Oxygen insertion in *Sesamum indicum* furanofuran lignans. Diastereoselective syntheses of enzyme substrate analogues

Patrice A. Marchand, Jaroslav Zajicek, and Norman G. Lewis

Abstract: The furofuran lignans in sesame seed have an unusual oxygen insertion between their furan and aryl rings. In our continuing investigations on the isolation and characterization of the enzyme(s) involved, the diastereoselective syntheses of various substrate analogues for the oxygen insertion step were developed for future substrate specificity and inhibitor studies. This synthetic strategy also provided entry to so-called furofuranone epoxy-lignans, such as salicifoliol from *Bupleurum* sp., and acuminatolide from *Helichrysum* sp.

Key words: furofuran lignans, sesame, epimers, salicifoliol, acuminatolide.

Resumé: On observe dans les lignanes furanofuraniques des graines de sésame, une insertion d'oxygène typique entre le cycle tétrahydrofuranique et le cycle aromatique. Comme avant propos à l'étude et l'isolation des enzymes responsables de la biosynthèse de la sésamoline à partir du pinorésinol, nous décrivons ici la synthèse diastéréosélective des épimères de ces intermédiaires oxygénés. Une extension de cette voie synthétique nous a fournit la possibilité d'accéder aux furolactones, le salicifoliol et l'acuminatolide.

Mots clés: lignanes furofuraniques, sésame, épimères, salicifoliol, acuminatolide.

Introduction

The furofuran lignans present in sesame (Sesamum indicum) seed have many important roles. They can function as potent antioxidants (1–5), either directly or through conversion to sesaminol **1** (4), thereby enhancing the quality and shelf life of sesame seed oil. Sesame lignans can also act as synergists with pyrethrum insecticides (6, 7), or as cyclic AMP phosphodiesterase (8) and Δ -5 desaturase inhibitors in mammalian species (9). Moreover, addition of sesamin **2** to the diet reduces serum and liver cholesterol levels in rats (10) and has a hypocholesterolemic effect in humans (11). Dietary sesame lignans can protect against EtOH or CCl₄ induced liver damage (12), and enhance vitamin E activities (13) and the bioavailability of γ -tocopherol (14) in vivo.

Chemotaxonomic analyses of *Sesame* seed has established its major lignans as (+)-sesamin 2 and (+)-sesamolin 4 (Scheme 1) with the latter differing only by an oxygen insertion between the furofuran and piperonyl (3,4)

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methylenedioxyphenyl) groups. Biochemical entry to these furofuran lignans occurs via stereoselective coupling of two molecules of *E*-coniferyl alcohol **5** to give (+)-pinoresinol **3**, an enzymatic conversion of particular interest being the first example of regio- and stereospecific control of bimolecular phenoxy radical coupling (15–17). Although its enzymology of formation was first delineated in *Forsythia* sp. (15–17), (+)-pinoresinol **3** biosynthesis from *E*-coniferyl alcohol **5** in *Sesame* sp. is apparently engendered in a comparable manner (unpublished data).

Depending upon the species involved, (+)-pinoresinol 3 (a central intermediate in lignan biosynthesis) can be metabolized into a wide variety of different furofuran, furano, dibenzyl butane, dibenzyl butyrolactone, and aryl (tetrahydro)-naphthalene lignans (18, 19). For example, in *Forsythia intermedia*, enantiospecific reduction of (+)-pinoresinol 3 gives rise to (+)-lariciresinol 6 and (-)-secoisolariciresinol 7 (20–22), the dehydrogenation of which then affords (-)-matairesinol 8 (23). The NADPH-dependent enzyme catalyzing the sequential reductive steps leading to (-)-secoisolariciresinol 7 has been purified to apparent homogeneity, and the gene encoding the protein cloned (24).

In contrast, the lignan pathway in *Sesame* sp. converts (+)-pinoresinol 3 into (+)-sesamolin 4 via introduction of methylenedioxy bridges and an oxygen insertion between its aryl and furan rings. However, the sequence of oxidative/cyclization steps, and its associated enzymology, are as yet unknown. Scheme 2 shows plausible biosynthetic intermediates in the direct conversion of (+)-pinoresinol 3 to (+)-sesamolin 4. At this juncture, it is emphazised that the nomenclature system adopted is that of the 8,8'-linked lignans; the other recommended systems (IUPAC and CAS) were not used since they lacked any internal numbering consistency between the

Scheme 1.

Legend: $Ar_1 = 3$ -methoxy-4-hydroxyphenyl; $Ar_2 = 3,4$ -methylenedioxyphenyl

different classes of lignans (and lignan-derived substances) described in this paper.

As can be seen, oxygen insertion could initially occur with (+)-pinoresinol 3 (route a) to give (+)-pinoresinolin 9, with sequential methylenedioxy bridge formation to afford (+)-piperitolin 10 and (+)-sesamolin 4, respectively. Oxygen insertion could also either follow or precede methylenedioxy bridge formation, i.e., from (+)-piperitol 11 to give (+)-sesamolinol 12 (route b) or from (+)-piperitol 11 to (+)-piperitolin 10 (route c), respectively. Perhaps, more unlikely, (+)-sesamin 2 may be directly converted into (+)-sesamolin 4 (route d). Synthetic methodology to pinoresinol 3 (22, 25), piperitol 11 (26), sesamin 2 (27), sesamolin 4 (27), pinoresinolin 9, piperitolin 10, and sesamolinol 12 (manuscript in preparation) has already been established, thus setting the stage for systematic study of the biosynthetic sequence involved.

The purpose of this investigation was to synthesize "epimers" 13–15 of the plausible aryl-O-furofuran biosynthetic intermediates 9, 10, and 12, which differ only in their stereochemistry at C-7' (Scheme 3). These are required to examine both the substrate specificity of the oxygen insertion reaction using epi-pinoresinol 16, epi-piperitol 17, and related compounds as enzyme substrate analogues, as well as determining whether they could function as inhibitors of the enzymatic transformation itself. Moreover, given our ongoing interest in the enantiospecificity of lignan biosynthetic conversions, and identification of the precise enzymology involved, it was important to obtain these putative substrate analogues in

Fig. 1. General strategy to "oxygenated" lignans.

racemic form. This is because the various enantiomers can readily be separated by chiral column HPLC and, hence, provide the means to probe in detail both the enantiospecificity and diastereoselectivity of the oxygen insertion step. This approach has already been successfully applied to the study of (+)-pinoresinol 3 (15–17), (+)-lariciresinol 6, (-)-secoisolariciresinol 7, (-)-matairesinol 8 (20–24), and (-)-arctigenin 18 (28) biosynthesis.

Results and discussion

The main goal of this study was to develop a facile synthesis of the "oxygenated" lignans, epi-pinoresinolin 13, epi-piperitolin 14, and epi-sesamolinol 15. To achieve this goal, we envisaged the best overall strategy to be that of synthesis of lactols of the type shown above, followed by aryl alkyl ether formation with inversion of configuration at C-7' (see Fig. 1).

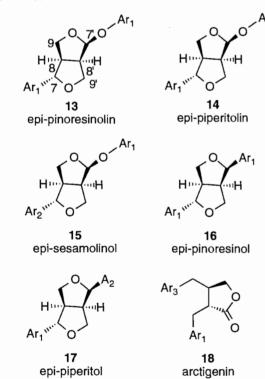
We sought first to establish the validity and usefulness of the proposed approach using, as synthon, the lactol samin 19, which can readily be obtained in either racemic (\pm) , (+), or (-) form (27, 29, 30). For this study, (+)-samin 19 (230 mg) was first conveniently prepared by acid hydrolysis (30, 31) of (+)-sesamolin 4 (527 mg), which in turn had previously been isolated from *S. indicum* seeds (Scheme 4).

In the ¹H NMR spectrum of the starting material, (+)-sesamolin 4, the H_{ax} proton at C-7' appears as a singlet resonance at δ 5.47 (32); its absence of coupling with the adjacent bridgehead proton at H-8' is in agreement with the projected dihedral angle of $\sim 90^{\circ}$. Importantly, the corresponding H_{ab} proton at C-7' of (+)-samin 19 also gave a singlet at δ 5.45 (data not shown), indicating that the relative stereochemistry in both molecules was the same at this position. Although the (+)-samin 19 thus generated had essentially an identical melting point, as well as IR, UV, and ¹H/¹³C NMR spectra, to that reported previously (29), additional verification of the configuration of the furofuranol skeleton of 19 was determined by its conversion via oxidation with pyridinium dichromate to give the lactone, (+)-acuminatolide 20. Its (-) antipode had previously been isolated from Helichrysum acuminatum (33), as well as obtained by total synthesis (27). Other than its optical rotation, (+)-acuminatolide 20 was identical in every respect to that of the natural product, thus verifying the overall configuration of (+)-samin 19 to be as shown. Moreover, to our knowledge, this is the first (hemi)synthesis of (+)-acuminatolide 20.

With the configuration of (+)-samin 19 so defined, attention was next directed to the synthesis of (+)-epi-sesamolinol 15. First, 4-benzyloxy-3-methoxyphenol 22 was obtained by Bayer-Villiger oxidation (34, 35) of O-benzyl vanillin 21 (23) via the action of m-chloroperoxybenzoic acid (m-CPBA) and subsequent NaOH hydrolysis. Phenol 22 was then condensed with (+)-samin 19, in the presence of PPh₃ and DEAD (36), to give 23, subsequent deprotection of which with $H_2/Pd/C$

Scheme 2. Proposed biosynthetic pathways from (+)-pinoresinol 3 to (+)-sesamolin 4.

Scheme 3.



Legend: $Ar_1 = 3$ -methoxy-4-hydroxyphenyl; $Ar_2 = 3$,4-methylenedioxyphenyl; $Ar_3 = 3$,4-dimethoxyphenyl

afforded (+)-epi-sesamolinol 15 in 60% overall yield. That the synthetic product so obtained had undergone an inversion of configuration at C-7' could readily be determined by inspection of the ¹H NMR spectrum. The most striking change was the appearance of a doublet (J = 5.6 Hz) at 5.59 ppm, corresponding to the H_{eq} proton at C-7' coupled to the C-8' bridgehead proton. (This contrasts with the comparable H-7'ax resonance in (+)-sesamolin 4, which has a singlet at δ 5.47 ppm.) This downfield shift of 0.15 ppm is consistent with related spectroscopic assignments for the naturally occurring lignan insecticidal agent, haedoxan A (H-7'ax), to that of its epimeric synthetic product, isohaedoxan A (H-7'eq), which showed a comparable downfield shift of 0.17 ppm (37). All other spectroscopic data supported epi-sesamolinol 15 to have the structure as shown. Consequently, not only does this represent the first (hemi)synthesis of (+)-epi-sesamolinol 15, but adaptation of this approach using (\pm) -samin 19 gives a convenient route to racemic (\pm)-epi-sesamolinol 15.

With the overall synthetic approach thus validated, attention was next directed to formation of the two other putative substrate analogues/inhibitors, epi-pinoresinolin 13 and epi-piperitolin 14. In an analogous manner, this required synthesis of the pivotal lactol intermediate 24 (Scheme 5), which differs only from samin 19 by having a 4-benzyloxy-3-methoxyphenyl rather than a piperonyl group. Condensation of lactol 24 with either phenol 22 or its corresponding piperonyl analog would therefore afford (following deprotection) epi-pinoresinolin 13 and epi-piperitolin 14, respectively (Schemes 5 and 6).

Scheme 4. Hemisynthesis of (+)-epi-sesamolinol 12 and (+)-acuminatolide 20.

Legend: $Ar_1 = 3$ -OMe-4-OH-Ph; $Ar_2 = 3,4$ -OCH₂O-Ph; $Bn = CH_2$ -Ph; $a: H_3O^+$, 65%; $b: PPh_3$, DEAD, THF, 80%; $c: H_2/Pd/EtOAc$, 75%; d: m-CPBA, CH_2Cl_2 , then NaOH, MeOH, 64%; e: PDC, AcOH, 4 Å, CH_2Cl_2 , 78%.

Lactol 24 was obtained from 4-vinylbutyrolactone 25 (38) as follows. Treatment of lactone 25 with LDA generated the corresponding lithiated form, which readily underwent diastereoselective addition with O-benzylvanillin 21. This afforded exclusively the *trans* lactonic alcohol 26, as a mixture of diastereoisomers in 92% yield (Scheme 5), which was used without further purification/resolution. (That the geometry of the pendant groups on the butyrolactone was indeed *trans* was based on a previous report of the configuration of the piperonyl analogue of 26, prepared in a similar manner, which in turn was employed in the synthesis of the lignan insecticidal agents, haedoxan A, D, E, and their stereoisomers (37).)

Subsequent reduction of lactone 26 with LiAlH₄ and reaction of corresponding triol 27 with bromotrimethylsilane gave tetrahydrofuran 28, presumed to possess both vinyl and hydroxymethyl substituents in a syn conformation, in 81% overall yield for both steps (37). Treatment of the vinyl group of alcohol 28 with OsO₄ in catalytic amount then afforded triol 29 as a mixture of diastereoisomers in 79% yield. Subsequent cleavage of its 1,2-diol by NaIO₄ gave the corresponding aldehyde, which underwent spontaneous cyclization, to afford lactol 24 in 84% yield. That this possessed a C-7' H_{ax} was again established by inspection of the ¹H NMR spectrum, which displayed a 1H singlet at δ 5.35 ppm. Additionally, verification that only one stereoisomer resulted from this synthetic scheme was established via two pieces of interlocking evidence. The first involved oxidative conversion to the known lignanderived furofuranone natural product, salicifoliol 31, and the second by determination of the configuration of epi-pinoresinolin 13 via a combination of NOESY and HETCOR analyses (see later).

It had previously been established that the aerial portions of Bupleurum salicifolium Soland (39) contain salicifoliol 31, in addition to the lignans (+)-pinoresinol 3, (+)-eudesmin, and (+)-medioresinol. Thus, to verify the configuration of racemic lactol 24, it was converted into salicifoliol 31 by oxidation with pyridinium dichromate (PDC) in the presence of AcOH and 4Å molecular sieves as before (40), with subsequent debenzylation (Scheme 5). That the synthetic product was indeed (±)-salicifoliol 31 was determined by its identical melting point to that of the natural product (101–102°C vs. 102-103°C (39)), as well as by comparison of its MS fragmentation patterns, ¹H/¹³C NMR, UV, and IR spectra to those reported for (+)-salicifoliol 31. Previous COSY experiments had shown that the pendant aryl group of salicifoliol 31 was exo, and that the bridgehead hydrogens were cis to each other. Thus, with all spectroscopic data identical to the naturally occurring product, with the exception of optical rotation, this represents the first synthesis of (\pm) -salicifoliol 31. More importantly, it provided strong evidence that the configuration of lactol 24 was as shown.

With good evidence supporting the structure and configuration of lactol 24 as shown, attention was directed to the formation of (\pm) -epi-pinoresinolin 13 (Scheme 6). Again, Obenzylvanillin 21 was converted into the corresponding phenol 22, via Bayer–Villiger oxidation with m-CPBA and subsequent NaOH hydrolysis. This was then reacted with lactol 24, in the presence of PPh₃ and DEAD, to give 32, subsequent

Scheme 5. Synthesis of (\pm) -lactol **24** and (\pm) -salicifoliol **31**.

Legend: $Ar_1 = 3$ -OMe-4-OH-Ph, $Bn = CH_2$ -Ph; a: LDA, THF, **21**, 92%; b: LiAlH₄, THF, 89%; c: TMSBr, THF, 89%; d: OsO₄, NMO, THF/H₂O, 79%; e: NalO₄, THF, 89%; f: PDC, AcOH, 4 Å, CH_2 Cl₂, 78%; g: H_2 /Pd/C, EtOAc, 90%.

Scheme 6. Synthesis of (\pm) -epi-pinoresinolin 13 and (\pm) -epi-piperitolin 14.

Legend: $Ar_1 = 3$ -OMe-4-OH-Ph; $Ar_2 = 3,4$ -OCH₂O-Ph; $Bn = CH_2$ -Ph; a: PPh₃, DEAD, THF, **22**: 89%; **33**: 85%; b: H_2 /Pd/C, EtOAc, **13**: 84%; **14**: 88%.

deprotection of which with $H_2/Pd/C$ gave (\pm)-epi-pinoresinolin 13 in 75% overall yield. Its configuration was next determined by a combination of HETCOR and NOESY spectral analyses, where the 1H NMR signals of epi-pinoresinolin 13

were assigned to individual protons on the basis of chemical shift, observed multiplicities, and decoupling experiments (Table 1), and all carbons (with directly attached protons) assigned using the HETCOR spectrum. Assignments of sig-

Table 1. NMR (600 MHz, ¹H; 150 MHz, ¹³C; CDCl₃) spectral data for epi-pinoresinolin **13**.

Assignment	¹³ C δ	$^{1}\mathrm{H}$	
		δ	J (Hz)
1	132.69	_	
2	108.53	6.93	d, $J_{2.6} = 1.9$
3	146.66°	_	2,0
4	145.23^{a}		
5	114.23	6.89	d, $J_{5.6} = 8.1$
6	118.90	6.83	d,d, $J_{2.6} = 1.9$, $J_{5.6} = 8.1$
7	85.30	4.80	d, $J_{7.8} = 7.1$
8	52.75	3.04	m, $J_{8.9ax} = 7.6$, $J_{8.9cq} = 4.7$
9	69.30	H_{ax} , 4.12	$m, J_{9ax,eq} = 9.1$
		H_{eq} , 4.03	
OH	_	5.43	br s
OCH_3	55.87	3.90	s, 3H
1'	150.80	_	
2'	101.75	6.69	d, $J_{2',6'} = 2.7$
3′	146.83°	_	
4'	140.91^{a}	_	
5′	114.17	6.84	d, $J_{5',6'} = 8.7$
6′	108.41	6.64	dd, $J_{6',2'} = 2.7$, $J_{6',5'} = 8.7$
7′	102.83	5.60	d, $J_{7',8'} = 5.6$
8′	51.10	3.33	$J_{8',9'ax} = 8.0, J_{8',9'eq} = 4.1, J_{8',8} = 10.0$
9′	67.00	H_{ax} , 4.42	$J_{9'ax,eq} = 9.1$
		$H_{eq}, 4.17$	•
OH	_	5.73	br s
OCH_3	55.91	3.87	S

^aAssigments are interchangeable for nonprotonated carbons.

nals derived from quaternary carbons were based on comparison with reported chemical shift data for (+)-sesamolin 4 (32).

To summarize, the NOESY spectra (mixing times 300 and 600 ms) showed a strong cross-peak between the H-8 and H-8' proton signals suggesting that both were cis to each other. Moreover, the H-8 signal exhibited a correlation with the H-2 and H-6 proton resonances of the pendant aryl (Ar) group at δ 6.93 and 6.83, respectively, indicating that the 3-OMe-4-OHphenyl group was attached to C-7 in a cis relationship to the H-8 and H-8' protons. Significantly, although the NOESY spectra also displayed cross peaks between the H-7' proton resonance and the H-2' and H-6' signals, none of the proton signals of the aryl group attached to the C-7' showed any correlation to the H-8' signal. In contrast, the H-7' signal exhibited a correlation to the H-8' signal, suggesting that the pendant aryl group attached to C-7' was trans to the H-8 and H-8' protons. The axial "β" configuration of the aryl group at C-7' was again further supported by the fact that the signal of the H-7'_{eq} proton was a downfield doublet $(J_{7',8'} = 5.6 \text{ Hz})$ at δ 5.6 ppm. Thus, via a combination of derivatization and spectroscopic analyses, the configuration of epi-pinoresinolin 13 was as shown.

Lastly, with the configuration of epi-pinoresinolin 13 unambiguously determined, sesamol 33 (3,4-methylenedioxy phenol) was condensed with lactol 24 to afford, after deprotection of 34, (\pm) -epi-piperitol 14 in 75% overall yield.

Concluding remarks

This study provides a convenient synthetic strategy to the "oxygenated" lignans epi-pinoresinolin 13, epi-piperitolin 14, and epi-sesamolinol 15 in good overall yield. The configuration of each molecule was determined via a number of spectroscopic means as well as by chemical derivatization, the latter proceeding in a manner that also permitted the synthesis of furofuranone lignan-derived substances, such as acuminatolide 20 and salicifoliol 31. Future studies will be directed to defining the enzymology of the "oxygen" insertion step, with the compounds so obtained used as substrate analogues or inhibitors.

Experimental

Unroasted and unbleached seeds of *Sesamum indicum* were purchased in a local grocery store. IR and UV spectra were recorded on Perkin–Elmer 1720-X FTIR and Lambda 6 UV/VIS spectrophotometers, respectively. NMR studies of epipinoresinolin **13** were performed on a Varian UNITY*plus* (599.89 MHz, ¹H) spectrometer, whereas all other ¹H/¹³C spectra were obtained on a Brüker AMX-300 (300.14 MHz, ¹H) instrument. NMR spectra were recorded in CDCl₃ solution with TMS as an internal reference, with chemical shifts (δ) expressed in ppm and coupling constants (*J*) in Hz.

NOESY spectra were recorded using the standard pulse sequence (41), with mixing times, t_m, of 300 and 600 ms. HETCOR spectra were measured using the pulse sequence previously described (42). The ¹H chemical shifts and coupling constants were obtained by first-order analysis of 1D spectra. EIMS were carried out using a Waters Integrity HPLC/MS, whereas HRMS employed a VG 7070 EHF mass spectrometer with a temperature source of 250°C at 70 eV with $Ar_1 = 3$ -methoxy-4-hydroxyphenyl and $Ar_2 = 3$,4-methylenedioxyphenyl for fragment interpretation. Solvents were either HPLC grade (CH3CN, MeOH, AcOH, CH2Cl2, THF) or ACS grade (EtOAc, hexanes) and were obtained from Baker. Tetrahydrofuran (THF) and methylene chloride (CH₂Cl₂) were distilled over LiAlH₄-triphenylmethane and CaH₂, and column and analytical thin-layer chromatographic separations were performed using silica gel 60 (230-400 mesh) and AL SI G/UV 254 (Whatman), respectively.

(+)-Samin 19

This was synthesized as outlined (30, 31) but modified as follows. A suspension of (+)-sesamolin 4 (527 mg, 1.42 mmol), DOWEX 50 X2-200 (545 mg), NaOAc (150 mg, 1.83 mmol), and HCl 10% (1 mL) in CH₃CN (10 mL) was stirred under reflux for 2 h. The suspension was then cooled to room temperature, the resin removed by filtration, and the filtrate evaporated in vacuo, to give a dry solid. The resulting residue was dissolved in a minimum amount of CHCl₃ and subjected to silica gel column chromatography, eluted with hexanes:EtOAc 3:1 to 1:1, to give (+)-samin 19 (230.8 mg, 65%); mp 108-109°C (EtOAc:cyclohexane) (lit. (31) mp 106°C; (29) mp 108–109°C); $[\alpha]_D$ +104.65 (c 0.516, CHCl₃) (lit. $[\alpha]_D$ (30) +103); IR (CHCl₃) ν_{max} : 3400 (br, OH), 1610, 1505, 1490, 1446, 1245 cm⁻¹; MS m/e (relative %): 250 (M⁺, 86), 232 $(M - H_2O^+, 6)$, 194 $(M - C_2O_2^+, 38)$, 176 (54), 149 $(Ar_2CO^+, 6)$ 100), 135 (Ar₂CH₂⁺, 58), 122 (Ar₂H⁺, 22); HRMS; calcd. for $C_{13}H_{14}O_5$: 250.0841 (M⁺), found: 250.0805; for ¹H and ¹³C NMR spectral analyses: see ref. 29.

(+)-Acuminatolide 20

To a solution of (+)-samin 19 (80 mg, 0.32 mmol) in dry CH₂Cl₂ (50 mL) was added pyridinium dichromate (PDC) (4 equiv., 481 mg), AcOH (1 equiv., 20 µL), and 4Å molecular sieves (500 mg). The resulting suspension was stirred for 6 h at room temperature (r.t.), then filtered through a plug of silica gel and the filtrate removed in vacuo to give a yellow solid. The residue was dissolved in a minimum amount of CHCl₃ and subjected to silica gel column chromatography, eluted with hexanes:EtOAc (2:1), to give 20 (61.84 mg, 78%); mp 118°C (EtOAc:hexanes) (lit. (27, 33) mp 118–119°C); $[\alpha]_D$ +100.2 (c 0.11, CHCl₃) (lit. $[\alpha]_D$ (27) -103.82 (c 0.31, CHCl₃); [α]_D (33) -37 (c 0.11, CHCl₃)); IR (CHCl₃) ν _{max}: 1775 (C=O), 1610, 1510, 1500 (C=CAr), 1245 (C-O) cm⁻¹; 1 H NMR δ: 6.80 (br s, 1H, H-2), 6.70 (br s, 2H, H-5 and H-6), 5.95 (s, 2H, OC H_2 O), 4.57 (d, 1H, $J_{7.8}$ = 6.8, H-7), 4.46 (dd, 1H, $J_{9eq,9ax} = 6.8$, $J_{8,9eq} = 9.7$, H-9eq), 4.35–4.25 (m, 2H, H-9'ax and H-9ax), 4.14 (dd, 1H, $J_{9'eq,9'ax} = 3.6$, $J_{8',9'eq} = 9.3$, H-9'eq), 3.40 (ddd, 1H, $J_{7,8'} = 3.6$, $J_{8',9'eq} = 9.3$, $J_{8',8} = 9.0$, H-8'), 3.05 (m, 1H, H-8); 13 C NMR δ : 178.16 (C-7'), 148.11 (C-3), 147.60 (C-4), 132.73 (C-1), 119.57 (C-6), 108.22 (C-5), 106.27 (C-2), 101.19 (OCH₂O), 85.92 (C-7), 69.93 and 69.77 (C-9/C-9'), 48.86 (C-8), 46.58 (C-8') ppm; MS m/e (relative

%): 248 (M⁺, 100), 218 (M - CH₂O⁺, 22), 163 (83), 150 (Ar₂CHO⁺, 80), 149 (Ar₂CO⁺, 100), 135 (Ar₂CH₂⁺, 82), 121 (Ar₂⁺, 38); HRMS, calcd. for C₁₃H₁₂O₅: 248.0685 (M⁺); found: 248.0683.

4-Benzyloxy-3-methoxy phenol 22

Phenol **22** (139.5 mg, 0.64 mmol) was prepared from *O*-benzylvanillin **21** (242 mg, 1 mmol) (23) in 64% overall yield exactly as described (34, 35).

O-Benzyl-epi-sesamolinol 23

To a solution of (+)-samin 19 (29.8 mg, 0.12 mmol) in dry THF (10 mL), PPh₃ (31.6 mg, 0.12 mmol), and protected phenol 22 (26.2 mg, 0.12 mmol) was added diethylazodicarboxylate (DEAD) (19 μ L, 1 equiv.). The mixture was stirred at r.t. for 24 h and the solvent removed in vacuo. The residue was next subjected to silica gel column chromatography, eluted with hexanes: EtOAc 3:1 (v/v), to give O-benzyl epi-sesamolinol **23** (44.8 mg, 80%). IR (CHCl₃) ν_{max} : 1600, 1510, 1450 (C=CAr), 1250, 1220 (C-O) cm⁻¹; ¹H NMR δ: 7.50–7.20 (m, 5H, Bn), 6.90–6.55 (m, 6H, CHAr), 5.94 (s, 2H, OCH₂O), 5.62 (d, 1H, $J_{7',8'}$ = 5.6, H-7'), 5.10 (s, 2H, C H_2 Ph), 4.80 (d, 1H, $J_{7.8} = 6.8$, H-7), 4.45–4.38 (m, 1H, H-9'ax), 4.20–4.10 (m, 1H, H-9'eq), 4.15-4.10 (m, 1H, H-9ax), 4.05-3.95 (m, 1H, H-9eq), 3.87 (s, 3H, OC H_3), 3.35–3.25 (m, 1H, H-8'), 3.05–2.90 (m, 1H, H-8); ¹³C NMR δ: 151.95, 151.14, 150.56, 147.92, 147.13 (C-3, C-4, C-1', C-3', C-4'), 134.79 (Cq/Bn), 143.31 (C-1), 128.42, 128.38, 127.37 (CH/Bn), 119.42 (C-6), 115.27 (C-5'), 108.13 (C-5), 107.03 (C-6'), 106.39 (C-2), 102.65 (C-7'), 102.41 (C-2'), 101.02 (OCH₂O), 85.31 (C-7), 71.83 (CH₂Ph), 69.40 (C-9'), 67.08 (C-9), 55.93 (OCH₃), 52.85 (C-8), 51.09 (C-8'); MS m/e (relative %): 462 (M+, 5), 243 (20), 230 (70), 203 (30), 165 (15), 149 (Ar₂CO⁺, 15), 135 (Ar₂CH₂⁺, 100), 121 (Ar_2^+ , 70), 105 (15); HRMS, calcd. for $C_{27}H_{26}O_7$: 462.1679; found: 462.1671.

(+)-Epi-sesamolinol 15

(+)-O-Benzyl epi-sesamolinol 23 (59.2 mg, 0.13 mmol) in EtOAc (5 mL) was treated with Pd/C 10% (15 mg). The solution was then stirred for 3 h under H₂ atmosphere, following which it was filtered over a short path silica gel column eluted with EtOAc. The filtrate was removed in vacuo, and the resulting residue purified by silica gel column chromatography, eluted with EtOAc:hexanes 2:1 (v/v), to give (+)-epi-sesamolinol 15 (29 mg, 75%). $[\alpha]_D$ +182 (c 0.996 g/100 mL in CHCl₃); IR (CHCl₃) ν_{max} : 3425 (br, OH), 1613, 1510, 1490, 1448 (C=CAr), 1234, 1196 cm⁻¹; UV λ_{max} : 230, 285 nm; ¹H NMR δ : 6.90–6.70 (m, 6H), 5.95 (s, 2H, OC H_2 O), 5.59 (d, 1H, $J_{7',8'} = 5.6$, H-7'), 5.38 (br s, 1H, OH), 4.78 (d, 1H, $J_{7.8} = 7$, H-7), 4.50-4.35 (m, 1H, H-9'ax), 4.20-4.10 (m, 1H, H-9'eq), 4.10–4.08 (m, 1H, H-9ax), 4.05–3.95 (m, 1H, H-9eq), 3.87 (s, 3H, OC H_3), 3.35–3.20 (m, 1H, H-8'), 3.05-2.95 (m, 1H, H-8); ¹³C NMR δ: 150.69 (C-1'), 147.88 (C-3), 147.07 (C-4), 146.80 (C-3'), 140.82 (C-4'), 134.90 (C-1), 119.36 (C-6), 114.13 (C-5'), 108.56 (C-5), 108.26 (C-6'), 106.54 (C-2), 101.92 (C-7'), 101.15 (OCH₂O), 100.98 (C-2'), 85.44 (C-7), 69.47 (C-9) 67.23 (C-9'), 56.08 (OCH₃), 53.08 (C-8'), 51.26 (C-8); MS m/e (relative %): 372 (M⁺, 5), 273 (5), 233 (28), 203 $(Ar_2C_5H_6O^+, 45)$, 149 $(Ar_2CHO^+, 30)$, 140 (70), 135 (Ar₂CH₂⁺, 100), 121 (Ar₂⁺, 25), 105 (18); HRMS, calcd. for C₂₀H₂₀O₇: 372.1209; found: 372.1194.

Lactone 26

To a solution of LDA (3.18 mmol, 1.1 equiv.), freshly prepared from diisopropyl amine (0.42 mL) and 1.6 M n-BuLi in hexanes (1.99 mL) at -15°C, in dry THF (10 mL) under argon at -78°C (acetone - Dry Ice bath), was added dropwise 4vinyl butyrolactone 25 (323.5 mg, 2.88 mmol, 1 equiv.) in dry THF (6 mL) over 30 min. The resulting mixture was stirred at -78° C for 30 min, following which O-benzylvanillin 21 (578.5 mg, 2.39 mmol) in dry THF (5 mL) was added dropwise over 30 min with the mixture allowed to stir at -78°C for an additional 3.5 h. A saturated solution of NH₄Cl_(aq) (0.5 mL) was added, and the resulting mixture was then allowed to warm to r.t. The mixture was sequentially extracted with diethyl ether (2 \times 15 mL) and EtOAc (2 \times 15 mL), with the combined organic layers dried (Na2SO4), evaporated in vacuo, and the resulting residue chromatographed on a silica gel column, eluted with hexanes: EtOAc 2:1 (v/v), to give the diastereoisomeric lactone 26 (779.7 mg, 92%) mixture, which was not resolved further. IR (CHCl $_3$) ν_{max} : 3492 (br, OH), 1762 (C=O), 1593, 1515, 1455 (C=CAr) cm $^{-1}$; 1 H NMR δ : 7.40– 7.18 (m, 5H, Bn), 6.88-6.85 (m, 3H, CHAr), 5.30-5.15 (m, 1H, =CH), 4.85-4.55 (m, 1H, CHOH), 4.35-4.15 (m, 2H, $=CH_2$), 4.15–3.90 (m, 2H, CH_2OCO), 3.74 (s, 3H, OCH_3), 3.25-3.10 and 2.85-2.70 (m, 1H, =CH-CH), 2.70-2.55 (m, 1H, CO-CH); ¹³C NMR δ: 177.72, 177.46 (CO), 149.17, 149.07, 147.42, 146.76 (Cq-O/Ar), 136.69, 136.65 (Cq/Bn), 135.78, 134.46 (CH₂=CH), 134.26, 131.92 (Cq/Ar), 128.04, 127.38, 126.92 (CH/Bn), 118.84, 117.46 (CH/Ar), 113.36, 113.16 (CH/Ar), 116.96, 116.38 (CH₂=), 110.05, 109.27 (CH/Ar), 72.90, 69.93 (CHOH), 70.44, 70.35 (CH₂O), 70.42 (CH₂Ph), 55.97 (OCH₃), 55.47, 51.58 (CH₂CO), 41.96, 38.58 (CH-CH=); MS m/e (relative %): 354 (M⁺, 90), 337 (65), 336 (27), 315 (30), 243 (100), 151 (Ar₁CO⁺, 80), 123 (Ar₁⁺, 50); HRMS, calcd. for C₂₁H₂₂O₅: 354.1467; found: 354.1486.

Triol 27

To a solution of LiAlH₄ (51.2 mg, 1.5 equiv.) in dry THF (10 mL) at 0°C, under an atmosphere of argon, was added dropwise a solution of lactone 26 (302.1 mg, 0.85 mmol) in dry THF (15 mL). The temperature was next raised to 50°C, this being maintained for an additional 1.5 h. After cooling to r.t., the solution was neutralized with saturated NH₄Cl_(aq) and extracted with CHCl₃ (3 × 10 mL). The organic solubles were combined, dried (Na₂SO₄), evaporated to dryness in vacuo, and the resulting residue purified by chromatography on silica gel, eluted sequentially with hexanes:EtOAc 2:1 (v/v) and EtOAc, to give the diastereoisomeric triol 27 (258 mg, 89%) mixture, which was not resolved further. IR (CHCl3) ν_{max} 3310 (br, OH), 1640 (C=C), 1606, 1593, 1510, 1455 (C=C, C=CAr), 1261, 1219 (C-O) cm⁻¹; ¹H NMR δ: 7.35–7.20 (m, 5H, Bn), 6.90-6.80 and 6.80-6.70 (m, 3H, CHAr), 5.90-5.50 (m, 1H, = CH), 5.20-5.00 (m, 2H, = CH₂), 5.07 (s, 1H) and5.06 (s, 1H, CH₂-Ph), 5.00–4.70 (m, 1H, CHOH), 3.81 (s, 3H, OCH_3), 3.80–3.30 (m, 4H, CH_2OH), 2.60–2.20 (m, 1H, CH_2OH) CHOH), 2.10–1.60 (m, 1H, =CH-CH); 13 C NMR δ : 149.29, 149.20 (C-4), 146.99, 146.90 (C-3), 138.52, 138.40 (CH=), 136.86, 136.05 (C-1, Cq/Bn), 128.32, 127.67, 127.15 (CH/ Bn), 118.40, 117.95 (C-6), 117.24, 116.30 (CH₂==), 113.59, 113.46 (C-5), 109.87, 109.49 (C-2), 74.55, 72.16 (CHOH), 70.82 (CH₂-Ph), 63.06, 62.50, 59.96, 59.53 ($2 \times CH_2OH$), 55.77 (OCH₃), 50.06, 48.07 (=CH-CH), 44.87, 43.05 (CH- CHOH); MS m/e (relative %): 340 (M⁺ - H₂O⁺, 19), 322 (100), 291 (19), 275 (34), 231 (85), 201 (81), 167 (87), 149 (79), 128 (81); HRMS, calcd. for $C_{21}H_{26}O_5$: 340.1675 (M - H₂O⁺); found: 340.1686.

Tetrahydrofuran 28

To a solution of the diastereoisomeric triol mixture 27 (189) mg, 0.55 mmol) in dry CH₂Cl₂ (5 mL) was added TMSBr (140 μL, 2 equiv.). The resulting solution was then stirred at r.t. for 15 min, neutralized with saturated NaHCO_{3(aq)} (0.5 mL), dried (Na₂SO₄), with the organic solvent evaporated in vacuo and the residue subjected to silica gel column chromatography, eluted with hexanes:EtOAc 2:1 (v/v), to give tetrahydrofuran **28** (159.3 mg, 89%). IR (CHCl₃) ν_{max} : 3300 (br, OH), 1640 (C=C), 1606, 1594, 1515, 1465 (C=CAr), 1263, 1225 (C-O) cm⁻¹; ¹H NMR δ : 7.45–7.30 (m, 5H, Bn), 6.93 (d, 1H, J = 1.7, CHAr), 6.84–6.78 (m, 1H, CHAr), 5.97–5.90 (m, 1H, =CH), 5.22-5.14 (m, 2H, $=CH_2$), 5.16 (s, 2H, CH_2 -Ph), 4.70 (d, 1H, J = 7.3, ArCHO), 4.23 and 3.83 (dd, 2H, J = 6.7, 8.6, CH₂OC), 3.90 (s, 3H, OCH₃), 3.75-3.69 (m, 1H) and 3.64-3.60 (m, 1H, CH_2OH), 3.15–3.05 (m, 1H, =-CH-CH), 2.40–2.30 (m, 1H, CH-CH₂OH); ¹³C NMR δ: 149.63 (C-4), 147.49 (C-3), 137.07 (Cq/Bn), 135.57 (CH), 135.38 (C-1), 128.44, 127.73, 127.19 (CH/Bn), 118.10 (C-6), 117.32 (CH₂==), 113.72 (C-5), 109.47 (C-2), 82.41 (C-7), 72.61 (C-9'), 70.94 (CH₂-Ph), 60.76 (C-9), 55.90 (OCH₃), 53.41 (C-8), 45.62 (C-8'); MS m/e (relative %): 322 (M - H_2O^+ , 100), 293 (10), 268 (5), 231 (95), 201 (73), 171 (55), 149 (63), 123 (Ar₁⁺, 65); HRMS, calcd. for $C_{21}H_{24}O_4$ (M - H_2O^+): 322.1569; found: 322.1558.

Triol 29

To a solution of tetrahydrofuran 28 (144 mg, 0.45 mmol) in THF (3 mL) was added OsO₄ (2.5% equiv., 108 μ L of a 2.5% soln. in t-BuOH) and N-methylmorpholine-N-oxide (NMO) (1.4 equiv., 124 μ L of a 60% soln. in H₂O). The suspension was stirred for 2 days at r.t., following which it was extracted with CHCl₃ (3×10 mL). The organic solubles were combined, dried (Na₂SO₄), the solvent removed in vacuo, and the resulting residue chromatographed on silica gel, eluted with hexanes:EtOAc 2:1 (v/v), to give the triol 29 (132.1 mg, 79%). IR $(CHCl_3)$ ν_{max} : 3390 (br, OH), 1607, 1594, 1515, 1455 (C=CAr), 1263, 1217 (C-O) cm⁻¹; ¹H NMR δ : 7.40–7.20 (m, 5H, Bn), 6.90–6.70 (m, 3H, CHAr), 5.14 (s, 2H, CH₂-Ph), 4.79 (d, J = 6.4) and 4.51 $(d, J = 3.8, 1H, OCH-Ar), <math>4.1\bar{0}-3.90$ (m, J = 6.4)1H, CHOH-CH₂OH), 3.90-3.30 (m, 6H, CH₂OH and CH₂O), 3.83 (s, 3H, OC H_3), 2.60-2.30 (m, 1H, CHCHOH), 2.60-2.35and 2.30-2.20 (m, 1H, CHCH₂OH); ¹³C NMR δ: 149.51 (C-4), 147.47 (C-3), 136.84 (Cq/Bn), 135.07, 134.98 (C-1), 128.38, 127.73, 127.20 (CH/Bn), 117.96, 117.81 (C-6), 113.62 (C-5), 109.41 (C-2), 82.81, 82.11 (C-7), 70.87 (CH₂-Ph), 70.46, 70.35 (CHOH), 68.93, 68.25 (C-7') 65.71, 65.39 (CH₂OH), 60.67, 59.53 (C-9), 55.91 (OCH₃), 51.15, 50.74 (C-8), 43.86, 42.54 (C-8'); MS m/e (relative %): 374 (M⁺, 83), 356 (80), 338 (50), 295 (50), 283 (20), 265 (90), 175 (78), 151 (Ar₁CO⁺, 80), 123 (Ar₁⁺, 100), 107 (90); HRMS, calcd. for C₂₁H₂₆O₆: 374.1726; found: 374.1741.

Furanofuran-1-ol 24

To a solution of triol **29** (125.1 mg, 0.334 mmol) in MeOH (3 mL) at 0° C was added dropwise a solution of NaIO₄ (85.7 mg, 1.2 equiv.) in MeOH–H₂O (6.5 mL, 10:3) over 10 min. The

temperature was raised to r.t., with the mixture allowed to stir for an additional 12 h, and then filtered. The resulting filtrate was extracted with CHCl₃ (3×10 mL) with the organic layers combined and dried (Na₂SO₄), and the solution evaporated to dryness in vacuo. The residue was subjected to chromatography on silica gel, eluted with hexanes: EtOAc 2:1 (v/v), to give the furanofuranol **24** (101.8 mg, 89%). IR (CHCl₃) ν_{max} : 3410 (br, OH), 1594, 1515, 1455 (C=CAr) cm⁻¹; ¹H NMR δ : 7.45– 7.20 (m, 5H, Bn), 6.95 (d, 1H, $J_{2.6} = 1.7$, H-2), 6.75–6.65 (m, 2H, H-5, H-6), 5.16 (s, 2H, CH₂-Ph), 5.35 (s, 1H, H-7'), 4.38 (d, 1H, $J_{7,8}$ = 7.1, H-7), 4.39 (dd, 1H, $J_{9ax,9eq}$ = 7, $J_{8,9ax}$ = 8.9, H-9ax), 4.25-4.10 (m, 1H, H-9'ax), 3.95–3.85 (m, 1H, H-9'ax) 9'eq), 3.56 (dd, 1H, $J_{9ax,9eq} = 7$, $J_{8,9eq} = 9.1$, H-9 eq), 3.91 (s, 3H, OC H_3), 3.76 (br s, 1H, OH), 3.11–2.87 (m, 2H, H-8, H-8'), ¹³C NMR δ: 149.78 (C-3), 147.82 (C-4), 136.99 (*C*q/Bn), 133.51 (C-1), 128.47, 127.78, 127.20 (CH/Bn), 118.47 (C-6), 113.69 (C-5), 109.61 (C-2), 102.02 (C-7'), 85.79 (C-7), 71.21 (C-9), 70.93 (CH₂Ph), 69.13 (C-9'), 55.95 (OCH₃), 53.51 (C-8'), 52.52 (C-8); MS m/e (relative %): 342 (M⁺, 20), 251 (10), 223 (10), 193 (5), 151 (Ar₁CO⁺, 6), 137 (Ar₁CH₂⁺, 5), 123 $(Ar_1^+, 5)$, 91 (100); HRMS, calcd. for $C_{20}H_{22}O_5$: 342.1467; found: 342.1475.

Furolactone 30

To a solution of 24 (34.41 mg, 0.1 mmol) in dry CH₂Cl₂ (5 mL) was added pyridinium dichromate (PDC) (4 equiv., 151.4 mg), AcOH (1 equiv., 5.76 μL), and 4Å molecular sieves (150 mg). Following stirring for 6 h at r.t., the suspension was filtered through a short-path silica gel plug and the filtrate removed in vacuo to give a residue that was then subjected to silica gel column chromatography, eluted with hexanes:EtOAc 2:1 (v/v), to give 30 (26.7 mg, 78%); mp 114-115°C; IR (CHCl₃) ν_{max} : 1772 (C=O), 1594, 1516, 1456 (C=CAr), 1244 (C-O) cm⁻¹; ¹H NMR δ : 7.45–7.25 (m, 5H, Bn), 6.93 (d, 1H, J = 1.8, H-2), 6.87 (d, 1H, $J_{5.6} = 8.2$, H-5), 6.79 (dd, 1H, $J_{2,6} = 1.8$, $J_{5,6} = 8.2$, H-6), 5.17 (s, 2H, Ar-C H_2), $4.62 \text{ (d, 1H, } J_{7,8} = 6.7, \text{H-7}), 4.50 \text{ (dd, 1H, } J_{9eq,8} = 9.8, J_{9eq,9ax} = 4.62 \text{ (do. 1H, } J_{9eq,9ax} = 9.8) \text{ (do. 1H, } J_{9eq,9ax} = 9.8 \text{ (do. 1H, } J_{9eq,9ax} = 9.8) \text{ (do. 1H, } J_{9eq,9ax} = 9.8 \text{ (do. 1H, } J_{9eq,9ax} = 9.8) \text{ (do. 1H, } J_{9eq,9ax} = 9.8 \text{ (do. 1H, } J_{9eq,9ax} = 9.8) \text{ (do. 1H, } J_{9eq,9ax} = 9.8 \text{ (do. 1H, } J_{9eq,9ax} = 9.8) \text{ (do. 1H, } J_{9eq,9ax} = 9.8 \text{ (do. 1H, } J_{9eq,9ax} = 9.8) \text{ (do. 1H, } J_{9eq,9ax} = 9.8 \text{ (do. 1H, } J_{9eq,9ax} = 9.8) \text{ (do. 1H, } J_{9eq,9ax} = 9.8 \text{ (do. 1H, } J_{9eq,9ax} = 9.8) \text{ (do. 1H, } J_{9eq,9ax} = 9.8 \text{ (do. 1H, } J_{9eq,9ax} = 9.8) \text{ (do. 1H, } J_{9eq,9ax} = 9.8 \text{ (do. 1H, } J_{9eq,9ax} = 9.8) \text{ (do. 1H, } J_{9eq,9ax} = 9.8 \text{ (do. 1H, } J_{9eq,9ax} = 9.8) \text{ (do. 1H, } J_{9eq,9ax} = 9.8 \text{ (do. 1H, } J_{9eq,9ax} = 9.8) \text{ (do. 1H, } J_{9eq,9ax} = 9.8) \text{ (do. 1H, } J_{9eq,9ax} = 9.8) \text{ (do. 1H, } J_{9eq,9ax} = 9.8)$ 6.8, H-9 eq), 4.40-4.30 (m, 2H, H-9ax and H-9'ax), 4.20 (dd, 1H, $J_{8',9'\text{eq}} = 9.2$, $J_{9'\text{eq},9'\text{ax}} = 3.8$, H-9' eq), 3.92 (s, 3H, OC H_3), 3.45 (ddd, 1H, $J_{8',9'\text{ax}} = 9.2$, $J_{8',8} = 9.2$, $J_{8',9'\text{eq}} = 3.8$, H-8'), 3.15–3.10 (m, 1H, H-8); ^{13}C NMR δ : 178.13 (C-7'), 149.94 (C-4), 148.17 (C-5), 136.81 (Cq/Bn), 131.81 (C-1), 128.49, 127.82, 127.13 (CH/Bn), 118.29 (C-6), 113.70 (C-5), 109.43 (C-2), 85.89 (C-7), 70.89 (CH₂Ph), 69.93 and 69.77 (C-9 and C-9'), 55.98 (OCH₃), 48.03 (C-8'), 45.92 (C-8); MS m/e (relative %): 340 (M⁺, 100), 250 (20), 221 (45), 149 (48); HRMS, calcd. for $C_{20}H_{20}O_5$: 340.1311; found: 340.1319.

(±)-Salicifoliol 31

To a solution of **30** (26.7 mg, 0.078 mmol) in EtOAc (5 mL) was added Pd/C 10% (15 mg). The suspension was then stirred under an H₂ atmosphere, as described for epi-(+)-sesamolinol **15**, with salicifoliol **31** purified by silica gel column chromatography, eluted with hexanes:EtOAc (2:1 \rightarrow 1:1 (v/v)), to give **31** (17.66 mg, 90%); mp 101–102°C (lit. (39) mp 102–103°C); IR (CHCl₃) ν_{max} : 3417 (br, OH), 1768 (C=O), 1605, 1519, 1466 (C=CAr), 1241 (C-O) cm⁻¹; UV λ_{max} : 234, 278 nm; ¹H NMR δ: 6.93 (d, 1H, J = 6.9, H-5), 6.89 (d, 1H, J = 1.6, H-2), 6.80 (dd, 1H, J_{2,6} = 1.6, J_{5,6} = 8, H-6), 5.74 (s, 1H, OH), 4.61 (d, 1H, J_{7,8} = 6.9, H-7), 4.51 (dd, 1H, J_{9eq,8} = 9.8, J_{9eq,9ax} = 6.8, H-9ax), 4.40–4.30 (m, 1H, H-9eq and H-9'ax), 4.19 (dd,

1H, $J_{8',9'eq} = 9.2$, $J_{9'eq,9'ax} = 3.8$, H-9' eq), 3.90 (s, 3H, OC H_3), 3.43 (ddd, $J_{8',9'eq} = 9.2$, $J_{8',8} = 9.2$, $J_{8',9'ax} = 3.8$, 1H, H-8'), 3.18–3.08 (m, 1H, H-8); 13 C NMR δ : 178.32 (C-7'), 146.98 (C-3), 145.88 (C-4), 130.63 (C-1), 119.15 (C-6), 114.47 (C-5), 105.51 (C-2), 86.13 (C-7), 70.01 and 69.91 (C-9/C-9'), 56.02 (OC H_3), 48.21 (H-8), 45.99 (H-8'); MS m/e (relative %): 250 (M⁺, 100), 233 (10), 219 (15), 205 (Ar₁C₅H₆O⁺, 10), 165 (50), 152 (Ar₁CHO⁺, 90), 151 (Ar₁CO⁺, 100), 137 (Ar₁CH₂⁺, 95), 123 (Ar₁⁺, 50), 109 (75); HRMS, calcd. for C₁₄H₁₄O₅: 250.0841; found: 250.0843.

Furofuran 32

To a solution of lactol 24 (101.1 mg, 0.3 mmol) in dry THF (30 mL), PPh₃ (79 mg, 0.3 mmol), and 4-benzyloxy-3-methoxyphenol 22 (65.4 mg, 0.3 mmol) was added diethylazodicarboxylate (DEAD) (47.5 µL, 1 equiv.), and the mixture was allowed to stir at r.t. for an additional 24 h. The solvent was then removed in vacuo and the residue subjected to silica gel column chromatography, eluted with hexanes:EtOAc (3:1 \rightarrow 2:1 (v/v)), to give the furofuran lignan 32 (147.9 mg, 89%). IR (CHCl₃) ν_{max} : 1597, 1510, 1455 (C=CAr) cm⁻¹; ¹H NMR δ : 7.43-7.25 (m, 10H, Bn), 6.95 (d, 1H, $J_{2,6} = 1.5$, H-2), 6.85– 6.70 (m, 2H, CHAr), 6.70 (d, 1H, $J_{2',6'}$ = 2.7, H-2'), 6.60 (dd, 1H, J = 2.7, H-6'), 5.59 (d, 1H, J = 5.6, H-7'), 5.13 (s, 2H, CH_2Ph), 5.08 (s, 2H, CH_2Ph), 4.78 (d, 1H, $J_{7',8'}$ = 6.9, H-7), 4.38 (dd, 1H, $J_{9'ax,8'} = 9.1$, $J_{9'eq,9'ax} = 4.7$, H-9'ax), 4.20–4.00 (m, 2H, H-9'eq and H-9ax), 3.95 (dd, $J_{8,9eq} = 9.1$, $J_{9eq,9ax} = 4.7$, H-9eq), 3.89 (s, 3H, OC H_3), 3.86 (s, 3H, OC H_3), 3.35–3.20 (m, 1H, H-8'), 3.04 (m, 1H, H-8); 13 C NMR δ : 151.95 (C-1'), 150.53 (C-3'), 149.72 (C-3), 147.63 (C-4), 143.27 (C-4'), 137.31 (Cq/Bn), 137.02 (Cq/Bn), 133.92 (C-1), 128.47, 128.42, 127.71, 127.70, 127.35, 127.20 (CH/Bn), 118.17 (C-6), 115.53 (C-5), 113.80 (C-5'), 109.63 (C-6'), 107.06 (C-2), 102.63 (C-7'), 102.47 (C-2'), 85.21 (C-7), 71.82 (CH₂Ph), 70.96 (CH₂Ph), 69.38 (C-9), 67.04 (C-9'), 55.93 (OCH₃), 55.92 (OCH₃), 52.78 (C-8), 51.15 (C-8'); MS m/e (relative %): 554 (M⁺, 5), 464 (3), 445 (2), 429 (2), 325 (10), 295 (30), 243 (50), 230 (100), 139 (75); HRMS, calcd. for $C_{34}H_{34}O_7$: 554.2304; found: 554.2318.

(±)-Epi-pinoresinolin 13

To a solution of the protected furolignan **32** (69.7 mg, 0.125 mmol) in EtOAc (5 mL) was added Pd/C 10% (20 mg). The solution was then stirred for 3 h under an H_2 atmosphere, as for epi-sesamolinol **15**, to give epi-pinoresinolin **13** (39.4 mg, 84%). IR (CHCl₃) ν_{max} : 3410 (br, OH), 1613, 1511, 1452 (C=CAr) cm⁻¹; ¹H and ¹³C NMR spectra (see Table 1); MS *m/e* (relative %): 374 (M⁺, 35), 275 (22), 235 (90), 205 (Ar₁C₅H₆O⁺, 95), 137 (Ar₁CH₂⁺, 100); HRMS, calcd. for $C_{20}H_{22}O_7$: 374.1366; found: 374.1387.

Furofuran 34

The furanofuranol **24** (17.7 mg, 0.05 mmol) was treated with sesamol **33** (7.01 mg, 0.05 mmol) as described above to give **34** (20.3 mg, 85%). IR (CHCl₃) ν_{max} : 1600, 1508, 1466 (C=CAr), 1245, 1217 (C-O) cm⁻¹; ¹H NMR δ : 7.45–7.20 (m, 5H, Bn), 6.90–6.50 (m, 6H, CHAr), 5.93 (s, 2H, OCH₂O), 5.56 (d, 1H, $J_{7',8'}$ = 5.6, H-7'), 5.15 (m, 2H, CH₂Ph), 4.77 (d, 1H, $J_{7,8}$ = 7, H-7), 4.38 (dd, 1H, $J_{9'\text{ax},8'}$ = 9.1, $J_{9'\text{eq},9'\text{ax}}$ = 4.2, H-9'ax), 4.20–4.05 (m, 2H, H-9'eq and H-9ax), 3.97 (dd, 1H, $J_{8,9\text{eq}}$ = 9.1, $J_{9\text{eq},9\text{ax}}$ = 4.8, H-9eq), 3.91 (s, 3H, OCH₃), 3.38–

3.25 (m, 1H, H-8'), 3.10–2.95 (m, 1H, H-8); 13 C NMR δ : 152.15 (C-1'), 149.75 (C-3'), 148.03 (C-3), 147.68 (C-4), 143.55 (C-4'), 136.95 (Cq/Bn), 133.87 (C-1), 128.46, 127.74, 127.16 (CH/Bn), 118.13 (C-6), 113.75 (C-5), 109.58 (C-6'), 108.93 (C-5'), 107.95 (C-2), 102.94 (C-7'), 101.18 (OCH_2O), 100.14 (C-2'), 85.17 (C-7), 70.95 (CH_2Ph), 69.30 (C-9), 66.96 (C-9'), 55.95 (OCH_3), 52.70 (C-8), 51.11 (C-8'); MS m/e (relative %): 462 (M^+ , 15), 325 (30), 295 (70), 243 (55), 227 (100), 203 ($Ar_2C_5H_6O^+$, 30), 179 (12), 137 ($Ar_1CH_2^+$, 55), 107 (15); HRMS, calcd. for $C_{27}H_{26}O_7$: 462.1678; found: 462.1671.

(±)-Epi-piperitolin 14

O-Benzyl piperitolin 34 (59.2 mg, 0.128 mmol) in EtOAc (5 mL) was treated with Pd/C 10% (15 mg), as described for furofuran 32, to give (\pm)-14 (41.4 mg, 95 %); mp 116–117°C; IR (CHCl₃) ν_{max} : 3446 (br, OH), 1612, 1547, 1488 (C=CAr), 1271, 1243 (C-O) cm⁻¹; UV λ_{max} : 230, 285 nm; ¹H NMR δ : 6.90 (d, 1H, $J_{2.6}$ = 1.8, H-2), 6.81 (dd, 1H, $J_{6.2}$ = 1.8, $J_{5.6}$ = 8.1, H-6), 6.8 (d, 1H, $J_{5,6}$ = 8.1, H-5), 6.72 (d, 1H, $J_{5',6'}$ = 8.4, H-5'), 6.69 (d, 1H, $J_{2',6}$ = 2.4, H-2'), 6.56 (dd, 1H, $J_{2',6'}$ = 2.4, $J_{5',6'}$ = 8.4, H-6'), 5.92 (s, 2H, OCH₂O), 5.74 (s, 1H, OH), 5.55 (d, 1H, $J_{7',8'} = 5.6$, H-7'), 4.47 (d, 1H, $J_{7.8} = 7$, H-7), 4.40–4.36 (m, 1H, H-9'ax), 4.17-4.07 (m, 2H, H-9'eq and H-9ax), 4.02-3.95 (m, 1H, H-9eq), 3.89 (s, 3H, OCH3), 3.33-3.28 (m, 1H, H-8'), 3.06–3.00 (m, 1H, H-8); ¹³C NMR δ: 152.28 (C-1'), 148.15 (C-3'), 146.77 (C-3), 145.33 (C-4'), 142.79 (C-4'), 132.79 (C-1), 118.99 (C-6), 114.34 (C-5), 109.14 (C-6'), 108.64 (C-2), 108.04 (C-5'), 103.07 (C-7'), 100.23 (OCH2O), 85.48 (C-7), 69.40 (C-9), 67.04 (C-9'), 55.97 (OCH₃), 52.87 (C-8'), 51.22 (C-8); MS m/e (relative %): 372 (M⁺, 15), 261 (5), 235 (60), 205 ($Ar_1C_5H_6O^+$, 65), 187 (5), 161 ($Ar_1C_3H_4^+$, 5), 159 (4), 153 (Ar₂CHO⁺, 30), 151 (Ar₁CHO⁺, 50), 138 (90), 137 (Ar₂CH₂⁺, 100), 115 (30), 107 (60); HRMS, calcd. for $C_{20}\tilde{H}_{20}\tilde{O}_7$: 372.1209; found: 372.1219.

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