.25 (4H, m), 4.45–4.60 (1H, m), 5.17 (2H, s), 5.42–5.67 (1H, m), 6.42 (1H, dd, *J* = 7.4, 0.9 Hz), 6.87 (1H, t, *J* = 7.5 Hz), 7.00 (1H, d, *J* = 8.1 Hz), 7.17 (1H, dd, *J* = 8.0, 2.1 Hz), 7.24 (1H, td, *J* = 7.8, 1.4 Hz), 7.33–7.44 (1H, m), 7.49–7.58 (2H, m), 7.64– 7.79 (4H, m), 8.24–8.30 (1H, m); MS *m/z* 984.4 (M+H)⁺; HRMS *m/z* (M+H)⁺ Calcd for

C₅₅H₆₆N₇O₈S: 984.4694, Found: 984.4626; Purity: 100% (HPLC); $[\alpha]_D^{25}$ -74.8 (*c* 0.205, methanol).

1-51. 2-(4-(2-Hydroxyethoxy)phenoxy)ethyl 4-methylbenzenesulfonate (44a).

To a mixture of **43a** (4.00 g, 20.2 mmol) in pyridine (50 mL) was added TsCl (3.85 g, 20.2 mmol) at 0 °C. The mixture was stirred at 0 °C for 2 h. The mixture was diluted with EtOAc, neutralized with 6 M HCl at 0 °C, and extracted with EtOAc. The organic layer was separated, washed with 6 M HCl, 5% aq. NaHCO₃, and brine, dried over Na₂SO₄, and concentrated *in vacuo*. The residue was purified by column chromatography on silica gel (eluted with 50–100% EtOAc in hexane) to give the title compound (3.68 g, 52%) as a colorless oil. ¹H NMR (300 MHz, CDCl₃) δ 1.98–2.05 (1H, m), 2.45 (3H, s), 3.88–4.13 (6H, m), 4.34 (2H, dd, *J* = 5.6, 4.0 Hz), 6.67–6.89 (4H, m), 7.34 (2H, d, *J* = 8.2 Hz), 7.82 (2H, d, *J* = 8.3 Hz).

1-52. 2-(3-(2-Hydroxyethoxy)phenoxy)ethyl 4-methylbenzenesulfonate (44b).

Compound **44b** was prepared from **43b** in a similar manner to that described for compound **44a** and obtained as a colorless oil (56%). ¹H NMR (300 MHz, CDCl₃) δ 1.97–2.03 (1H, m), 2.45 (3H, s), 3.89–4.20 (6H, m), 4.36 (2H, dd, J = 5.6, 3.9 Hz), 6.34–6.46 (2H, m), 6.49–6.59 (1H, m), 7.15 (1H, t, J = 8.1 Hz), 7.34 (2H, d, J = 8.1 Hz), 7.82 (2H, d, J = 8.3Hz); MS *m/z* 353.2 (M+H)⁺.

1-53. 2-(2-(2-Hydroxyethoxy)phenoxy)ethyl 4-methylbenzenesulfonate (44c).

Compound **44c** was prepared from **43c** in a similar manner to that described for compound **44a** and obtained as a colorless amorphous solid (55%). ¹H NMR (300 MHz, CDCl₃) δ 2.44 (3H, s), 2.66 (1H, t, *J* = 6.5 Hz), 3.86–3.97 (2H, m), 4.05–4.12 (2H, m), 4.22 (2H, dd, *J* = 5.6, 3.7 Hz), 4.39 (2H, dd, *J* = 5.7, 3.8 Hz), 6.78–7.01 (4H, m), 7.34 (2H, d, *J* = 8.0 Hz), 7.82 (2H, d, *J* = 8.4 Hz); MS *m/z* 353.2 (M+H)⁺.

1-54.

4-(4-Cyanophenyl)-1-(4-(2-(4-(2-hydroxyethoxy)phenoxy)ethoxy)benzyl)-2,5-dimethyl-

H-pyrrole-3-carbonitrile (45a).

A mixture of **33a** (327 mg, 1.00 mmol), **44a** (423 mg, 1.20 mmol), and K₂CO₃ (193 mg, 1.40 mmol) in DMF (4 mL) was stirred at 60 °C overnight. After cooling, the mixture was diluted with EtOAc, washed with 5% aq. NaHCO₃ and brine, dried over Na₂SO₄, and concentrated *in vacuo*. The residue was purified by column chromatography on silica gel (eluted with 50–100% EtOAc in hexane) to give the title compound (418 mg, 82%) as a colorless solid. ¹H NMR (300 MHz, CDCl₃) δ 2.04 (1H, t, *J* = 6.0 Hz), 2.20 (3H, s), 2.38 (3H, s), 3.88–4.08 (4H, m), 4.28 (4H, s), 5.05 (2H, s), 6.79–6.99 (8H, m), 7.52 (2H, d, *J* = 8.4 Hz); MS *m/z* 508.3 (M+H)⁺.

1-55.

4-(4-Cyanophenyl)-1-(4-(2-(3-(2-hydroxyethoxy)phenoxy)ethoxy)benzyl)-2,5-dimethyl-1*H*-pyrrole-3-carbonitrile (45b).

Compound **45b** was prepared from **44b** and **33a** in a similar manner to that described for compound **45a** and obtained as a colorless amorphous solid (90%). ¹H NMR (300 MHz, CDCl₃) δ 2.01 (1H, t, *J* = 6.1 Hz), 2.20 (3H, s), 2.38 (3H, s), 3.89–4.00 (2H, m), 4.02–4.10

(2H, m), 4.30 (4H, s), 5.05 (2H, s), 6.49–6.61 (3H, m), 6.83–6.99 (4H, m), 7.14–7.24 (1H,

m), 7.52 (2H, d, J = 8.3 Hz), 7.70 (2H, d, J = 8.4 Hz); MS m/z 508.3 (M+H)⁺.

1-56.

4-(4-Cyanophenyl)-1-(4-(2-(2-(2-hydroxyethoxy)phenoxy)ethoxy)benzyl)-2,5-dimethyl-1*H*-pyrrole-3-carbonitrile (45c).

Compound **45c** was prepared from **44c** and **33a** in a similar manner to that described for compound **45a** and obtained as a colorless amorphous solid (99%). ¹H NMR (300 MHz, CDCl₃) δ 2.20 (3H, s), 2.38 (3H, s), 2.79 (1H, t, *J* = 6.6 Hz), 3.85 (2H, dt, *J* = 6.3, 4.6 Hz), 4.07–4.17 (2H, m), 4.28–4.41 (4H, m), 5.05 (2H, s), 6.82–7.02 (8H, m), 7.49–7.57 (2H, m), 7.67–7.74 (2H, m); MS *m/z* 508.4 (M+H)⁺.

1-57.

2-(4-(2-(4-((3-Cyano-4-(4-cyanophenyl)-2,5-dimethyl-1*H*-pyrrol-1-yl)methyl)phenoxy) ethoxy)phenoxy)ethyl 4-methylbenzenesulfonate (46a).

To a mixture of **45a** (0.42 g, 0.82 mmol) in pyridine (6 mL) was added TsCl (471 mg, 2.47 mmol) at 0 °C. The mixture was stirred at room temperature for 2 h. The mixture was poured into 5% aq. NaHCO₃ and extracted with EtOAc. The organic layer was separated, washed with 5% aq. NaHCO₃ and brine, dried over Na₂SO₄, and concentrated *in vacuo*. The residue was purified by column chromatography on silica gel (eluted with 30–80% EtOAc in hexane) to give the title compound (450 mg, 83%) as a colorless amorphous powder. ¹H NMR (300 MHz, CDCl₃) δ 2.20 (3H, s), 2.38 (3H, s), 2.45 (3H, s), 4.11 (2H, dd, *J* = 5.6, 3.9 Hz), 4.20–4.30 (4H, m), 4.33 (2H, dd, *J* = 5.6, 3.9 Hz), 5.05 (2H, s), 6.68–6.98 (8H, m), 7.34 (2H, d, *J* = 8.2 Hz), 7.52 (2H, d, *J* = 8.3 Hz), 7.66–7.74 (2H, m), 7.81 (2H, d, *J* = 8.3 Hz); MS *m*/z 662.4 (M+H)⁺.

1-58.

2-(3-(2-(4-((3-Cyano-4-(4-cyanophenyl)-2,5-dimethyl-1*H*-pyrrol-1-yl)methyl)phenoxy) ethoxy)phenoxy)ethyl 4-methylbenzenesulfonate (46b).

Compound **46b** was prepared from **45b** in a similar manner to that described for compound **46a** and obtained as a colorless amorphous solid (87%). ¹H NMR (300 MHz, CDCl₃) δ 2.20 (3H, s), 2.38 (3H, s), 2.44 (3H, s), 4.09–4.15 (2H, m), 4.29 (4H, s), 4.35 (2H, dd, J = 5.6, 3.9 Hz), 5.05 (2H, s), 6.37–6.46 (2H, m), 6.51–6.59 (1H, m), 6.81–6.99 (4H, m), 7.08–7.22 (1H, m), 7.34 (2H, d, J = 8.1 Hz), 7.52 (2H, d, J = 8.2 Hz), 7.70 (2H, d, J = 8.2 Hz), 7.81 (2H, d, J = 8.2 Hz); MS m/z 662.4 (M+H)⁺.

1-59.

2-(2-(2-(4-((3-Cyano-4-(4-cyanophenyl)-2,5-dimethyl-1*H*-pyrrol-1-yl)methyl)phenoxy) ethoxy)phenoxy)ethyl 4-methylbenzenesulfonate (46c).

Compound **46c** was prepared from **45c** in a similar manner to that described for compound **46a** and obtained as a colorless amorphous solid (85%). ¹H NMR (300 MHz, CDCl₃) δ 2.21 (3H, s), 2.38 (3H, s), 2.41 (3H, s), 4.17–4.24 (2H, m), 4.27–4.38 (6H, m), 5.06 (2H, s), 6.78–7.02 (8H, m), 7.29 (2H, d, J = 7.9 Hz), 7.49–7.56 (2H, m), 7.65–7.73 (2H, m), 7.74–7.82 (2H, m); MS *m/z* 662.5 (M+H)⁺.

 1-60.
 tert-Butyl

 ((2S)-1-(((1S)-2-((2S)-2-(4-(3-(2-(4-((2-(4-((3-cyano-4-(4-cyanophenyl)-2,5-dimethyl-1H

 pyrrol-1-yl)methyl)phenoxy)ethoxy)phenoxy)benzoyl)-1,3-thiazol-2-yl)pyrrolid

 in-1-yl)-1-cyclohexyl-2-oxoethyl)amino)-1-oxopropan-2-yl)methylcarbamate (47a).

A mixture of 46a (0.45 g, 0.68 mmol), 19 (0.41 g, 0.68 mmol), and K₂CO₃ (113 mg, 0.818 mmol) in DMF (3 mL) was stirred at room temperature overnight and then heated at 50 °C for 24 h. After cooling, the mixture was diluted with EtOAc, washed with 5% aq. NaHCO₃ and brine, dried over Na₂SO₄, and concentrated in vacuo. The residue was purified by column chromatography on silica gel (eluted with 50-100% EtOAc in hexane), and then on NH silica gel (eluted with 50–100% EtOAc in hexane) to give the title compound (546 mg, 74%) as a colorless solid. ¹H NMR (300 MHz, CD₃OD) δ 0.96–1.37 (8H, m), 1.38–1.48 (9H, m), 1.49–1.83 (6H, m), 2.03–2.46 (10H, m), 2.62–2.93 (3H, m), 3.75–4.42 (10H, m), 4.43–4.73 (2H, m), 5.17 (2H, s), 5.38–5.68 (1H, m), 6.80–7.05 (8H, m), 7.25 (1H, dd, J = 8.3, 1.5 Hz), 7.44 (1H, t, J = 8.0 Hz), 7.57 (2H, d, J = 8.2 Hz), 7.66–7.88 (4H, m), 8.31 $(1H, s); MS m/z 1088.8 (M+H)^+$.

tert-Butyl

1-61. ((2S)-1-(((1S)-2-((2S)-2-(4-(3-(2-(3-(2-(4-((3-cyano-4-(4-cyanophenyl)-2,5-dimethyl-1Hpyrrol-1-yl)methyl)phenoxy)ethoxy)phenoxy)ethoxy)benzoyl)-1,3-thiazol-2-yl)pyrrolid

in-1-yl)-1-cyclohexyl-2-oxoethyl)amino)-1-oxopropan-2-yl)methylcarbamate (47b).

Compound 47b was prepared from 46b and 19 in a similar manner to that described for compound 47a and obtained as a colorless amorphous solid (82%). ¹H NMR (300 MHz, CD₃OD) & 0.94–1.38 (8H, m), 1.39–1.48 (9H, m), 1.49–1.82 (6H, m), 2.03–2.41 (10H, m), 2.63-2.91 (3H, m), 3.74-4.01 (2H, m), 4.21-4.66 (10H, m), 5.16 (2H, s), 5.39-5.66 (1H, m), 6.51–6.63 (3H, m), 6.87–7.00 (4H, m), 7.12–7.21 (1H, m), 7.25 (1H, dd, J = 8.3, 2.4Hz), 7.39–7.49 (1H, m), 7.57 (2H, d, J = 8.4 Hz), 7.67–7.86 (4H, m), 8.31 (1H, s); MS m/z $1088.8 (M+H)^+$.

1-62.

tert-Butyl

((2S)-1-(((1S)-2-((2S)-2-(4-(3-(2-(2-(2-(4-((3-cyano-4-(4-cyanophenyl)-2,5-dimethyl-1H-

pyrrol-1-yl)methyl)phenoxy)ethoxy)phenoxy)ethoxy)benzoyl)-1,3-thiazol-2-yl)pyrrolid in-1-yl)-1-cyclohexyl-2-oxoethyl)amino)-1-oxopropan-2-yl)methylcarbamate (47c). Compound **47c** was prepared from **46c** and **19** in a similar manner to that described for compound **47a** and obtained as a colorless amorphous solid (79%). ¹H NMR (300 MHz, CD₃OD) δ 0.96–1.37 (8H, m), 1.38–1.48 (9H, m), 1.49–1.82 (6H, m), 2.03–2.38 (10H, m), 2.60–2.90 (3H, m), 3.71–4.00 (2H, m), 4.20–4.40 (8H, m), 4.43–4.68 (2H, m), 5.10 (2H, s), 5.38–5.65 (1H, m), 6.79–7.10 (8H, m), 7.19 (1H, dd, *J* = 8.0, 2.2 Hz), 7.36 (1H, t, *J* = 7.9 Hz), 7.50–7.60 (2H, m), 7.63–7.82 (4H, m), 8.23–8.29 (1H, m); MS *m/z* 1088.9 (M+H)⁺.

1-63.

N-((1*S*)-2-((2*S*)-2-(4-(3-(2-(4-((3-Cyano-4-(4-cyanophenyl)-2,5-dimethyl-1*H*-pyrr ol-1-yl)methyl)phenoxy)ethoxy)phenoxy)ethoxy)benzoyl)-1,3-thiazol-2-yl)pyrrolidin-1-vl)-1-cyclohexyl-2-oxoethyl)- N^2 -methyl-L-alaninamide (48a).

Compound **48a** was prepared from **47a** in a similar manner to that described for compound **42a** and obtained as a colorless amorphous powder (85%). ¹H NMR (300 MHz, CDCl₃) δ

ACS Paragon Plus Environment

0.93–1.38 (8H, m), 1.51–1.89 (7H, m), 2.05–2.54 (13H, m), 2.83–3.12 (1H, m), 3.72–4.01 (2H, m), 4.25–4.42 (8H, m), 4.63 (1H, dd, J = 9.3, 6.2 Hz), 5.05 (2H, s), 5.55 (1H, dd, J = 7.7, 2.7 Hz), 6.82–6.99 (8H, m), 7.19 (1H, ddd, J = 8.3, 2.6, 0.9 Hz), 7.40 (1H, t, J = 8.0 Hz), 7.49–7.56 (2H, m), 7.66–7.73 (3H, m), 7.77 (1H, dd, J = 2.5, 1.5 Hz), 7.83 (1H, dt, J = 7.7, 1.1 Hz), 8.03–8.17 (1H, m); MS *m*/*z* 988.4 (M+H)⁺; HRMS *m*/*z* (M+H)⁺ Calcd for C₅₇H₆₂N₇O₇S: 988.4431, Found: 988.4403; Purity: 99.8% (HPLC); $[\alpha]_{D}^{25}$ –68.2 (*c* 0.201, methanol).

1-64.

N-((1*S*)-2-((2*S*)-2-(4-(3-(2-(3-(2-(4-((3-Cyano-4-(4-cyanophenyl)-2,5-dimethyl-1*H*-pyrr ol-1-yl)methyl)phenoxy)ethoxy)phenoxy)ethoxy)benzoyl)-1,3-thiazol-2-yl)pyrrolidin-1yl)-1-cyclohexyl-2-oxoethyl)-*N*²-methyl-L-alaninamide (48b).

Compound **48b** was prepared from **47b** in a similar manner to that described for compound **42a** and obtained as a colorless amorphous solid (86%). ¹H NMR (300 MHz, CDCl₃) δ 0.92–1.40 (8H, m), 1.48–1.87 (7H, m), 2.06–2.54 (13H, m), 2.81–3.14 (1H, m), 3.71–4.01

(2H, m), 4.25–4.44 (8H, m), 4.62 (1H, dd, J = 9.2, 6.3 Hz), 5.05 (2H, s), 5.55 (1H, dd, J = 7.7, 2.7 Hz), 6.51–6.64 (3H, m), 6.81–6.98 (4H, m), 7.13–7.25 (2H, m), 7.40 (1H, t, J = 7.9 Hz), 7.48–7.56 (2H, m), 7.65–7.73 (3H, m), 7.76 (1H, dd, J = 2.4, 1.5 Hz), 7.84 (1H, dt, J = 7.7, 1.0 Hz), 8.04–8.17 (1H, m); MS *m/z* 988.4 (M+H)⁺; HRMS *m/z* (M+H)⁺ Calcd for C₅₇H₆₂N₇O₇S: 988.4431, Found: 988.4432; Purity: 97.8% (HPLC); $[\alpha]_{D}^{25}$ –71.0 (*c* 0.200, methanol).

1-65.

N-((1*S*)-2-((2*S*)-2-(4-(3-(2-(2-(2-(4-((3-Cyano-4-(4-cyanophenyl)-2,5-dimethyl-1*H*-pyrr ol-1-yl)methyl)phenoxy)ethoxy)phenoxy)ethoxy)benzoyl)-1,3-thiazol-2-yl)pyrrolidin-1vl)-1-cyclohexyl-2-oxoethyl)-*N*²-methyl-L-alaninamide (48c).

Compound **48c** was prepared from **47c** in a similar manner to that described for compound **42a** and obtained as a colorless amorphous solid (89%). ¹H NMR (300 MHz, CDCl₃) δ 0.93–1.39 (8H, m), 1.52–1.87 (7H, m), 2.06–2.54 (13H, m), 2.81–3.13 (1H, m), 3.70–4.02 (2H, m), 4.24–4.46 (8H, m), 4.63 (1H, dd, *J* = 9.3, 6.2 Hz), 5.01 (2H, s), 5.54 (1H, dd, *J* =

7.7, 2.6 Hz), 6.75–7.06 (8H, m), 7.16 (1H, ddd, J = 8.2, 2.6, 0.8 Hz), 7.36 (1H, t, J = 7.9 Hz), 7.47–7.57 (2H, m), 7.64–7.86 (5H, m), 8.02–8.15 (1H, m); MS *m/z* 988.4 (M+H)⁺; HRMS *m/z* (M+H)⁺ Calcd for C₅₇H₆₂N₇O₇S: 988.4431, Found: 988.4393; Purity: 99.6% (HPLC); $[\alpha]_{D}^{25}$ –72.7 (*c* 0.202, methanol).

1-66. NMR SPECTRA

Compound 4a





Compound 4b



Compound 4c



Compound 4d







Compound 18



Compound 23



Compound 24







Compound 32b



Compound 42a






Compound 42b



Compound 42c





Compound 42d

¹H NMR (300 MHz, CD₃OD)



Compound 42e



Compound 48a







Compound 48c



1-67. HRMS CHART

Compound 42a

1	
2	
2	
1	
4	
5	
6	
7	
8	
9	
10	
11	
12	
13	
14	
15	
16	
17	
18	
19	
20	
20	
21	
22	
23	
24	
25	
26	
27	
28	
29	
30	
31	
32	
33	
34	
35	
36	
37	
38	
30	
10	
40	
41	
42	
43	
44	
45	
46	
47	
48	
49	
50	
51	
52	
53	
54	
55	
56	
57	
50	
20 50	
59	
60	







1-68. HPLC CHART

Compound 42a



2. Reagents.

Tissue culture plastics were purchased from Greiner Bio-One (Tokyo, Japan). Compound

and methyl
(3*S*)-5-fluoro-3-(((2*S*)-2-(((2*S*)-3-methyl-2-(phenylmethoxycarbonylamino)butanoyl)amino
)propanoyl)amino)-4-oxopentanoate **78** were purchased from Peptide Institute (Osaka,
Japan). 5α-Dihydrotestosterone (DHT) was purchased from Tokyo Chemical Industry Co.,
Ltd. (Tokyo, Japan).

3. Cell culture.

Human PC cell lines (22Rv1, VCaP, LNCaP and PC-3) and human lung adenocarcinoma cell line A549 were maintained in Roswell Park Memorial Institute (RPMI)-1640 medium (Sigma-Aldrich) supplemented with 10% fetal bovine serum (Gibco) and 50 µg/mL of kanamycin (Sigma-Aldrich). Human fibrosarcoma cell line HT1080 was maintained in Dulbecco's modified Eagle's medium (Sigma-Aldrich) supplemented with 10% fetal bovine serum and 50 µg/mL of kanamycin. To examine the effects of androgen on VCaP cells, cells were cultured in phenol red-free RPMI-1640 medium (Gibco) containing 5%

charcoal-stripped fetal bovine serum (HyClone Laboratories, Logan, UT, USA) for 2 days, and then DHT was added to the medium.

4. Western blotting.

Cells were lysed with SDS lysis buffer (0.1 M Tris-HCl at pH 8.0, 10% glycerol, 1% SDS). Protein concentration was measured by the BCA method (Pierce) and an equal amount of protein lysate was separated by SDS-PAGE, transferred to PVDF membranes (Millipore), and analyzed by Western blot using the appropriate antibodies. The immunoreactive proteins were visualized using Clarity Western ECL substrate (Bio-Rad, Hercules, CA, USA), and their light emission was quantified with a LAS-3000 lumino-image analyzer (Fuji, Tokyo, Japan). The following antibodies were used: anti-AR (5153), anti-PARP (9532), anti-cleaved Caspase-3 (9664) (Cell Signaling Technology), anti-cIAP1 (AF8181) (R&D systems), anti-GAPDH (sc-25778 HRP) (Santa Cruz).

5. Measurement of binding affinity.

After FreeStyle293F (Invitrogen) cells were transfected with pcDNA3.1 containing an AR gene (wild-type AR) by using 293fectin transfection reagent (Invitrogen), these cells were

seeded into Erlenmeyer flask (Corning, 1L, 430518) at 1.1×10^6 cells/mL in FreeStyle293 Expression Medium (Invitrogen). After a 48 h shaking incubation (125 rpm) at 37 °C in an 8% CO₂ atmosphere, these cells were washed with TEG Buffer (10 mM Tris-HCl (pH 7.2), 50 mM EDTA, 10% Glycerol), and suspended with TEGM Buffer (10 mM Tris-HCl (pH 7.2), 1 mM EDTA, 10% glycerol, 10 mM Na₂MoO₄, 1 mM DTT, 1 mM 2-ME, 1 \times Complete protease inhibitor tablet (Roche)). After freezing and thawing to lyse cells, lysate was centrifuged at 228,000 × g at 4 °C for 20 min. The supernatant was stored at -80 °C as AR cell lysate. To cell lysate solution containing an AR were added $[17-\alpha-methyl-^{3}H]$ mibolerone (final 3 nM, PerkinElmer NET-919) and a compound, and the mixture was incubated at 4 °C for 3 h. B (Bound)/ F (Free) were separated by the dextran/charcoal method.⁵¹ The label count of B was measured, and the inhibitory rate of the compound was calculated.

6. RNA isolation and quantitative PCR.

Total RNA was prepared from cells with RNeasy (Qiagen). First-strand cDNA was synthesized from 1 µg total RNA with an oligo-dT primer using the SuperScript

> First-Strand Synthesis System (Invitrogen). Quantitative real-time PCR was performed with an ABI Prism 7300 sequence detection system (Applied Biosystems, Foster City, CA) using SYBR GreenER (Invitrogen) with gene-specific primers. Human 36B4 mRNA was used as an invariant control. The following PCR primers were used (5' to 3'): 36b4, GGC CCG AGA AGA CCT CCT T and CCA GTC TTG ATC AGC TGC ACA; Psa, CAG TCT GCG GCG GTG TTC and TGC CGA CCC AGC AAG ATC; Tmprss2, GAC CAG GAG TGT ACG GGA ATG T and AGG ACG AAG ACC ATG TGG ATT AG; Klk2, GCT GCC CAT TGC CTA AAG AA and TGG GAA GCT GTG GCT GAC A; Nkx3-1, GGA GAG AGG GAA AAT CAA GTG GTA and GTG ACA CAG GAG GAT GGA GTT G. The relative amounts of each mRNA were calculated using the comparative Ct method.⁵²

7. Cell viability assay.

Cell viability was determined using water-soluble tetrazolium WST-8 (4-[3-(2-methoxy-4-nitrophenyl)-2-(4-nitrophenyl)-2H-5-tetrazolio]-1,3-benzene

disulfonate) for the spectrophotometric assay according to the manufacturer's instructions (Dojindo, Tokyo, Japan). Cells were seeded at a concentration of 5×10^3 cells per well in a

96-well culture plate. After 24 h, the cells were treated with the indicated compounds for 48 h. The WST-8 reagent was added and the cells were incubated for 0.5 h at 37 °C in a humidified atmosphere of 5% CO₂. The absorbance at 450 nm of the medium was measured using an EnVision Multilabel Plate Reader (PerkinElmer).

8. Microscopy.

After treatment with the tested compounds, phase-contrast images of the cells were obtained by a BZ-9000 (Keyence, Osaka, Japan).

9. Measurement of apoptosis by flow cytometry.

Apoptosis was analyzed with an Annexin V-FITC Apoptosis Detection Kit (BioVision, Milpitas, CA, USA), as previously described.^{17, 53} Briefly, after being treated with the tested compounds, the cells were harvested and resuspended in binding buffer. The cells were stained with annexin V-FITC and propidium iodide (PI) at room temperature for 5 min in the dark, according to the manufacturer's instructions, and analyzed in a FACScan flow cytometer (Becton Dickinson, Braintree, MA, USA).

10. Statistical analysis

Student's t-test was used to determine the significance of differences among the

experimental groups. Values of P<0.05 were considered significant.

2
3
Δ
-
5
6
7
8
0
9
10
11
12
13
14
15
16
17
10
10
19
20
21
22
22
23
24
25
26
20
21
28
29
30
21
51
32
33
34
35
00
36
37
38
30
40
40
41
42
43
44
 15
40
46
47
48
10
49
50
51
52
53
50
5 4
55
56
57
50
00
59

ASSOCIATED CONTENT

Supporting Information

Schemes and experimental procedures for synthesis of AR and IAP ligand parts.

Molecular formula strings.

AUTHOR INFORMATION

Corresponding Author

*E-mail: miki-naito@nihs.go.jp. Phone: +81-3-3700-9428. Fax: +81-3-3707-6950.

ORCID

Hiroshi Nara: 0000-0001-6770-2750

Mikihiko Naito: 0000-0003-0451-1337

Notes

K Nagai, Y Morita, O Ujikawa, R Koyama, O Sano, Y Imaeda, H Nara, and N Cho are employees of Takeda Pharmaceutical Co., Ltd. (Osaka, Japan). M Naito received a research fund from Takeda Pharmaceutical Co., Ltd. The other authors declare no conflict of interest.

ACKNOWLEDGMENTS

This study was supported, in part, by Japan Society for the Promotion of Science (KAKENHI Grant Numbers 26860050 to N.S., 26860049 to N.O., 16H05090 to T.H. and M.N. and 16K15121 to N.O. and M.N.), by the Project for Cancer Research And Therapeutic Evolution (P-CREATE) (16cm0106124j0001 to N.O.) and the Research on Development of New Drugs (15ak0101029h1402 and 16ak0101029j1403 to M.N.) from the Japan Agency for Medical Research and Development (AMED), by the Ministry of Health and Labor Welfare, Japan (to M.N.), by Takeda Science Foundation (to N.O.) and by Takeda Pharmaceutical Co. Ltd. (to M.N.). The authors thank Mariko Seki for measurement of the protein knockdown activities.

ABBREVIATIONS USED

SNIPERs, Specific and Non-genetic IAP-dependent Protein Erasers; cIAP, cellular inhibitor of apoptosis protein; AR, androgen receptor; PC, prostate cancer; CRPC, castration-resistant prostate cancer; UPS, ubiquitin proteasome system; PROTACs, Proteolysis Targeting

ACS Paragon Plus Environment

Journal of Medicinal Chemistry

Chimeras; POI, protein of interest; PEG, polyethylene glycol; DHT, 5α-dihydrotestosterone; VHL, von Hippel-Lindau. MeOH, methanol; CD₃OD, deuterated methanol; EtOH, ethanol; CDCl₃, deuterated chloroform; DMSO-d₆, deuterated dimethyl sulfoxide; MeCN, acetonitrile; CPME, cyclopentyl methyl ether; EtOAc, ethyl acetate; *m*-CPBA, *m*-chloroperbenzoic acid; DIPEA, *N*,*N*'-diisopropylethylamine; EDC, 1-(3-(dimethylamino)propyl)-3-ethylcarbodiimide; Et₃N, triethylamine; HATU, *O*-(7-azabenzotriazol-1-yl)-*N*,*N*,*N'*,*N'*-tetramethyluronium hexafluorophosphate; HOBt. 1-hydroxybenzotriazole; HOBt H_2O , 1-hydroxy-1*H*-benzotriazole hydrate; SEMCl, 2-(trimethylsilyl)ethoxymethyl chloride; MsCl, methanesulfonyl chloride; TsCl, p-toluenesulfonyl chloride; TrCl, triphenylmethyl chloride; TsOH·H₂O, p-toluenesulfonic acid monohydrate; IPE, diisopropyl ether; TBAC, tetrabutylammonium chloride; Me₂NH, dimethylamine; Pd-C, palladium on carbon; Pd(OAc)₂, palladium(II) acetate; Pd(Ph₃P)₄, tetrakis(triphenylphosphine)palladium(0); Ph₃P, triphenylphosphine; Boc₂O, di-tert-butyl dicarbonate.

Aragon-Ching, J. B. The evolution of prostate cancer therapy: targeting the androgen receptor. *Front. Oncol.* 2014, *4*, 295.
 Heinlein, C. A.; Chang, C. Androgen receptor in prostate cancer. *Endocr. Rev.* 2004, *25*, 276-308.
 Mashima, T.; Okabe, S.; Seimiya, H. Pharmacological targeting of constitutively

active truncated androgen receptor by nigericin and suppression of hormone-refractory prostate cancer cell growth. *Mol. Pharmacol.* **2010**, *78*, 846-854.

(4) Mashima, T.; Soma-Nagae, T.; Migita, T.; Kinoshita, R.; Iwamoto, A.; Yuasa, T.; Yonese, J.; Ishikawa, Y.; Seimiya, H. TRIB1 supports prostate tumorigenesis and tumor-propagating cell survival by regulation of endoplasmic reticulum chaperone expression. *Cancer Res.* **2014**, *74*, 4888-4897.

(5) Tian, X.; He, Y.; Zhou, J. Progress in antiandrogen design targeting hormone binding pocket to circumvent mutation based resistance. *Front. Pharmacol.* **2015**, *6*, 57.

(6) Chen, C. D.; Welsbie, D. S.; Tran, C.; Baek, S. H.; Chen, R.; Vessella, R.;

Rosenfeld, M. G.; Sawyers, C. L. Molecular determinants of resistance to antiandrogen therapy. *Nat. Med.* **2004**, *10*, 33-39.

(7) Harris, W. P.; Mostaghel, E. A.; Nelson, P. S.; Montgomery, B. Androgen deprivation therapy: progress in understanding mechanisms of resistance and optimizing androgen depletion. *Nat. Clin. Pract. Urol.* **2009**, *6*, 76-85.

(8) Tan, M. H.; Li, J.; Xu, H. E.; Melcher, K.; Yong, E. L. Androgen receptor:

structure, role in prostate cancer and drug discovery. Acta Pharmacol. Sin. 2015, 36, 3-23.

- (9) Kim, D. H.; Rossi, J. J. Strategies for silencing human disease using RNA interference. *Nat. Rev. Genet.* 2007, *8*, 173-184.
- (10) Davidson, B. L.; McCray, P. B., Jr. Current prospects for RNA interference-based therapies. *Nat. Rev. Genet.* **2011**, *12*, 329-340.
- (11) Ohoka, N.; Shibata, N.; Hattori, T.; Naito, M. Protein knockdown technology:

application of ubiquitin ligase to cancer therapy. Curr. Cancer Drug Targets 2016, 16, 136-146.

(12) Itoh, Y.; Ishikawa, M.; Naito, M.; Hashimoto, Y. Protein knockdown using methyl

bestatin-ligand hybrid molecules: design and synthesis of inducers of ubiquitination-mediated degradation of cellular retinoic acid-binding proteins. J. Am. Chem.

Soc. 2010, 132, 5820-5826.

(13) Okuhira, K.; Ohoka, N.; Sai, K.; Nishimaki-Mogami, T.; Itoh, Y.; Ishikawa, M.;
Hashimoto, Y.; Naito, M. Specific degradation of CRABP-II via cIAP1-mediated
ubiquitylation induced by hybrid molecules that crosslink cIAP1 and the target protein. *FEBS Lett.* 2011, 585, 1147-1152.

(14) Demizu, Y.; Okuhira, K.; Motoi, H.; Ohno, A.; Shoda, T.; Fukuhara, K.; Okuda,

H.; Naito, M.; Kurihara, M. Design and synthesis of estrogen receptor degradation inducer based on a protein knockdown strategy. *Bioorg. Med. Chem. Lett.* **2012**, *22*, 1793-1796.

(15) Itoh, Y.; Ishikawa, M.; Kitaguchi, R.; Okuhira, K.; Naito, M.; Hashimoto, Y.

Double protein knockdown of cIAP1 and CRABP-II using a hybrid molecule consisting of

ATRA and IAPs antagonist. Bioorg. Med. Chem. Lett. 2012, 22, 4453-4457.

(16) Okuhira, K.; Demizu, Y.; Hattori, T.; Ohoka, N.; Shibata, N.; Nishimaki-Mogami,

T.; Okuda, H.; Kurihara, M.; Naito, M. Development of hybrid small molecules that induce

degradation of estrogen receptor-alpha and necrotic cell death in breast cancer cells. *Cancer Sci.* **2013**, *104*, 1492-1498.

(17) Ohoka, N.; Nagai, K.; Hattori, T.; Okuhira, K.; Shibata, N.; Cho, N.; Naito, M. Cancer cell death induced by novel small molecules degrading the TACC3 protein via the ubiquitin-proteasome pathway. *Cell. Death. Dis.* **2014**, *5*, e1513.

(18) Tomoshige, S.; Naito, M.; Hashimoto, Y.; Ishikawa, M. Degradation of HaloTag-fused nuclear proteins using bestatin-HaloTag ligand hybrid molecules. *Org. Biomol. Chem.* **2015**, *13*, 9746-9750.

(19) Demizu, Y.; Shibata, N.; Hattori, T.; Ohoka, N.; Motoi, H.; Misawa, T.; Shoda, T.;
Naito, M.; Kurihara, M. Development of BCR-ABL degradation inducers via the conjugation of an imatinib derivative and a cIAP1 ligand. *Bioorg. Med. Chem. Lett.* 2016, 26, 4865-4869.

(20) Okuhira, K.; Demizu, Y.; Hattori, T.; Ohoka, N.; Shibata, N.; Kurihara, M.; Naito,
M. Molecular design, synthesis, and evaluation of SNIPER(ER) that induces proteasomal degradation of ERalpha. *Methods Mol. Biol.* 2016, *1366*, 549-560.

(21) Okuhira, K.; Shoda, T.; Omura, R.; Ohoka, N.; Hattori, T.; Shibata, N.; Demizu, Y.; Sugihara, R.; Ichino, A.; Kawahara, H.; Itoh, Y.; Ishikawa, M.; Hashimoto, Y.; Kurihara, M.; Itoh, S.; Saito, H.; Naito, M. Targeted degradation of proteins localized in subcellular compartments by hybrid small molecules. *Mol. Pharmacol.* 2017, *91*, 159-166.
(22) Ohoka, N.; Okuhira, K.; Ito, M.; Nagai, K.; Shibata, N.; Hattori, T.; Ujikawa, O.; Shimokawa, K.; Sano, O.; Koyama, R.; Fujita, H.; Teratani, M.; Matsumoto, H.; Imaeda, Y.; Nara, H.; Cho, N.; Naito, M. In vivo knockdown of pathogenic proteins via specific and nongenetic inhibitor of apoptosis protein (IAP)-dependent protein erasers (SNIPERs). *J. Biol. Chem.* 2017, *292*, 4556-4570.
(23) Sakamoto, K. M.; Kim, K. B.; Verma, R.; Ransick, A.; Stein, B.; Crews, C. M.;

Deshaies, R. J. Development of PROTACs to target cancer-promoting proteins for ubiquitination and degradation. *Mol. Cell. Proteomics* **2003**, *2*, 1350-1358.

(24) Schneekloth, J. S., Jr.; Fonseca, F. N.; Koldobskiy, M.; Mandal, A.; Deshaies, R.;
Sakamoto, K.; Crews, C. M. Chemical genetic control of protein levels: selective in vivo
targeted degradation. *J. Am. Chem. Soc.* 2004, *126*, 3748-3754.

(25) Puppala, D.; Lee, H.; Kim, K. B.; Swanson, H. I. Development of an aryl hydrocarbon receptor antagonist using the proteolysis-targeting chimeric molecules approach: a potential tool for chemoprevention. *Mol. Pharmacol.* 2008, *73*, 1064-1071.
(26) Rodriguez-Gonzalez, A.; Cyrus, K.; Salcius, M.; Kim, K.; Crews, C. M.; Deshaies, R. J.; Sakamoto, K. M. Targeting steroid hormone receptors for ubiquitination and degradation in breast and prostate cancer. *Oncogene* 2008, *27*, 7201-7211.
(27) Schneekloth, A. R.; Pucheault, M.; Tae, H. S.; Crews, C. M. Targeted intracellular

protein degradation induced by a small molecule: En route to chemical proteomics. *Bioorg. Med. Chem. Lett.* , *18*, 5904-5908.

(28) Bondeson, D. P.; Mares, A.; Smith, I. E.; Ko, E.; Campos, S.; Miah, A. H.;

Mulholland, K. E.; Routly, N.; Buckley, D. L.; Gustafson, J. L.; Zinn, N.; Grandi, P.;

Shimamura, S.; Bergamini, G.; Faelth-Savitski, M.; Bantscheff, M.; Cox, C.; Gordon, D.

A.; Willard, R. R.; Flanagan, J. J.; Casillas, L. N.; Votta, B. J.; den Besten, W.; Famm, K.;

Kruidenier, L.; Carter, P. S.; Harling, J. D.; Churcher, I.; Crews, C. M. Catalytic in vivo

protein knockdown by small-molecule PROTACs. Nat. Chem. Biol. 2015, 11, 611-617.

(29) Lu, J.; Qian, Y.; Altieri, M.; Dong, H.; Wang, J.; Raina, K.; Hines, J.; Winkler, J.

D.; Crew, A. P.; Coleman, K.; Crews, C. M. Hijacking the E3 ubiquitin ligase cereblon to efficiently target BRD4. *Chem. Biol.* **2015**, *22*, 755-763.

(30) Winter, G. E.; Buckley, D. L.; Paulk, J.; Roberts, J. M.; Souza, A.; Dhe-Paganon,

S.; Bradner, J. E. Phthalimide conjugation as a strategy for in vivo target protein degradation. *Science* **2015**, *348*, 1376-1381.

(31) Lai, A. C.; Toure, M.; Hellerschmied, D.; Salami, J.; Jaime-Figueroa, S.; Ko, E.;

Hines, J.; Crews, C. M. Modular PROTAC design for the degradation of oncogenic BCR-ABL. Angew. Chem. Int. Ed. Engl. 2016, 55, 807-810.

(32) Toure, M.; Crews, C. M. Small-molecule PROTACS: New approaches to protein degradation. *Angew. Chem. Int. Ed. Engl.* **2016**, *55*, 1966-1973.

(33) Sekine, K.; Takubo, K.; Kikuchi, R.; Nishimoto, M.; Kitagawa, M.; Abe, F.;

Nishikawa, K.; Tsuruo, T.; Naito, M. Small molecules destabilize cIAP1 by activating auto-ubiquitylation. *J. Biol. Chem.* **2008**, *283*, 8961-8968.

(34) Charest, M. G.; Chen, C. H.-T.; Chen, Z.; Dai, M.; He, F.; Lei, H.; Pham, L. L.;

Sharma, S. K.; Straub, C. S.; Wang, R.-M. D.; Yang, F.; Zawel, L. Smac peptidomimetics useful as IAP inhibitors. WO/2008/016893A1, February 7, 2008.

(35) Yamamoto, S.; Matsunaga, N.; Hitaka, T.; Yamada, M.; Hara, T.; Miyazaki, J.;

Santou, T.; Kusaka, M.; Yamaoka, M.; Kanzaki, N.; Furuya, S.; Tasaka, A.; Hamamura, K.;

Ito, M. Design, synthesis, and biological evaluation of 4-phenylpyrrole derivatives as novel androgen receptor antagonists. *Bioorg. Med. Chem.* **2012**, *20*, 422-434.

(36) Bohl, C. E.; Gao, W.; Miller, D. D.; Bell, C. E.; Dalton, J. T. Structural basis for antagonism and resistance of bicalutamide in prostate cancer. *Proc. Natl. Acad. Sci. USA* 2005, *102*, 6201-6206.

(37) Yamamoto, S.; Tomita, N.; Suzuki, Y.; Suzaki, T.; Kaku, T.; Hara, T.; Yamaoka,

M.; Kanzaki, N.; Hasuoka, A.; Baba, A.; Ito, M. Design, synthesis, and biological evaluation of 4-arylmethyl-1-phenylpyrazole and 4-aryloxy-1-phenylpyrazole derivatives

as novel androgen receptor antagonists. Bioorg. Med. Chem. 2012, 20, 2338-2352.

(38) Varfolomeev, E.; Blankenship, J. W.; Wayson, S. M.; Fedorova, A. V.; Kayagaki,

N.; Garg, P.; Zobel, K.; Dynek, J. N.; Elliott, L. O.; Wallweber, H. J.; Flygare, J. A.;

Fairbrother, W. J.; Deshayes, K.; Dixit, V. M.; Vucic, D. IAP antagonists induce autoubiquitination of c-IAPs, NF-kappaB activation, and TNFalpha-dependent apoptosis.

Cell **2007,** *131*, 669-681.

(39) Umezawa, H.; Aoyagi, T.; Suda, H.; Hamada, M.; Takeuchi, T. Bestatin, an inhibitor of aminopeptidase B, produced by actinomycetes. *J. Antibiot. (Tokyo)* **1976**, *29*, 97-99.

(40) Orning, L.; Krivi, G.; Fitzpatrick, F. A. Leukotriene A4 hydrolase. Inhibition by bestatin and intrinsic aminopeptidase activity establish its functional resemblance to metallohydrolase enzymes. *J. Biol. Chem.* **1991**, *266*, 1375-1378.

(41) Tsubuki, S.; Kawasaki, H.; Saito, Y.; Miyashita, N.; Inomata, M.; Kawashima, S. Purification and characterization of a Z-Leu-Leu-Leu-MCA degrading protease expected to regulate neurite formation: a novel catalytic activity in proteasome. *Biochem. Biophys. Res.*

Commun. 1993, 196, 1195-1201.

(42) Liu, W.; Xie, C. C.; Zhu, Y.; Li, T.; Sun, J.; Cheng, Y.; Ewing, C. M.; Dalrymple,

S.; Turner, A. R.; Sun, J.; Isaacs, J. T.; Chang, B. L.; Zheng, S. L.; Isaacs, W. B.; Xu, J.

Homozygous deletions and recurrent amplifications implicate new genes involved in prostate cancer. Neoplasia 2008, 10, 897-907. Murtha, P.; Tindall, D. J.; Young, C. Y. Androgen induction of a human (43)prostate-specific kallikrein, hKLK2: characterization of an androgen response element in the 5' promoter region of the gene. *Biochemistry* 1993, 32, 6459-6464. Prescott, J. L.; Blok, L.; Tindall, D. J. Isolation and androgen regulation of the (44)human homeobox cDNA, NKX3.1. Prostate 1998, 35, 71-80. Lin, B.; Ferguson, C.; White, J. T.; Wang, S.; Vessella, R.; True, L. D.; Hood, L.; (45)Nelson, P. S. Prostate-localized and androgen-regulated expression of the membrane-bound serine protease TMPRSS2. Cancer Res. 1999, 59, 4180-4184. Zhu, H.; Fearnhead, H. O.; Cohen, G. M. An ICE-like protease is a common (46)mediator of apoptosis induced by diverse stimuli in human monocytic THP.1 cells. FEBS

Lett. 1995, 374, 303-308.

(47) Hunter, A. M.; LaCasse, E. C.; Korneluk, R. G. The inhibitors of apoptosis

(IAPs) as cancer targets. Apoptosis 2007, 12, 1543-1568.

(48) Tamm, I.; Kornblau, S. M.; Segall, H.; Krajewski, S.; Welsh, K.; Kitada, S.; Scudiero, D. A.; Tudor, G.; Qui, Y. H.; Monks, A.; Andreeff, M.; Reed, J. C. Expression and prognostic significance of IAP-family genes in human cancers and myeloid leukemias. *Clin. Cancer Res.* **2000**, *6*, 1796-1803.

(49) Tucker, H.; Crook, J. W.; Chesterson, G. J. Nonsteroidal antiandrogens. Synthesis and structure-activity relationships of 3-substituted derivatives of 2-hydroxypropionanilides.
 J. Med. Chem. 1988, *31*, 954-959.

(50) Ito, M.; Tomita, N.; Kaku, T.; Suzuki, T. Substituted pyrazole derivatives and use thereof. WO/2009/119880, October 1, 2009.

(51) Boesel, R. W.; Shain, S. A. A rapid, specific protocol for determination of available androgen receptor sites in unfractionated rat ventral prostate cytosol preparations. *Biochem. Biophys. Res. Commun.* **1974**, *61*, 1004-1011.

(52) Livak, K. J.; Schmittgen, T. D. Analysis of relative gene expression data using real-time quantitative PCR and the 2(-delta delta C(T)) Method. *Methods* **2001**, *25*, 402-408.

(53) Shibata, N.; Ohoka, N.; Sugaki, Y.; Onodera, C.; Inoue, M.; Sakuraba, Y.;

Takakura, D.; Hashii, N.; Kawasaki, N.; Gondo, Y.; Naito, M. Degradation of stop codon

read-through mutant proteins via the ubiquitin-proteasome system causes hereditary

disorders. J. Biol. Chem. 2015, 290, 28428-28437.

Table of Contents Graphic

