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Articles

Activated Ketone Based Inhibitors of Human Renin

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Application of the concept of activated ketones to the design of novel and potent transition-state analog inhibitors of the aspartyl protease renin is described. Three different classes of peptidic activated ketones were synthesized: 1,1,1-trifluoromethyl ketones, α -keto esters, and α -diketones. The corresponding alcohols were also evaluated as renin inhibitors in each series. While the trifluoromethyl alcohol 12 ($I_{50} = 4000 \text{ nM}$) was equipotent to the simple methyl alcohol 7 ($I_{50} =$ 3200 nM), the structurally similar α -hydroxy esters (32 and 30, I_{50} 's = 5.3 and 4.7 nM, respectively) and α -hydroxy ketones (41 and 42, I_{50} = 23 and 15 nM, respectively) were 150–300-fold more active. The hydrating capability of the activated ketone functionality was important for intrinsic potency in the case of trifluoromethyl ketones, as illustrated by the significantly better activity of trifluoromethyl ketone 13 ($I_{50} = 250 \text{ nM}$) compared to its alcohol analog 12 ($I_{50} = 4000 \text{ nM}$). It was however unimportant for the α -keto ester (20 and 31, $I_{50} = 15$ and 4.1 nM, respectively) and α -diketone (43 and 44, $I_{50} = 52$ and 28 nM, respectively) based inhibitors, since their activity was essentially similar to that of the corresponding alcohols. These results collectively suggest that, whereas the trifluoromethyl ketones derive their renin inhibitory potency primarily from their ability to become hydrated, this is not a critical feature for the activity of α -dicarbonyl-based inhibitors. The α -keto ester and α -diketone based renin inhibitors benefit predominantly from the hydrophobic and/or H-bonding type binding interactions of the neighboring ester or acyl group itself, rather than the ability of this group to deactivate the adjacent ketone group and thereby make it susceptible to hydration.

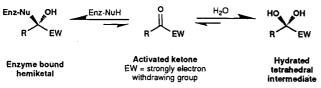
Introduction

Renin is an aspartyl protease that cleaves angiotensinogen to the decapeptide angiotensin I, which in itself is inactive but is hydrolyzed by angiotensin converting enzyme (ACE) to the octapeptide angiotensin II, a potent vasoconstrictor and stimulant of aldosterone secretion. Captopril, the first orally active ACE inhibitor, has demonstrated that interruption of the renin-angiotensin system is of therapeutic benefit in hypertension and congestive heart failure.¹ Inhibitors of renin would be expected to produce the same result and therefore might constitute a novel alternative to ACE inhibitors. This has resulted in intensive research in this area in several laboratories over the last decade.²

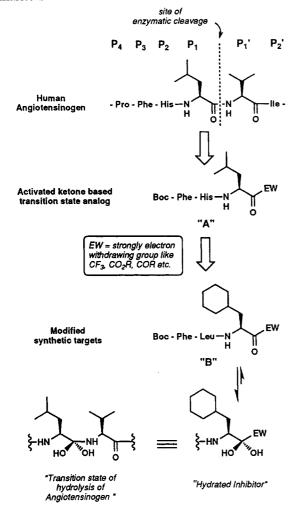
Activated ketone based inhibitors have found application for all of the four different classes of proteases.³ This versatility arises from the fact that they exist as hydrates in aqueous media and can thus directly serve as transitionstate analogs and/or they can react with a nucleophilic residue (e.g. serine hydroxyl or cysteine thiol) to form a reversible, hemiacetal-type intermediate (Scheme I). Either pathway leads to mimics of the tetrahedral intermediate formed during peptide-bond hydrolysis and thus, such compounds can be viewed as transition-state-analog inhibitors. Very elegant applications of this simple yet powerful concept were originally demonstrated by Abeles

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Scheme I



Scheme II. Design of Activated Ketone Based Renin Inhibitors



and co-workers by making fluoro ketone based proteinase inhibitors.⁴

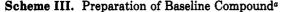
During our efforts in the renin program, we investigated the application of this principle in the design of novel renin inhibitors. Thus, insertion of an activated ketone functionality at the site of enzymatic cleavage $(P_1-P_1'Leu-$ Val linkage) in a human renin substrate analog could potentially mimic the transition state of this reaction (Scheme II). Specifically, the investigation of three classes of activated ketones, namely the 1,1,1-trifluoromethyl ketones, α -keto esters, and α -diketones was undertaken in this study. For synthetic simplicity, histidine at the P_2 position of human renin sequence was replaced by leucine in our initial studies since the activated ketone functionality may not be compatible with a basic functional group within the same molecule (vide infra). Such a replacement has now been reported to be acceptable for several classes of renin inhibitors.^{2,5-8} For enhanced potency, the isobutyl side chain of leucine at P_1 was replaced by a cyclohexylmethyl group, a substitution that usually leads to dramatic levels of improvement in the potency of renin inhibitors.^{2,9}

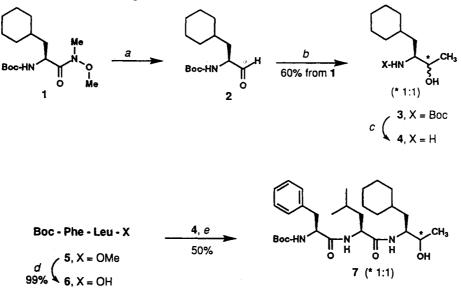
Chemistry

Preparation of the Baseline Compound 7. The α -amino aldehyde 2 was best prepared by LiAlH₄ treatment of the Weinreb amide 1¹⁰ and used directly without any further purification¹¹ (Scheme III). Prolonged storage of 2 was avoided to prevent possible epimerization of this sensitive molecule.¹² Treatment of 2 with MeMgBr afforded a 1:1 mixture of the amino alcohols 3. Deprotection to 4 followed by coupling with the dipeptide 6 yielded the alcohol 7.

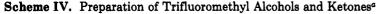
Preparation of the Trifluoromethyl Ketone 13.13 At the outset of this project, no general method existed for the synthesis of peptidic trifluoromethyl ketones like 13. Having decided to prepare 13 from its alcohol precursor 12, the main task was to prepare the trifluoromethyl amino alcohol 11. We reasoned that 11 could be prepared by Curtius rearrangement of the appropriately protected carboxylic acid such as 9. Alternatively, one can also utilize the nitro group as an amine equivalent to arrive at an intermediate such as 11, an approach that has been independently disclosed by Abeles¹⁴ and Trainor.¹⁵ Thus, condensation of the dianion of 3-cyclohexyl propionic acid with trifluoroacetaldehyde generated in situ from its ethyl hemiacetal¹⁶ yielded the β -hydroxy acid 8 as a racemic mixture of erythro and threo isomers in 67% yield (Scheme IV). The alcohol was protected as the silvl ether 9 in essentially quantitative yields. The Curtius rearrangement of 9 utilizing diphenylphosphoryl azide (DPPA) deserves some comments.¹⁷ In our observation, it proceeds more efficiently in a nonpolar solvent such as hexane despite the initial immiscibility of the carboxylic acid. For example, under identical conditions, rearrangement of acid 9 in hexane and THF afforded the amine 10 in 75%and 41% yields, respectively. A second critical feature of this reaction is the presence of excess base. Thus, a mixture of the acid 9, DPPA, and triethylamine (1.0 equiv) in hexane was refluxed for 3 h, at which stage complete isocyanate formation was indicated by IR (C=O at 2250 cm^{-1}). After addition of benzyl alcohol to the refluxing solution, very little reaction was observed even after an additional 2 h. However, upon the addition of another equivalent of triethylamine, immediate product formation was revealed by TLC. The reaction was left for overnight reflux and after purification, afforded the rearranged product in 75% overall yield. Intermediate 10 was completely deprotected¹⁸ and the amine was cleanly isolated as its tosylate salt 11 in 85% overall yield from 10. DCC/HOBt coupling of 11 with the dipeptide acid 6 afforded the trifluoromethyl alcohol 12 in 75% yield as a 1:1.5:2.7:2.7 mixture of four diastereomers as determined by HPLC. Dess-Martin periodinane¹⁹ proved to be the reagent of choice for the oxidation of tripeptidic trifluoromethyl alcohol 12 to the ketone 13.20

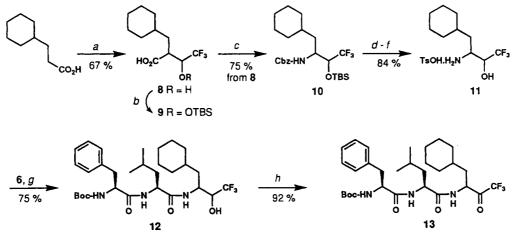
Preparation of α -Keto Esters. Preparation of peptidic α -keto esters was pursued by two routes (Scheme V). First, a nonchiral synthesis via a modified Dakin-West reaction²¹ was undertaken and this was later followed by an investigation of new routes for the chiral synthesis of such compounds. Thus, the tripeptide 19, prepared by straightforward coupling and deprotections, was subjected to the modified Dakin-West reaction.²² Although our initial attempts were less rewarding (11% yield of 20), substantial improvement was realized when freshly distilled oxalyl chloride and an excess of base (5.0 equiv pyridine) were employed for the reaction, and if the enol anhydride formed





^a Reagents: (a) LiAlH₄, Et₂O; (b) MeMgBr, THF; (c) anhydrous HCl in EtOAc; (d) 1 N NaOH, MeOH; (e) 4 and 6, DCC, HOBt, iPr₂NEt, THF.



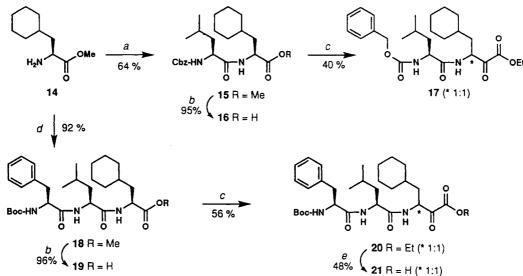


^a Reagents: (a) LDA, [CF₃CHO], THF, -20 °C; (b) 1. Et₃N, TBSOTf; 2. K₂CO₃; (c) Et₃N, DPPA, BnOH, hexane, reflux 14 h; (d) H₂, PD(OH)₂/C; (e) TBAF; (f) pT₈OH; (g) Boc-Phe-Leu-OH 6, DCC, HOBt, iPr₂NEt, THF; (h) Dess-Martin [O].

employed for the reaction, and if the enol anhydride formed during the course of the reaction was hydrolyzed by stirring with NaHCO₃/EtOH at room temperature rather than reflux temperature (56%). The dipeptidic α -keto ester 17 was synthesized in an analogous manner starting from 14. Attempts were also made at synthesizing α -keto esters bearing histidine instead of leucine at the P₂ position via this route.²³ Mild hydrolysis of **20** provided the α -keto acid **21** in moderate yields (48%).

The encouraging biological data on the tripeptidic α -keto ester 20 (vide infra) prompted us to develop a chiral route for these molecules. While several retrosynthetic disconnections were envisioned and attempted,²⁴ only the most successful strategy is discussed below. An obvious advantage of this route is that it provides an easy access to the corresponding α -hydroxy esters, which can also be considered as transition-state analogs.²⁵ Condensation of the ester anion synthon tris(methylthio)methane²⁶ with the aldehyde 2 gave a diastereomeric mixture (4:1) of the α -hydroxy ortho thioesters 22 and 23 in 63% overall yield. Most of the major diastereomer 22 (34%) crystallized cleanly from the crude reaction mixture and chromatographic purification of the rest of the material gave a further 16% of 22 and 13% of the minor, slow-moving diastereomer 23. Mercury-catalyzed hydrolysis²⁷ of diastereomers 22 and 23 afforded the corresponding α -hydroxy esters 24 and 25, respectively. At this stage, stereochemical correlations were made in the following manner.²⁸ The two diastereomers were individually deprotected with 1:1 TFA/CH₂Cl₂ and the amine salts coupled with CDI to yield the cyclic carbamates 26 and 27. In the ¹H NMR, the H_1 proton of *cis* oxazolidinone is expected to appear downfield and have a larger coupling constant when compared to the trans isomer.²⁹ A remote substituent effect in ¹³C NMR is the steric compression effect wherein sterically perturbed carbon atoms are expected to appear at higher field than similar carbons that are not crowded.³⁰ The spectral data summarized in Scheme VI led to the unambiguous assignment of trans and cis stereochemistry for oxazolidinones 26 and 27, respectively. This in turn translated to anti stereochemistry for the major diastereomer 22 and syn for the minor isomer 23. It was gratifying to realize that for peptidic α -hydroxy esters, superior potency was demonstrated by the trans diastereomers derived from the major isomer 22 (vide infra).

Scheme V. Preparation of α -Keto Esters via Modified Dakin-West Reaction^{α}



^a Reagents: (a) Cbz-Leu-OH, DCC, HOBt, iPr₂NEt; (b) 1 N NaOH, MeOH; (c) EtOC(O)C(O)Cl, pyridine, DMAP, THF/DMF (1:1), reflux, 2 h; (d) Boc-Phe-Leu-OH 6, DCC, HOBt, iPr₂NEt, THF/DMF (1:1); (e) K₂CO₃, EtOH.

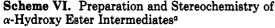
The trans- α -hydroxy ester 24 was deprotected and coupled with Boc-L-leucine to give 28 (Scheme VII). Coupling of the free amine derived after removal of the Boc group from 28 with the morpholino acid 29³¹ provided the anti-hydroxy ester 30. Oxidation of tripeptide 30 with the Dess-Martin periodinane reagent proceeded smoothly to provide the chiral α -keto ester 31 in 90% yield. Thus, our objective of achieving a chiral synthesis was accomplished, but, unfortunately, we found that the α -keto ester 31 slowly epimerized on standing. Thus, a ¹³C NMR of 31 run immediately after its synthesis showed it to be a single diastereomer, whereas a sample run after 1 week revealed a roughly 1:1 mixture of two diastereomers, thereby highlighting the relative susceptibility of peptidic α -keto esters toward epimerization.³² The free amine of 24 was also coupled with the dipeptide acid 6 to provide the trans hydroxy ester tripeptide 32. The corresponding syn diastereomer 33 was prepared in an analogous manner starting with the erythro intermediate 25.

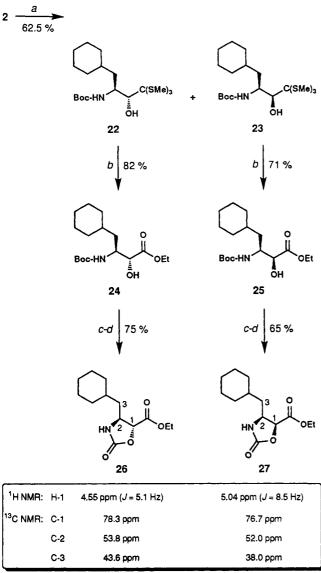
It was interesting to compare the hydrating ability of the two classes of compounds. Thus, while the trifluoromethyl ketone 13 was readily hydrated and was isolable only as a hydrate, the keto ester 31 existed mainly in the free ketone form. The ¹³C NMR of 31 in CD₃OD initially showed only the ketone carbonyl (194 ppm) and the peak due to hemiketal formation (100 ppm) appeared gradually and intensified with time at the expense of the carbonyl peak (Scheme VIII). This suggested that the hemiketal formation was a slow equilibrium process in the case of the α -keto ester 31.

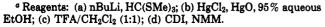
Preparation of α -diketones. Compared to the fluoro ketones and α -keto esters, the α -diketones are a much less well-studied class of inhibitors.^{33,34} To date, there has been only one specific report on the preparation of β -amino α -diketones, but it is restricted to the preparation of methyl diketones.³⁴ We adopted a strategy similar to the one utilized for α -keto esters for preparing this class of compounds, utilizing 1,3-dithiane as a formyl anion synthon.³⁵ If successful, this would constitute a versatile route for their preparation and enable access to a wide variety of peptidic diketones in a straightforward manner. An additional advantage of this route would be that the penultimate α -hydroxy ketone intermediates would also

be available for evaluation as proteinase inhibitors. Thus, reaction of the lithio anion of 2-n-propyl-1,3-dithiane (36)³⁶ with aldehyde 2 gave the single diastereomer 37 in 35%yield (Scheme IX). Although a definitive assignment of the absolute stereochemistry of 37 cannot be made, on the basis of our own experience with related nucleophiles like ortho thioesters, the reaction was expected to favor the formation of the desired anti diastereomer.³⁷ This is further supported by the relative potency of the corresponding hydroxy ketones (vide supra, see the biological section). Removal of the Boc group with anhydrous HCl in ethyl acetate followed by coupling of the amine salt 38 with appropriate dipeptide acids 6 and 35 gave the tripeptides 39 and 40, respectively. Oxidative cleavage of the dithiane group in 39 with ceric ammonium nitrate³⁸ furnished the hydroxy ketone 41 in 48% yield. Subsequently, we have realized that this deprotection can in general be accomplished very efficiently and cleanly when thallium nitrate³⁹ is employed as the oxidant. Thus, removal of the dithiane group from the tripeptide 40 with the latter reagent gave the desired α -hydroxy ketone 42 in 83% yield. The hydroxy ketones 41 and 42 were smoothly oxidized in high yields to α -diketones using Dess-Martin periodinane.¹⁹

Preparation of histidine-containing diketones required a judicious choice of complementary protecting groups for the imidazole ring of histidine and for the diketone moiety, and coupling procedures that would be tolerated by these protecting groups. Among several histidineprotecting groups that are available, the tosyl group appeared to be most compatible with our route for preparing α -diketones, i.e. the reaction conditions needed for regeneration of the diketone functionality from the hydroxy dithiane fragment. Since the tosyl group cannot survive the standard DCC/HOBt coupling conditions, Boc-N-tosylhistidine $(45)^{40}$ was coupled with 38 using triethylamine and diphenylphosphoryl azide yielding 71% of the desired product 46⁴¹ (Scheme X). A substantial amount (21%) of the deprotected compound 47 resulting from removal of the tosyl group was also obtained which fortunately could be easily reprotected using TsCl/Et₃N to 46 in essentially quantitative yields. Treatment of 46 with thallium nitrate afforded the hydroxy ketone 48,







which after removal of the tosyl group with HOBt/ MeOH^{40b} furnished the histidine-containing dipeptidic α -hydroxy ketone 49. On the other hand, oxidation of 48 followed by HOBt treatment of the intermediate diketone 50 afforded the histidine bearing 1,2-diketone 51. As expected, both 49 and 51 were found to be water soluble. To our disappointment, the diketone 51 was found to be unstable to storage, and it was therefore not tested for renin inhibitory activity.

Biology

The I_{50} values of various activated ketones and their precursor alcohols against human renin are summarized in Table I.

Since alcohols can also be viewed as transition-state analogs, both the activated ketones and the corresponding alcohols were tested for inhibitory potency against human renin. The 1:1 diastereomeric mixture of the simple baseline alcohol 7 was only weakly active when tested against human renin ($I_{50} = 3200$ nM). In the tripeptide alcohol 12, the methyl group of 7 is replaced by a trifluoromethyl group. This substitution increases the

 Table I. In Vitro Activity of Various Classes of Renin Inhibitors^a

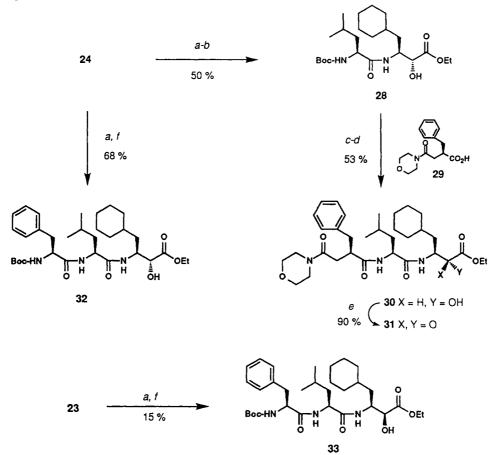
compd no.	I ₅₀ (nM, human renin)
Baseline Compound	
7	3200 ± 750 (2)
Trifluoromethyl Alcohol and Ketone	
12	4000 ± 750 (2)
13	250 • 32 (3)
α -Hydroxy Esters and Ketones	
17	52 000 (1)
20 ^b	$15 \pm 2 \ (4)^{b}$
21	64 000 (1)
30	4.7 ± 1.1 (3)
31	4.1 ± 0.3 (3)
32	5.3 ± 0.5 (3)
33	110 • 19 (3)
α -Hydroxy Ketones and Diketones	
39	2200 ± 500 (2)
41	23 ± 2 (2)
42	14 🛳 2 (3)
43	52 ± 7 (3)
44	28 ± 2 (2)
49	420 000 (1)

^a See the general Experimental Section for a description of the method for determining the I_{50} values of these inhibitors. Number of determinations is indicated in parentheses. ^b Compound 20: I_{50} (porcine pepsin) = 2700 nM; I_{50} (bovine cathepsin D) = 1300 nM.

acidity of the adjacent hydroxyl group by ca. 4 pKa units, thereby making it a better hydrogen atom donor. Additionally, the fluorine atoms are themselves capable of weak H-bond acceptor interactions. The nearly equal activity of 12 (I_{50} = 4000 nM) to 7 suggests that the beneficial effect from such binding contributions, if any, was minimal. The 1,1,1-trifluoromethyl ketone 13 ($I_{50} = 250 \text{ nM}$), which existed completely in its hydrated form on the basis of ¹³C NMR data (see Experimental Section), was about 10-fold more active than the corresponding alcohol 12. This difference in activity demonstrates the importance of the hydrated carbonyl group in binding to the enzyme. After the completion of our work, a similar magnitude of activity difference was reported in the literature between various peptidic perfluoroalkyl alcohols and ketones as renin inhibitors.⁴² A similar activity trend has also been observed with various difluorostatine alcohols and the corresponding difluorostatone-based renin inhibitors.43-46 However, this is not a general rule since examples with minimal difference between the activity of fluoro alcohols and the corresponding ketones have also been reported.48

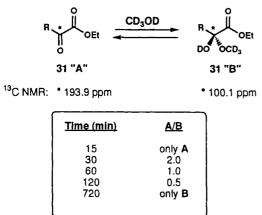
The inhibitory potency of α -hydroxy ester 32 ($I_{50} = 5.3$ nM) compares well with that of similar renin inhibitors reported in the literature.²⁵ The corresponding syn analog 33 was substantially less active $(I_{50} = 110 \text{ nM})$, indicating the preference for anti, statine-like stereochemistry of the hydroxyl group for this class of renin inhibitors.^{25,48} Interestingly enough, the α -hydroxy ester 32 was over 100fold more active than the simple tripeptide alcohols 7 and 12, suggesting a significant contribution toward binding from the ester group. Participation in productive Hbonding interactions by the ethyl ester moiety and/or hydrophobic interaction by its ethyl group at the S_1 subsite are the most likely explanations for this impressive 2 orders of magnitude enhancement in activity. This observation is in accordance with the concept of extended binding for renin inhibitors. In general, various tripeptide renin inhibitors have exhibited significant levels of improvement in activity when modified by introduction of groups capable of interacting at the S_1' subsite of the enzyme.⁴²⁻⁴⁷ For example, while our trifluoromethyl alcohol 12 was only

Scheme VII. Preparation of α -Keto Esters via Oxidation of α -Hydroxy Esters^a



^a Reagents: (a) TFA/CH₂Cl₂ (1:1); (b) DCC, HOBt, iPr₂NEt, Boc-Leu-OH; (c) HCl/AcOH, EtOAc; (d) DCC, HOBt, iPr₂NEt, **29**; (e) Dess-Martin [O]; (f) DCC, HOBt, iPr₂NEt, Boc-Phe-Leu-OH 6.

Scheme VIII. Hemiketal Formation of α -Keto Esters

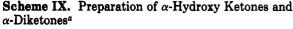


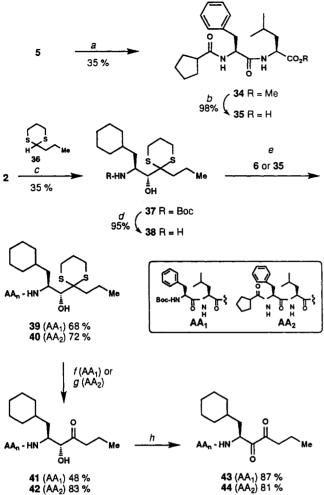
moderately active $(I_{50} = 4000 \text{ nM})$, the structurally related perfluoropropyl alcohol has been reported to possess 75fold better potency $(I_{50} = 52 \text{ nM})$.⁴² Similarly, a difluoro ketone bearing a retroamide bond and an isovaleryl side chain has also been found to be 70-fold more potent than the trifluoromethyl ketone 13.⁴⁶ However, it is important to note that precise assay conditions, especially the pH, can also markedly influence the observed intrinsic activity of renin inhibitors.

Correcting for the diastereomeric contents, the α -keto ester 20 and the related analog 31 ($I_{50} = 15$ and 4.1 nM respectively), wherein the Boc-Phe group of 18 is replaced by a morpholino succinamide moiety, were essentially equipotent to their hydroxy analogs 32 and 30 ($I_{50} = 5.3$ and 4.7 nM, respectively). This contradicts the 10-fold superior activity observed by us and others for fluoro ketone based activated ketones in comparison to the corresponding fluoro alcohols. Additionally, compared to the trifluoromethyl ketone 13, the keto esters 20 and 31 had 25-fold better potency. These results collectively suggest that contributions from H-bonding and/or hydrophobic binding interactions of an ethyl ester group are superior to that of a trifluoromethyl group (32 and 30 vs 12 for alcohols, 20 and 31 vs 13 for ketones). Secondly, it is these interactions of the ethyl ester group that are more critical for the intrinsic activity of α -keto esters, rather than the ability of the ester group to activate and thereby hydrate the adjacent ketone functionality (20 vs 32, 31 vs 30).

Truncation of the tripeptide moiety of α -keto ester 20 to the dipeptide analog 17 ($I_{50} = 52\,000$ nM) proved to be detrimental toward potency, illustrating the requirement of a P₄-P₃ amide group for good binding. Not unexpectedly, the α -keto acid 21 ($I_{50} = 64\,000$ nM) was more than 3 orders of magnitude weaker in potency than the corresponding ethyl ester 20, highlighting the incompatibility of a negatively charged carboxylic acid function in close proximity with the two catalytic aspartic acid residues residing at the active site of the enzyme. The keto ester 20 showed a very high level of specificity toward human renin. For example, its I_{50} values against the aspartic proteinases pepsin and cathepsin D were 2700 and 1300 nM, respectively, making it roughly 2 orders of magnitude more selective for the targeted enzyme, renin.

The α -diketones are a relatively much less well-studied class of inhibitors.³³ After the completion of our own work,





^a Reagents: (a) (i) HCl, AcOH; (ii) DCC, HOBt, iPr_2NEt , C_5H_9 -CO₂H; (b) NaOH, MeOH; (c) nBuLi, 36; (d) anhydrous HCl, EtOAc; (e) DCC, HOBt, iPr_2NEt , 6 or 35; (f) Ce(NH₄)₂(NO₃)₆; (g) Tl(NO₃)₃· 3H₂O, 3:1 MeOH/H₂O; (h) Dess-Martin [O].

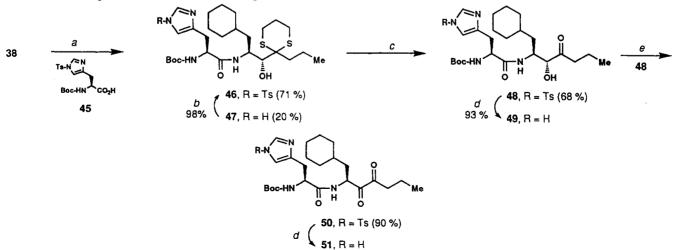
their application as inhibitors of cysteine and serine proteinases was recently reported in the literature.⁴⁸ The tripeptidic α -diketones 43 and 44 ($I_{50} = 52$ and 28 nM, respectively) exhibited nanomolar levels of activity when evaluated for inhibition of human renin. Unlike the α -keto esters, the α -diketones did not epimerize and retained

Scheme X. Preparation of Histidine-Bearing α -Diketones^a

their chirality upon storage. While we did not evaluate the hydrating ability of this class of compounds, NMR studies in D₂O on related compounds reported in the literature suggest that one of the carbonyl groups is indeed hydrated.⁴⁸ The inhibitory potency of corresponding α -hydroxy ketones 41 and 42 ($I_{50} = 23$ and 14 nM, respectively) was actually 2-fold better than the corresponding diketones 43 and 44, once again highlighting the predominance of binding interactions associated with the acyl group, as was observed for the α -hydroxy and ketoester class of compounds. An attempt to truncate the size of these inhibitors was unsuccessful, as evidenced by the poor activity of dipeptidic α -hydroxy ketone 49 (I_{50} = 420 000 nM). Not unexpectedly, the dithiane intermediate 39 ($I_{50} = 2.200 \text{ nM}$) of analogs 41 and 42 displayed inferior potency, highlighting the important contribution of the free carbonyl group and/or the poor fit of the bulky dithiane residue to that region of the enzyme.

Summary

The synthesis and biological evaluation of three different classes, namely the 1,1,1-trifluoromethyl ketones, α -keto esters, and α -diketone type of activated ketone based tripeptides as novel and potent ($I_{50} = 4-250$ nM) renin inhibitors is reported. In each series, the corresponding alcohols were also evaluated for their ability to inhibit human renin. The hydrating capability of the activated ketone functionality was important for intrinsic potency in the case of trifluoromethyl ketones, as illustrated by the 10-fold better activity of trifluoromethyl ketone analog 13 compared to its alcohol 12. It was however unimportant for the α -keto ester (20, 31) and α -diketone (43, 44) based renin inhibitors, since they were found to be essentially equipotent to their corresponding alcohols (32, 30 and 41, 42, respectively). This suggests that potential side chain hydrophobic and/or H-bonding interactions of the adjacent ester or acyl group was more critical for *in vitro* activity than its ability to deactivate and thereby hydrate the neighboring ketone functionality. The general synthetic method developed for preparing peptidic α -diketones should facilitate the application of this relatively less wellstudied class of activated ketones as novel inhibitors of various proteolytic enzymes.



^a Reagents: (a) Et₃N, DPPA, CH₂Cl₂; (b) Et₃N, T₈Cl; (c) Tl(NO₃)₃·3H₂O; (d) HOBt, MeOH; (e) Dess-Martin [O].

Experimental Section

General. All reactions were carried out under a positive pressure of dry argon, unless otherwise specified. Tetrahydrofuran (THF) and diethyl ether were distilled from sodium or potassium/benzophenone ketyl prior to use. Acetonitrile, benzene, dichloromethane, diisopropylamine, hexane, methanol, pyridine, and toluene were distilled from calcium hydride prior to use.

TLC was performed using EM Science (E. Merck) 5- × 10-cm plates precoated with silica gel 60 F_{254} (0.25 mm thickness), and the spots were visualized by any of the following: UV, iodine, phosphomolymdic acid (PMA), ceric ammonium sulfate, anisaldehyde, vanillin, or Rydons stain. EM Science's silica gel 60 (230-400 mesh ASTM) was used for flash chromatography. A ratio of 25-100:1 silica gel/crude product by weight and a nitrogen pressure of 5-25 psi was normally employed for flash columns. Reverse-phase chromatographic purification of final compounds was carried out using CHP20P gel, a 75-150 μ m polystyrenedivinyl benzene copolymer purchased from Mitsubishi Chemical Industries. Analytical HPLC was performed using two Shimadzu LC-6A pumps with an SCL-6B system controller and a C-R4AX chromatopac, and an SPD-6AV UV-VIS spectrophotometric detector. HPLC columns were commercially available from either Whatman or YMC Corporation.

Melting points were determined on an electrothermal Thomas Hoover capillary melting point apparatus and are uncorrected. NMR spectra were recorded on one of the following instruments: JOEL GX-400 operating at 400 (1H) or 100 MHz (13C), JOEL FX -270 operating at 270 (1H) or 67.8 (13C) MHz, and JOEL FX-60Q operating at 15 MHz (13C). Chemical shifts are reported in parts per million (ppm) downfield from tetramethylsilane (TMS) and coupling constants (J) are in hertz (Hz). ¹⁹F NMR spectra were recorded on a JEOL FX-90Q using CFCl₃ ($\delta = 0$ ppm) as an external reference. IR spectra were recorded on a Mattson Sirius 100 FT-IR spectrophotometer, and the absorption maxima are reported in cm⁻¹. Mass spectra were recorded on a Finnigan MAT TSQ-4600 mass spectrometer (chemical ionization, CI) or a VG-ZAB-2F mass spectrometer (fast-atom bombardment, FAB). High-resolution mass spectra (HRMS) were determined using peak matching techniques versus PEG standards on a VG-ZAB-2F spectrometer. Optical rotations were measured using a Perkin-Elmer model 241 polarimeter and a 10-cm path length optical cell. Microanalysis results were adjusted to obtain the best fit assuming nonstoichiometric hydration.

In Vitro Inhibition Studies. The human kidney renin used for assay of inhibitor potency was a partially purified preparation (no. 216, 2.4 μ g AI/h/mg) generously provided by Dr. E. Haas (Mt. Sinai Medical Center, Cleveland, OH). The source of angiotensinogen substrate in renin incubation mixtures was human plasma (Mercer Regional Blood Center, Trenton, NJ). Incubation mixtures of 0.5 mL were buffered with 0.2 M TES, pH 7.0, and contained 0.10 mM EDTA, 0.10 mM sodium tetrathionate, and 0.04 mM phenylmethanesulfonyl fluoride. Renin concentrations in the mixtures were adjusted to generate angiotensin I (AI) at rates, constant with time, of 20-80 ng AI/mL/h. Human plasma was added at concentrations (10-50%) sufficient to provide a final angiotensinogen concentration of 0.5 μ M. Inhibition test compounds were dissolved, serially diluted, and added to incubation mixtures in dimethyl sulfoxide (DMSO), with the final DMSO concentration fixed at either 0.5% or 1.0%. Incubations were conducted for 30 min at 37 °C. After the reactions were terminated by reduction of the temperature to 0 °C, AI concentrations were measured by radioimmunoassay. Inhibitor potencies were expressed as I_{50} values, the interpolated concentrations corresponding to 50% inhibition of renin activity. For the 13 compounds demonstrating I_{50} concentrations below 10 000 nM, I_{50} concentrations are mean values determined from 2-4 experiments; standard errors for the mean I_{50} concentrations ranged from 7% to 23%

Inhibition of the activities of porcine pepsin (Sigma Chemical Co., St. Louis, MO) and bovine cathepsin D (Sigma) were measured in assays based upon the hydrolysis of 1.6% bovine hemoglobin (Sigma) at pH 1.6 (0.05 M HCl) and pH 3.5 (0.2 M sodium formate buffer), respectively. After incubation periods of up to 75 min at 37 °C, protein in the incubation mixtures was

precipitated with 3.1% trichloroacetic acid (final concentration), and concentrations of peptidic products in the supernatant fractions were measured by rates of increase in absorbance at 280 nm to determine protease activity. Test compound was dissolved and diluted in DMSO and added to incubation mixtures at final DMSO concentrations of 2.0% or lower. Inhibitory potencies were expressed as I_{50} concentrations. Pepstatin A, employed as a standard aspartyl protease inhibitor in these assays, displayed an I_{50} concentration of <5 nM against both pepsin and cathepsin D.

(S)-a-[[(1,1-Dimethylethoxy)carbonyl]amino]-N-methoxy-N-methylcyclohexanepropanamide (1).¹⁰ 1.1'-Carbonyldiimidazole (CDI, 9.95 g, 61.1 mmol) was added over 10 min to an efficiently stirring solution of Boc-L-cyclohexylalanine (13.9 g, 51.2 mmol) in THF (150 mL) at 0 °C. After 30 min of stirring, N,O-dimethylhydroxylamine hydrochloride (5.49 g, 56.3 mmol) was added followed by triethylamine (7.9 mL, 56.3 mmol). After stirring at room temperature for 2.5 h, the reaction mixture was filtered and the filtrate was concentrated in vacuo. The ambercolored residue was dissolved in Et₂O and poured into 1 N HCl (1000 mL). The two layers were separated, and the aqueous solution was further extracted with Et₂O. The combined ethereal extracts were washed sequentially with H₂O, saturated aqueous NaHCO₃, and saturated NaCl. Drying (MgSO₄) and concentration in vacuo gave 1 (16.1 g, 99%) as a colorless oil. TLC: R_f = 0.58 (1:1 hexane/ethyl acetate). MS: $(M + H)^+$ 315. $[\alpha]_D =$ -12.9° (c = 1.0, MeOH). ¹H NMR (CDCl₃): (s, 0.8–2.1 (m, 13 H), 1.44 (s, 9 H), 3.20 (s, 3 H), 3.78 (s, 3 H), 4.75 (m, 1 H), 5.06 (d, 1 H, J = 9.4). ¹³C NMR (CDCl₃): 26.0, 26.2, 26.4, 28.3, 32.2, 34.0, 40.4, 48.3, 61.4, 79.6, 155.6, 173.9. Anal. Calcd for C₁₆H₃₀N₂O₄: C, 61.11; H, 9.62; N, 8.91. Found: C, 61.35; H, 9.54; N, 9.10.

(S)-α-[[(1,1-Dimethylethoxy)carbonyl]amino]cyclohexanepropanal (2). A 1 M THF solution of LiAIH₄ (Aldrich, 85.4 mL, 85.4 mmol) was added dropwise over a period of 20 min to a solution of the Weinreb amide 1 (17.88 g, 56.94 mmol) in 350 mL of ether at 0 °C (caution, exothermic!). After an additional 30 min at 0-2 °C, the reaction mixture was quenched with 250 mL of 5% KHSO4 (caution, exothermic!) and warmed to room temperature, and the aqueous and organic layers were separated. The aqueous layer was diluted with 250 mL of H_2O and reextracted with ether $(2 \times 150 \text{ mL})$. The combined organic extracts were washed sequentially with 5% HCl $(1 \times 150 \text{ mL})$, saturated aqueous $NaHCO_3$ (1 \times 150 mL) and saturated aqueous NaCl $(2 \times 150 \text{ mL})$. After the extracts were dried over Na₂SO₄, the ethereal solution was filtered through Celite and concentrated in vacuo to give 12.56 g (85.8%) residue which looked homogeneous by TLC, $R_f = 0.53$ (1:1 hexane/ethyl acetate). $[\alpha]_D =$ -37.2° (c = 5.6, CH₃OH). IR: 3075, 3050, 695. ¹³C NMR (CDCl₃): 25.9, 26.0, 26.2, 28.2, 32.5, 33.7, 36.6, 57.7, 79.9, 155.5, 200.4.

 (βS) - β -[[(1,1-Dimethylethoxy)carbonyl]amino]- α -methylcyclohexanepropanol (3). Methyl magnesium bromide (3.0 M, 17.9 mL, 53.7 mmol) was added to a solution of 2 (4.57 g, 17.9 mmol) in THF (90 mL) at -78 °C. After stirring for 5 min at -78 °C, the reaction mixture was warmed to 0 °C and then stirred overnight with gradual warming to room temperature. After 16 h, the reaction was quenched with saturated aqueous NH4Cl (90 mL) and THF was removed in vacuo. The residue was partitioned between EtOAc (100 mL) and water $H_2O(75 \text{ mL})$, and the aqueous layer was reextracted with EtOAc $(2 \times 100 \text{ mL})$. The combined organic extracts were washed with saturated aqueous NaCl (100 mL), dried (Na₂SO₄), and concentrated. Chromatographic purification of the crude product (4.4 g, 4:1 hexane/ethyl acetate) yielded 2.87 g (60%) of 3. TLC: $R_f = 0.53$ (1:1 hexane/ethyl acetate). MS: $(M + H)^+ 272$. $[\alpha]_D = -25.5^\circ$ (c = 1.32, MeOH). ¹H NMR (CDCl₃): 1.2 (d, 3 H, J = 6), 0.7–2.5 (m, 13 H), 1.45 (s, 9 H), 3.45-3.95 (m, 2 H), 4.45-4.7 (m, 1 H). ¹³C NMR (CDCl₃): 20.1, 26.0, 26.2, 26.4, 27.9, 28.2, 32.3, 32.6, 33.8, 34.1, 37.2, 39.8, 52.9, 53.3, 69.4, 70.5, 78.8, 79.2, 156.4, 156.5. Anal. Calcd for C₁₅H₂₉N₁O₃: C, 66.38; H, 10.77; N, 5.16. Found: C, 66.63; H, 10.58; N, 5.20.

 (βS) - β -Amino- α -methylcyclohexanepropanol (4). 3 (1.71 g, 6.3 mmol) was dissolved in a solution of ethyl acetate saturated with anhydrous HCl (app 1.5 M, 30 mL) and stirred for 1 h at room temperature. Concentration followed by trituration of the residue with hexane gave 1.4 g of solid hydrochloride salt, which was directly employed for subsequent reactions. MS (CI, +/-

ions): $(M + H)^+$ 172, $(M + Cl)^-$ 206. Anal. Calcd for C₁₀H₂₂NClO 0.27H₂O: C, 56.48; H, 10.69; N, 6.59; Cl, 16.67. Found: C, 56.68; H, 10.41; N, 6.42; Cl, 16.40.

N-[N-[(1,1-Dimethylethoxy)carbonyl]-L-phenylalanyl]-L-leucine, Methyl Ester (5). DCC (10.315 g, 50 mmol) was added to a solution of Boc-L-phenylalanine (13.265 g, 50 mmol), L-leucine methyl ester (9.085 g, 50 mmol), HOBT (7.65 g, 50 mmol), and iPr₂NEt (8.7 mL, 50 mmol) in THF (150 mL) at 0 °C, and the reaction was left for overnight stirring with gradual warming to room temperature. The next day (total reaction time, 14 h), the precipitated urea was filtered off and the filtrate concentrated in vacuo. The residue was taken up in EtOAc (500 mL) and washed sequentially with saturated aqeuous NaHCO₃ $(2 \times 200 \text{ mL})$ and saturated aqueous NaCl $(1 \times 100 \text{ mL})$, dried (Na₂SO₄), and concentrated to give a crude solid. Crystallization from ethyl ether gave pure 5 (11.61 g). Concentration of the mother liquor followed by chromatographic purification of the residue eluting with 4:1 hexane/ethyl acetate gave additional 5 (1.35 g, total 12.96 g, 66.1%). TLC: $R_f = 0.57 (1.1 \text{ hexane/ethyl})$ acetate). Mp: 104-105 °C. MS: $(M + H)^+$ 393. $[\alpha]_D = -17.5^\circ$ (c = 1.2, MeOH). ¹H NMR (CDCl₈): 0.9 (m, 6 H), 1.15-1.7 (m, 3 H), 1.42 (s, 9 H), 3.08 (d, 2 H, J = 7), 3.7 (s, 3 H), 4.35 (m, 1 H), 4.58 (m, 1 H), 5.0 (br s, 1 H), 6.3 (d, 1 H, J = 7), 7.1–7.35 (m, 5 H). ¹³C NMR (CDCl₃): 21.9, 22.7, 24.6, 28.2, 38.1, 41.6, 50.7, 52.2, 55.0, 80.2, 126.9, 128.6, 129.3, 136.6, 155.3, 170.9, 172.8. Anal. Calcd for C21H32N2O5: C, 64.3; H, 8.15; N, 7.14. Found: C, 64.12; H. 8.16; N. 7.02.

N-[N-[(1,1-Dimethylethoxy)carbonyl]-L-phenylalanyl]-L-leucine (6). NaOH (1 N, mL, 6 mmol) was added to a solution of Boc-Phe-Leu-OMe prepared above (1.92g, 5 mmol) in methanol (20 mL). After 1 h, the solution was concentrated and the residue was suspended in EtOAc (50 mL) and H₂O (20 mL). While stirring, the biphasic solution was acidified to pH 3.5 with concentrated HCl and the two layers were separated. The aqueous solution was reextracted with EtOAc $(2 \times 50 \text{ mL})$. The combined organic extracts were dried and concentrated to give a residue which after trituration with ethyl ether gave 6 (1.87 g, 99%). Mp: 143-146 °C. TLC: $R_f = 0.4$ (9:1:0.1 CHCl₃/MeOH/ AcOH). MS: $(M + H)^+ 379$. $[\alpha]_D = -13.98^\circ$ (c = 1.48, MeOH). IR (KBr): 3327, 2959, 1728, 1670, 1534, 1173, 698. ¹H NMR (CD₃OD): 0.93 (d, 3 H, J = 7), 0.95 (d, 3 H, J = 7), 1.35 (s, 9 H), 1.5–1.8 (m, 3 H), 2.79 (dd, 1 H, J = 9.4, 13.7), 3.12 (dd, 1 H, J= 4.7, 13.7), 4.33 (q, 1 H, J = 5.1, 9.4), 4.45 (t, 1 H, J = 7.3), 7.25 (s, 5 H). ¹³C NMR (CD₃OD): 21.9, 23.4, 25.8, 28.6, 39.0, 41.8, 52.0, 57.1, 80.5, 127.6, 129.3, 130.4, 157.3, 174.3, 175.7. Anal. Calcd for C₂₀H₃₀N₂O₅: C, 63.47; H, 7.99; N, 7.40. Found: C, 63.31; H, 7.78; N, 7.37.

N-[1-(Cyclohexylmethyl)-2-hydroxypropyl]-N²-[N-[(1,1dimethylethoxy)carbonyl]-L-phenylalanyl]-L-leucinamide (7). The hydrochloride salt 4 (414 mg, 2 mmol) was added to a solution of BOC-Phe-Leu-OH 6 (756 mg, 2 mmol) in THF (8 mL) and cooled to 0 °C. HOBt (306 mg, 2 mmol), iPr₂NEt (383 μ L, 2.2 mmol) and DCC (412.6 mg, 2 mmol) were added sequentially, and the reaction was stirred overnight with gradual warming to room temperature. After 16 h at room temperature, the reaction mixture was concentrated and the residue was treated with CH₂Cl₂ and filtered. The organic filtrate was washed sequentially with 10% citric acid $(2 \times 10 \text{ mL})$, saturated aqueous NaHCO₃ (2×10 mL), and saturated aqueous NaCl (10 mL). Drying and concentration gave 0.95 g of crude product which was purified by flash chromatography (1:1 hexane/ether to 1:1 hexane/ethyl acetate) to afford 514.8 mg (48.5%) of 7. Mp: 145– 147 °C. MS (CI, +/- ions): (M + H)⁺ 532, (M + Cl)⁻ 566, M - Cl)⁻ 530. TLC: $R_f = 0.42$ (90/10/1/0.1 CHCl₃/MeOH/H₂O/ AcOH). $[\alpha]_D = -37.8^\circ$ (c = 1.26, MeOH). ¹H (CDCl₃): 0.92 (d, 6 H, J = 6), 1.12 and 1.2 (d, 3 H, J = 6), 0.75 - 1.85 (m, 16 H), 1.42 (s, 9 H), 3.0-3.17 (m, 2 H), 3.65-4.05 (m, 2 H), 4.25-4.4 (m, 2 H), 4.95 (m, 1 H), 6.38 (m, 2 H), 7.15–7.35 (m, 5 H). ¹³C (CDCl₃): 18.4, 20.0, 22.2, 22.5, 24.5, 25.9, 26.0, 26.1, 26.3, 28.0, 32.3, 32.8, 33.5, 33.8, 34.1, 36.3, 37.9, 38.8, 40.9, 41.2, 52.0, 52.2, 52.6, 55.4, 68.6, 70.3, 79.8, 126.5, 128.2, 129.1, 130.7, 136.4, 155.5, 171.7, 172.1. Anal. Calcd for C₃₀H₄₉N₃O₅ 0.24H₂O: C, 67.22; H, 9.30; N, 7.84. Found: C, 67.34; H, 9.20; N, 7.78.

α-(2,2,2-Trifluoro-1-hydroxyethyl)cyclohexanepropanoic Acid (8). nBuLi (2.5 M, 152.68 mL, 382 mmol) was added dropwise to a solution of iPr₂NH (53.5 mL, 382 mmol) in THF (250 mL) at -78 °C. After sirring for 5 min at -78 °C and 30 min at -10 °C to ensure complete formation of LDA, a solution of cyclohexane propionic acid (30 mL, 173.5 mmol) in THF (30 mL) was added, and stirring was continued for 1 h at -10 °C. The reaction was then cooled to -50 °C and freshly generated trifluoroacetaldehyde was cannulated into the reaction flask. [Trifluoroacetaldehyde was prepared by refluxing a 100-g 1:1 mixture of trifluoroacetaldehyde ethyl hemiacetal and concentrated sulfuric acid and condensing the liberated gaseous product in a side arm flask precooled to $-78 \,^{\circ}C^{16}$]. After stirring for 2 h with gradual warming to room temperature, the reaction was quenched with water (10 mL), concentrated to remove THF, and partitioned between EtOAc (250 ml) and 10% HCl (350 mL). The aqueous layer was reextracted with EtOAc (3×250 mL), and the combined organic extracts were washed with saturated aqueous NaCl ($1 \times 200 \text{ mL}$). Drying (Na₂SO₄) and concentration afforded 66.6 g of crude product. Purification by flash chromatography (30:1:0.1 to 19:1:0.1 CHCl₃/MeOH/AcOH) followed by trituration with toluene to remove traces of acetic acid, afforded 29.26 g (66.7%) of pure 8. MS: $(M + H)^+$ 255, $(M - H)^-$ 253. TLC: $R_f = 0.4$ (9:1:0.1 CHCl₃/MeOH/AcOH). ¹H (CDCl₃): 0.75-2.0 (m, 13 H), 2.6-2.87 (m, 1 H), 3.95-4.15 (m, 1 H), 4.97 (br S, 1 H). ¹³C (CD₃OD): 27.2, 27.3, 27.4, 27.6, 33.2, 33.6, 34.9, 35.5, 36.7, 37.0, 38.3, 44.8, 45.6, 71.8, 72.3, 72.7, 126.6 (q, J = 283), 175.6, 177.1.

 α -[1-[[(1,1-Dimethylethyl)dimethysilyl]oxy]-2,2,2-trifluoroethyl]cyclohexanepropanoic Acid (9). TBDMS-OTf (73.8 mL, 321 mmol) was added dropwise to a solution of 8 (27.17 g, 107 mmol) and Et₃N (44.7 mL, 321 mmol) in CH₂Cl₂ (430 mL) at 0 °C. After overnight stirring with gradual warming to room temperature, the reaction was quenched with H_2O (500 mL) and extracted with additional CH_2Cl_2 (3 × 350 mL). The combined organic extracts were concentrated, and the crude disilylated product was stirred with K₂CO₃ (1 M, 340 mL) in MeOH (500 mL) for 1.5 h at room temperature. MeOH was removed in vacuo. and the aqueous solution was acidified in the presence of EtOAc (600 mL) to pH = 5.3 using 10% aqueous HCl. The two layers were separated, and the aqueous layer was reextracted with EtOAc $(3 \times 250 \text{ mL})$. The combined organic extracts were dried (Na₂SO₄) and concentrated, and the crude product was flash chromatographed (2:1 hexane/EtOAc) to afford 38.6 g (98%) of pure 9. TLC: $R_f = 0.62$ (1:1 hexane/ethyl acetate). MS: $(M + H)^+ 369$, $(M - H)^{-}$ 367, $(2 M - H)^{-}$ 735. ¹H (CDCl₃): 0.1 (s, 6 H), 0.65–1.9 (m, 22 H), 2.7–2.92 (m, 1 H), 4.05–4.4 (m, 1 H), 9.05 (br S, 1 H). ¹³C (CD₃OD): -4.5, -4.8, 19.0, 26.2, 27.0, 27.3, 27.6, 33.2, 35.4, 35.6, 36.7, 45.2, 73.5 (q, J = 30.3), 126.0 (q, J = 284), 176.2 Anal. Calcd for C₁₇H₃₁O₃F₃Si: C, 55.41; H, 8.48; F, 15.46. Found: C, 55.25; H, 8.55; F, 15.25.

[1-(Cyclohexylmethyl)-2-[[(1,1-dimethylethyl)dimethylsilyl]oxy]-3,3,3-trifluoropropyl]carbamic Acid, Phenylmethyl Ester (10). Diphenylphosphoryl azide (DPPA 1.243 mL, 5.775 mmol) was added to a premixed and vigorously stirring solution of 9 (1.93 g, 5.25 mmol) and Et₃N (0.768 mL, 5.512 mmol) in hexane (21 mL). After 3 h reflux, complete disappearance of the acyl azide intermediate (2150 cm⁻¹) and formation of the rearranged isocyanate (2250 cm⁻¹) was revealed by IR. At this stage, benzyl alcohol (0.598 mL, 5.775 mmol) and additional Et₈N (0.768 mL, 5.512 mmol) were added, and the reaction mixture was vigorously stirred and refluxed for an additional 18 h. It was then taken up in EtOAc (50 mL) and washed sequentially with H_2O (1 × 25 mL), saturated aqueous NaHCO₃ (1 × 25 mL), 1% aqueous HCl $(1 \times 25 \text{ mL})$, and saturated NaCl $(1 \times 25 \text{ mL})$. Drying (Na₂SO₄) and concentration gave 2.448 g of crude product, which was purified by flash chromatography (9:1 hexane/ Et_2O) to give 1.837 g (75%) pure 10. The reaction was repeated on a 50 mmol scale to provide 15.76 g (66.7%) 10. TLC: $R_f = 0.43$ (4:1 hexane/Et₂O). MS: $(M + H)^+$ 474, $(M - H)^-$ 472. ¹H NMR (CDCl₃): 0.1 (s, 3 H), 0.17 (s, 3 H), 0.9 and 0.95 (2s, 9 H), 0.7-1.9 (m, 13 H), 3.8-4.27 (m, 2 H), 4.6 and 4.9 (2d, 1 H, J = 8), 5.13(m, 2 H), 7.35 (m, 5H) ¹³C NMR (CDCl₃) : -5.3, -5.1, -4.9, 18.1, 25.6, 25.9, 26.1, 26.2, 26.3, 26.4, 31.8, 32.6, 33.6, 33.8, 34.0, 34.4, 35.4, 39.5, 48.6, 49.3, 66.7, 66.8, 71.9 (q, J = 28.4), 73.0 (q, J =28.4), 124.3 (q, J = 282), 127.9, 128.0, 128.2, 128.5, 136.3, 136.5, 155.5, 155.7. Anal. Calcd for C24H38NO3F3Si: C, 60.86; H, 8.09; N, 2.96; F, 12.03. Found: C, 60.85; H, 7.97; N, 3.11; F, 11.96.

 β -Amino- α -(trifluoromethyl)cyclohexanepropanol, 4-Methylphenyl) Sulfonate Salt (11). Pd(OH)₂/C (Pearlman's catalyst, 3.6 g) was added to a solution of 10 (5.728 g, 12.1 mmol) in EtOAc (48 mL). After stirring under an atmosphere of hydrogen (balloon) for 20 h, the reaction mixture was filtered through Celite and the catalyst was washed with additional EtOAc. The combined filtrate was concentrated to give 3.43 g of the crude amine which was dissolved in THF (40 mL) and treated with nBu₄N⁺F⁻ (1 M in THF, 12.1 mL, 12.1 mmol). After 1 h, additional nBu₄N+F⁻ (1 M in THF, 6 mL, 6 mmol) was added to drive the reaction to completion. After 20 min, the reaction was judged to be complete by TLC. Concentration in vacuo gave a residue which was dissolved in EtOAc (300 mL) and washed sequentially with H_2O (1 × 150 mL), saturated Na_2CO_3 (2 × 150 mL), and saturated aqueous $NaCl (1 \times 150 \text{ mL})$. Drying (Na_2SO_4) and concentration gave the free amino alcohol 11 as a white solid (2.13 g, 10.24 mmol) which looked clean by TLC , $R_f = 0.27$ (9: 1:0.05 CHCl₈/MeOH/NH₄OH). It was dissolved in MeOH (100 mL) and treated with pTsOH (1.9 g, 10.24 mmol). After stirring for 5 min, the tosylate salt of 11 precipitated out which was filtered to provide 2.84 g of the product. The mother liquor was concentrated, the residue suspended in Et₂O, and filtered to provide an additional 1.18 g of 11 (overall 83.7% yield from 10). MS: $(M + H)^+ 474$, $(M - H)^- 472$. ¹H NMR (CD₃OD): 0.65–1.9 (m, 13 H), 2.33 (s, 3 H), 3.42-3.65 (m, 1 H), 4.13 and 4.35 (dd, 1 H, J = 2, 8), 7.22 (d, 2 H, J = 9), 7.7 (d, 2 H, J = 9). ¹³C NMR (CD₃OD): 21.3, 26.7, 26.8, 26.9, 27.1, 27.3, 32.9, 33.8, 34.1, 34.4, 35.0, 35.5, 38.8, 50.4, 69.0 (q, J = 32), 70.2 (q, J = 30), 125.7 (qJ = 282), 126.9, 129.8, 141.7, 143.4. Anal. Calcd for C₁₇H₂₈NO₄F₈S: C, 51.37; H, 6.59; N, 3.53; F, 14.34; S, 8.07. Found: C, 51.38; H, 6.62; N, 3.51; F, 14.49; S, 8.17.

N-[1-(Cyclohexylmethyl)-3,3,3-trifluoro-2-hydroxypropyl]-N²-[N-[(1,1-dimethylethoxy)carbonyl]-L-phenylalanyl]-Lleucinamide (12). HOBt (1.09 g, 7.15 mmol) and iPr2NEt (1.36 mL, 7.865 mmol) were sequentially added to a solution of 6 (2.7 g, 7.15 mmol) and 11 (2.84 g, 7.15 mmol) in DMF (35 mL) at 0°. DCC (1.47 g, 7.15 mmol) was added after 5 min, and the reaction was left for overnight stirring with gradual warming to room temperature. After 12 h, the reaction mixture was filtered and the filtrate was concentrated in vacuo. The residue was taken up in EtOAc (250 mL) and washed sequentially with H_2O (2 × 150 mL), saturated aqueous NaHCO₃ (1 × 150 mL), 5% aqueous HCl $(1 \times 100 \text{ mL})$, and saturated NaCl $(1 \times 100 \text{ mL})$. Drying (Na₂SO₄) and concentration gave 4.35 g of crude product which was purified by flash chromatography (2:1 hexane/EtOAc) to give 3.12 g (75%) pure 12. Mp: 123-126 °C. HPLC: $t_R = 22.82$ (18.4%), 23.12 (12.3%), 24.4 (33.3%), and 26.6 min (33.5%); 50-90% aqueous CH_3CN 50-min gradient, flow rate = 1 mL/min; YMC 20% C₁₈ column. TLC: $R_f = 0.3$ (2:1 hexane/ethyl acetate). MS: $(M + H)^+ 586$, $(M + H - Boc)^+ 486$. $[\alpha]_D = -16.6^\circ$ (c = 1.32, CH₃OH). IR (KBr): 3306, 2956, 2927, 2853, 1690, 1650, 1524, 1451, 1391, 1368, 1273, 1252, 1169, 1143, 1094, 699. ¹H NMR (CDCl₃): 1.4 and 1.42 (s, 9 H), 0.75-1.9 (m, 22 H), 3.1 (m, 2 H), 3.8-4.7 (m, 4 H), 4.9 and 5.05 (m, 1 H), 6.2-6.4 (m, 1 H), 6.7 (m, 1 H), 7.15-7.4 (m, 5 H). ¹³C NMR (CDCl₃): 22.0, 22.4, 22.6, 24.5, 24.6, 25.9, 26.1, 26.3, 28.1, 31.7, 31.9, 33.9, 34.0, 35.1, 40.5, 40.8, 47.9, 52.0, 52.4, 80.6, 126.9, 128.5, 128.6, 129.1, 136.2, 171.9, 172.2, 172.6, 172.8. Anal. Calcd for C₃₀H₄₈N₃O₅F₃: C, 61.52; H, 7.92; N, 7.18; F, 9.73. Found: C, 61.66; H, 8.04; N, 7.11; F, 9.56.

N-[1-(Cyclohexylmethyl)-3,3,3-trifluoro-2-oxopropyl]-N²-[N-[(1,1-dimethylethoxy)carbonyl]-L-phenylalanyl]-L-leucinamide (13). A solution of the alcohol 12 (585 mg, 1 mmol) in CH₂Cl₂ was added to a solution of Dess-Martin periodinane reagent (1.275 g, 3 mmol) and tBuOH (0.3 mL, 4 mmol) in CH₂Cl₂ (40 mL). After 12 h at room temperature, the reaction mixture was filtered through Celite and the filtrate concentrated *in vacuo*. The residue was suspended in EtOAc (15 mL) and the excess reagent was removed by filtration through Celite. Reconcentration of the filtrate gave the crude product which was purified by flash chromatography (3:1 hexane/EtOAc) to give 539 mg (92.5%) pure 13. Mp: 148-153 °C. TLC: $R_f = 0.27$ (2:1 hexane/ ethyl acetate). HPLC: $t_R = 20.91$ (47.2%), 22.8 (44.5%), 50 min; 50-90% aqueous CH₃CN gradient, flow rate 1 mL/min; YMC 20% C₁₈ column. MS: (M + H)⁺ 584. $[\alpha]_D = -20.4^{\circ}$ (c = 1.1, CH₃OH). IR (KBr): 3301, 2957, 2927, 1690, 1651, 1525, 1451, 1392, 1368, 1251, 1173, 1143, 398. ¹H NMR (DMSO- d_{6}): 0.95 (m, 6 H), 1.42 (s, 9 H), 0.75–1.85 (m, 16 H), 3.1 (m, 2 H), 4.1–4.5 (m, 2 H), 4.85 (m, 1 H), 7.15–7.37 (m, 5 H), 7.5 and 7.72 (2d, 1 H, J = 7), 7.95 and 8.02 (2d, 1 H, J = 7), 8.68 (d, 1 H, J = 7). ¹³C NMR (CD₃OD): 22.2, 22.3, 22.6, 23.1, 23.3, 25.5, 25.7, 27.0, 27.4, 27.5, 28.6, 32.8, 32.9, 34.8, 35.0, 35.2, 35.5, 36.3, 36.7, 39.1, 41.8, 41.9, 51.5, 51.8, 52.9, 53.0, 53.6, 57.0, 80.5, 97.1, 94.8 (q, J = 30), 97.3 (q, J = 28), 125.7 (q, J = 297), 127.5, 129.3, 130.2, 138.4, 157.4, 173.9, 174.1, 174.2, 174.4. Anal. Calcd for C₃₀H₄₄N₃O₅F₃· 0.5H₂O: C, 60.79; H, 7.65; N, 7.09; F, 9.62. Found: C, 60.94; H, 7.65; N, 7.00; F, 9.71.

(R*)-N-[1-(Cyclohexylmethyl)-2-methoxy-2-oxoethyl]-N²-[(phenylmethoxy)carbonyl]-L-leucinamide (15). Amine 149 (891 mg, 2.98 mmol) was added to a solution of Cbz-Leu-OH (789 mg, 2.98 mmol) in THF (12 mL). The solution was cooled to 0 °C, and HOBt (456 mg, 2.98 mmol) was added. After 2 min, iPr_2NEt (518.5 μ L, 2.98 mmol) was added and this was followed immediately by addition of DCC (614 mg, 2.98 mmol). The reaction was stirred at 0 °C and left for overnight stirring at room temperature. Next day, the precipitated urea was filtered off and the filtrate concentrated to give an oil which was taken up in EtOAc (40 mL) and washed sequentially with $NaHCO_3$ (1) \times 20 mL) and NaCl (1 \times 20 mL). Drying and concentration gave an oily residue which after flash chromatographic purification (silica gel, 4:1 Hex/EtOAc) yielded 15 (832 mg, 64%). TLC: $R_f = 0.48$ (1:1 hexane/ethyl acetate). Mp: 76-79 °C. $[\alpha]_D = -33.8^\circ$ (c = 1.34, CH₃OH). MS: (M + H)⁺ 433. ¹H NMR (CDCl₃): 0.7-1.8 (m, 22 H), 3.7 (s, 3 H), 4.1-4.25 (br m, 1 H), 4.6 (m, 1 H), 5.12 (s, 2 H), 5.25 (d, 1 H, J = 8), 6.38 (d, 1 H, J = 8), 7.33 (br s, 5 H). ¹³C NMR (CDCl₃): 22.1, 22.8, 24.6, 26.0, 26.1, 26.3, 32.5, 33.4, 34.9, 39.9, 41.3, 50.1, 52.2, 53.5, 67.9, 127.9, 128.1, 128.2, 128.5, 136.1, 156.1, 171.8, 173.1.

(R*)-N-[1-(Cyclohexylmethyl)-2-hydroxy-2-oxoethyl]-N²-[(phenylmethoxy)carbonyl]-L-leucinamide (16). NaOH (1 N, 2 mL, 2 mmol) was added to solution of 15 in methanol (13 mL). A TLC check after 5 h at room temperature revealed completion of reaction. Solvents were stripped down on a rotary evaporator and the residue was partitioned between 50 mL of 1:1 EtOAc/1 N HCl. The aqueous layer was reextracted with EtOAc $(2 \times 25 \text{ mL})$. The combined organic extracts were dried (Na₂SO₄) and concentrated, and the residue was purified by flash chromatography eluting with 9:1:0.1 CHCl₃/MeOH/AcOH to give 16 (688 mg, 94.8%). TLC: $R_f = 0.35 (9:1:0.1 \text{ CHCl}_8/\text{MeOH}/\text{AcOH})$. Mp: 55-66 °C. $[\alpha]_D = -23.7^\circ$ (c = 2.0, MeOH). MS: (M + H)⁺ 419. ¹H NMR (CD₃OD): 0.7-1.9 (m, 22 H), 3.7 (s, 3 H), 4.2 (br m, 1 H), 4.48 (br m, 1 H), 4.9 (s, 2 H), 7.3 (br s, 5 H). ¹³C NMR (CD₃OD): 22.1, 23.4, 25.8, 27.1, 27.3, 27.5, 33.3, 34.8, 35.3, 40.2, 42.1, 42.4, 51.3, 54.7, 67.6, 128.7, 129.1, 129.4, 138.1, 158.2, 175.1, 175.9.

N-[1-(Cyclohexylmethyl)-3-ethoxy-2,3-dioxopropyl]-N²-[(phenylmethoxy)carbonyl]-L-leucinamide (17). DMAP (8.5 mg) was added to a 1:1 THF/DMF solution (8.4 mL) of the acid 16 (585.2 mg, 1.4 mmol) and this was followed by addition of pyridine (453 μ L, 5.6 mmol). After 5 min, ethyl oxalyl chloride $(313.6 \,\mu\text{L}, 2.8 \,\text{mmol})$ was added and the reaction mixture set to reflux for 2 h, then cooled to room temperature, and quenched with 1 mL of H₂O. After an additional 15 min of stirring, THF was stripped down and the residue taken in H_2O (25 mL) and extracted with EtOAc (3×30 mL). The combined organic extracts were washed with NaCl $(1 \times 25 \text{ mL})$, dried (Na₂SO₄), and concentrated to give 881 mg of residue which was stirred overnight in 20 mL of EtOH with 1 g of solid NaHCO₃. Next day, the solid was filtered off and the filtrate concentrated to give an oily residue which after chromatographic purification yielded 17 (270 mg, 40.6%) as a 1:1 mixture of diastereomers. TLC: $R_f = 0.47$ (1:1 hexane/ethyl acetate). $[\alpha]_D = -6.1^\circ$ (c = 1.44, MeOH). MS: (M + H)⁺ 475. IR: 3310, 3303, 2955, 2926, 1730, 1661, 1535, 1449, 1285, 1262, 1244, 1049, 1029, 412, 405, 400. ¹H NMR (CDCl₃): 0.7–1.9 (m, 25 H), 4.25 (br m, 3 H), 5.1 (s, 2 H), 5.0–5.5 (m, 2 H), 6.7–7.0 (m, 1 H), 7.3 (s, 5 H). ¹³C NMR (CD₃OD): 13.9, 22.0, 22.7, 24.5, 24.7, 25.8, 26.0, 26.2, 32.1, 33.7, 34.2, 34.3, 37.9, 38.0, 40.9, 41.1, 41.3, 41.4, 53.2, 53.6, 62.5, 67.0, 127.8, 128.0, 128.4, 136.0, 156.2, 160.3, 172.2. Anal. Calcd for $C_{26}H_{38}N_2O_6$: C, 65.80; H, 8.07; N, 5.90. Found: C, 65.43; H, 8.03; N, 5.90.

 (R^*) -N-[1-(Cyclohexylmethyl)-2-methoxy-2-oxoethyl]-N²-[N-[(1,1-dimethylethoxy)carbonyl]-L-phenylalanyl]-L-leucinamide (18). 6 (2.017 g, 5.33 mmol) was dissolved in 20 mL of 1:1 DMF/THF and amine 14 (1.595 g, 5.33 mmol) was added. The solution was cooled to 0 °C and HOBt (0.815 g, 5.33 mmol) was added. After 2 min, iPr₂NEt (0.927 mL, 5.33 mmol) was added and this was followed immediately by addition of DCC (1.099 g, 5.33 mmol). The reaction was stirred at 0 °C for 2 h and left for stirring overnight at room temperature. Next day, the precipitated urea was filtered off and the filtrate concentrated to give an oil which was taken up in EtOAc (100 mL). The precipitated urea was refiltered and the filtrate was washed sequentially with NaHCO₃ (1×50 mL), 10% aqueous citric acid $(2 \times 50 \text{ mL})$, and NaHCO₈ $(1 \times 50 \text{ mL})$, dried; and concentrated to give 3.08 g crude product. Flash chromatographic purification eluting with 4:1 Hex/EtOAc yielded 18 (2.673 g, 92.2% yield). The reaction was repeated on a 30 mmol scale to provide additional amounts of 18 (15.89 g, 97%). Mp: 134-138 °C. TLC: $R_f = 0.45$ (1:1 hexane/ethyl acetate). $[\alpha]_D = -31.4^\circ$ (c = 1.02 MeOH). MS: (M + H)⁺ 546⁺. ¹H NMR (CDCl₃): 0.9 (d, 6 H, J = 6, 1.42 (s, 9 H), 0.8–1.8 (m, 16 H), 2.95–3.15 (m, 2 H), 3.71 (s, 3 H), 4.25-4.65 (m, 3 H), 5.08 (d, 1 H, J = 8), 6.58 (d, 1 H, J = 8), 6.65 (d, 1 H, J = 8), 7.15–7.35 (m, 5 H). ¹³C NMR (CDCl₈): 22.1, 22.8, 24.5, 26.0, 26.3, 25.9, 28.2, 32.5, 33.3, 34.0, 37.9, 39.7, 41.0, 50.2, 51.7, 52.1, 55.7, 80.3, 126.8, 128.5, 129.2, 136.5, 155.4, 171.3, 173.0. Anal. Calcd for C₃₀H₄₇N₃O₆: C, 66.03; H, 8.68; N, 7.70. Found: C, 65.79; H, 8.61; N, 7.61.

(R*)-N-[1-(Cyclohexylmethyl)-2-hydroxy-2-oxoethyl]-N²-[N-[(1,1-dimethylethoxy)carbonyl]-L-phenylalanyl]-L-leucinamide (19). NaOH (1 N,35 mL, 35 mmol) was added to a solution of methyl ester 18 (15.26 g, 28 mmol) in methanol (100 mL). After refluxing for 1 h, the reaction was judged to be complete by TLC. It was cooled to room temperature and acidified to pH = 4.5 using 10% HCl. MeOH was removed on the rotary evaporator and the remainder diluted with $H_{2}O$ (250 mL) and extracted with EtOAc (3×200 mL). The combined organic extracts were washed with saturated NaCl $(1 \times 100 \text{ mL})$, dried, and concentrated to give the crude acid which upon trituration with ether yielded pure 19 (14.35 g, 96.5%). Mp: 58-66 °C. TLC: $R_f = 0.45$ (1:1 hexane/ethyl acetate). $[\alpha]_D =$ -25.8° (c = 1.23, CH₃OH). MS: (M + H) + 532. ¹H NMR (CDCl₃): 0.88 (m, 6 H), 1.38 (s, 9 H), 0.7-1.82 (m, 16 H), 2.8-3.15 (m, 2 H), 4.3-4.6 (m, 3 H), 5.25 (br s, 1 H), 6.95 (br s, 1 H), 7.05-7.35 (m, 5 H), 10.0 (br s, 1 H). ¹³C NMR (CDCl₃): 22.1, 22.7, 24.5, 26.0, 26.1, 26.3, 28.2, 32.5, 33.4, 34.1, 38.2, 39.5, 41.0, 50.5, 51.9, 55.5, 80.4, 126.9, 128.5, 129.3, 136.4, 155.6, 171.9, 172.1, 175.8. Anal. Calcd for C₂₉H₄₅N₃O₆: C, 65.51; H, 8.93; N, 7.90. Found: C, 64.98; H, 8.55; N, 7.87.

N-[1-(Cyclohexylmethyl)-3-ethoxy-2,3-dioxopropyl]-N²-[N-[(1,1-dimethylethoxy)carbonyl]-L-phenylalanyl]-L-leucinamide (20). DMAP (73 mg, 0.6 mmol) and pyridine (4.86 mL, 60 mmol) were added to a solution of acid 19 (6.372 g, 12 mmol) in 1:1 THF/DMF (72 mL). After 5 min, freshly distilled ethyl oxalyl chloride (2.688 mL, 24 mmol) was added, and the solution was heated to reflux for 2 h. After cooling to room temperature, the reaction was quenched with H_2O (5 mL) and stirred for 30 min. The solvents were stripped down and the residue was partitioned between 250 mL of H₂O and 250 mL of EtOAc. The aqueous layer was reextracted with EtOAc $(2 \times 250 \text{ mL})$, and the combined organic extracts were washed sequentially with H₂O $(1 \times 100 \text{ mL})$, 1% HCl $(1 \times 100 \text{ mL})$, and saturated NaCl $(1 \times 100 \text{ mL})$ 100 mL). Drying and concentration gave 7.21 g of residue which was dissolved in EtOH (200 mL) and stirred very rapidly overnight with solid NaHCO₃ (10 g). The next day, the reaction mixture was filtered and the filtrate concentrated to give a residue which upon flash chromatographic purification yielded 20 (3.93 g, 56 %)as a diastereomeric mixture. Mp: 102-128 °C. TLC: $R_f = 0.38$ (1:1 hexane/ethyl acetate). $[\alpha]_D = -14.2^\circ$ (c = 0.8, MeOH). MS: (M + H)⁺ 588. IR: 3289, 2976, 2957, 2927, 2853, 1731, 1694, 1646, 1539, 1527, 1499, 1450, 1391, 1368, 1252, 1172, 1050, 700, 405, 400. ¹H NMR (CDCl₃): 0.7-1.9 (m, 34 H), 3.05 (m, 2 H), 4.3 (m, 2 H), 4.2-4.55 (m, 2 H), 5.1 (br m, 2 H), 6.45 and 6.6 (d, 1 H, J = 8), 6.9 and 7.05 (d, 1 H, J = 1 H), 7.1–7.32 (m, 5 H). ¹³C NMR (CDCl₃): 13.8, 21.7, 21.9, 22.0, 22.7, 22.8, 24.2, 24.4, 24.6, 25.8, 26.0, 26.1, 26.2, 28.1, 32.0, 32.2, 33.6, 34.1, 34.2, 37.4, 37.5,

37.57, 37.7, 37.8, 38.0, 40.4, 40.7, 40.8, 51.5, 53.5, 53.6, 56.1, 62.4, 62.5, 80.4, 80.8, 126.8, 126.9, 127.0, 128.6, 128.7, 129.1, 129.2, 136.3, 160.3, 160.4, 171.4, 171.5, 171.6, 192.5. Anal. Calcd for $C_{32}H_{49}N_3O_7$: C, 65.39; H, 8.40; N, 7.15. Found: C, 65.12; H, 8.45; N, 7.14.

N-[1-(Cyclohexylmethyl)-3-hydroxy-2,3-dioxopropyl]-N²-[N-[(1.1-dimethylethoxy)carbonyl]-L-phenylalanyl]-L-leucinamide (21). The α -keto ester 20 (171 mg, 0.29 mmol) was dissolved in ethanol (2 mL) and treated with 1 M K₂CO₃ (0.3 mL. 0.3 mmol). The reaction mixture was stirred at room temperature and a TLC check after 1 h revealed completion of reaction. The solvents were stripped down, the residue was taken up in 20 mL of 1:1 H₂O/EtOAc and acidified to pH = 4.0 with 10% HCl, and the two layers were separated. The aqueous layer was reextracted with EtOAc (2×25 mL). The combined organic extracts were dried (Na₂SO₄) and concentrated to give 117 mg of crude product which after flash chromatographic purification eluting with 9:1: 0.1 CHCl₃/MeOH/AcOH yielded pure acid 21 (77 mg, 48%). Mp: 97-110 °C. TLC: $R_f = 0.43$ (9:1:0.1 CHCl₃/MeOH/AcOH). $[\alpha]_D$ $=-11.4^{\circ}$ (c = 0.42, MeOH). MS: (M + H) + 560. IR: 3424, 3415, 3409, 3404, 3396, 3385, 3321, 3312, 3302, 2956, 2927, 2853, 1650, 1525, 1499, 1451, 1421, 1416, 1392, 1368, 1252, 1170, 422, 412, 404. ¹H NMR (CDCl₃): 0.7-1.9 (m, 31 H), 2.7-3.15 (m, 2 H), 4.2-4.5 (m, 3 H), 7.25 (br s, 5 H). ¹³C NMR (CDCl₃): 22.2, 23.4, 25.7, 27.2, 27.3, 27.5, 28.6, 33.2, 33.3, 34.1, 34.8, 35.3, 35.5, 39.0, 40.4, 42.1, 51.0, 51.7, 52.8, 53.0, 57.2, 57.5, 57.9, 80.6, 116.8, 127.6, 127.7, 129.3, 130.3, 138.5, 138.6, 157.5, 166.5, 174.1, 174.2. Anal. Calcd for C₃₀H₄₅N₃O₇: C, 64.38; H, 8.10; N, 7.51. Found: C, 64.56; H, 8.50; N, 7.74.

 $[R \cdot (R^*, S^*)]$ - and $[S \cdot (R^*, R^*)] - \beta \cdot [[(1, 1-\text{Dimethylethoxy}) - \beta \cdot [[(1, 1-\text{Dimethylethoxy}) - \beta \cdot [(1, 1-\text{Di$ carbonyl]amino]-a-[tris(methylthio)methyl]cyclohexanepropanol (22 and 23). nBuLi (50 mL of a 2.5 M solution, 125 mmol) was added over a period of 10 min to a solution of tris(methylthio)methane (20.37 g, 132 mmol) in 200 mL THF at -75 °C. A white precipitation occurs but smooth stirring could be maintained. After 20 min, aldehyde 2 (13.9 g, 54.5 mmol) was added as 150 mL of a THF solution at -70 to -65 °C. During the later half of the addition, the precipitation disappeared and a clear yellow solution resulted after the addition was complete. The reaction was continued for 3 h at -50 to -40 °C after which it was quenched with 100 mL of saturated NH_4Cl and warmed to room temperature. The two layers were separated; the aqueous layer was diluted with 200 mL of H₂O to dissolve the precipitated NH₄Cl and extracted with ether $(2 \times 100 \text{ mL})$. The organic solutions were combined, washed with saturated NaCl (1 \times 150 mL), dried (Na₂SO₄), and concentrated to give 34.63 g of solid. The crude product yielded 7.57 g of pure diastereomer 22 (33.65%) upon crystallization from 6:1 hexane/ethyl ether. Mp: 147-149 °C. TLC: $R_f = 0.29$ (1:1 hexane/Et₂O). $[\alpha]_D = -12.4^\circ$ (c = 1.33, MeOH). MS: (M + H)⁺ 410. ¹H NMR (CDCl₃): 0.7-2.3 (m, 13 H), 1.45 (s, 9 H), 2.22 (s, 9 H), 3.5 (s, 1 H), 3.7 (br s, 1 H), 4.15 (m, 1 H), 5.1 (br s, 1 H). 13 C NMR (CDCl₃): 13.9, 26.2, 26.3, 26.6, 28.5, 33.1, 33.4, 34.2, 43.7, 47.9, 74.5, 78.8, 155.4. Anal. Calcd for C₁₈H₃₅NO₃S₃: C, 52.77; H, 8.61; N, 3.42; S, 23.48. Found: C, 52.60; H, 8.56; N, 3.36; S, 23.38.

The mother liquor was concentrated and the residue chromatographed by eluting with 4:1 to 2:1 hexane/Et₂O to give an additional 6.5 g of diastereomeric mixture of α -hydroxy ortho thioesters 22 and 23 (overall yield, 62.5%).

 $[R-(R^*,S^*)]-\beta-[[(1,1-Dimethylethoxy)carbony]]amino]-\alpha$ hydroxycyclohexanebutanoic Acid, Ethyl Ester (24). HgCl₂ (6.278 g, 23.1 mmol) and HgO (2.50 g, 11.56 mmol) were added to a solution of α -hydroxy ortho thioester 22 (1.891 g, 4.625 mmol) in 95% aqueous EtOH (115 mL). After stirring for 72 h at room temperature, the reaction mixture was filtered through Celite and the solid was washed with dichloromethane (50 mL). The filtrate was treated with H_2O (100 mL), and the two layers were separated. The aqueous layer was reextracted with dichloromethane $(2 \times 100 \text{ mL})$. The combined organic extracts were washed sequentially with saturated NH4Cl (2 \times 100 mL), 75% NH₄OAc (1×100 mL), and saturated NaCl (1×100 mL), dried over Na₂SO₄, and filtered through Celite. The filtrate was concentrated and the residue chromatographed by eluting with 4:1 to 1:1 Hex/Et₂O to give pure 24 (1.246 g, 82%). TLC: $R_f =$ 0.27 (1:1 hexane/Et₂O). $[\alpha]_D = -42.6^\circ$ (c = 2.39, MeOH). MS: $(M + H)^+$ 330. ¹H NMR (CDCl₃): 0.8–1.95 (m, 13 H), 1.3 (t, J

= 7, 3 H) 1.4 (s, 9 H), 3.15 (d, 1 H, J = 7), 4.0-4.3 (m, 4 H), 4.63 (d, J = 9, 1 H). ¹³C NMR (CDCl₃): 14.0, 26.1, 26.3, 26.4, 28.2, 32.8, 33.5, 34.2, 39.8, 50.3, 62.0, 72.4, 79.0, 155.2, 173.7.

 $[R-(R^*,S^*)]$ - and $[S-(R^*,R^*)]-\beta-[[(1,1-Dimethylethoxy)$ carbonyl]amino]-a-hydroxycyclohexanebutanoic Acid, Ethyl Ester (24 and 25). Mercury(II) chloride (7.35 g, 27.08 mmol) and mercury(II) oxide (2.93 g, 13.54 mmol) were added to a solution of diastereomeric mixture of α -hydroxy ortho thioesters 22 and 23 (2.77 g, 6.77 mmol) in 95% aqueous EtOH (175 mL). After stirring for 36 h at 25 °C, the reaction mixture was filtered through Celite and the solids were washed with additional EtOH (30 mL). The filtrate was diluted with chloroform (250 mL), and the organic solution was washed with saturated aqueous NH₄Cl $(2 \times 100 \text{ mL})$ and 75% aqueous NH₄OAc (1 × 100 mL), dried, and concentrated to give 2.56 g of crude product. Chromatographic purification eluting with 4:1 to 2:1 hexane/ether yielded 24 as the fast moving isomer $(1.225 \text{ g}, 55\%, R_f = 0.27 \text{ in } 1:1 \text{ hexane}/$ ether) and 25 as the slow moving Isomer (356 mg, 16%, $R_f = 0.17$ in 1:1 hexane/ether). The data for 25 is as follows. MS: (M + H)⁺ 330. ¹H NMR (CDCl₃): 0.8–1.85 (m, 13 H), 1.23 (t, J = 7, 3 H) 1.38 (s, 9 H), 3.18 (d, J = 5, 1 H), 4.0–4.3 (m, 4 H), 4.75 (d, J = 9, 1 H). ¹³C NMR (CDCl₃): 14.2, 26.0, 26.3, 26.4, 28.3, 32.3, 33.8, 34.2, 36.7, 50.5, 61.8, 73.4, 79.5, 155.6, 173.0.

(4S-trans)-4-(Cyclohexylmethyl)-2-oxo-5-oxazolidinecarboxylic Acid, Ethyl Ester (26). Anhydrous HCl in dioxane (4 M, 0.5 mL) was added to a solution of 24 (31 mg, 0.094 mmol) in EtOAc (1 mL). TLC check after 4 h revealed total disappearance of starting material. The reaction mixture was concentrated in vacuo, and the residue was dissolved in THF (1 mL). $iPr_2NEt (30 \mu L)$ and CDI (25 mg) were added to the solution, and the reaction was left for overnight stirring at room temperature. The next day, the reaction mixture was concentrated to give a residue which was dissolved in EtOAc (30 mL) and washed sequentially with saturated NaHCO₃ (1×15 mL), 10%HCl $(2 \times 15 \text{ mL})$, and saturated NaCl $(1 \times 15 \text{ mL})$. Drying and concentration gave a residue which was purified by flash chromatography to give pure 26 (18 mg, 75%). TLC: $R_f = 0.37$ (1:1 hexane/EtOAc). ¹H NMR (CDCl₃): 0.8-1.8 (m, 13 H), 1.32 (t, J = 7, 3 H) 3.95 (m, 1 H), 4.3 (q, J = 7, 2 H), 4.55 (d, J = 5, 3 H)1 H), 6.31 (s, 1 H). ¹³C NMR (CDCl₃): 14.1, 25.9, 26.0, 26.2, 32.7, 33.3, 34.0, 43.7, 53.8, 62.2, 78.3, 158.1, 168.8.

(4S-cis)-4-(Cyclohexylmethyl)-2-oxo-5-oxazolidinecarboxylic Acid, Ethyl Ester (27). Anhydrous HCl in dioxane (4 M, 0.75 mL) was added to a solution of 25 (60 mg, 0.182 mmol) in EtOAc (1 mL). TLC check after 1 h revealed total disappearance of starting material. The reaction mixture was concentrated in vacuo, and the residue was dissolved in THF (1.5 mL). iPr₂NEt (48 µL, 0.273 mmol) and CDI (38.3 mg, 0.237 mmol) were added to the solution, and the reaction was left for overnight stirring at room temperature. The next day, the reaction mixture was concentrated to give a residue which was dissolved in EtOAc (45 mL) and washed sequentially with saturated NaHCO₃ (1 \times 15 mL), 10% HCl (2 \times 20 mL), and saturated NaCl (1×15 mL). Drying and concentration gave 55 mg of residue which was purified by flash chromatography to give pure 27 (30 mg, 65%). ¹H NMR (CDCl₃): 0.8-1.8 (m, 13 H), 1.35 (t, J = 7, 3 H) 4.1–4.3 (m, 3 H), 5.04 (d, J = 8.5, 1 H), 6.4 (s, 1 H). ¹³C NMR (CDCl₃): 14.1, 25.7, 26.0, 26.2, 31.9, 34.0, 34.2, 38.0, 52.0, 61.8, 76.7, 158.3, 167.2.

(S*,R*)-N-[1-(Cyclohexylmethyl)-3-ethoxy-2-hydroxy-3 $oxopropyl]-N^2-[(1,1-dimethylethoxy)carbonyl]-L-leucina$ mide (28). The α -hydroxy ester 24 (1.245 g, 3.787 mmol) was treated with 1:1 TFA/CH₂Cl₂ (20 mL) at room temperature, and a TLC check after 15 min revealed completion of the reaction. The reaction mixture was filtered through a short cotton plug and the filtrate concentrated to give a residue which was taken up in MeOH (15 mL) and refiltered through a short pad of Celite to remove the precipitated impurities. The filtrate was reconcentrated to give the amine salt as a yellow-colored clear oil (1.285 g), which was dissolved in THF (10 mL) and treated with iPr₂NEt (717 μ L, 3.75 mmol). In a separate flask, HOBt (574 mg, 3.75 mmol) and DCC (774 mg, 3.75 mmol) were sequentially added to a solution of BOC-Leu-OH hydrate (935 mg, 3.75 mmol) in DMF (10 mL) at 0 °C. After 20 min of stirring at room temperature, a premixed THF solution of the amine salt and

iPr2NEt prepared above was added to the reaction mixture. The reaction was stirred at 0 °C for 2 h and then left for overnight stirring at room temperature. Next day, the precipitated urea was filtered and the filtrate concentrated to give a residue that was taken up in EtOAc (100 mL) and washed sequentially with $H_2O(2 \times 25 \text{ mL}), 10\%$ citric acid $(1 \times 25 \text{ mL}),$ saturated NaHCO₈ $(1 \times 25 \text{ mL})$, and saturated NaCl $(1 \times 25 \text{ mL})$. Drying (Na_2SO_4) and concentration afforded 1.39 g crude product which after chromatographic purification eluting with 3:1 hexane/EtOAc yielded pure 28 (823 mg, 50%). TLC: $R_f = 0.25$ (2:1 hexane/ EtOAc). MS: (M + H)⁺ 443. ¹H NMR (CDCl₃): 0.7-1.9 (m, 16 H), 0.9 (m, 6 H), 1.3 (t, J = 7, 3 H) 1.45 (s, 9 H), 3.42 (d, J = 5, 1 H), 4.0 (m, 1 H), 4.15 (m, 1 H), 4.2 (q, J = 7, 2 H), 4.47 (m, 1 H), 4.95 (br s, 1 H), 6.27 (d, J = 10, 1 H). ¹³C NMR (CDCl₈): 14.0, 22.0, 22.7, 24.6, 26.0, 26.2, 26.4, 28.2, 32.7, 33.5, 34.8, 39.4, 41.2, 48.8, 53.2, 62.1, 72.1, 79.9, 155.5, 171.9, 173.4. Anal. Calcd for C23H42N2O6: C, 62.42; H, 9.57; N, 6.33. Found: C, 62.45; H, 9.53; N, 6.29.

(S*,R*)-N-[1-(Cyclohexylmethyl)-3-ethoxy-2-hydroxy-3oxopropyl]-N²-[4-morpholino-1,4-dioxo-2-(phenylmethyl)butyl]-L-leucinamide (30). 28 (395 mg, 0.892 mmol) was dissolved in EtOAc (5 mL) and treated for 30 min with 3 N HCl/AcOH. The solvents were removed on a rotary evaporator to give a residue which upon crystallization from MeOH/AcOH (1:8) gave the amine hydrochloride (240 mg, 71%). DCC (232 mg, 1.126 mmol), iPr₂NEt (216 mL, 1.126 mmol), and HOBt (172 mg, 1.126 mmol) were sequentially added to a solution of acid 29³¹ (312 mg, 1.126 mmol) and amine salt prepared above (427 mg, 1.126 mmol) in DMF (10 mL) at 0 °C. The reaction was stirred at 0 °C for 2 h and then left for overnight stirring at room temperature. The next day, the precipitated urea was filtered off, the filtrate was diluted with EtOAc (30 mL), and the organic solution was washed sequentially with H_2O (2 × 25 mL), 10% citric acid (1 \times 25 mL), saturated NaHCO₃ (1 \times 25 mL) and saturated NaCl $(1 \times 20 \text{ mL})$. Drying (Na_2SO_4) and concentration afforded 555 mg of crude product which upon flash chromatographic purification eluting with 19:19:1:0.05 Et₂O/CHCl₃/MeOH/ NH4OH yielded pure 30 (505 mg, 74.5%, overall yield from 28, 53%). Mp: 75–87 °C. TLC: $R_f = 0.55$ (9:1:0.05 CHCl₃/MeOH/ NH₄OH). $[\alpha]_D = -50.4^{\circ} (c = 1.02, MeOH)$. MS: $(M + H)^+ 602$. IR: 3311, 2955, 2924, 2853, 1741, 1640, 1540, 1447, 1387, 1367, 1270, 1235, 1216, 1203, 1148, 1116, 1030, 700. ¹H NMR (CDCl₃): 0.7-1.9 (m, 25 H), 2.35 (s, 1 H), 2.6-2.8 (m, 2 H), 2.95-3.2 (m, 2 H), 3.2-3.4 (m, 2 H), 3.45-3.7 (m, 6 H), 3.8 (d, J = 7, 1 H), 4.0-4.25(m, 4 H), 4.45 (m, 1 H), 6.5–6.65 (m, 2 H), 7.0–7.4 (m, 5 H). ^{13}C NMR (CDCl₃): 14.0, 21.7, 22.8, 24.5, 26.0, 26.2, 26.4, 32.8, 33.4, 34.0, 34.2, 37.8, 39.0, 40.3, 42.0, 43.8, 45.6, 49.8, 52.6, 61.8, 66.3, 66.6, 72.1, 126.5, 128.5, 128.8, 138.5, 170.0, 171.3, 173.2, 174.7. Anal. Calcd for C₃₃H₅₁N₃O₇: C, 65.86; H, 8.54; N, 6.98. Found: C, 65.73; H, 8.64; N, 6.75.

(S*,R*)-N-[1-(Cyclohexylmethyl)-3-ethoxy-2,2-dihydroxy-3-oxopropyl]-N²-[4-morpholino-1,4-dioxo-2-(phenylmethyl)butyl]-L-leucinamide (31). Dess-Martin reagent (310 mg, 0.75 mmol) was added in one portion to a solution of the α -hydroxy ester 30 (294 mg, 0.488 mmol) and tert-butyl alcohol (55 mg, 0.75 mmol) in dichloromethane 924 mL), and the reaction mixture was stirred overnight at room temperature. The next day, the reaction was judged to be complete by TLC, (total reaction time = 19 h). The solvents were removed on rotary evaporator, and the residue was dissolved in ethyl acetate (25 mL) and washed $(3 \times 10 \text{ mL})$ with an aqueous solution of NaHCO₃/Na₂S₂O₃ (500 mg of Na₂S₂O₃ dissolved in 30 mL of saturated NaHCO₃) to remove the excess reagent. Drying (Na₂SO₄), concentration, and chromatographic purification eluting with 40:1:0.05 CHCl₃/ MeOH/NH4OH afforded pure keto ester 31 (262 mg, 89.6%). TLC: $R_f = 0.65 (9:1:0.05 \text{ CHCl}_{g}/\text{MeOH}/\text{NH}_4\text{OH})$. $[\alpha]_D = -19.24^{\circ}$ (c = 1.0, MeOH). MS: $(M + H)^+ 600$. IR: 3427, 3423, 3405, 3395, 3295, 2955, 2925, 2854, 1731, 1641, 1539, 1498, 1463, 1448, 1297, 1271, 1236, 1116, 1068, 1048, 1031, 449, 434, 416, 410, 402, 399. ¹³C NMR (CD₃OD): 14.3, 22.2, 23.4, 25.5, 27.0, 27.2, 27.4, 33.1, 34.8, 35.2, 38.1, 39.5, 41.7, 43.2, 45.5, 47.0, 52.9, 53.1, 54.7, 63.4, 67.5, 127.5, 129.4, 130.1, 140.0, 162.1, 171.9, 174.6, 176.8, 193.9. Anal. Calcd for C₃₃H₄₉N₃O₇: C, 66.08; H, 8.23; N, 7.01. Found: C, 66.34; H, 8.43; N, 7.06.

(S*,R*)-N-[1-(Cyclohexylmethyl)-3-ethoxy-2-hydroxy-3oxopropyl]-N²-[N-[(1,1-dimethylethoxy)carbonyl]-L-pheny**lalanyl]-t-leucinamide (32).** The α -hydroxy ester 24 (625 mg, 1.89 mmol) was treated with 1:1 TFA/CH₂Cl₂ (20 mL) at room temperature, and a TLC check after 15 min revealed completion of the reaction. The reaction mixture was filtered through a short cotton plug, and the filtrate was concentrated to give a residue which was taken up in MeOH (10 mL), refiltered through a short pad of Celite to remove precipitated impurities, and reconcentrated to give the free amine salt as a yellow-colored clear oil (609 mg, 94%). iPr₂NEt (309.7 µL, 1.78 mmol) was added to a solution of the crude amine in THF (5 ml) at 0 °C. After 5 min, it was transferred to a flask containing a 0 °C DMF (5 mL) solution of 6 (672.8 mg, 1.78 mmol) and HOBt (272.3 mg, 1.78 mmol). After 5 min, DCC (367.3 mg, 1.78 mmol) was added, and the reaction mixture was stirred at 0 °C for 2 h, and then left for overnight stirring at room temperature. The next day, the reaction mixture was filtered through Celite, and concentrated, and the resulting residue was taken up in EtOAc (50 mL) and refiltered. The filtrate was washed sequentially with saturated NaHCO₃ (2×20 mL), 10% citric acid (1×20 mL), and saturated NaCl $(1 \times 25 \text{ mL})$, dried (Na₂SO₄), and concentrated to give 960 mg of residue. Chromatographic purification yielded pure 32 (752 mg, 71.7%, overall yield from 24, 68%). Mp: 164-166 °C. TLC: $R_f = 0.35$ (1:1 hexane/EtOAc). $[\alpha]_D =$ -61.2° (c = 1.37, MeOH). MS: (M + H)⁺ 590. IR (KBr): 3308, 2957, 2926, 2853, 1731, 1691, 1648, 1526, 1450, 1390, 1367, 1253, 1202, 1171. ¹H NMR (CDCl₃): 0.8–1.9 (m, 16 H), 0.88 (d, J =7, 6 H), 1.3 (t, J = 7, 3 H), 1.42 (s, 9 H), 3.08 (m, 2 H), 3.68 (d, J = 5, 1 H), 4.0-4.5 (m, 6 H), 5.1 (d, J = 7, 1 H), 6.5 (m, 2 H), 7.1-7.4 (m, 5 H). ¹³C NMR (CDCl₃): 14.1, 22.0, 22.8, 24.6, 26.1, 26.2, 26.4, 28.2, 33.0, 33.3, 34.1, 37.7, 39.2, 41.2, 49.1, 52.1, 55.6, 55.8, 62.0, 72.0, 80.5, 126.9, 128.6, 129.2, 136.3, 155.6, 171.0, 171.2, 173.2. Anal. Calcd for C₃₂H₅₁N₃O₇: C, 65.17; H, 8.72; N, 7.12. Found: C, 65.25; H, 8.72; N, 7.09.

(R*,R*)-N-[1-(Cyclohexylmethyl)-3-ethoxy-2-hydroxy-3oxopropyl]-N²-[N-[(1,1-dimethylethoxy)carbonyl]-L-phenylalanyl]-L-leucinamide (33). 23 (334 mg, 1.02 mmol) was dissolved in a solution of TFA/CH₂Cl₂ (1:1, 10 mL). After 15 min, the reaction mixture was filtered and concentrated, yielding the free amine (181 mg). A portion of the free amine (151 mg, 0.44 mmol) was treated with iPr₂ NEt (84.2 μ L, 0.44 mmol) in DMF (3 mL). In a separate flask, HOBt (67 mg, 0.44 mmol) and DCC (90.7 mg. 0.44 mmol) were added sequentially to a 0 °C solution of 6 (166.8 mg, 0.44 mmol) in DMF (3 mL). After 5 min, the amine solution was transferred to this flask. After 16 h at 0 °C, the reaction mixture was filtered and concentrated to give a residue which was dissolved in EtOAc (20 mL) and washed sequentially with H_2O (3 × 10 mL), saturated aqueous NaHCO₃ $(2 \times 10 \text{ mL})$, 10% citric acid (10 mL), and saturated aqueous NaCl (10 mL). Drying and concentration gave 236 mg of crude product which after chromatographic purification and preparatory TLC afforded pure 33 (38 mg, 15%). Mp: 95-98 °C. TLC: $R_f = 0.50$ (9:1 CHCl₃/MeOH). $[\alpha]_D = -21.6^\circ$ (c = 0.44, MeOH). MS: (M + H)⁺ 590. IR (KBr): 3431, 2926, 2852, 1727, 1690, 1647, 1527, 1467, 1450, 1391, 1367, 1251, 1172, 1125, 1095, 1048, 1023, 699, 565, 536, 506, 490, 474, 459. ¹H NMR (CDCl₃): 0.8-1.85 (m, 16 H), 0.9 (d, J = 7, 6 H), 1.3 (t, J = 7, 3 H), 1.5 (s, 9 H), 3.1 (m, 2 H), 3.25 (br s, 1 H), 4.15-4.45 (m, 6 H), 4.9 (m, 1 H), 6.2-6.4 (m, 2 H), 7.15-7.35 (m, 5 H). ¹³C NMR (CDCl₃): 14.2, 22.0, 22.9, 24.7, 26.0, 26.2, 26.4, 28.2, 32.3, 33.9, 34.0, 36.3, 37.5, 40.9, 49.6, 52.4, 56.0, 61.8, 73.0, 80.6, 127.0, 128.7, 129.3, 136.5, 155.5, 171.6, 172.6. Anal. Calcd for C32H51N3O7.1.2H2O: C, 62.86; H, 8.80; N, 6.87. Found: C, 63.24; H, 9.01; N, 6.48.

N-[N-(Cyclopentylcarbonyl)-L-phenylalanyl]-L-leucine, Methyl Ester (34). Compound 5 (12.01 g, 30.64 mmol) was treated with a solution of HCl in acetic acid (62 mL) for 1 h and concentrated to give an oily residue. It was triturated with toluene and concentrated to yield the dipeptide amine hydrochloride salt (10 g, 98%). A portion of this compound (5.0 g, 15.2 mmol), cyclopentane carboxylic acid (1.65 mL, 15.2 mmol), iPr₂NEt (2.93 mL, 17 mmol), and HOBt (2.33 g, 15.2 mmol) were dissolved in DMF (60 mL) and cooled to 0 °C. DCC (3.14 g, 15.2 mmol) was finally added, and after 16 h at 0 °C, the reaction mixture was filtered and the filtrate concentrated. The residue was taken up in EtOAc (250 mL) and washed sequentially with H₂O (3 × 150 mL), and saturated NaHCO₈ (1 × 150 mL). 5% HCl (1 × 150 mL), and saturated NaCl (1 × 150 mL). Drying and concentration gave 6.0 g of crude product which after chromatographic purification eluting with 4:1 to 1:1 hexane/EtOAc afforded pure 34 (3.4 g, 57.6%). Mp: 170–171 °C. TLC: $R_f = 0.41$ (1:1 hexane/EtOAc). $[\alpha]_D = -23.9^\circ$ (c = 1.18, MeOH). MS: (M-H)⁺ 387. ¹H NMR (CDCl₃): 0.9 (d, 6 H, J = 6), 1.4–2.0 (m, 11 H), 2.5 (m, 1 H), 3.08 (d, 2 H, J = 8), 3.7 (s, 3 H), 4.5 (m, 1 H), 4.68 (m, 1 H), 6.05 (d, 1 H, J = 8), 6.28 (d, 1 H, J = 8), 7.15–7.35 (m, 5 H). ¹³C NMR (CDCl₃): 21.8, 22.6, 24.7, 25.8, 29.9, 30.5, 38.1, 41.0, 45.4, 50.9, 52.1, 53.9, 126.7, 128.3, 129.4, 136.6, 171.4, 172.7, 176.3. Anal. Calcd for C₂₂H₃₂N₂O₄: C, 68.01; H, 8.30; N, 7.21. Found: C, 67.57; H, 8.31; N, 7.20.

N-[N-(Cyclopentylcarbonyl)-L-phenylalanyl]-L-leucine (35). NaOH (1 N, 12.36 mL, 12.36 mmol) was added to a solution of 34 (2.04 g, 5.3 mmol) in MeOH (20 mL). After 5 h at room temperature, the reaction mixture was concentrated, and the residue was taken up in a mixture of H₂O (20 mL) and EtOAc (50 mL) and acidified to pH = 2. The layers were separated, and the aqueous solution was reextracted with EtOAc $(3 \times 75 \text{ mL})$. The combined organic extracts were dried and concentrated to yield 35 (1.84 g, 98.4%). Mp: 148–151 °C. $[\alpha]_D = -12.9^\circ$ (c = 1.19, MeOH). MS: $[(M + H)^+ - H_2O]$ 357. ¹H NMR (CD₃OD): 0.95 (m, 6 H), 1.35-1.9 (m, 11 H), 2.6 (m, 1 H), 2.87 (dd, 1 H, J = 9, 14, 3.18 (dd, 1 H, J = 5, 14), 4.45 (m, 1 H), 4.71 (m, 1 H), 7.25 (m, 5 H). ¹³C NMR (CDCl₃): 22.0, 23.4, 25.9, 26.7, 26.8, 30.9, 31.6, 38.9, 41.7, 46.1, 52.0, 55.4, 127.5, 129.2, 130.4, 138.4, 173.9, 175.7, 178.8. Anal. Calcd for C₂₁H₃₀N₂O₄·1.34H₂O: C, 63.28; H, 8.26; N, 7.03. Found: C, 63.32; H, 7.87; N, 7.01.

 $[R-(R^*,S^*)]-\beta-[[(1,1-Dimethylethoxy)carbonyl]amino]-\alpha$ (2-propyl-1,3-dithian-2-yl)cyclohexanepropanol (37). nBuLi (2.5 M, 11.76 mL, 29.4 mmol) was added dropwise at -25 °C to a solution of 2-n-propyl-1,3-dithiane (36, 4.536 g, 28.0 mmol; D. Seebach et al. Synthesis, 1976, 477) in THF (40 mL). After stirring for 2 h at -20 to -25 °C, the solution was cooled to -78°C and aldehyde 2 (3.57 g, 14.0 mmol) in THF (20 mL) was added. The reaction mixture was warmed gradually from -78 to -50 °C over a period of 18 h and quenched with saturated NH4Cl (50 mL), and the two layers were separated upon warming to room temperature. The aqueous layer was diluted with water $(75\,\mathrm{mL})$ to dissolve the precipitated NH4Cl and reextracted with EtOAc $(2 \times 50 \text{ mL})$. The combined organic extracts were washed with saturated NaCl $(1 \times 75 \text{ mL})$, dried (Na_2SO_4) , and concentrated to give the crude product (7.9 g) which, upon flash chromatographic purification eluting from 9:1 to 2:1 hexane/ EtOAc, yielded pure 37 (1.517 g, 26%). The product appeared to be a single diastereomer by ¹³C. TLC: $R_f = 0.56$ (1:1 hexane/ ether). MS: (M + H)⁺ 418. ¹H NMR (CDCl₃): 1.44 (m, 9 H), 0.8-2.15 (m, 22 H), 2.5-3.05 (m, 4H), 3.88 (s, 1 H), 4.37 (m, 1 H), 5.05 (d, 1 H, J = 8). ¹³C NMR (CDCl₃): 14.5, 18.2, 24.3, 25.0, 26.0, 26.39, 26.47, 26.6, 28.4, 33.3, 34.5, 36.2, 44.9, 45.5, 59.1, 71.3, 78.6, 154.8. Anal. Calcd for C21H39NO3S2: C, 60.39; H, 9.41; N, 3.35. Found: C, 60.88; H, 9.31; N, 3.50.

[*R*-(*R**,*S**)]- β -Amino- α -(2-propyl-1,3-dithian-2-yl)cyclohexanepropanol (38). 37 (2.37 g, 5.7 mmol) was dissolved in a saturated solution anhydrous HCl in EtOAc (20 mL) and stirred for 1 hat 0 °C and 3 hat room temperature. The reaction mixture was concentrated *in vacuo* and the residue triturated with Et₂O. Filtration of the precipitated product and drying gave pure hydrochloride salt (1.493 g, 75%). TLC: $R_f = 0.15$ (9:1:0.05 CHCl₃/MeOH/NH₄OH). ¹H NMR (CD₃OD): 0.8-2.10 (m, 22 H), 2.75 (m, 2 H), 3.10 (m, 2 H), 3.76 (t, 1 H, *J* = 7), 3.93 (s, 1 H). ¹³C NMR (CD₃OD): 14.6, 18.8, 25.6, 27.9, 27.3, 27.4, 27.7, 33.9, 34.4, 34.8, 40.0, 41.7, 50.3, 57.7, 75.6. Anal. Calcd for C₁₆H₃₁NO₂S·1.0HCl: C, 54.29; H, 9.11; N, 3.96; S, 18.11; Cl, 10.01. Found: C, 54.21; H, 9.17; N, 3.98; S, 18.03; Cl, 10.00.

 (S^*, \mathbb{R}^*) -N-[1-(Cyclohexylmethyl)-2-hydroxy-2-(2-propyl-1,3-dithian-2-yl)ethyl]-N²-[N-[(1,1-dimethylethoxy)carbonyl]-L-phenylalanyl]-L-leucinamide (39). The amine hydrochloride salt 38 (388 mg, 1.1 mmol) and acid 6 (415.8 mg, 1.1 mmol) were dissolved in DMF (5 mL) and cooled to 0 °C. iPr₂NEt (191.4 μ L, 1.1 mmol) was added and after 5 min, this was followed by sequential addition of HOBt (168.3 mg, 1.1 mmol) and DCC (227 mg, 1.1 mmol). The reaction mixture was left for overnight stirring with gradual warming to room temperature. The next day (total reaction time, 15 h), the reaction mixture was diluted with EtOAc (35 mL), the urea was filtered off, and the filtrate was washed sequentially with H_2O (2 × 20 mL), saturated NaHCO₃ $(2 \times 20 \text{ mL})$, 10% citric acid $(1 \times 20 \text{ mL})$, and saturated NaCl $(1 \times 20 \text{ mL})$. Drying (Na_2SO_4) and concentration afforded crude product (821 mg) which, upon chromatographic purification eluting with 4:1 Hex/EtOAc, yielded (621 mg, 83.3%) pure 39. Mp: 87-92 °C. TLC: $R_f = 0.50$ (1:1 hexane/EtOAc). $[\alpha]_D =$ -35.0° (c = 0.34, MeOH). MS: (M + H)⁺ 678. IR (KBr):3421, 2957, 2924, 2870, 2851, 1651, 1513, 1468, 1451, 1388, 1367, 1275, 1250, 1169, 1081, 1050, 1023, 699. ¹H NMR (CDCl₃): 1.45 (m, 9 H), 0.8-2.15 (m, 31 H), 2.65 (m, 2 H), 2.9 (m, 2 H), 3.08 (m, 3 H), 3.95 (m, 1 H), 4.35 (m, 2 H), 4.55 (m, 1 H), 4.92 (m, 1 H), 6.35(d, 1 H, J = 7), 6.45 (d, 1 H, J = 7), 7.15-7.35 (m, 5 H).¹³C NMR (CDCl₃): 14.3, 18.0, 21.6, 22.8, 23.9, 24.3, 24.9, 25.8, 26.1, 26.3, 28.0, 32.5, 33.5, 34.3, 35.7, 37.2, 41.2, 43.8, 44.6, 51.6, 55.5, 59.0, 70.7, 79.7, 126.5, 128.2, 129.1, 136.4, 155.1, 169.7, 170.9. Anal. Calcd for C₃₆H₅₉N₃O₅S₂.0.6H₂O: C, 62.82; H, 8.81; N, 6.11; S, 9.32. Found: C, 62.84; H, 8.60; N, 6.00; S, 9.13.

(S*,R*)-N-[1-(Cyclohexylmethyl)-2-hydroxy-2-(2-propyl-1,3-dithian-2-yl)ethyl]-N2-[N-(cyclopentylcarbonyl)-L-phenylalanyl]-L-leucinamide (40). The amine hydrochloride 38 (707 mg, 2 mmol) was added to a solution of acid 35 (748 mg, 2 mmol) in THF (8 mL) and cooled to 0 °C. HOBt (306 mg, 2 mmol), iPr₂NEt (383 mL, 2.2 mmol), and DCC (413 mg, 2 mmol) were then sequentially added, and the reaction was stirred for 16 h at 0 °C. The reaction mixture was filtered and the filtrate concentrated to give a residue which was taken up in EtOAc (50 mL) and washed sequentially with H_2O (2 × 30 mL), saturated NaHCO₃ (2×40 mL), 10% citric acid (40 mL), and saturated NaCl (40 mL). Drying and concentration gave the crude product (1.2 g) which was purified by flash chromatography eluting from 9:1 to 4:1 Hex/EtOAc to give 40 (936 mg, 72%). Mp: 135-142 °C. TLC: $R_f = 0.36$ (1:1 hexane/EtOAc). $[\alpha]_D = -30.4^\circ$ (c = 1.16, MeOH). MS: (M + H)⁺ 674. ¹H NMR (CD₃OD): 0.9 (m, 9 H), 1.05–2.05 (m, 30 H), 2.5–2.7 (m, 3 H), 2.8–3.0 (m, 2 H), 3.2 (m, 2 H), 3.95 (s, 1 H), 4.4 (m, 1 H), 4.6-4.8 (m, 3 H), 7.20 (m, 1 H), 7.25 (s, 5 H), 7.4 (d, 1 H, J = 9), 7.88 (d, 1 H, J = 8). ¹³C NMR (CDCl₃): 15.1, 19.2, 22.0, 23.6, 25.6, 25.7, 26.2, 26.9, 27.0, 27.4, 27.7, 30.9, 31.7, 34.5, 35.5, 37.7, 39.0, 41.8, 45.3, 46.1, 46.7, 49.6, 53.2, 55.5, 60.0, 73.5, 127.6, 129.3, 130.3, 138.6, 172.7, 173.9, 178.8. Anal. Calcd for $C_{37}H_{59}N_3O_4S_2$: C, 65.93; H, 8.82; N, 6.24; S, 9.51. Found: C, 65.70; H, 8.98; N, 5.98; S, 9.45.

(S*,R*)-N-[1-(Cyclohexylmethyl)-2-hydroxy-3-oxohexyl]-N²-[N-[(1,1-dimethylethoxy)carbonyl]-L-phenylalanyl]-Lleucinamide (41). Dithiane 39 (338.5 mg, 0.5 mmol) was dissolved in CH₃CN (12 mL) and diluted with H_2O (3 mL). $Ce(NH_4)_2(NO_3)_6$ (1.096 g, 2.0 mmol) was added to this solution, and after 20 min, the reaction mixture was diluted with water (40 mL) and extracted with ether $(3 \times 30 \text{ mL})$. The combined organic extracts were dried (Na₂SO₄) and concentrated to give 378 mg of residue which, upon chromatographic purification eluting with 3:1 hexane/EtOAc, yielded hydroxy ketone 41 (140 mg, 47.7%). Mp: 149–152 °C. TLC: $R_f = 0.51$ (1:1 hexane/ EtOAc). MS: $(M + H)^+$ 588. ¹H NMR (CDCl₃): 0.87 (d, 6 H, J = 6, 0.93 (t, 3 H, J = 8), 1.38 (s, 9 H), 0.7–1.9 (m, 18 H), 2.47 (m, 1 H), 2.68 (m, 1 H), 3.07 (m, 2 H), 3.95 (d, 1 H, J = 4), 4.12(d, 1 H, J = 4), 4.32 (m, 2 H), 4.55 (m, 1 H), 5.05 (d, 1 H, J =7), 6.36 (d, 1 H, J = 9), 6.42 (d, 1 H, J = 8), 7.15–7.35 (m, 5 H). ¹³C NMR (CDCl₃): 13.6, 16.8, 22.0, 22.6, 24.6, 26.0, 26.1, 26.4, 28.1, 33.0, 33.3, 34.1, 37.7, 39.7, 39.9, 41.2, 48.1, 52.0, 55.7, 77.7, 80.4, 126.9, 128.6, 129.2, 136.4, 155.5, 171.1, 210.2. Anal. Calcd for C₃₃H₅₃N₃O₃: C, 67.43; H, 9.09; N, 7.15. Found: C, 67.14; H, 9.40; N, 7.03.

(S*,R*)-N-[1-(Cyclohexylmethyl)-2-hydroxy-3-oxohexyl]-N²-[N-(cyclopentylcarbonyl)-L-phenylalanyl]-L-leucinamide (42). A solution of Tl(NO₃)₃·3H₂O (1.05 g, 1.18 mmol) in methanol (20 mL) was added to a solution of 40 (799 mg, 1.18 mmol) in MeOH (40 mL) and Et₂O (20 mL). After 10 min, the reaction mixture was filtered and concentrated, and the residue was partitioned between EtOAc (150 mL) and H₂O (100 mL). The aqueous layer was reextracted with EtOAc (2 × 120 mL). The combined organic extracts were washed with 10% citric acid (150 mL), dried, and concentrated, affording 2.5 g of crude compound. Purification by flash chromatography, eluting with 2:1 Hex/EtOAc, yielded 42 (573 mg, 83.4%). Mp: 169 °C decomposition. TLC: $R_f = 0.40$ (1:1 hexane/EtOAc). $[\alpha]_D =$ -61.3° (c = 1.19, MeOH). MS: (M + H)⁺ 584. IR (KBr): 3290, 2956, 2925, 2869, 2853, 1714, 1640, 1541, 1449, 1398, 1369, 1230, 698, 422. ¹H NMR (CDCl₉): 0.88 (m, 6 H), 0.92 (t, 3 H, J = 7), 0.75–1.9 (m, 26 H), 2.35–2.55 (m, 2 H), 2.65–2.75 (m, 1 H), 3.07 (m, 2 H), 3.85 (m, 1 H), 4.12 (s, 1 H), 4.2 (m, 1 H), 4.47–4.7 (m, 2 H), 5.97 (d, 1 H, J = 7), 6.1 (d, 1 H, J = 9), 6.38 (d, 1 H, J = 8), 7.15–7.35 (m, 5 H). ¹³C NMR (CDCl₉): 13.6, 16.8, 22.0, 22.7, 24.5, 25.9, 26.0, 26.3, 29.6, 30.8, 33.0, 33.2, 34.1, 38.4, 39.5, 39.9, 40.9, 45.1, 48.4, 52.1, 53.5, 77.6, 126.5, 128.2, 129.5, 136.6, 171.4, 171.9, 176.3, 210.6. Anal. Calcd for C₃₄H₅₈N₃O₅-0.63H₂O: C, 68.62; H, 9.19; N, 7.06. Found: C, 68.79; H, 8.92; N, 6.99.

 (R^*) -N-[1-(Cyclohexylmethyl)-2,3-dioxohexyl]-N²-[N-[(1,1dimethylethoxy)carbonyl]-L-phenylalanyl]-L-leucinamide (43). A solution of the hydroxy ketone 41 (58.7 mg, 0.1 mmol) in CH₂Cl₂ (3 mL) was added to a suspension of Dess-Martin periodinane (65 mg, 0.15 mmol) and tBuOH (12 mg, 0.15 mmol) in CH₂Cl₂ (2 mL). The reaction mixture was stirred vigorously, and after 6 h, additional Dess-Martin reagent (153 mg, 0.45 mmol) and tBuOH (35 mg, 0.45 mmol) were added along with 5 mL of CH₂Cl₂ and the reaction mixture left for overnight stirring. The next day, the reaction mixture was filtered through Celite, the filtrate was concentrated, and the residue was chromatographed, eluting with 2:1 hexane/EtOAc, to give pure 43 (51 mg, 87.2%). Mp: 134-143 °C. TLC: $R_f = 0.44$ (2:1 hexane/ EtOAc). $[\alpha]_D = -36.6^\circ$ (c = 0.61, MeOH). MS: (M + H)⁺ 586. IR (KBr): 3294, 3065, 2960, 2928, 2873, 2854, 1713, 1691, 1645, 1526, 1451, 1390, 1367, 1315, 1266, 1252, 1170, 699, 422. ¹H NMR (CDCl₃): 0.88 (m, 9 H), 1.38 (s, 9 H), 0.7-1.85 (m, 18 H), 2.73 (m, 2 H), 3.07 (m, 2 H), 4.2–4.45 (m, 2 H), 4.75–5.0 (m, 2 H), 6.35 (m, 1 H), 6.78 (m, 1 H), 7.1-7.35 (m, 5 H). ¹³C NMR (CDCl₃): 13.5, 16.3, 21.8, 22.8, 24.5, 25.9, 26.0, 26.3, 28.2, 32.3, 33.6, 34.2, 37.7, 38.1, 40.5, 51.3, 51.8, 56.6, 80.6, 127.0, 128.7, 129.2, 136.2, 171.5, 197.2, 199.3. Anal. Calcd for C₃₃H₅₁N₃O₆: C, 67.66; H, 8.78; N, 7.18. Found: C, 67.52; H, 8.71; N, 7.13.

(*R**)-*N*-[1-(Cyclohexylmethyl)-2,3-dioxohexyl]-*N*²-[*N*-(cyclopentylcarbonyl)-L-phenylalanyl]-L-leucinamide (44). A solution of the hydroxy ketone 42 (268 mg, 0.46 mmol) was added to a suspension of Dess-Martin reagent (389 mg, 0.91 mmol) and tBuOH (68 mg, 0.92 mmol) in CH₂Cl₂ (40 mL). After 5 h, the reaction mixture was filtered and concentrated, yielding 666 mg of crude product which was subjected to chromatographic purification, eluting with 2:1 Hex/EtOAc, to afford 44 (217 mg, 81.2%). Mp: 152-161 °C. TLC: $R_f = 0.37$ (1:1 hexane/EtOAc). $[\alpha]_D = -59.1^{\circ} (c = 0.45, CHCl_3)$. MS: (M + H)⁺ 582. IR (KBr): 3278, 2957, 2927, 2870, 2854, 1715, 1638, 1542, 1449, 1398, 1227. ¹³C NMR (CDCl_3): 13.5, 16.3, 21.8, 22.9, 24.5, 25.9, 26.1, 26.3, 29.8, 30.8, 32.2, 33.3, 34.2, 37.5, 38.1, 38.8, 41.2, 44.9, 51.4, 51.8, 53.5, 126.4, 128.1, 129.3, 136.8, 172.0, 172.2, 176.3, 197.4, 199.4. Anal. Calcd for C₃₄H₅₁N₃O₅-1.0H₂O: C, 70.19; H, 8.84; N, 7.22. Found: C, 70.12; H, 8.79; N, 7.12.

N-[(1,1-Dimethylethoxy)carbonyl]-1-[(4-methylphenyl)sulfonyl]-L-histidine (45). $N-\alpha$ -[Boc-L-Histidine (12.7 g; 50 mmol) was dissolved in a solution of Na₂CO₃ (10.6 g, 100 mmol) in 150 mL of H₂O and cooled to 10 °C. Tosyl chloride (12.8 g, 67 mmol) was added in very small portions over a period of 30 min while maintaining vigorous stirring and controlling the temperature between 10 and 15 °C. After the addition was complete, the reaction mixture was warmed to room temperature and stirring was continued for an additional 4 h. The reaction mixture was extracted with Et_2O (2 × 75 mL), and the organic portions were discarded. The aqueous layer was acidified to pH = 3.3 with 1 N HCl and extracted with EtOAc (2×150 mL), and the combined organic extracts were dried and concentrated to give an oily residue. Crystallization from EtOAc afforded pure 45 (9.42 g, 46%). Mp: 120 °C decomposition. TLC: $R_f = 0.38$ $(9:1:0.1 \text{ CHCl}_{3}/\text{MeOH}/\text{AcOH})$. $[\alpha]_{D} = +15.3^{\circ} (c = 1.66, \text{CH}_{3}\text{OH})$. MS: (M + H)+ 410. ¹H NMR (CDCl₃): 1.46 (s, 9 H), 2.45 (s, 3 H), 3.18 (s, 2 H), 4.45 (m, 1 H), 5.37 (m, 1 H), 7.1 (s, 1 H), 7.37 (d, 2H, J = 8), 7.8 (d, 2H, J = 8), 8.08 (s, 1H). ¹³C NMR (CDCl₈): 21.6, 28.2, 29.6, 52.5, 79.7, 115.6, 127.4, 130.4, 134.3, 136.8, 138.1, 146.6, 155.1, 172.7. Anal. Calcd for C₁₈H₂₃N₃O₆S-0.14H₂O: C, 52.48; H, 5.69; N, 10.20; S, 7.78. Found: C, 52.72; H, 5.70; N, 9.72; S. 8.09.

(S*,R*)-N-[1-(Cyclohexylmethyl)-2-hydroxy-2-(2-propyl-1,3-dithian-2-yl)ethyl]-N²-[(1,1-dimethylethoxy)carbonyl]-1-[(4-methylphenyl)sulfonyl]-L-histidinamide (46). Trieth-

Activated Ketone Based Inhibitors of Human Renin

ylamine (108 μ L, 0.775 mmol) was added dropwise to a solution of the acid 45 (133 mg, 0.325 mmol) and the amine hydrochloride salt 38 (88.4 mg, 0.25 mmol) in CH_2Cl_2 (2.5 mL) at 0 °C. After 5 min, diphenylphosphoryl azide (DPPA, 70 µL, 0.325 mmol) was added, and the reaction mixture was stirred for 2 h at 0 °C and then overnight at room temperature The next day, the reaction mixture was concentrated on a rotary evaporator to remove excess Et₃N, and the residue was partitioned between CH₂Cl₂ (15 mL) and saturated NaHCO₃ (15 mL). The aqueous layer was reextracted $(2 \times 15 \text{ mL of CH}_2\text{Cl}_2)$, and the combined organic extracts were dried (Na₂SO₄) and concentrated to give 292 mg residue. Chromatographic purification eluting with CHCl₃/MeOH/NH₄OH (19:1:0.05) gave pure 46 (125 mg, 71%) followed by the corresponding deprotected compound 47 (27 mg, 20%, $R_f = 0.25$, 9:1:0.05 CHCl₃/MeOH/NH₄OH). 47 could be conveniently converted to 46 (98% yield) by treatment with triethylamine (1.6 equiv) and p-toluenesulfonyl chloride (1.2 equiv) in dioxane for 15 min, followed by concentration and flash chromatography of the crude product. Characterization of 46: Mp: 83-88°C. TLC: $R_f = 0.72$ (9:1:0.05 CHCl₃/MeOH/NH₄OH). $[\alpha]_{\rm D} = -16.5^{\circ} (c = 1.3, \text{MeOH}).$ ¹H NMR (CDCl₃): 1.38 (s, 9 H), 0.8-2.1 (m, 22 H), 2.45 (s, 3 H), 2.47-3.1 (m, 6 H), 3.25 (s, 1 H), 3.9 (s, 1 H), 4.35 (m, 1 H), 4.55 (m, 1 H), 5.85 (m, 1 H), 6.93 (d, 1 H, J = 8), 7.1 (s, 1 H), 7.35 (d, 2 H, J = 7), 7.8 (d, 2 H, J = 7), 7.92 (s, 1 H). ¹³C NMR (CDCl₃): 13.8, 14.1, 17.8, 21.3, 23.9, 24.7, 25.6, 25.9, 26.2, 27.9, 29.9, 32.5, 33.0, 34.0, 35.5, 43.7, 44.4, 53.5, 58.9, 71.0, 79.2, 114.4, 126.9, 130.0, 134.6, 135.8, 140.6, 145.8, 154.9, 169.0. Anal. Calcd for $C_{34}H_{52}N_4S_3O_6$: C, 57.60; H, 7.39; N, 7.90; S, 13.57. Found: C, 57.71; H, 7.48; N, 7.64; S, 13.71

Characterization of 47: TLC: $R_f = 0.25$ (9:1:0.05 CHCl₃/ MeOH/NH4OH). MS: (M + H)+ 555. 1H NMR (CDCl₃): 1.44 (s, 9 H), 0.7-2.1 (m, 22 H), 2.4-3.2 (m, 6 H), 3.9 (s, 1 H), 4.32 (m, 1 H), 4.62 (m, 1 H), 5.63 (br m, 1 H), 6.85 (s, 1 H), 7.1 (m, 1 H), 7.55 (s, 1 H).

(S*,R*)-N-[1-(Cyclohexylmethyl)-2-hydroxy-3-oxohexyl]-N²-[(1,1-dimethylethoxy)carbonyl]-1-[(4-methylphenyl)sulfonyl]-L-histidinamide (48). Tl(NO₃)3³H₂O (844 mg, 1.9 mmol) was added in one portion to a solution of the dithiane 46 (674 mg, 0.95 mmol) in MeOH/Et₂O (1:1, 40 mL) at 0 °C. After 15 min. the reaction mixture was filtered through Celite, and the solids were washed with EtOAc $(2 \times 25 \text{ mL})$. Concentration of the solid gave a residue which upon chromatographic purification eluting from 2:1 to 1:1 hexane/EtOAc yielded 48 (399 mg, 67.9%). Mp: 67-76 °C. TLC: $R_f = 0.24$ (1:2 hexane/EtOAc). $[\alpha]_D =$ -42.6° (c = 1.21, MeOH). ¹H NMR (CDCl₃): 0.93 (t, 3 H, J = 7), 1.43 (s, 9 H), 0.8–1.85 (m, 15 H), 2.43 (s, 3 H), 2.3–3.0 (m, 4 H), 3.7-3.95 (m, 1 H), 4.05 (s, 1 H), 4.27 (m, 1 H), 4.52 (m, 1 H), 5.77 (d, 1 H, J = 7), 6.7 (m, 1 H), 7.12 (s, 1 H), 7.37 (d, 2 H, J= 8), 7.84 (d, 2 H, J = 8), 7.98 (s, 1 H). ¹³C NMR (CDCl₃): 13.6, 16.9, 21.6, 25.9, 26.1, 26.4, 28.2, 30.4, 32.8, 33.3, 33.9, 39.7, 40.0, 48.0, 53.7, 77.9, 79.9, 114.8, 127.4, 130.3, 134.7, 136.1, 140.3, 146.3, 155.2, 170.7, 210.1. Anal. Calcd for C31H46N4O7S.2.2H2O: C, 58.85; H, 7.57; N, 8.86. Found: C, 59.09; H, 7.47; N, 8.56.

(S*,R*)-N-[1-(Cyclohexylmethyl)-2-hydroxy-3-oxohexyl]-N²-[(1,1-dimethylethoxy)carbonyl]-L-histidinamide (49). HOBt (93 mg, 0.6 mmol) was added in one portion to a solution of hydroxy ketone 48 (93 mg, 0.15 mmol) in MeOH (3 mL). After 4 h, methanol was removed on a rotary evaporator and the residue chromatographed (20 g silica gel, 40:1:0.5 to 19:1:0.05 CHCl₃/ MeOH/NH₄OH) to yield pure 49 (65 mg, 93%). HCl (1.0 N, 106.4 μ L, 0.106 mmol) was added to a solution of 49 (52 mg, 0.112 mmol) in methanol (10 mL). After 5 min, the solvents were stripped down and the residue was lyophilized to give the hydrochloride salt of 49. Mp: 95–150 °C. TLC: $R_f = 0.16$ (9: 1:0.05 CHCl₃/MeOH/NH₄OH). $[\alpha]_D = -72.8^\circ$ (c = 0.64, MeOH). MS: $(M + H)^+$ 465. IR (KBr): 3282, 2967, 2925, 2852, 1710, 1668, 1624, 1525, 1449, 1392, 1367, 1250, 1168, 1114, 1086, 1049, 1021, 627. ¹H NMR (CDCl₃): 0.95 (t, 3 H, J = 7), 1.43 (s, 9 H), 0.75-1.9 (m, 15 H), 2.4-3.0 (m, 4 H), 3.1-3.2 (m, 1 H), 4.3 (s, 1 H), 4.45-4.75 (m, 3 H), 5.65 (m, 1 H), 7.1 (m, 1 H), 7.35 (m, 1 H), 8.0 (m, 1 H), 8.9 (m, 1 H). ¹³C NMR (CDCl₃): 13.7, 17.0, 26.0, 26.2, 26.4, 28.3, 29.5, 32.8, 33.5, 33.9, 39.9, 40.2, 48.2, 53.7, 78.3, 80.1, 135.0, 155.5, 171.6, 210.4. Anal. Calcd for C24H41ClN4O5.0.8 H₂O: C, 55.92; H, 8.33; N, 10.87, Cl, 6.88. Found: C, 56.18; H, 8.08; N, 10.71; Cl, 7.07.

 (R^*) -N-[1-(Cyclohexylmethyl)-2,3-dioxohexyl]-N²-[(1,1dimethylethoxy)carbonyl]-1-[(4-methylphenyl)sulfonyl]-Lhistidinamide (50). The Dess-Martin periodinane reagent (308.5 mg, 0.728 equiv) was added to a solution of hydroxy ketone 48 (300 mg, 0.485 mmol) and tBuOH (53.4 mg, 0.728 equiv) in dichloromethane (5 mL). After 2 h, the reaction mixture was diluted with dichloromethane (20 mL) and washed with aqueous $NaHCO_3/Na_2S_2O_3$ (15 mL, 500 mg of $Na_2S_2O_3$ dissolved in 30 mL of saturated NaHCO₃) to remove the excess reagent. Drying and concentration gave a residue which was purified by flash chromatography, eluting with 1:1 hexane/ethyl acetate, to yield 50 (288 mg, 96%). Mp: 60-67 °C. TLC: $R_f = 0.71$ (9:1:0.05 CHCl₃/MeOH/NH₄OH). $[\alpha]_{D} = -21.6^{\circ}$ (c = 0.44, MeOH). MS: $(M + H)^+$ 617. ¹H NMR (CDCl₃): 0.95 (t, 3 H, J = 7), 1.48 (s, 9 H), 0.8–2.0 (m, 15 H), 2.45 (s, 3 H), 2.73 (t, 2 H, J = 7), 2.9 (m, 2 H), 4.4 (m, 1 H), 4.95 (m, 1 H), 5.88 (d, 1 H, J = 7), 7.12 (s, 1 H), 7.37 (d, 2 H, J = 8), 7.83 (d, 2 H, J = 8), 7.95 (s, 1 H). ¹³C NMR (CDCl₃): 13.5, 16.2, 21.5, 25.7, 25.9, 26.1, 27.8, 28.1, 30.0, 32.0, 33.6, 34.1, 38.0, 38.2, 51.6, 53.3, 80.1, 114.8, 127.3, 130.3, 134.7, 136.0, 140.4, 146.2, 155.4, 171.1, 197.0, 199.1.

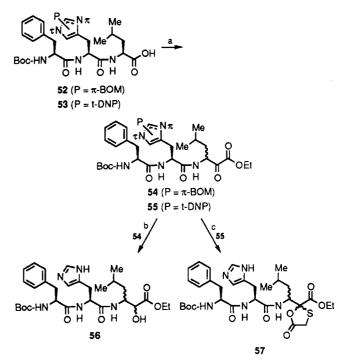
(R*)-N-[1-(Cyclohexylmethyl)-2,3-dioxohexyl]-N²-[(1,1dimethylethoxy)carbonyl]-L-histidinamide (51). HOBt (144 mg, 0.936 mmol) was added in one portion to a solution of the diketone 45 (144 mg, 0.234 mmol) in methanol (10 mL). After 3 h at room temperature, the solvents were removed in vacuo, and the residue was purified by flash chromatography, eluting with 19:1:0.05 CHCl₈/MeOH/NH4OH, to provide 51 (90 mg, 83%). TLC: $R_f = 0.20$ (9:1:0.05 CHCl₃/MeOH/NH₄OH). MS: (M + H)⁺ 463. ¹H NMR (CDCl₃, 270 MHz): 0.97 (t, 3 H, J = 7), 1.46 (s, 9 H), 0.8-1.9 (m, 15 H), 2.73 (t, 2 H, J = 7), 2.9-3.25 (m, 2 H),4.4 (m, 1 H), 5.0 (m, 1 H), 5.75 (m, 1 H), 7.38 (s, 1 H), 7.6 (s, 1 H).

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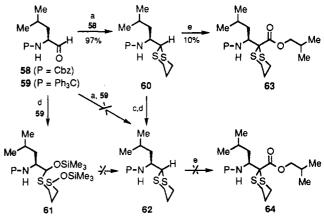
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Reagents: (a) ClC(O)CO₂Et, pyridine, DMAP; (b) H₂, Pd(OH)₂/C; (c) HSCH₂CO₂H.

(24) Our initial attempts centered on converting a suitably protected α -amino aldehyde into a dithiane and then acylating it with an alkyl chloroformate. Thus, N-Cbz-L-Leucinal (58) was efficiently converted to the dithiane 60, but acylation of 60 under various reaction conditions gave only modest yields of the desired ester 63. Alternatively, we decided to investigate this chemistry utilizing the N-trityl group. As expected, preparation of 62 directly from N-trityl-L-leucinal (59) using BF₃:Et₂O was unsuccessful. Attempted conversion under neutral conditions gave 61 as the only isolable intermediate, which could not be converted to 62. The trityl-protected dithiane intermediate 62 was eventually prepared from 60 by hydrogenolysis followed by N-tritylation. Unfortunately, acylation of the dithiane 62 to 64 was unsuccessful under a variety of conditions.



- Reagents: (a) BF₃·Et₂O, HS(CH₂)₂SH; (b) ZnI₂, Me₃SiS-(CH₂)₂SSiMe₈; (c) Me₃Si, (d) Ph₃CCl, iPr₂NEt; (e) nBuLi, ClCO₂iBu.
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