

Steroids

Steroids 65 (2000) 529-535

Synthesis of 7α -hydroxy derivatives of regulatory oxysterols

Dansu Li, Thomas A. Spencer*

Department of Chemistry, Dartmouth College, Hanover, NH 03755, USA

Received 22 March 2000; received in revised form 15 May 2000; accepted 26 May 2000

Abstract

 7α -Hydroxy derivatives of oxysterols are of considerable interest because of their possible involvement in regulation of cholesterol metabolism. This paper describes stereoselective syntheses and complete characterization of the 7α -hydroxy derivatives of four key oxysterols: 25-hydroxycholesterol, 27-hydroxycholesterol, 24(*S*)-hydroxycholesterol, and 24(*S*),25-epoxycholesterol. © 2000 Elsevier Science Inc. All rights reserved.

Keywords: Oxysterols; 7*α*-Hydroxyoxysterols; Synthesis

1. Introduction

Recent evidence indicates that oxysterols are involved in regulation of cholesterol 7α -hydroxylase, the enzyme that catalyzes the rate-limiting step in the conversion of cholesterol to bile acids. Mangelsdorf's group [1] reported that specific oxvsterols activate transcription of an LXRE-luciferase construct through the orphan nuclear transcription factor $LXR\alpha$, and suggested that this might be part of a feedback regulation of crucial metabolic pathways, such as steroid hormone or bile acid biosynthesis. Shortly thereafter, Lehmann et al. [2], reported that the receptors LXR α and LXR β are activated by specific oxysterols, and that they induce transcription of a cholesterol 7α -hydroxylase promoter, implicating oxysterols as regulators of this key enzyme. Mangelsdorf et al. have also described an extended investigation of structural requirements for oxysterol ligands for the LXR α and LXR β receptors [3]. It has been observed that some of these same oxysterols are also subject to enzymatic 7α -hydroxylation [4]. It has, in fact, been suggested that such 7α -hydroxylation of 25-hydroxycholesterol (1) (Scheme 1) may be important to inactivate 1 as a regulator of HMG-CoA reductase and the LDL receptor [5], in another putative regulatory role for oxysterols. In support of this possibility, it has been reported that the 7α -hydroxy derivative of 27-hydroxycholesterol (2) has no inhibitory effect

on cholesterol biosynthesis [6]. However, it has also been reported that in human diploid fibroblasts the 7α -hydroxy derivatives of both **1** and **2** do repress HMG-CoA reductase activity [7].

Further studies of the effects of 7α -hydroxylation of oxysterols on different aspects of cholesterol regulation are clearly needed. As part of such studies, it is obviously important to have available samples of the pertinent 7α -hydroxylated oxysterols. This paper describes the synthesis and complete characterization of the 7α -hydroxy derivatives of four key oxysterols: 25-hydroxycholesterol (1), 27-hydroxycholesterol (2), 24(S)-hydroxycholesterol (3), and 24(S), 25-epoxycholesterol (4). These were selected on the basis of their previous implication as possible regulators of cholesterol metabolism. 25-Hydroxycholesterol (1) is one of the most potent oxysterols in HMG-CoA reductase repression assays [8,9] and because it is commercially available, it has been by far the most widely used "representative" oxysterol. Observations consistent with a regulatory role for **1** have recently been reported [10,11]. 27-Hydroxycholesterol (2) has been identified in mouse liver [12], and has been proposed as a regulator of cholesterol metabolism [13-15], although not without dispute [16]. 24(S)-Hydroxycholesterol (3) and 24(S),25-epoxycholesterol (4) appeared to be particularly promising candidate oxysterols on the basis of our earlier investigations [17,18] and the recent finding of their high affinities as ligands for the LXR receptors [1,2].

The 7α -hydroxy derivative **5** (Scheme 2) of 25-hydroxycholesterol (**1**) has been previously prepared by a different route from that described herein, requiring isolation from a mixture with its 7β -isomer [19]. The 7α -hydroxy derivative

[☆] This research was supported by NIH grant HL52069.

^{*} Corresponding author. Tel.: +1-603-646-2805; fax: +1-603-646-3946.

E-mail address: taspen@dartmouth.edu (T.A. Spencer).

⁰⁰³⁹⁻¹²⁸X/00/\$ – see front matter © 2000 Elsevier Science Inc. All rights reserved. PII: S0039-128X(00)00131-8



Scheme 1.

6 of 27-hydroxycholesterol (**2**) has also been reported, but without complete separation from its 7β -isomer and complete characterization [20]. During the course of this work, Corey and Grogan [21] reported preparation of 7α -hydroxy-24(*S*),25-epoxycholesterol (**8**) (Scheme 3), by a synthesis slightly different from that reported herein. This paper presents full details of stereoselective syntheses of the 7α -hydroxy derivatives **5–8** of all four key oxysterols **1–4**.

2. Experimental

2.1. General

NMR spectra, unless otherwise noted, were taken in $CDCl_3$ on a 300-MHz Varian spectrometer. The chemical

shifts are reported in units of δ . Melting points (m.p.) were determined using a Thomas-Hoover apparatus in capillary tubes and are uncorrected. Thin-layer chromatography (TLC) was carried out on EM plastic sheets pre-coated with silica gel 60 F-254 (Whatman, Tewksbury, MA, USA). Visualization was obtained by exposure to 5% phosphomolybdic acid in 2-propanol. Flash chromatography was carried out on EM Reagent silica gel 60 (230–400 mesh), unless otherwise noted. MgSO₄ was used to dry all organic layers.

CH₂Cl₂ was distilled from calcium hydride. Tetrahydrofuran and ether were distilled from sodium/benzophenone. Pyridine was distilled from calcium hydride onto 3Å molecular sieves. Acetonitrile was dried over 3Å molecular sieves for 24 h and then distilled onto 3Å molecular sieves. CrO₃ was dried in vacuo for 12 h over KOH before use. All



Scheme 2.





Scheme 3.

reagents, unless otherwise noted, were obtained from Aldrich Chemical Co. (Milwaukee, WI, USA).

2.2. Cholest-5-ene-3 β ,25-diol 3 β ,25-Dibenzoate (9)

According to a procedure by Kim et al. [22], to a solution of 43.6 mg (0.108 mmol) of **1** (Steraloids Inc.) in 2.0 ml of dry pyridine was added 0.5 ml (4 mmol) of benzoyl chloride. The mixture was stirred at room temperature for 3 h under N₂, diluted with 10 ml of MeOH and 20 ml of hexane, washed with 10 ml each of 10% HCl, H₂O, saturated NaHCO₃ solution and H₂O, dried, filtered, evaporated to give 65.4 mg (98%) of **9**: m.p. 100–103°C (lit. [23] m.p. 100–102°C); ¹H NMR 8.03 (m, 4H), 7.54 (m, 2H), 7.45 (m, 4H), 5.43 (m, 1H), 4.87 (m, 1H), 1.26 (s, 6H), 1.07 (s, 3H), 0.93 (d, J = 6.6 Hz, 3H), 0.69 (s, 3H); ¹³C NMR 166.1, 165.8, 139.8, 132.9, 132.6, 132.2, 131.0, 129.7, 129.6, 128.4, 128.3, 122.9, 83.5, 74.7, 56.8, 56.3, 50.2, 42.5, 41.7, 39.9, 38.4, 37.2, 36.8, 36.4, 35.8, 32.1, 32.0, 28.4, 28.1, 26.3, 26.3, 24.5, 21.2, 20.6, 19.6, 18.8, 12.1.

2.3. 3β,25-Dihydroxycholest-5-en-7-one 3β,25-Dibenzoate (12)

According to a procedure by Parish and Wei [24], to a solution of 26.3 mg (0.0431 mmol) of **9** in 5 ml of dry benzene were added 35.0 mg of 3A molecular sieves and 232 mg (1.08 mmol) of PCC. After the mixture was refluxed for 24 h under N₂, 10 ml of brine was added carefully and the resulting mixture was extracted with 3×10 ml of CH₂Cl₂. The combined organic layers were evaporated to give a brown residue, which was taken up in 50 ml of ether and filtered. The filtrate was dried, filtered, and evaporated to give 28.2 mg of residue that was chromatographed (1:9 EtOAc:hexane) to give 23.4 mg (87%) of **12**. Recrystallization from CH₂Cl₂-hexane gave colorless **12**: m.p. 148.9–149.4°C; ¹H NMR 8.02 (m, 4H), 7.56 (m, 2H), 7.44 (m, 4H), 5.75 (m, 1H), 4.98 (m, 1H), 1.27 (s, 6H), 1.07 (s, 3H),

0.93 (d, J = 6.6 Hz, 3H), 0.70 (s, 3H); ¹³C NMR 202.2, 166.1, 166.0, 164.1, 133.3, 132.7, 132.3, 130.5, 129.9, 129.6, 128.6, 128.5, 127.1, 83.6, 73.1, 55.1, 50.2, 50.1, 45.7; 43.4, 41.8, 38.9, 38.7, 38.1, 36.5, 36.3, 35.9, 28.8, 27.8, 26.6, 26.4, 26.4, 21.4, 20.7, 19.1, 17.6, 12.3. Analysis Calculated for C₄₁H₅₂O₅: C, 78.80; H, 8.39. Found: C, 78.58; H, 8.55.

2.4. Cholest-5-ene-3 β ,7 α ,25-triol 3 β ,25-Dibenzoate (15)

According to a modification of a procedure by Kumar et al. [25], to a solution of 18 mg (0.035 mmol) of 12 in 1 ml of dry THF at -78°C was added 0.1 ml of 1 M L-Selectride in THF. The resulting solution was stirred at -78° C for 5 h under N₂ and 0.1 ml each of water, 6 N NaOH and 30% H₂O₂ were added. Most of the THF was evaporated, 10 ml of water was added, and the mixture was extracted with 3 \times 10 ml of CH₂Cl₂. The organic layers were dried, filtered, and evaporated to give 17.0 mg of residue, which was chromatographed (3:17 EtOAc:hexane) to afford 12.8 mg (71%) of 15. Recrystallization from ether-hexane gave colorless 15: m.p. 136.5–137.2°C; ¹H NMR 8.03 (m, 4H), 7.55 (m, 2H), 7.44 (m, 4H), 5.69 (d, J = 5.1 Hz, 1H), 4.93 (m, 1H), 3.87 (m, 1H), 1.26 (s, 6H), 1.07 (s, 3H), 0.94 (d, J =6.6 Hz, 3H), 0.70 (s, 3H); ¹³C NMR 166.1, 165.9, 145.4, 133.0, 132.6, 132.3, 130.9, 129.8, 129.6, 128.5, 128.4, 125.1, 83.5, 74.2, 65.5, 56.1, 49.6, 42.4, 42.4, 41.7, 39.4, 38.2, 37.8, 37.7, 37.0, 36.4, 35.9, 28.5, 27.8, 26.4, 24.5, 20.9, 20.6, 18.9, 18.5, 11.9. Analysis Calculated for C₄₁H₅₄O₅: C, 78.54; H, 8.69. Found: C, 78.46; H, 8.62.

2.5. Cholest-5-ene-3 β , 7 α , 25-triol (5)

A solution of 12.8 mg (0.0204 mmol) of **15** in 10 ml of MeOH containing 0.5 g of KOH was heated at reflux for 1 h. Most of MeOH was evaporated, 10 ml of H₂O was added, and the resulting mixture was extracted with 3×10 ml of CH₂Cl₂. The combined organic layers were washed

with 10 ml of brine, dried, filtered, and evaporated to give a yellow residue, which was chromatographed (4:1 EtOAc: hexane) twice to give 10 mg of white solid. Further purification by HPLC on a 5- μ m Ultrasphere ODS semi-preparative HPLC column (10 × 300 mm, Beckman) with 1:9 H₂O:MeOH as efluant at a flow rate of 3 ml/min and recrystallization from CH₂Cl₂-acetone afforded 4.3 mg (51%) of **5**: m.p. 234 to 235°C (lit. [19] m.p. 234.5–235.5°C); ¹H NMR (CD₃OD) 5.53 (d, J = 5.1 Hz, 1H), 3.75 (m, 1H), 3.45 (m, 1H), 1.16 (s, 6H), 0.99 (s, 3H), 0.96 (d, J = 6.3 Hz, 3H), 0.71 (s, 3H). (lit. [19] ¹H NMR (pyridine-d₅) 5.57 (m, 1H), 5.3–4.5 (m, 1H), 3.92 (m, 1H), 1.28 (s, 6H), 0.93 (s, 3H), 0.61 (s, 3H)).

2.6. (25R)-Cholest-5-ene-3β,26-diol 3β,26-Dibenzoate (10)

As in the preparation of 9 from 1, 76.4 mg (0.190 mmol) of 2, prepared from diosgenin by the method of Kim et al. [22], was converted to crude 10, which was chromatographed (1:19 EtOAc:hexane) to give 102 mg (87%) of 10. Recrystallization from methanol-CH₂Cl₂ gave colorless 10: m.p. 136–137°C (lit. [22] m.p. 136.5–137.5°C); ¹H NMR 8.07 (m, 4H), 7.56 (m, 2H), 7.46 (m, 4H), 5.43 (m, 1H), 4.87 (m, 1H), 4.13 (m, 2H), 1.08 (s, 3H), 1.04 (d, J = 6.6Hz, 3H), 0.95 (d, J = 6.3 Hz, 3H), 0.70 (s, 3H) (lit. [22] ¹H NMR 8.05 (m, 4H), 7.54 (m, 2H), 7.44 (m, 4H), 5.42 (m, 1H), 4.86 (m, 1H), 4.21 (m, 1H), 4.12 (m, 1H), 1.07 (s, 3H), 1.02 (s, 3H), 0.93 (d, J = 6.5 Hz, 3H), 0.69 (s, 3H)); ¹³C NMR 166.8, 166.1, 139.8, 132.9, 132.9, 131.0, 130.7, 129.7, 128.5, 128.4, 122.9, 74.7, 70.1, 56.8, 56.2, 50.1, 42.5, 39.9, 38.4, 37.2, 36.8, 36.2, 35.9, 34.1, 32.9, 32.1, 32.0, 28.4, 28.1, 24.4, 23.5, 21.2, 19.5, 18.9, 17.2, 12.0; (lit. [22] ¹³C NMR 166.6, 166.0, 139.6, 132.8, 132.7, 130.7, 130.5, 129.6, 129.5, 128.3, 128.2, 122.7, 74.5, 70.0, 56.6, 56.0, 50.0, 42.2, 39.7, 38.2, 37.0, 36.6, 36.0, 35.7, 33.9, 32.7, 31.9, 28.2, 27.8, 24.2, 23.3, 21.0, 19.4, 18.7, 17.0, 11.8.)

2.7. (25R)-3β,26-Dihydroxycholest-5-en-7-one 3β,26-Dibenzoate (13)

According to a procedure by Blair et al. [26], to a solution of 101.5 mg (0.163 mmol) of **10** in 0.6 ml of dry pyridine and 20 ml of dry CH_2Cl_2 was added 0.6 g of CrO_3 . The mixture was heated at reflux overnight under N₂, cooled, and decanted. After addition of 3×10 ml of CH_2Cl_2 and 10 ml of ether, the resulting mixture was washed with 4×7 ml of saturated NaHCO₃ solution and the combined aqueous layers were extracted with 15 ml of ether. The combined organic layers were washed with 2×10 ml of 5% HCl, 10 ml of saturated NaHCO₃ solution and 10 ml of brine, dried, filtered, and evaporated to give 215.6 mg of brown residue, which was chromatographed (1:9 EtOAc:hexane) to afford 56.0 mg (54%) of **13**: m.p. 167.5–168.3°C; ¹H NMR 8.06 (m, 4H), 7.57 (m, 2H), 7.46 (m, 4H), 5.76 (m, 1H), 4.98 (m, 1H), 4.17 (m, 2H), 1.27 (s, 3H),

1.03 (d, J = 6.9 Hz, 3H), 0.94 (d, J = 6.3 Hz, 3H), 0.70 (s, 3H); ¹³C NMR 202.1, 166.9, 166.0, 164.0, 133.2, 133.0, 130.7, 130.5, 129.8, 129.7, 128.6, 128.5, 127.0, 73.0 70.1, 54.9, 50.1, 50.0, 45.6, 43.3, 38.9, 38.6, 38.0, 36.2, 35.9, 34.1, 32.9, 29.9, 28.8, 27.7, 26.5, 23.5, 21.4, 19.0, 17.5, 17.5, 12.2. FAB-HRMS (M-H⁺) Analysis Calculated for C₄₁H₅₁O₅: 623.3737. Found: 623.3735. Analysis Calculated for C₄₁H₅₂O₅: C, 78.80; H, 8.39. Found: C, 78.83; H, 8.38.

2.8. (25R)-*Cholest-5-ene-3β*,7α,26-*triol 3β*,26-*Dibenzoate* (16)

As in the preparation of **15** from **12**, 38.6 mg (0.0619 mmol) of **13** was converted to 45.0 mg of crude **16**, which was recrystallized from acetone-CH₂Cl₂ three times to give 35.0 mg (93%) of **16**: m.p. 143.4–144.4°C; ¹H NMR 8.06 (m, 4H), 7.55 (m, 2H), 7.46 (m, 4H), 5.69 (d, J = 5.1 Hz, 1H), 4.92 (m, 1H), 4.17 (m, 2H), 3.87 (m, 1H), 1.07 (s, 3H), 1.03 (d, J = 6.9 Hz, 3H), 0.95 (d, J = 6.3 Hz, 3H), 0.70 (s, 3H); ¹³C NMR 166.9, 166.1, 145.4, 133.0, 130.8, 130.7, 129.8, 129.7, 128.5, 128.5, 125.1, 74.2, 70.2, 65.5, 56.0, 49.6, 42.4, 42.3, 39.3, 38.2, 37.7, 37.7, 37.0, 36.2, 35.9, 34.1, 32.9, 28.5, 27.8, 24.5, 23.4, 20.9, 19.0, 18.5, 17.2, 11.9. Analysis Calculated for C₄₁H₅₄O₅: C, 78.54; H, 8.68. Found: C, 78.03; H, 8.78.

2.9. (25R)-Cholest-5-ene-3β,7α,26-triol (6)

As in the preparation of **5** from **15**, 39.1 mg (0.0625 mmol) of **16** was converted to crude **6**, which was chromatographed (7:3 EtOAc:hexane) to give 20.9 mg (80%) of **6**. Recrystallization from CH₂Cl₂-acetone gave colorless **6**: m.p. 214–216°C; ¹H NMR (CD₃OD) 5.52 (d, J = 5.4 Hz, 1H), 3.74 (m, 1H), 3.47 (m, 1H), 3.34 (m, 2H), 0.99 (s, 3H), 0.94 (d, J = 6.6 Hz, 3H), 0.88 (d, J = 6.6 Hz, 3H), 0.70 (s, 3H); ¹³C NMR (CD₃OD) 146.8, 125.1, 72.2, 68.7, 66.0, 57.6, 50.8, 43.5, 43.4, 43.0, 40.8, 39.1, 38.6, 37.6, 37.3, 37.0, 34.5, 32.3, 30.9, 29.6, 25.3, 24.7, 22.0, 19.4, 18.8, 17.2, 12.3. Analysis Calculated for C₂₇H₄₆O₃: C, 77.45; H, 11.08. Found: C, 77.20; H, 11.06.

2.10. Cholest-5-ene-3 β ,24(S)-diol 3 β ,24-Dibenzoate (11)

To a solution of 21.6 mg (0.0537 mmol) of **3**, prepared by the procedures of Zhang Z, Li D, Blanchard DE, Lear SE, Erickson SK, Spencer TA (being prepared for publication), in 1.0 ml of dry pyridine was added 0.5 ml (4 mmol) of benzoyl chloride. After the mixture was stirred at room temperature overnight, 5 ml of MeOH was added carefully, and the resulting mixture was extracted with 2×20 ml of EtOAc. The combined organic layers were washed with 10 ml each of 10% HCl, H₂O, saturated NaHCO₃ solution, and H₂O, dried, filtered, and evaporated to give 145 mg of yellow solid, which was chromatographed (1:19 EtOAc: hexane) to give 28.3 mg (86%) of **11**: m.p. 183.5–184.1°C (lit. [27] m.p. 182–184°C); ¹H NMR 8.06 (m, 4H), 7.56 (m, 2H), 7.46 (m, 4H), 5.42 (m, 1H), 4.96 (m, 1H), 4.87 (m, 1H), 1.07 (s, 3H), 1.00 (d, J = 6.9 Hz, 3H), 0.98 (d, J = 6.6 Hz, 3H), 0.95 (d, J = 6.6 Hz, 3H), 0.66 (s, 3H); ¹³C NMR 166.7, 166.3, 139.9, 133.0, 133.0, 131.1, 130.9, 129.8, 128.6, 128.5, 123.0, 80.0, 74.8, 56.9, 56.0, 50.3, 42.6, 40.0, 38.5, 37.3, 36.9, 36.0, 32.1, 31.9, 31.5, 28.4, 28.1, 27.8, 24.5, 21.3, 19.7, 19.2, 19.0, 17.6, 12.1 (lit. [28] ¹³C NMR (25.2 MHz) 165.5, 139.4, 132.7, 130.8, 129.6, 128.3, 122.5, 79.6, 74.5, 56.6, 55.6, 50.0, 42.2, 39.7, 38.2, 37.0, 36.6, 35.6, 31.8, 31.8, 31.2, 31.1, 28.1, 27.9, 27.3, 24.3, 21.1, 19.3, 18.9, 18.7, 17.3, 11.8.

2.11. 3β,24(S)-Dihydroxycholest-5-en-7-one 3β,24-Dibenzoate (**14**)

According to a procedure by Ratcliffe and Rodehorst [29], to a solution of 16.0 mg (0.0256 mmol) of **11** in 0.3 ml of pyridine and 10 ml of dry CH₂Cl₂ was added 0.3 g of CrO₃. The mixture was heated at reflux overnight under N₂, decanted and evaporated to give a black residue, which was taken up in 50 ml of ether and filtered. The filtrate was washed with 10 mL of 5% NaHCO₃ solution and 10 ml of brine, dried, filtered, and evaporated to give 38.0 mg of yellow oil which was chromatographed (1:9 EtOAc:hexane) to afford 12.7 mg (78%) of 14: m.p. 143-147°C. Recrystallization from CH₂Cl₂-hexane three times gave 14: m.p. 151.6 to 152.4°C; ¹H NMR 8.05 (m, 4H), 7.56 (m, 2H), 7.45 (m, 4H), 5.75 (s, 1H), 4.96 (m, 2H), 1.27 (s, 3H), 1.00 (d, J = 6.9 Hz, 3H), 0.98 (d, J = 6.9 Hz, 3H), 0.95 (d, J = 6.3 Hz, 3H), 0.67 (s, 300)3H); ¹³C NMR 202.2, 166.7, 166.0, 164.1, 133.3, 133.0, 131.1, 130.5, 129.9, 129.8, 128.6, 128.6, 127.1, 80.0, 73.1, 54.6, 50.2, 50.0, 45.7, 43.4, 38.9, 38.7, 38.1, 36.3, 35.9, 31.8, 31.4, 28.7, 27.8, 27.7, 26.5, 21.4, 19.2, 19.2, 17.6, 17.6, 12.2. Analysis Calculated for C₄₁H₅₂O₅: C, 78.80; H, 8.39. Found: C, 78.56, H, 8.37.

2.12. *Cholest-5-ene-3β*,7α,24(S)*-triol 3β*,24*-Dibenzoate* (17)

As in the preparation of **15** from **12**, except for a reaction time of three rather than 5 h, 34.9 mg (0.0558 mmol) of **14** was converted to 34.6 mg of crude **17**, which was chromatographed (3:17 EtOAc:hexane) to give 24.2 mg (69%) of **17**: m.p. 171–173°C; ¹H NMR 8.05 (m, 4H), 7.56 (m, 2H), 7.45 (m, 4H), 5.67 (d, J = 5.1 Hz, 1H), 4.95 (m, 2H), 3.86 (m, 1H), 1.06 (s, 3H), 1.00 (d, J = 6.9 Hz, 3H), 0.97 (d, J = 6.6 Hz, 3H), 0.95 (d, J = 5.7 Hz, 3H), 0.67 (s, 3H); ¹³C NMR 166.6, 166.1, 145.4, 133.0, 132.9, 131.1, 130.8, 129.8, 129.7, 128.5, 125.1, 80.0, 74.2, 65.4, 55.6, 49.6, 42.4, 42.3, 39.3, 38.2, 37.7, 37.7, 37.0, 35.8, 31.8, 31.4, 28.4, 27.8, 27.6, 24.4, 20.9, 19.1, 19.0, 18.5, 17.5, 11.8. FAB-HRMS (M-H⁺) Analysis Calculated for C₄₁H₅₃O₅: 625.3893. Found: 625.3894.

2.13. Cholest-5-ene-3β,7α,24(S)-triol (7)

As in the preparation of **5** from **15**, 17.5 mg (0.0280 mmol) of **17** was converted to crude **7**, which was chromatographed (4:1 EtOAc:hexane) to give 11.4 mg (97%) of colorless **7** that was homogeneous by TLC. Recrystallization twice from acetone-hexane gave **7**: m.p. 203–204°C; ¹H NMR (CD₃OD) δ 5.53 (d, J = 5.4 Hz, 1H), 3.76 (m, 1H), 3.47 (m, 1H), 3.21 (m, 1H), 1.00 (s, 3H), 0.97 (d, J = 6.6 Hz, 3H), 0.91 (d, J = 6.0 Hz, 3H), 0.89 (d, J = 6.0 Hz, 3H), 0.72 (s, 3H). ¹³C NMR (CD₃OD) 146.8, 125.1, 78.3, 72.2, 66.0, 57.5, 50.8, 43.5, 43.4, 43.0, 40.8, 39.1, 38.6, 38.2, 37.6, 34.7, 33.8, 32.3, 31.8, 29.5, 25.2, 22.0, 19.7, 19.6, 18.8, 17.7, 12.3; EI-HRMS (M⁺-H₂O) Analysis Calculated for C₂₇H₄₂O: 400.3341. Found: 400.3349.

2.14. 24(S),25-Epoxycholesteryl-3β-acetate (18)

To a solution of 40.4 mg (0.101 mmol) of 4, prepared as previously described [30], in 1.5 ml of dry pyridine was added 0.7 ml of acetic anhydride. The resulting mixture was stirred under N₂ for 12 h, 10 ml of saturated NaHCO₃ solution was added, and the mixture was extracted with 3 \times 10 ml of CH₂Cl₂. The combined organic layers were dried, filtered, and evaporated to give 109.7 mg of residue, which was chromatographed on aluminum oxide (active neutral, gamma, -60 mesh, AlfaAesar; 1:19 EtOAc:hexane) to give 36.7 mg (82%) of 18. Recrystallization from CH₂Cl₂-hexane afforded colorless 18: m.p. 113 to 115°C (lit. [31] m.p. 114.5–116.5°C); ¹H NMR 5.38 (m, 1H), 4.60 (m, 1H), 2.68 (m, 1H), 2.03 (s, 3H), 1.30 (s, 3H), 1.27 (s, 3H), 1.02 (s, 3H), 0.94 (d, J = 6.3 Hz, 3H), 0.70 (s, 3H); ¹³C NMR 170.7, 139.8, 122.8, 74.2, 65.1, 58.3, 56.9, 56.2, 50.2, 42.6, 39.9, 38.3, 37.2, 36.8, 35.9, 32.8, 32.0, 29.9, 28.4, 28.0, 25.9, 25.2, 24.5, 21.7, 21.2, 19.5, 18.9, 18.8, 12.1.

2.15. 24(S),25-*Epoxycholest-5-en-3β-ol-7-one, 3β-Acetate* (**19**)

According to a procedure by Salvador et al. [32], to a solution of the crude product (40.6 mg) containing 18, prepared as described above from 28.5 mg (0.0713 mmol) of 4, and 8.1 mg of CuI in 2 ml of dry CH₃CN was added 0.2 ml (5.0-6.0 M solution in decane, 1.0-1.2 mmol) of t-BuOOH. The resulting mixture was stirred at 50°C for 25 h under N₂, 10 ml of 10% Na₂SO₃ was added, and the mixture was extracted with 3×10 ml of CH₂Cl₂. The combined organic layers were washed with 10 ml of saturated NaHCO₃ solution, 10 ml of H₂O, and 10 ml of brine, dried, filtered, and evaporated to give 60.4 mg of yellow residue that was crystallized twice from CH₂Cl₂-hexane to give 22.3 mg (69% from 4) of 19: m.p. 175.5–177.5°C; ¹H NMR 5.70 (s, 1H), 4.72 (m, 1H), 2.69 (m, 1H), 2.05 (s, 3H), 1.31 (s, 3H), 1.27 (s, 3H), 1.21 (s, 3H), 0.95 (d, J = 6.6 Hz, 3H), 0.69 (s, 3H); ¹³C NMR 202.2, 170.6, 164.2, 127.0, 72.5, 65.1, 54.8, 50.2, 50.0, 45.7, 43.4, 38.9, 38.6, 38.0,

36.3, 35.8, 32.8, 28.8, 27.6, 26.6, 25.9, 25.2, 21.5, 21.4, 19.0, 18.9, 17.5, 12.2. Analysis Calculated for $C_{29}H_{44}O_4$: C, 76.27; H, 9.71. Found: C, 76.40; H, 9.63.

2.16. 24(S),25-*Epoxycholest-5-ene-3β*,7α-*diol 3β*-Acetate (**20**)

As in the preparation of **15** from **12**, 24.3 mg (0.0533 mmol) of **19** was converted to 20.4 mg of crude **20**, which was crystallized from ether-hexane to afford 17.3 mg (71%) of **20**: m.p. 185 to 186°C; ¹H NMR 5.63 (d, J = 4.8 Hz, 1H), 4.65 (m, 1H), 3.85 (bs, 1H), 2.69 (m, 1H), 2.04 (s, 3H), 1.31 (s, 3H), 1.27 (s, 3H), 1.01 (s, 3H), 0.95 (d, J = 6.6 Hz, 3H), 0.69 (s, 3H); ¹³C NMR 170.7, 145.4, 124.9, 73.6, 65.4, 65.2, 58.4, 55.9, 49.6, 42.4, 39.3, 38.1, 37.7, 37.7, 37.0, 35.9, 32.8, 28.5, 27.7, 25.8, 25.2, 24.5, 21.6, 20.9, 18.9, 18.8, 18.4, 11.9. Analysis Calculated for C₂₉H₄₆O₄: C, 75.94; H, 10.11. Found: C, 75.75; H, 10.08.

2.17. 24(S),25-Epoxycholest-5-ene-3β,7α-diol (8)

According to a modification of a procedure by Corey and Grogan [21], a solution of 17.3 mg of 20 in 4 ml of 1:4 THF-MeOH containing 0.04 g of NaOH was stirred at rt for 2.5 h, 10 ml of water was added, and the resulting mixture was extracted with 3×10 ml of CH₂Cl₂. The combined organic layers were washed with 10 ml of brine, dried, filtered and evaporated to give 12.6 g of yellow residue that was chromatographed on aluminum oxide (active neutral, gamma, 60 mesh, AlfaAesar; 6:4 EtOAc:hexane) to give 8.3 mg (54%) of colorless 8 that was homogeneous by TLC: m.p. 149.4–150.5°C; ¹H NMR 5.61 (d, J = 4.8 Hz, 1H), 3.86 (bs, 1H), 3.59 (m, 1H), 2.69 (m, 1H), 1.31 (s, 3H), 1.27 (s, 3H), 1.00 (s, 3H), 0.95 (d, J = 6.6 Hz, 3H), 0.70 (s, 3H); ¹³C NMR 146.5, 124.1, 71.6, 65.6, 65.2, 58.4, 56.0, 49.7, 42.5, 42.4, 42.3, 39.4, 37.8, 37.7, 37.3, 36.0, 32.8, 31.7, 28.6, 25.9, 25.2, 24.6, 21.0, 19.0, 18.9, 18.5, 11.9; EI-HRMS (M⁺-H₂O) Analysis Calculated for $C_{27}H_{40}O$: 398.3184. Found: 398.3184.

3. Results and discussion

25-Hydroxycholesterol (1) is commercially available, but adequate amounts of the other three oxysterols had to be synthesized for conversion to their 7α -hydroxy derivatives. 27-Hydroxycholesterol (2) was prepared from diosgenin by the method reported by Schroepfer and co-workers [22]. 24(*S*)-Hydroxycholesterol (3) was prepared from stigmasterol by our recently developed route (by the procedures of Zhang, Li, Blanchard, Lear, Erickson, Spencer, unpublished results) and 24(*S*),25-epoxycholesterol (4) was synthesized using procedures we have recently reported [30].

The strategy for the preparation of the 7α -hydroxy derivatives is illustrated for diols **1** to **3** in Scheme 2. Initial protection of the free hydroxyl groups was accomplished by conversion to the known dibenzoates 9 to 11 [22,25,27] Allylic oxidation at C7 can be effected to afford 7-hydroxy derivatives directly, but produces a mixture of 7α and 7β isomers [20]. Alternatively, oxidation can be effected to form a 7-keto derivative which can be stereoselectively reduced to the axial 7α -hydroxy compound by use of a hindered complex metal hydride, such as L-selectride [25]. Accordingly, each of 9 to 11 was subjected to allylic oxidation (CrO₃/pyridine for 9 [24] and 11 [29]; PCC for 10) [27] to afford enones 12 to 14 in good yield, and these were in turn reduced using L-selectride to yield the 7α -hydroxy steroids 15 to 17 with complete stereoselectivity. The 1 H NMR spectra of each of the crude hydride reduction products showed only the relatively deshielded doublet for a 6β-H and none of the relatively shielded singlet expected for a 6α -H [25]. Removal of the dibenzoate protecting groups from 15 to 17 by treatment with KOH in MeOH then produced the desired triols 5 to 7 in excellent yield.

Synthesis of 7α -hydroxy-24(S),25-epoxycholesterol (8) (Scheme 3) required a modified reaction sequence because of the acid liability of the epoxide function. The known [31] acetate 18 of 24(S), 25-epoxycholesterol (4) was in this case used as the protected substrate for allylic oxidation. Several methods, including those used for the conversion of 9 to 11 to 12 to 14, caused destruction of the epoxide group as well as allylic oxidation. Attempted direct allylic oxidation of 4 was also unsuccessful. The conversion of 18 to 19 was achieved, however, by the procedure of Salvador et al. [32], with t-BuOOH and CuI and CH₃CN, as had been employed by Corey and Grogan [21] for the analogous reaction on 24(S),25-epoxycholesteryl benzoate. Reduction with L-selectride yielded 20 and cleavage of the acetate protecting group then afforded 8. Thus, all four 7α -hydroxylated oxysterols 5 to 8 are now available as reference standards and procedures have been developed for their synthesis in larger amounts as required for further experimentation.

References

- Janowski BA, Willy PJ, Devi TR, Falck JR, Mangelsdorf DJ. An oxysterol signalling pathway mediated by the nuclear receptor LXRα. Nature 1996;383:728–31.
- [2] Lehmann JM, Kliewer SA, Moore LB, Smith-Oliver TA, Oliver BB, Su J-L, Sundseth SS, Winegar DA, Blanchard DE, Spencer TA, Willson TM. Activation of the nuclear receptor LXR by oxysterols defines a new hormone response pathway. J Biol Chem 1997;272: 3137–40.
- [3] Janowski BA, Grogan MJ, Jones SA, Wisely GB, Kliewer SA, Corey EJ, Mangelsdorf DJ. Structural requirements of ligands for the oxysterol liver X receptors LXRα and LXRβ. Proc Natl Acad Sci USA 1999;96:266–71.
- [4] Axelson M, Shoda J, Sjövall J, Toll A, Wikvall K. Cholesterol is converted to 7α-hydroxy-3-oxo-4-cholestenoic acid in liver mitochondria. J Biol Chem 1992;267:1701–4.
- [5] Dueland S, Nenseter MS, MacPhee AA, Davis RA, Trawick JD. Expression of 7α-hydroxylase in non-hepatic cells results in liver phenotypic resistance of the low density lipoprotein receptor to cholesterol repression. J Biol Chem 1992;267:22695–8.

- [6] Martin KO, Reiss AB, Lathe R, Javitt NB. 7α-Hydroxylation of 27-hydroxycholesterol: biological role in the regulation of cholesterol synthesis. J Lipid Res 1997;38:1053–8.
- [7] Zhang J, Dricu A, Sjövall J. Studies on the relationships between 7α-hydroxylation and the ability of 25- and 27-hydroxycholesterol to suppress the activity of HMG-CoA reductase. Biochim Biophys Acta 1997;1344:241–9.
- [8] Kandutsch AA, Chen HW. Inhibition of sterol synthesis in cultured mouse cells by cholesterol derivatives oxygenated in the side chain. J Biol Chem 1974;249:6057–61.
- [9] Taylor FR, Saucier SE, Shown EP, Parish EJ, Kandutsch AA. Correlation between oxysterol binding to a cytosolic binding protein and potency in the repression of hydroxymethylglutaryl coenzyme A reductase. J Biol Chem 1984;259:12382–7.
- [10] Johnson KA, Morrow CJ, Knight GD, Scallen TJ. In vivo formation of 25-hydroxycholesterol from endogenous cholesterol after a single meal, dietary cholesterol challenge. J Lipid Res 1994;35: 2241–53.
- [11] Lala DS, Syka PM, Lazarchik SB, Mangelsdorf DJ, Parker KL, Heyman RA. Activation of the orphan nuclear receptor steroidogenic factor 1 by oxysterols. Proc Natl Acad Sci USA 1997;94:4895–4900.
- [12] Saucier SE, Kandutsch AA, Gayen AK, Swahn DK, Spencer TA. Oxysterol regulators of 3-hydroxy-3-methylglutaryl-coA reductase in liver. Effect of dietary cholesterol. J Biol Chem 1989;264:6863–9.
- [13] Esterman AL, Baum H, Javitt NB, Darlington GJ. 26-Hydroxycholesterol: regulation of hydroxymethylglutaryl-coA reductase activity in Chinese hamster ovary cell culture. J Lipid Res 1983;24:1304–9.
- [14] Andersson S, Davis DL, Dahlbäck H, Jörnvall H, Russell DW. Cloning, structure, and expression of the mitochondrial cytochrome P-450 sterol 26-hydroxylase, a bile acid biosynthesis enzyme. J Biol Chem 1989;264:8222–9.
- [15] Rennert H, Fischer RT, Alvarez JG, Trzaskos JM, Strauss JF III. Generation of regulatory oxysterols: 26-hydroxylation of cholesterol by ovarian mitochondria. Endocrinology 1990;127:738–46.
- [16] Lund E, Breuer O, Bjorkhem I. Evidence that 24- and 27-hydroxylation are not involved in the cholesterol-induced down-regulation of hydroxymethylglutaryl-CoA reductase in mouse liver. J Biol Chem 1992;267:25092–7.
- [17] Erickson SK, Lear SR, Gayen AK, Spencer TA. Oxysterol regulation of cholesterol metabolism. Circulation 1994;90:420.
- [18] Spencer TA. The squalene dioxide pathway of steroid biosynthesis. Acc Chem Res 1994;27:83–90.

- [19] Nagano H, Poyser JP, Cheng K-P, Luu B, Ourisson G, Beck JPL. Chimie et biochimie de drogues chinoises II-stérols hydroxylés cytotoxiques sur des cellules cancéreuses. Synthèse et activité biologique. J Chem Res(m) 1977;9:2522–71.
- [20] Shoda J, Axelson M, Sjövall J. Synthesis of potential C₂₇-intermediates in bile acid biosynthesis and their deuterium-labeled analogs. Steroids 1993;58:119–25.
- [21] Corey EJ, Grogan MJ. Stereocontrolled syntheses of 24(S),25-epoxycholesterol and related oxysterols for studies on the activation of LXR receptors. Tetrahedron Lett 1998;39:9351–4.
- [22] Kim H-S, Wilson WK, Needleman DH, Pinkerton FD, Wilson DK, Quiocho FA, Schroepfer GJ Jr. Inhibitors of sterol synthesis. Chemical synthesis, structure, and biological activities of (25*R*)-3β,26dihydroxy-5α-cholest-8(14)-en-15-one, a metabolite of 3β-hydroxy-5α-cholest-8(14)-en-15-one. J Lipid Res 1989;30:247–61.
- [23] Beckwith ALJ. Hydroxylation of 5α,6β-dibromocholestan-3β-yl acetate by chromic acid. J Chem Soc 1961:3162–4.
- [24] Parish EJ, Wei TY. Allylic oxidation of Δ⁵-steroids with pyridinium chlorochromate (PCC) and pyridinium dichromate (PDC). Synth Commun 1987;17:1227–33.
- [25] Kumar V, Amann A, Ourisson G, Luu B. Stereospecific synthesis of 7β- and 7α-hydroxycholesterols. Synth Commun 1987;17:1279–86.
- [26] Blair IA, Phillipou G, Seaborn C. Synthesis of C-7–2H2 steroids for human metabolism studies. J Labelled Compd Radiopharm 1978;15: 645–55.
- [27] Ikekawa N, Morisaki M, Koizumi N, Sawamura M, Tanaka Y, De-Luca HF. Synthesis and biological activity of $24\xi^1$ - and $24\xi^2$ -hydroxyvitamin D₃. Biochem Biophys Res Commun 1975;62:485–91.
- [28] Koizumi N, Fujimoto Y, Takeshita T, Ikekawa N. Carbon-13 nuclear magnetic resonance of 24-substituted steroids. Chem Pharm Bull 1979;27:38–42.
- [29] Ratcliffe R, Rodehorst R. Improved procedure for oxidation with the chromium-trioxide-pyridine complex. J Org Chem 1970;35:4000–2.
- [30] Tomkinson NCO, Willson TM, Russel JS, Spencer TA. Efficient, stereoselective synthesis of 24(S),25-epoxycholesterol. J Org Chem 1998;63:9919–23.
- [31] Emmons GT, Wilson WK, Schroepfer GJ Jr. 24,25-Epoxysterols. Differentiation of 24R- and 24S-epimers by ¹³C nuclear magnetic resonance spectroscopy. J Lipid Res 1989;30:133–8.
- [32] Salvador JAR, Melo MLS, Campos Neves AS. Copper-catalysed allylic oxidation of Δ^5 -steroids by *t*-butyl hydroperoxide. Tetrahedron Lett 1997;38:119–22.