

RESEARCH ARTICLE

Fluorinated pyrrolidines and piperidines incorporating tertiary benzenesulfonamide moieties are selective carbonic anhydrase II inhibitors

Alexandre Le Darz^{1,2}, Agnès Mingot², Fodil Bouazza¹, Ugo Castelli², Omar Karam¹, Muhammet Tanc³, Claudiu T. Supuran³, and Sébastien Thibaudeau²

¹rtMolecule, Poitiers, France, ²Superacid group in "Organic Synthesis" Team, Université de Poitiers, Poitiers Cedex 09, France, and

³Laboratorio di Chimica Bioinorganica, Università degli Studi di Firenze, Sesto Fiorentino, Firenze, Italy

Abstract

A series of substituted pyrrolidines and piperidines were synthesized using superacid HF/SbF₅ chemistry. Investigated as inhibitors of several human carbonic anhydrase (hCA, EC 4.2.1.1) isoforms, i.e. the cytosolic hCA I and II as well as the tumor-associated transmembrane isoforms hCA IX and XII, these compounds showed a never yet reported selectivity toward the human carbonic anhydrase hCA II. In the tertiary benzenesulfonamide family, this class of inhibitors points out a new mechanism of action for human carbonic anhydrase II inhibition.

Keywords

Carbonic anhydrase, fluorine, selectivity, superacid, tertiary benzenesulfonamides

History

Received 26 August 2014

Revised 3 September 2014

Accepted 3 September 2014

Published online 27 November 2014

Introduction

Carbonic anhydrases (CAs, EC 4.2.1.1) are widely distributed metalloenzymes in the living world and are involved in many biochemical processes that depend on the hydration/dehydration of carbon dioxide/bicarbonate^{1,2}. Human CAs (hCAs) exist in 15 isoforms, belonging to the α -class and differ in their cellular localization (cytosol, mitochondria and cell membrane) and catalytic activity³. They are involved in numerous cellular and physiological processes such as respiration, electrolyte secretion, pH homeostasis and biosynthetic reactions which require bicarbonate as substrate including lipogenesis, gluconeogenesis and ureagenesis, as well as tumorigenicity among others⁴. As a consequence, CAs are well-established therapeutic targets to treat a wide range of disorders through the use of inhibitors or activators^{5–7}. Classical CA inhibitors (CAIs) act by complexing the metal ion from the enzyme active site, as anions, and among the numerous families of CAIs, the primary sulfonamide is one of the most largely exploited, mainly considering the aromatic/heteroaromatic representatives of such compounds^{8–11}. To prevent undesired side effects, designing selective CAIs is now largely

accepted to be a privileged strategy^{12,13}. In this context, the so-called family of "non-zinc bonding inhibitors" has deeply increased over the last few years^{14–16}, especially in the quest of potent and selective anti-cancer agents (targeting the cancer-associated isoforms hCA IX and XII)^{17–21}. Our recent studies, focused on the synthesis of fluorinated compounds in superacid^{22–26}, allowed to identify a series of fluorinated tertiary benzenesulfonamides acting as selective hCA IX inhibitors^{27–29}. The peculiar, never reported behavior of these tertiary benzenesulfonamides was attributed to a new mode of inhibition, comparable to that of non-zinc binding inhibitors (e.g. the coumarins)^{30–32}, and the evaluation of their ¹⁸F-labeled analogue for imaging CA IX expression in tumors through PET imaging was recently reported³³. As a consequence, this new class of hCA inhibitors might now be considered as an innovative tool in the design of selective CAIs for further applications in medicinal chemistry. A strong impact of the modification of the geometrical shape of the 4-aminobenzenesulfonamides on the selectivity between cytosolic (hCA I and II) and transmembrane isoforms (hCA IX and XII) was identified with compounds **A** and **B** (Table 1). The impact of the presence of fluorine atom(s) substituents and sterically crowded aromatic rings on hCA inhibition selectivity was also predominant (compounds **C** and **D**). Applied to *N*-substituted saccharin derivatives as selective hCA XII inhibitors, studies based on similar strategies were also recently conducted by one of our groups, showing a dramatic effect of the nitrogen atom substitution pattern on CA inhibition and selectivity towards various CA isoforms (compounds **E** and **F**)³⁴.

In this context, the X-ray crystallographic structure of the adduct of human isozyme II with the antipsychotic drug sulpiride

Address for correspondence: S. Thibaudeau, Superacid group in "Organic Synthesis" Team, Université de Poitiers, CNRS UMR 7285 IC2MP, Bât. B28, 4 rue Michel Brunet, TSA 51106, 86073 Poitiers Cedex 09, France. Tel: +33-5-49454588. Fax: +33-5-49453501. E-mail: sebastien.thibaudeau@univ-poitiers.fr

C. T. Supuran, Università degli Studi di Firenze, Laboratorio di Chimica Bioinorganica, Rm 188, Via della Lastruccia 3, I-50019 Sesto Fiorentino (Firenze), Italy. Tel: + 39-055-4573005. Fax: + 39-055-4573385. E-mail: claudiu.supuran@unifi.it

Table 1. Inhibition data of selected tertiary benzenesulfonamides and saccharins against isoforms hCA I, II, IX and XII.

Entry	Substrate	K _i (μM)*			
		hCA I†	hCA II†	hCA IX‡	hCA XII‡
1¶	A	0.637	0.431	0.136	0.298
2¶	B	6.796	4.995	0.073	0.080
3§	C	0.073	>50	0.009	0.033
4	D	>100	>100	0.451	>100
5#	E	>50	>50	>50	1.780
6#	F	>50	0.042	0.150	0.440

*Errors in the range of $\pm 5\%$ of the reported data from three different assays.

†Recombinant isoforms^{35–37}.

‡Catalytic domain.

¶Values from Ref. ²⁹.

§Values from Ref. ²⁷.

||Values from Ref. ²⁸.

#Values from Ref. ³⁴.

strongly inspired this work³⁸. Sulpiride has been shown to selectively inhibit hCA II versus hCA I and the membrane-bound hCA IV. Furthermore, the binding of sulpiride to hCA II revealed strong stacking interactions between the *N*-methyl-pyrrolidine ring and the Phe 131 residue of the carbonic anhydrase II, a moiety known to be critical for the binding of inhibitors possessing cyclic side chains (Figure 1)³⁹.

In addition, the role of fluorine in medicinal chemistry is well recognized^{40,41}, and the evaluation of its impact on modification of nitrogen-containing proximal functionalities behavior is considered as a common strategy in drug design studies^{42–45}. Furthermore, recent studies revealed a strong effect of fluorine atom(s) on nitrogen containing heterocycles preferred conformations, with possible effect on bioactivities^{46–48}. Thus, we report here the synthesis of new fluorinated *N*-pyrrolidino and piperidino tertiary benzenesulfonamides, and their screening for the inhibition of cytosolic CA isoforms hCA I and II and transmembrane isoforms IX and XII. For the first time, hCA II

selective inhibitors with no measurable affinity for hCA I, IX and XII have been identified.

Materials and methods

Chemistry

The authors draw the reader's attention to the dangerous features of superacidic chemistry. Handling of hydrogen fluoride and antimony pentafluoride must be done by experienced chemists with all the necessary safety arrangements in place. Reactions performed in superacid were carried out in a sealed Teflon[®] flask with a magnetic stirrer. No further precautions have to be taken to prevent mixture from moisture (test reaction worked out in anhydrous conditions leads to the same results as expected). Yields refer to isolated pure products. ¹H, ¹³C and ¹⁹F NMR were recorded on a 400-MHz Bruker Advance DPX spectrometer using CDCl₃ as solvent and C₆F₆ as external reference. COSY ¹H–¹H and ¹H–¹³C experiments were used to confirm the NMR

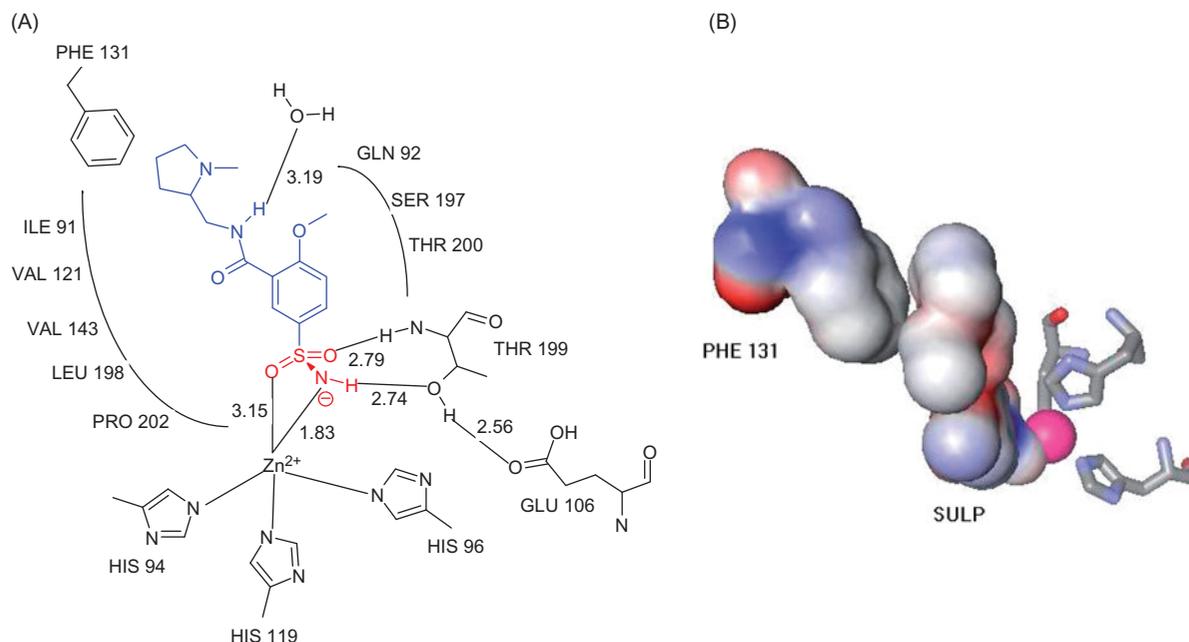


Figure 1. (A) Binding of sulphiride to hCA II active site. Figures represent distances in Å. (B) Representation of the π -stacking between the phenyl ring of Phe131 and the N-methylpyrrolidone fragment of the drug.

peaks assignments. Melting points were determined in a capillary tube with a device Büchi melting point B-545 and were uncorrected. All separations were done under flash-chromatography conditions on silica gel (15–40 μ m). High-resolution mass spectrometry (HRMS) spectra were performed at the Institut Lavoisier de Versailles (ILV) of the University of Versailles St Quentin (LTQ-Orbitrap Velos Pro, THERMOFISHER SCIENTIFIC), at Centre Régional de Mesures Physiques de l'Ouest (CRMPO) of the University of Rennes (Q-TOF 2, Waters), at the Institut de Chimie Organique et Analytique (ICOA) of the University of Orléans (Q-ToF MaXis, Bruker), at the Institut de Chimie des Milieux et des Matériaux de Poitiers (IC2MP) of the University of Poitiers (LC-QTOF MaXis Impact, Bruker). Optical activities (α^D) were analyzed on a Schmidt+Haensch polatron HH8 polarimeter in MeOH at 20 °C.

Methyl-(2S)-1-(4-nosyl)pyrrolidine-2-carboxylate (**2**)

Into a round-bottom flask, methyl ester L-proline (165 mg, 1 mmol) was solubilized in 10 mL of dichloromethane then pyridine (1 mL, 12 eq.) was added followed by 4-nosyl chloride (665 mg, 3 eq.). The mixture was magnetically stirred at room temperature overnight, then water was added and the aqueous layer was extracted with dichloromethane ($\times 3$). The combined organic phases were dried over magnesium sulfate, filtered and concentrated *in vacuo*. The reaction crude was purified over silica gel with the eluent ethyl acetate/petroleum ether: 30/70, thereby obtaining compound **2** (290 mg, 91%). Aspect: white solid. Mp: 111.5 °C. ^1H NMR (CDCl_3 , 400 MHz, ppm) δ : 8.35 (dm, 2H, $J = 9.0$ Hz), 8.06 (dm, 2H, $J = 9.0$ Hz), 4.44 (m, 1H), 3.69 (s, 3H), 3.44 (m, 2H), 2.15 (m, 1H), 2.08–1.84 (m, 3H). ^{13}C NMR (CDCl_3 , 100 MHz, ppm) δ : 172.2, 150.2, 144.7, 128.7, 124.3, 60.7, 52.6, 48.4, 31.0, 24.8. $\alpha^D = -107.9^\circ$.

(2S)-1-(4-nosyl)pyrrolidine-2-carbaldehyde (**3**)

In an argon-flushed round-bottom flask, compound **2** (5.66 g, 18 mmol) was solubilized in 45 mL of dichloromethane, cooled to -78°C then DIBALH (1 M in CH_2Cl_2 , 25.2 mL, 1.4 eq.) was slowly added and magnetically stirred for 1 h 40 min at the same

temperature. MeOH was then added and stirred overnight at room temperature after which the mixture was concentrated *in vacuo*. At the crude residue, an aqueous solution of HCl (1 M) and ice were added, the mixture was stirred 15 min then ethyl acetate was added, followed by Na_2CO_3 up to pH 9–10 and extracted with ethyl acetate ($\times 3$). The combined organic phases were dried over magnesium sulfate, filtered and concentrated *in vacuo*. The crude residue was purified by column chromatography over silica gel with the eluent ethyl acetate/petroleum ether: 30/70 thereby obtaining compound **3** (4.67 g, 91%). Aspect: pale yellow solid. Mp: degradation. ^1H NMR (CDCl_3 , 400 MHz, ppm) δ : 9.66 (d, 1H, $J = 2.0$ Hz), 8.41 (d, 2H, $J = 9.0$ Hz), 8.05 (d, 2H, $J = 9.0$ Hz), 4.01 (ddd, $J = 8.1$ Hz, $J = 4.9$ Hz, $J = 2.0$ Hz, 1H), 3.60 (m, 1H), 3.28 (m, 1H), 2.12 (m, 1H), 2.00–1.85 (m, 2H), 1.78 (m, 1H). ^{13}C NMR (CDCl_3 , 100 MHz, ppm) δ : 198.8, 150.6, 143.1, 129.0, 124.7, 66.9, 49.3, 27.7, 24.8.

(2S)-1-(4-nosyl)-2-vinylpyrrolidine (**4**)

In an argon-flushed double-necked round-bottom flask, methyl-triphenylphosphine bromide (11.29 g, 31.6 mmol) was dissolved in THF (100 mL), cooled to -78°C , then n-BuLi (1.6 M in hexane, 20 mL, 32.0 mmol) was slowly added. The reaction mixture was stirred 1 h at the same temperature, then a solution of compound **3** (4.49 g, 15.8 mmol) in THF (100 mL) was slowly added via cannula. The mixture was allowed to warm to room temperature and stirred for 1 h, after which pentane (110 mL) was added, followed by MeOH (220 mL). Solvents were removed *in vacuo*, then water was added and the aqueous layer was extracted with ethyl acetate ($\times 3$). The combined organic phases were washed with brine, dried over magnesium sulfate, filtered and concentrated *in vacuo*. The crude residue was purified by column chromatography over silica gel with the eluent ethyl acetate/petroleum ether 20/80 thereby obtaining compound **4** (2.43 g, 54%). Aspect: pale yellow solid. Mp: 82 °C. ^1H NMR (CDCl_3 , 400 MHz, ppm) δ : 8.35 (d, 2H, $J = 9.0$ Hz), 8.01 (d, 2H, $J = 9.0$ Hz), 5.71 (ddd, 1H, $J = 16.8$ Hz, $J = 10.2$ Hz, $J = 6.4$ Hz), 5.25 (dt, 1H, $J = 16.9$ Hz, $J = 1.2$ Hz), 5.13 (dt, 1H, $J = 10.2$ Hz, $J = 1.2$ Hz), 4.24 (m, 1H), 3.47 (m, 1H), 3.33 (m, 1H), 1.81 (m, 4H). ^{13}C NMR (CDCl_3 , 100 MHz, ppm) δ : 150.1, 144.7, 137.8,

128.7, 124.4, 116.4, 62.5, 48.9, 32.6, 24.0. $\alpha^D = -75.8^\circ$. HRMS (Q-TOF 2, ES⁺, MeOH): *m/z* calc. for [M+H]⁺: C₁₂H₁₅N₂O₄S 283.0747 found 283.0745.

Procedure A: general procedure for selective reduction of aromatic nitro compound to aromatic amine

Into a round-bottom flask, the aromatic nitro containing compound was solubilized in absolute ethanol. SnCl₂ · 2H₂O (5 eq.) was added. The mixture was magnetically stirred at 70 °C for 30 min, then poured into ice. A solution of Na₂CO₃ (5% aq.) was slowly added up to pH 7 then extracted with ethyl acetate (×3). The combined organic phases were treated with activated charcoal, dried over magnesium sulfate, filtered and concentrated *in vacuo*. The reaction crude was purified over silica gel.

(2S)-1-(4-aminobenzenesulfonyl)-2-vinylpyrrolidine (5)

This compound was obtained from compound **4** (34 mg, 0.12 mmol) following the general procedure A. The reaction crude was purified over silica gel with the eluent ethyl acetate/petroleum ether 40/60 thereby obtaining compound **5** (28 mg, 93%). Aspect: pale yellow solid. MP: 109.5 °C. ¹H NMR (CDCl₃, 400 MHz, ppm) δ : 7.56 (d, 2H, *J* = 8.7 Hz), 6.66 (d, 2H, *J* = 8.7 Hz), 5.80 (ddd, 1H, *J* = 16.9 Hz, *J* = 10.2 Hz, *J* = 5.9 Hz), 5.24 (dt, 1H, *J* = 17.0 Hz, *J* = 1.4 Hz), 5.08 (dt, 1H, *J* = 10.2 Hz, *J* = 1.3 Hz), 4.21 (s_{broad}, 2H), 4.05 (m, 1H), 3.39 (ddd, 1H, *J* = 10.0 Hz, *J* = 7.1 Hz, *J* = 4.4 Hz), 3.20 (dt, 1H, *J* = 10.0 Hz, *J* = 7.4 Hz), 1.81–1.54 (m, 4H). ¹³C NMR (CDCl₃, 100 MHz, ppm) δ : 150.8, 139.1, 129.6, 125.8, 115.1, 114.1, 61.9, 48.9, 32.3, 23.8. $\alpha^D = -128.4^\circ$. HRMS (Q-TOF 2, ES⁺, MeOH): *m/z* calc. for [M+H]⁺: C₁₂H₁₇N₂O₂S 253.1005 found 253.1005.

(2S)-1-(4-aminobenzenesulfonyl)-2-ethylpyrrolidine (6)

In a round-bottom flask, compound **4** (100 mg, 0.35 mmol) was solubilized in dichloromethane (10 mL), the mixture was placed under vacuum before being placed under inert atmosphere, then palladium on carbon 5% (catalytic amount) was added and stirred under H₂ atmosphere (1 atm.) for 5 h. The reaction mixture was filtered on celite powder and concentrated *in vacuo*, thereby obtaining compound **6** without further purification (87 mg, 96%). Aspect: brown solid. MP: 145.3 °C. ¹H NMR (CDCl₃, 400 MHz, ppm) δ : 7.54 (d, *J* = 8.6 Hz, 2H), 6.65 (d, *J* = 8.6 Hz, 2H), 4.27 (s_{broad}, 2H), 3.46 (m, 1H), 3.30 (ddd, 1H, *J* = 10.5 Hz, *J* = 7.0 Hz, *J* = 5.4 Hz), 3.13 (dt, 1H, *J* = 10.4 Hz, *J* = 7.3 Hz), 1.80–1.69 (m, 2H), 1.57–1.35 (m, 4H), 0.86 (t, 3H, *J* = 7.5 Hz). ¹³C NMR (CDCl₃, 100 MHz, ppm) δ : 150.7, 129.5, 126.1, 114.1, 61.8, 49.1, 30.2, 29.3, 24.2, 10.5. $\alpha^D = -91.5^\circ$.

(2S)-1-(4-nosyl)-2-((1S)-fluoroethyl)pyrrolidine/(2R)-1-(4-nosyl)-2-((1R)-fluoroethyl)pyrrolidine (7)

To a HF/SbF₅ mixture (8 mL, 3.8 mol% SbF₅) cooled at –20 °C, the substrate **4** (100 mg, 0.35 mmol) was slowly added. The mixture was magnetically stirred at the same temperature for 30 min, after which the reaction mixture was neutralized with water/ice/Na₂CO₃ up to pH 9–10 and extracted with ethyl acetate (×3). The combined organic phases were dried over magnesium sulfate, filtered and concentrated *in vacuo*. The crude residue was purified by column chromatography over silica gel with the eluent diethyl ether/petroleum ether/triethylamine: 29/70/1 thereby obtaining compound **7** as a racemic mixture of enantiomers mixed with compound **8** 1/1, also obtained as a racemic mixture of enantiomers (473 mg, 78%). Aspect: white solid. MP: 95.7 °C. ¹H NMR (CDCl₃, 400 MHz, ppm) δ : 8.38 (m, 2H), 8.03 (m, 2H), 5.00 (dq, 1H, *J* = 49.1 Hz, *J* = 6.5 Hz, *J* = 2.6 Hz), 3.73 (dm, 1H, *J* = 26.1 Hz), 3.41 (m, 1H), 3.31 (m, 1H), 2.05–1.85 (m, 2H),

1.72–1.55 (m, 2H), 1.34 (dd, 3H, *J* = 23.6 Hz, *J* = 6.5 Hz). ¹³C NMR (CDCl₃, 100 MHz, ppm) δ : 150.3, 144.4, 128.6, 124.6, 91.5 (d, *J* = 174.1 Hz), 64.0 (d, *J* = 21.2 Hz), 49.4, 25.4 (d, *J* = 4.3 Hz), 25.0 (d, *J* = 1.4 Hz), 18.07 (d, *J* = 22.0 Hz). ¹⁹F {1H} NMR (CDCl₃, 376 MHz, ppm) δ : –190.7. HRMS (Q-TOF 2, ES⁺, MeOH): *m/z* calc. for [M+Na]⁺: C₁₂H₁₅FN₂NaO₄S 325.0629 found 325.0631.

(2R)-1-(4-nosyl)-2-((1S)-fluoroethyl)pyrrolidine/(2S)-1-(4-nosyl)-2-((1R)-fluoroethyl)pyrrolidine (8)

Aspect: white solid. MP: 87.7 °C. ¹H NMR (CDCl₃, 400 MHz, ppm) δ : 8.38 (m, 2H), 8.04 (m, 2H), 4.82 (dq, 1H, *J* = 46.5 Hz, *J* = 6.3 Hz, *J* = 4.9 Hz), 3.92 (m, 1H), 3.39 (m, 2H), 1.87 (m, 2H), 1.65 (m, 2H), 1.38 (dd, 3H, *J* = 24.4 Hz, *J* = 6.4 Hz). ¹³C NMR (CDCl₃, 100 MHz, ppm) δ : 150.3, 143.9, 128.9, 124.5, 91.6 (d, *J* = 171.1 Hz), 62.5 (d, *J* = 25.0 Hz), 49.7, 27.0 (d, *J* = 2.3 Hz), 24.8, 16.5 (d, *J* = 22.3 Hz). ¹⁹F {1H} NMR (CDCl₃, 376 MHz, ppm) δ : –180.0. HRMS (Q-TOF 2, ES⁺, MeOH): *m/z* calc. for [M+Na]⁺: C₁₂H₁₅FN₂NaO₄S 325.0629 found 325.0631.

(2S)-1-(4-aminobenzenesulfonyl)-2-((1S)-fluoroethyl)pyrrolidine and (2R)-1-(4-aminobenzenesulfonyl)-2-((1R)-fluoroethyl)pyrrolidine (9)

This compound was obtained from compound **7** (13 mg, 43 μmol) following the general procedure A. The reaction crude was purified over silica gel with the eluent ethyl acetate/petroleum ether: 30/70 thereby obtaining compound **9** (7 mg, 60%). Aspect: orange oil. ¹H NMR (CDCl₃, 400 MHz, ppm) δ : 7.61 (d, 2H, *J* = 6.1 Hz), 6.71 (d, 2H, *J* = 6.1 Hz), 4.94 (dq, 1H, *J* = 49.0 Hz, *J* = 6.5 Hz, *J* = 3.2 Hz), 3.60 (dm, 1H, *J* = 24.7 Hz), 3.34 (ddd, 1H, *J* = 10.5 Hz, *J* = 7.3 Hz, *J* = 5.2 Hz), 3.20 (dt, 1H, *J* = 10.4 Hz, *J* = 7.1 Hz), 1.93 (m, 1H), 1.83 (m, 1H), 1.53 (m, 2H), 1.34 (dd, 3H, *J* = 23.8 Hz, *J* = 6.5 Hz). ¹³C NMR (CDCl₃, 100 MHz, ppm) δ : 150.5, 129.7, 126.4, 114.4, 92.1 (d, *J* = 172.9 Hz), 63.6 (d, *J* = 21.9 Hz), 49.3, 25.4 (d, *J* = 4.3 Hz), 24.8 (d, *J* = 1.2 Hz), 18.3 (d, *J* = 22.1 Hz). ¹⁹F {1H} NMR (CDCl₃, 376 MHz, ppm) δ : –189.7. HRMS (Q-TOF 2, ES⁺, MeOH): *m/z* calc. for [M+H]⁺: C₁₂H₁₈FN₂O₂S 273.1068 found 273.1076.

(2R)-1-(4-aminobenzenesulfonyl)-2-((1S)-fluoroethyl)pyrrolidine and (2S)-1-(4-aminobenzenesulfonyl)-2-((1R)-fluoroethyl)pyrrolidine (10)

This compound was obtained from compound **8** (21 mg, 69 μmol) following the general procedure A. The reaction crude was purified over silica gel with the eluent ethyl acetate/petroleum ether: 30/70 thereby obtaining compound **10** (17 mg, 90%). Aspect: orange solid. MP: 116.2 °C. ¹H NMR (CDCl₃, 400 MHz, ppm) δ : 7.61 (m, 2H), 6.70 (m, 2H), 4.93 (dq, 1H, *J* = 46.3 Hz, *J* = 6.4 Hz, *J* = 4.2 Hz), 3.80 (ddt, 1H, *J* = 12.7 Hz, *J* = 7.8 Hz, *J* = 3.7 Hz), 3.35 (m, 1H), 3.22 (m, 1H), 1.86 (m, 1H), 1.76 (m, 1H), 1.60 (m, 1H), 1.49 (m, 1H), 1.37 (dd, 3H, *J* = 24.5 Hz, *J* = 6.4 Hz). ¹³C NMR (CDCl₃, 100 MHz, ppm) δ : 150.7, 129.8, 125.6, 114.4, 91.9 (d, *J* = 169.0 Hz), 61.5 (d, *J* = 26.6 Hz), 49.7, 26.5 (d, *J* = 1.2 Hz), 24.7, 15.9 (d, *J* = 22.1 Hz). ¹⁹F {1H} NMR (CDCl₃, 376 MHz, ppm) δ : –181.1. HRMS (Q-TOF 2, ES⁺, MeOH): *m/z* calc. for [M+H]⁺: C₁₂H₁₈FN₂O₂S 273.1068 found 273.1076.

(2S)-1-(4-nosyl)-2-(2-chloro-(1R)-fluoroethyl)pyrrolidine and (2R)-1-(4-nosyl)-2-(2-chloro-(1S)-fluoroethyl)pyrrolidine (11)

To a HF/SbF₅ mixture (1.3 mL, 21.6 mol% SbF₅) was added NCS (140 mg, 1.05 mmol). After 1 min of stirring at –10 °C, the substrate **4** (100 mg, 0.35 mmol) was slowly added. The mixture

was magnetically stirred at the same temperature for 1 h, then neutralized with water/ice/ Na_2CO_3 up to pH 10 and extracted with ethyl acetate ($\times 3$). The combined organic phases were dried over magnesium sulfate, filtered and concentrated *in vacuo*. The reaction crude was purified over silica gel with the eluent diethyl ether/pentane/diethylamine: 20/79/1, thereby obtaining compounds **11** and **12** (3/2) as a racemic mixture of their enantiomers (84 mg, 71%). Aspect: brown oil. ^1H NMR (CDCl_3 , 400 MHz, ppm) δ : 8.39 (d, 2H, $J = 8.8$ Hz), 8.04 (d, 2H, $J = 8.8$ Hz), 4.67 (ddt, 1H, $J = 46.4$ Hz, $J = 7.6$ Hz, $J = 3.7$ Hz), 4.10 (ddt, 1H, $J = 21.7$ Hz, $J = 9.0$ Hz, $J = 3.1$ Hz), 3.89 (ddd, 1H, $J = 26.7$ Hz, $J = 12.3$ Hz, $J = 3.9$ Hz), 3.76 (ddd, 1H, $J = 14.9$ Hz, $J = 12.3$ Hz, $J = 7.7$ Hz), 3.41 (m, 2H), 1.93 (m, 1H), 1.93 (m, 1H), 1.70 (m, 1H), 1.56 (m, 1H). ^{13}C NMR (CDCl_3 , 100 MHz, ppm) δ : 150.3, 143.4, 128.8, 124.5, 94.6 (d, $J = 182.7$ Hz), 60.3 (d, $J = 21.2$ Hz), 49.8, 43.1 (d, $J = 25.2$ Hz), 28.2, 24.5. ^{19}F {1H} NMR (CDCl_3 , 376 MHz, ppm) δ : -192.0. HRMS (Q-TOF 2, ES^+ , MeOH): m/z calc. for $[\text{M}+\text{Na}]^+$: $\text{C}_{12}\text{H}_{14}\text{ClFN}_2\text{NaO}_4\text{S}$ 359.0239 found: 359.0239.

(2*S*)-1-(4-nosyl)-2-(2-chloro-(1*S*)-fluoroethyl)pyrrolidine and (2*R*)-1-(4-nosyl)-2-(2-chloro-(1*R*)-fluoroethyl)pyrrolidine (**12**)

Aspect: brown oil. ^1H NMR (CDCl_3 , 400 MHz, ppm) δ : 8.41 (d, 2H, $J = 9.0$ Hz), 8.04 (d, 2H, $J = 9.0$ Hz), 4.92 (dm, 1H, $J = 47.8$ Hz), 3.95 (dm, 1H, $J = 20.6$ Hz), 3.77 (ddd, 1H, $J = 16.3$ Hz, $J = 12.1$ Hz, $J = 6.3$ Hz), 3.71 (ddd, 1H, $J = 16.1$ Hz, $J = 12.1$ Hz, $J = 5.5$ Hz), 3.48 (ddd, 1H, $J = 10.3$ Hz, $J = 7.2$ Hz, $J = 4.5$ Hz), 3.26 (dt, 1H, $J = 10.2$ Hz, $J = 7.2$ Hz), 2.02 (m, 1H), 1.93 (m, 1H), 1.62 (m, 1H), 1.59 (m, 1H). ^{13}C NMR (CDCl_3 , 100 MHz, ppm) δ : 150.3, 143.1, 128.8, 124.5, 92.7 (d, $J = 182.8$ Hz), 60.4 (d, $J = 22.9$ Hz), 49.4, 42.8 (d, $J = 25.6$ Hz), 26.0 (d, $J = 4.7$ Hz), 24.5. ^{19}F {1H} NMR (CDCl_3 , 376 MHz, ppm) δ : -193.8.

(2*S*)-1-(4-nosyl)-2-((1*R*)-chloro-2-fluoroethyl)pyrrolidine and (2*R*)-1-(4-nosyl)-2-((1*S*)-chloro-2-fluoroethyl)pyrrolidine (**13**)

To a HF/SbF_5 mixture (2 mL, 12.2 mol% SbF_5) was added NCS (208 mg, 1.56 mmol). After 1 min of stirring at -20°C , the substrate **4** (122 mg, 0.52 mmol) was slowly added. The mixture was magnetically stirred at the same temperature for 10 min, then neutralized with water/ice/ Na_2CO_3 up to pH 10 and extracted with ethyl acetate ($\times 3$). The combined organic phases were dried over magnesium sulfate, filtered and concentrated *in vacuo*. The reaction crude was purified over silica gel with the eluent diethyl ether/pentane/diethylamine: 20/79/1, thereby obtaining compounds **13** and **13'** (7/3) as a racemic mixture of their enantiomers (154 mg, 88%), **13'** could not be isolated. Aspect: yellow solid. Mp: 125.4°C . ^1H NMR (CDCl_3 , 400 MHz, ppm) δ : 8.40 (d, 2H, $J = 9.0$ Hz), 8.04 (d, 2H, $J = 9.0$ Hz), 4.64 (ddd, 1H, $J = 47.5$ Hz, $J = 5.2$ Hz, $J = 8.9$ Hz), 4.64 (m, 1H), 4.51 (ddd, 1H, $J = 47.5$ Hz, $J = 9.6$ Hz, $J = 6.6$ Hz), 4.06 (ddd, 1H, $J = 4.0$ Hz, $J = 5.0$ Hz, $J = 8.5$ Hz), 3.42 (dt, 1H, $J = 10.6$ Hz, $J = 6.9$ Hz), 3.36 (ddd, 1H, $J = 10.6$ Hz, $J = 7.3$ Hz, $J = 5.7$ Hz), 2.08 (m, 1H), 1.96 (m, 1H), 1.76 (m, 1H), 1.52 (dt, 1H, $J = 12.5$ Hz, $J = 7.3$ Hz, $J = 7.2$ Hz). ^{13}C NMR (CDCl_3 , 100 MHz, ppm) δ : 150.3, 143.3, 128.7, 124.5, 83.1 (d, $J = 176.6$ Hz), 60.9 (d, $J = 19.8$ Hz), 60.8 (d, $J = 3.1$ Hz), 49.6, 26.7, 24.7. ^{19}F {1H} NMR (CDCl_3 , 376 MHz, ppm) δ : -219.2. HRMS (Q-TOF 2, ES^+ , MeOH): m/z calc. for $[\text{M}+\text{Na}]^+$: $\text{C}_{12}\text{H}_{14}\text{ClFN}_2\text{NaO}_4\text{S}$ 359.0239 found: 359.0239.

1-(4-nosyl)piperidine-4-ol (**15**)

In a round-bottom flask, 4-nosyl chloride (512 mg, 2.3 mmol) was solubilized in dichloromethane (8 mL), 4-hydroxypiperidine (404 mg, 4.0 mmol) and triethylamine (0.4 mL, 3.0 mmol)

were added. The mixture was magnetically stirred at room temperature overnight, then water was added and extracted with ethyl acetate ($\times 3$). The combined organic phases were dried over magnesium sulfate, filtered and concentrated *in vacuo*, thereby obtaining compound **15** without further purification (575 mg, 87%). Aspect: yellow solid. Mp: 177.2°C . ^1H NMR (CD_3OD , 400 MHz, ppm) δ : 8.44 (d, 2H, $J = 9.0$ Hz), 8.03 (d, 2H, $J = 9.0$ Hz), 3.66 (m, 1H), 3.40 (m, 2H), 2.88 (ddd, 2H, $J = 12.0$ Hz, $J = 8.8$ Hz, $J = 3.4$ Hz), 1.88 (m, 2H), 1.57 (m, 2H). ^{13}C NMR (CDCl_3 , 100 MHz, ppm) δ : 150.3, 142.7, 128.9, 124.5, 65.3, 43.0, 33.2.

1-(4-nosyl)-4-methanesulfonatepiperidine (**16**)

Into a round-bottom flask, compound **15** (400 mg, 1.4 mmol) was solubilized in acetonitrile (5 mL), triethylamine (0.38 mL, 2.8 mmol) and mesyl chloride (0.16 mL, 2.1 mmol) were added. The mixture was magnetically stirred at 80°C overnight, then water and an aqueous solution of NaOH 2M were added. The mixture is stirred 5 min and extracted with ethyl acetate ($\times 3$). The combined organic phases were dried over magnesium sulfate, filtered and concentrated *in vacuo*, thereby obtaining compound **16** without further purification (504 mg, 99%). Aspect: yellow solid. ^1H NMR (CD_3OD , 400 MHz, ppm) δ : 8.38 (d, 2H, $J = 9.0$ Hz), 7.98 (d, 2H, $J = 9.0$ Hz), 4.70 (m, 1H), 3.24–3.05 (m, 4H), 2.97 (s, 3H), 2.03 (m, 2H), 1.88 (m, 2H). ^{13}C NMR (CD_3OD , 100 MHz, ppm) δ : 151.5, 143.1, 129.9, 125.5, 77.0, 43.6, 38.7, 31.7. HRMS (Q-TOF 2, ES^+ , MeOH): m/z calc. for $[\text{M}+\text{H}]^+$: $\text{C}_{12}\text{H}_{17}\text{N}_2\text{O}_7\text{S}_2$ 365.0471 found 365.0473.

1-(4-nosyl)-1,2,5,6-tetrahydropyridine (**17**)

Into a round-bottom flask, compound **16** (300 mg, 0.82 mmol) was solubilized in DBU (5 mL). The mixture was magnetically stirred at 80°C for 2 h, then water was added and extracted with ethyl acetate ($\times 3$). The combined organic phases were washed with an aqueous solution of HCl 0.5M and brine, dried over magnesium sulfate, filtered and concentrated *in vacuo*. The reaction crude was purified over silica gel with the eluent ethyl acetate/petroleum ether: 30/70 thereby obtaining compound **17** (86 mg, 39%). Aspect: yellow solid. Mp: 134.3°C . ^1H NMR (CDCl_3 , 400 MHz, ppm) δ : 8.37 (d, 2H, $J = 9.0$ Hz), 7.98 (d, 2H, $J = 9.0$ Hz), 5.78 (dm, 1H, $J = 10.2$ Hz), 5.63 (dm, 1H, $J = 10.2$ Hz), 3.67 (m, 2H), 3.28 (t, 2H, $J = 5.7$ Hz), 2.21 (m, 2H). ^{13}C NMR (CDCl_3 , 100 MHz, ppm) δ : 150.3, 143.2, 128.8, 125.4, 124.4, 122.5, 44.8, 42.7, 25.2. HRMS (Q-TOF 2, ES^+ , MeOH): m/z calc. for $[\text{M}+\text{H}]^+$: $\text{C}_{11}\text{H}_{13}\text{N}_2\text{O}_4\text{S}$ 269.0590 found 269.0593.

(3*S*,4*S*)-1-(4-nosyl)-4-chloro-3-fluoropiperidine and (3*R*,4*R*)-1-(4-nosyl)-4-chloro-3-fluoropiperidine (**18**)

To a HF/SbF_5 mixture (1 mL, 12.2 mol% SbF_5) was added NCS (72 mg, 0.54 mmol). After 1 min of stirring at -20°C , compound **17** (50 mg, 0.18 mmol) was slowly added. The mixture was magnetically stirred at the same temperature for 10 min, then neutralized with water/ice/ Na_2CO_3 up to pH 10 and extracted with ethyl acetate ($\times 3$). The combined organic phases were dried over magnesium sulfate, filtered and concentrated *in vacuo*. The reaction crude was purified over silica gel with the eluent petroleum ether/ethyl acetate: 90/10, thereby obtaining compound **18** (30 mg, 50%) and **19** (4 mg, 6%) as a racemic mixture of enantiomers. Aspect: white solid. Mp: 157.1°C . ^1H NMR (CDCl_3 , 400 MHz, ppm) δ : 8.39 (d, 2H, $J = 8.9$ Hz), 7.98 (d, 2H, $J = 8.9$ Hz), 4.63 (dm, 1H, $J = 45.9$ Hz), 4.09 (m, 1H), 3.58 (broad d, 1H, $J = 13.7$ Hz, $J = 4.7$ Hz), 3.50 (ddd, 1H, $J = 27.3$ Hz, $J = 13.5$ Hz, $J = 2.5$ Hz), 3.38 (m, 1H), 3.26 (m, 1H), 2.30 (m,

1H), 1.92 (dtd, 1H, $J = 14.2$ Hz, $J = 5.3$ Hz, $J = 3.3$ Hz). ^{13}C NMR (CDCl_3 , 100 MHz, ppm) δ : 150.4, 143.4, 128.7, 124.6, 87.1 (d, $J = 184.4$ Hz), 53.7 (d, $J = 26.7$ Hz), 45.5 (d, $J = 23.6$ Hz), 41.3, 29.3. ^{19}F {1H} NMR (CDCl_3 , 376 MHz, ppm) δ : -178.8. HRMS (Q-TOF 2, ES^+ , CH_3CN): m/z calc. for $[\text{M}+\text{H}]^+$: $\text{C}_{11}\text{H}_{13}\text{ClFN}_2\text{O}_4\text{S}$ 323.0263 found: 323.0264.

(3*S*,4*S*)-*N*-(4-nosyl)-3-chloro-4-fluoropiperidine and (3*R*,4*R*)-*N*-(4-nosyl)-3-chloro-4-fluoropiperidine (**19**)

Aspect: white solid. Mp: 126.6 °C. ^1H NMR (CDCl_3 , 400 MHz, ppm) δ : 8.40 (d, 2H, $J = 9.0$ Hz), 7.98 (d, 2H, $J = 9.0$ Hz), 4.58 (dtd, 1H, $J = 47.0$ Hz, $J = 6.4$ Hz, $J = 3.4$ Hz), 4.07 (m, 1H), 3.62 (dt, 1H, $J = 13.2$ Hz, $J = 3.1$ Hz), 3.35 (dd, 1H, $J = 13.1$ Hz, $J = 6.2$ Hz), 3.25 (m, 2H), 2.31 (m, 1H), 1.93 (m, 1H). ^{13}C NMR (CDCl_3 , 100 MHz, ppm) δ : 150.5, 143.2, 128.8, 124.7, 89.2 (d, $J = 180.6$ Hz), 53.9 (d, $J = 24.5$ Hz), 48.7 (d, $J = 2.8$ Hz), 42.0 (d, $J = 6.5$ Hz), 27.76 (d, $J = 20.0$ Hz). ^{19}F {1H} NMR (CDCl_3 , 376 MHz, ppm) δ : -179.0.

CA inhibition assay

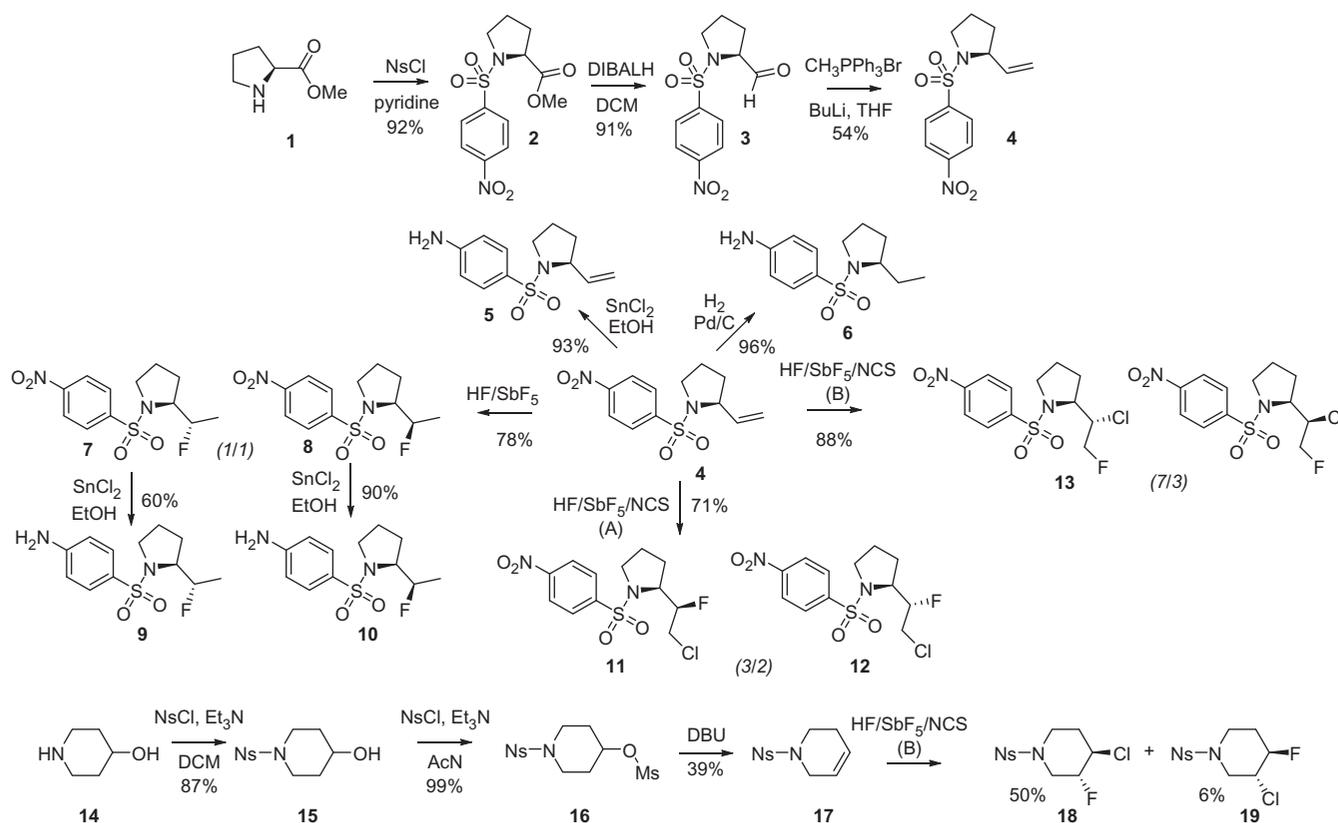
An applied photophysics stopped-flow instrument has been used for assaying the CA catalyzed CO_2 hydration activity⁴⁹. Phenol red (at a concentration of 0.2 mM) has been used as indicator, working at the absorbance maximum of 557 nm, with 20 mM Hepes (pH 7.5) as buffer, and 20 mM Na_2SO_4 (for maintaining constant the ionic strength), following the initial rates of the CA-catalyzed CO_2 hydration reaction for a period of 10–100 s. The CO_2 concentrations ranged from 1.7 to 17 mM for the determination of the kinetic parameters and inhibition constants. For each inhibitor, at least six traces of the initial 5–10% of the reaction have been used for determining the initial velocity. The uncatalyzed rates were determined in the same manner and subtracted from the total observed rates. Stock solutions of

inhibitor (0.1 mM) were prepared in distilled–deionized water and dilutions up to 0.01 nM were done thereafter with distilled–deionized water. Inhibitor and enzyme solutions were preincubated together for 15 min at room temperature prior to assay, in order to allow for the formation of the E-I complex. The inhibition constants were obtained by non-linear least-squares methods using the Cheng–Prusogg equation, as reported earlier, and represent the mean from at least three different determinations^{50–55}. CA isoforms were recombinant ones obtained in house as reported earlier^{50–55}.

Results and discussion

Chemistry

The synthesis of the unsaturated product **4** was first achieved (Scheme 1). Starting from L-proline methyl ester derivative **1**, a three steps procedure, including a nosyl protection of the nitrogen atom, a selective reduction of the methyl ester to the corresponding aldehyde, and a Wittig reaction led to the synthesis of the tertiary benzenesulfonamide **4** in 46% overall yield. This procedure could be conducted on a gram scale. This product served as the key intermediate for the divergent synthesis of a wide range of (fluorinated) pyrrolidines containing tertiary benzenesulfonamides. The tin (II) chloride selective reduction of the nitro group of substrate **4** to the corresponding amino-benzenesulfonamide **5** was performed in 93% yield. A complete reduction of the same substrate was also achieved in 96% yield after hydrogenation over Pd/C catalyst (formation of product **6**). The hydrofluorination of substrate **4** in superacid HF/SbF_5 (% mol $\text{SbF}_5 = 3.8$, -20°C , 10 min) led to the formation of a mixture of *like* and *unlike* diastereoisomers **7** and **8**. By adopting the same reduction procedure than the previously described, the corresponding 4-aminobenzenesulfonamides **9** and **10** have been synthesized in good yields. Regioselective superacid catalyzed



Scheme 1. Synthesis of carbonic anhydrase inhibitors.

Table 2. Inhibition data of tertiary (fluorinated) benzenesulfonamides.

Entry	Compound	Ki (μM)*			
		hCA I†	hCA II†	hCA IX‡	hCA XII‡
1	2	>100	>100	>100	>100
2	4	>100	0.5	>100	>100
3	5	>100	6.3	>100	>100
4	6	>100	7.3	>100	>100
5	7	>100	4.7	>100	>100
6	8	>100	3.9	>100	>100
7	9	>100	4.8	>100	>100
8	10	>100	3.4	>100	>100
9	11	>100	7.6	>100	>100
10	12	>100	6.3	>100	>100
11	13	>100	6.6	>100	>100
12	18	>100	11.0	>100	>100
13	19	>100	10.0	>100	>100

*Errors in the range of $\pm 5\%$ of the reported data from three different assays.

†Recombinant isoforms.

‡Catalytic domain.

chlorofluorination of substrate **4** in conditions A (% mol $\text{SbF}_5 = 21.7$, NCS (3eq.), -10°C , 60 min) allowed the synthesis of the β -fluoro- γ -chlorinated diastereoisomers **11** and **12** in overall 71% yield. An adapted procedure (conditions B: % mol $\text{SbF}_5 = 12.1$, NCS (3eq.), -20°C , 10 min) led to the synthesis of the corresponding regioisomers in 86% yield, the isomer **13** being cleanly separated from the mixture. It has to be noted that unfortunately these superacid catalyzed reactions suffer from epimerization of the pyrrolidines chiral carbon center, thus delivering in all cases only racemic mixture of *like* and *unlike* isomers. The relative configuration of the products was determined by careful analysis of the ^1H and ^{19}F NMR spectrum. To evaluate the impact of the ring shape on carbonic anhydrases inhibition selectivity, analogous piperidines were synthesized. Starting from piperidin-4-ol **14**, after successive protection of the amino group and activation of the alcohol through mesylation, elimination with DBU led to the synthesis of the unsaturated substrate **17**. The chlorofluorination of this compound led to the formation of products **18** and **19** showing a relative anticonfiguration.

Carbonic anhydrase inhibition

All the tested tertiary benzenesulfonamides were found to be ineffective as hCA I, hCA IX and hCA XII inhibitors (Table 2). Exceptionally, whereas they do not inhibit these isoforms, all the tested (despite compound **2**) substrates inhibited the highly catalytically active hCA II at the micromolar level. When these tertiary benzenesulfonamides act as selective hCA II inhibitor, the compound **2** was found to be inactive toward all the tested isoforms, revealing a strong impact of the pyrrolidine substituents on the inhibition profile. The amino group located in the position 4 of the aromatic benzenesulfonamides inhibitors was previously shown to be essential for the inhibition selectivity of primary benzenesulfonamides⁵⁵. In the case of these tertiary benzenesulfonamides, the impact of the presence of the amino group on the inhibition potential was difficult to predict. Indeed when compound **4** was around 10 times more efficient than its amino analogue **5**, nitroderivative **7** (or **8**) and its amino analogue **9** (or **10**) showed similar activities. In addition, the unsaturation was found not to be primordial for the selectivity of the inhibition, as compounds **5** and **6** showed a similar inhibition profile. The introduction of the fluorine atom in the β position of the nitrogen atom was found to increase the inhibition potency (Table 2,

entries 4, 7 and 8), but this effect was very small, or absent for the chlorofluorinated analogues. The disparity of this effect confirms the impact of the geometrical shape of the molecule on the hCA II selective inhibition and reinforced the hypothesis of a non-zinc binding mode of action of these inhibitors and a strong influence of the structural shape on the inhibition efficiency. In addition, no significant effect of the inversion of the relative configuration of the two stereogenic centers on the inhibition potency appeared. Interestingly, the piperidine analogues **18** and **19** also act as hCA II selective inhibitors, confirming the importance of a cyclic core for the inhibition selectivity.

Conclusions

In conclusion, through the use of superacid chemistry, a set of variously substituted pyrrolidine and piperidine were synthesized and evaluated as specific carbonic anhydrase inhibitors. This work allowed to identify a new family of hCA II inhibitors in the tertiary benzenesulfonamides family, a class of inhibitors which is expanding rapidly. This class of CAIs points out a new mechanism of action for inhibition of hCA II, the physiologically most dominant isoform, being present in virtually all human cells. Indeed, CA II is a cytoplasmic enzyme, which is the most widespread of all CA isozymes and the catalytically fastest member of the α -class family (together with CA IX) and thus implicated in numerous physiological functions. To get further insights into various pathologies implicating CA II in auto-immune reactions⁵⁶ or deficiency⁵⁷, it makes no doubt that specific hCA II inhibitors are needed and should be welcome for further advances in these fields.

Declaration of interest

The authors report no conflicts of interest. The authors alone are responsible for the content and writing of this article. We are indebted to the Université de Poitiers, the CNRS, @rtMolecule and ANRT (CIFRE scholarship for A.LD) and an EU FP7 grant (Dynano to CTS) for financial support.

References

- Krishnamurthy VM, Kaufman GK, Urbach, AR, et al. Carbonic anhydrase as a model for biophysical and physical-organic studies of proteins and protein-ligand binding. *Chem Rev* 2008;108:946–1051.
- Supuran CT. Carbonic anhydrase inhibitors and activators for novel therapeutic applications. *Future Med Chem* 2011;3:1165–80.
- Supuran CT. Carbonic anhydrases as drug targets: general presentation. In: Supuran CT, Winum JY, eds. *Drug design of zinc-enzyme inhibitors: functional, structural, and disease applications*. Hoboken (NJ): Wiley; 2009:15–38.
- Supuran CT. Carbonic anhydrases: novel therapeutic applications for inhibitors and activators. *Nat Rev Drug Discovery* 2008;7:168–81.
- Winum JY, Rami M, Scozzafava A, et al. Carbonic anhydrase IX: a new druggable target for the design of antitumor agents. *Med Res Rev* 2008;28:445–63.
- Supuran CT, Scozzafava A, Casini A. Carbonic anhydrase inhibitors. *Med Res Rev* 2008;23:146–89.
- Winum JY, Supuran CT. Recent advances in the discovery of zinc-binding motifs for the development of carbonic anhydrase inhibitors. *J Enzyme Inhib Med Chem* 2014;18:1–4.
- Supuran CT, Casini A, Scozzafava A. Development of sulfonamide carbonic anhydrase inhibitors. In: Supuran CT, Scozzafava A, Conway J, eds. *Carbonic anhydrase: its inhibitors and activators*. Boca Raton (FL): CRC Press; 2004:67–147.
- Chohan ZH, Scozzafava A, Supuran CT. Zinc complexes of benzothiazole-derived Schiff bases with antibacterial activity. *J Enzyme Inhib Med Chem* 2003;18:259–63.
- Supuran CT. Structure-bases drug discovery of carbonic anhydrase inhibitors. *J Enzyme Inhib Med Chem* 2012;27:759–72.
- Capasso C, Supuran CT. Anti-infective carbonic anhydrase inhibitors: a patent and literature review. *Expert Opin Ther Pat* 2013;23:693–704.

12. Swenson ER. Safety of carbonic anhydrase inhibitors. *Expert Opin Drug Saf* 2014;13:459–72.
13. Supuran CT, Mincione F, Scozzafava A, et al. Metal complexes of heterocyclic sulfonamides: a new class of strong topical intraocular pressure-lowering agents with potential use as antiglaucoma drugs. *Eur J Med Chem* 1998;33:247–54.
14. Alterio V, Di Fiore A, D'Ambrosio K, Supuran CT. Multiple binding modes of inhibitors to carbonic anhydrases: how to design specific drugs targeting 15 different isoforms? *Chem Rev* 2012;112:4421–68.
15. Wilkinson BL, Bornaghi LF, Houston TA, et al. Carbonic anhydrase inhibitors: inhibition of isozymes I, II and IX with triazole-linked O-glycosides of benzene sulfonamides. *J Med Chem* 2007;50:1651–7.
16. Chohan ZH, Scozzafava A, Supuran CT. Unsymmetrical 1,1'-disubstituted ferrocenes: synthesis of Co(ii), Cu(ii), Ni(ii) and Zn(ii) chelates of ferrocenyl-1-thiadiazolo-1'-tetrazole, -1-thiadiazolo-1'-triazole and -1-tetrazolo-1'-triazole with antimicrobial properties. *J Enzyme Inhib Med Chem* 2002;17:261–6.
17. Neri D, Supuran CT. Interfering with pH regulation in tumours as a therapeutic strategy. *Nat Rev Drug Discov* 2011;10:767–77.
18. McDonald PC, Winum JY, Supuran CT, Dedhar S. Recent development in targeting carbonic anhydrase IX for cancer therapeutics. *Oncotarget* 2012;3:84–97.
19. Guler OO, De Simone G, Supuran CT. Drug design studies of the novel antitumor targets carbonic anhydrase IX and XII. *Curr Med Chem* 2010;17:1516–26.
20. Tars K, Vullo D, Kazaks A, et al. Sulfocoumarins (1,2-benzoxathiine-2,2-dioxides): a class of potent and isoform-selective inhibitors of tumor-associated carbonic anhydrases. *J Med Chem* 2013;56:293–300.
21. Ceruso M, Bragagni M, AlOthman Z, et al. New series of sulfonamides containing amino acid moiety act as a effective and selective inhibitors of tumor-associated carbonic anhydrase XII. *J Enzyme Inhib Med Chem* 2014;4:1–5.
22. Thibaudeau S, Martin-Mingot A, Jouannetaud M-P, et al. A novel, facile route to β -fluoroamines by hydrofluorination using superacid HF-SbF₅. *Chem Commun* 2007;3:198–200.
23. Vardelle E, Gamba-Sanchez D, Martin-Mingot A, et al. Cyclisation/fluorination of nitrogen containing dienes in superacid HF-SbF₅: a new route to 3- and 4-fluoropiperidines. *Chem Commun* 2008:1473–5.
24. Liu F, Martin-Mingot A, Jouannetaud M-P, et al. Superelectrophilic activation in superacid HF/SbF₅ and synthesis of benzofused sultams. *Org Lett* 2010;12:868–71.
25. Compain G, Martin-Mingot A, Frapper G, et al. Anti-Markovnikov additions to N-allylic derivatives involving ammonium-carbenium superelectrophiles. *Chem Commun* 2012;48:5877–9.
26. Compain G, Jouvin K, Martin-Mingot A, et al. Stereoselective hydrofluorination of ynamides: a straightforward synthesis of novel α -fluoroenamides. *Chem Commun* 2012;48:5196–8.
27. Métayer B, Mingot A, Vullo D, et al. New superacid synthesized (fluorinated) tertiary benzenesulfonamides acting as selective hCA IX inhibitors: toward a new mode of carbonic anhydrase inhibition by sulfonamides. *Chem Commun* 2013;49:6015–17.
28. Métayer B, Mingot A, Vullo D, et al. Superacid synthesized tertiary benzenesulfonamides and benzofused sultams act as selective hCA IX inhibitors: toward understanding a new mode of inhibition by tertiary sulfonamides. *Org Biomol Chem* 2013;43:7540–9.
29. Compain G, Martin-Mingot A, Maresca A, et al. Superacid synthesis of halogen containing N-substituted-4-aminobenzene sulfonamides: new selective tumor-associated carbonic anhydrase inhibitors. *Bioorg Med Chem* 2013;21:1555–63.
30. Maresca A, Scozzafava A, Supuran CT. 7,8-disubstituted- but not 6,7-disubstituted coumarins selectively inhibit the transmembrane, tumor-associated carbonic anhydrase isoforms IX and XII over the cytosolic ones I and II in the low nanomolar/subnanomolar range. *Bioorg Med Chem Lett* 2010;20:7255–8.
31. Maresca A, Supuran CT. Coumarins incorporating hydroxyl- and chloro-moities selectively inhibit the transmembrane tumor-associated carbonic anhydrase isoforms IX and XII over the cytosolic ones I and II. *Bioorg Med Chem Lett* 2010;20:4511–14.
32. Grandane A, Tanc M, Zalubovskis R, Supuran CT. Synthesis of 6-tetrazolyl-substituted sulfocoumarins acting as highly potent and selective inhibitors of the tumor-associated carbonic anhydrase isoforms IX and XII. *Bioorg Med Chem* 2014;22:1522–8.
33. Lau J, Pan J, Zhang Z, et al. Synthesis and evaluation of (18)F-labeled tertiary benzenesulfonamides for imaging carbonic anhydrase IX expression in tumours with positron emission tomography. *Bioorg Med Chem Lett* 2014;24:3064–8.
34. D'Ascenzio M, Carradori S, De Monte C, et al. Design, synthesis and evaluation of N-substituted saccharin derivatives as selective inhibitors of tumor-associated carbonic anhydrase XII. *Bioorg Med Chem* 2014;22:1821–31.
35. Maresca A, Temperini C, Vu H, et al. Non-zinc mediated inhibition of carbonic anhydrases: coumarins are a new class of suicide inhibitors. *J Am Chem Soc* 2009;131:3057–62.
36. Temperini C, Innocenti A, Scozzafava A, et al. The coumarin-binding site in carbonic anhydrase accomodates structurally diverse inhibitors: the antiepileptic lacosamide as an example and lead molecule for novel classes of carbonic anhydrase inhibitors. *J Med Chem* 2010;53:850–4.
37. Biswas S, Aggarwal M, Güzel Ö, et al. Conformational variability of different sulfonamide inhibitors with thienyl-acetamido moieties attributes to different binding in the active site of the cytosolic human carbonic anhydrase isoforms. *Bioorg Med Chem* 2011;19:3732–8.
38. Abbate F, Coetzee A, Casini A, et al. Carbonic anhydrase inhibitors: X-ray crystallographic structure of the adduct of human isozyme II with the antipsychotic drug sulpiride. *Bioorg Med Chem Lett* 2004;14:337–41.
39. Abbate F, Casini A, Scozzafava A, Supuran CT. Carbonic anhydrase inhibitors: X-ray crystallographic structure of the adduct of human isozyme II with the perfluorobenzoyl analogue of methazolamide: implications for the drug design of fluorinated inhibitors. *J Enzyme Inhib Med Chem* 2003;18:303–8.
40. Begue J-P, Bonnet-Delpon D. *Bioorganic and medicinal chemistry of fluorine*. Hoboken (NJ): Wiley; 2008.
41. Wang J, Sanchez-Rosello M, Acena JL, et al. Fluorine in pharmaceutical industry: fluorine-containing drugs introduced to the market in the last decade (2001–2011). *Chem Rev* 2014;114:2432–506.
42. Morgenthaler M, Schweiser E, Hoffman-Röder A, et al. Predicting and tuning physicochemical properties in lead optimization: amine basicities. *Chem Med Chem* 2007;2:1100–15.
43. Müller K, Faeh C, Diederich F. Fluorine in pharmaceuticals: looking beyond intuition. *Science* 2007;317:1881–6.
44. Böhm H-J, Banner D, Bendels S, et al. Fluorine in medicinal chemistry. *Chem Bio Chem* 2004;5:637–43.
45. Hu X-G, Hunter L. Stereoselectively fluorinated N-heterocycles: a brief survey. *Beilstein J Org Chem* 2013;9:2696–708.
46. For a recent review, please see: Hunter L. The C-F bond as a conformational tool in organic and biological chemistry. *Beilstein J Org Chem* 2010;6:38.
47. For a recent article, please see: Yamamoto I, Jordan MJT, Gavande N, et al. The enantiomers of *syn*-2,3-difluoro-4-aminobutyric acid elicit opposite responses at the GABA_c receptor. *Chem Commun* 2012;48:829–31.
48. Ceruso M, Vullo D, Scozzafava A, Supuran CT. Sulfonamides incorporating fluorine and 1,3,5-triazine moieties are effective inhibitors of three β -class carbonic anhydrases from *Mycobacterium tuberculosis*. *J Enzyme Inhib Med Chem* 2014;25:686–9.
49. Khalifah RG. Carbon dioxide hydration activity of carbonic anhydrase: kinetics of alkylated anhydrases B and C from humans. *J Biol Chem* 1971;246:2561–73.
50. Rami M, Maresca A, Smaie F-Z, et al. Sulfonamides incorporating boroxazolidone moieties are potent inhibitors of the transmembrane, tumor-associated carbonic anhydrase isoforms IX and XII. *Bioorg Med Chem Lett* 2011;21:2975–9.
51. Capkauskaite E, Barauskiene L, Golovenko D, et al. Indapamide-like benzenesulfonamides as inhibitors of carbonic anhydrases I, II, VII, and XIII. *Bioorg Med Chem* 2010;18:7357–64.
52. Mader P, Brynda J, Gitto R, et al. Structural basis for the interaction between carbonic anhydrase and 1,2,3,4-tetrahydroisoquinolin-2-ylsulfonamides. *J Med Chem* 2011;54:2522–6.
53. Mincione F, Benedini F, Biondi S, et al. Synthesis and crystallographic analysis of new sulfonamides incorporating NO-donating moieties with potent antiglaucoma action. *Bioorg Med Chem Lett* 2011;21:3216–21.
54. Bertucci A, Innocenti A, Scozzafava A, et al. Carbonic anhydrase inhibitors. Inhibition studies with anions and sulfonamides of a new cytosolic enzyme from the scleractinian coral *Stylophora pistillata*. *Bioorg Med Chem Lett* 2011;21:710–14.

55. Hen N, Bialer M, Yagen B, et al. Anticonvulsant 4-aminobenzene-sulfonamide derivatives with branched-alkylamide moieties: X-ray crystallography and inhibition studies of human carbonic anhydrase isoforms I, II, VII, and XIV. *J Med Chem* 2011;54: 3977–81.
56. Botrè F, Botrè C, Podestà E, et al. Effect of anti-carbonic anhydrase antibodies on carbonic anhydrases I and II. *Clin Chem* 2003;49: 1221–3.
57. Bosley TM, Salih MA, Alorainy IA, et al. The neurology of carbonic anhydrase type II deficiency syndrome. *Brain* 2011;134: 3499–512.