

Full Paper

Long-Chain 3-Acyl-4-hydroxycoumarins: Structure and Antibacterial Activity***Giancarlo Cravotto¹, Silvia Tagliapietra¹, Rossella Cappello¹, Giovanni Palmisano², Massimo Curini³, Marco Boccalini⁴**¹ Dipartimento di Scienza e Tecnologia del Farmaco, Università di Torino, Torino, Italy² Dipartimento di Scienze Chimiche e Ambientali; Università dell'Insubria, Como, Italy³ Dipartimento di Chimica e Tecnologia del Farmaco, Università di Perugia, Perugia, Italy⁴ Dipartimento Chimica Organica, Università di Firenze, Firenze, Italy

By reacting 4-hydroxycoumarin with long-chain acyl chlorides, a number of new 3-acyl derivatives were prepared and the relationship was studied between their structures and activities against *Staphylococcus aureus* ATCC 6538, *Staphylococcus epidermidis* ATCC 12228, and *Propionibacterium acnes* ATCC 11827. Antibacterial activity was associated with enolic tautomers of a tricarbonylmethane group bearing a lipophilic side chain (undec-10-enoyl, undec-10-ynoyl, palmitoyl or octadec-9-enoyl). A newly synthesized 2-acyl derivative of 2H-indene-1,3-dione containing the same tricarbonylmethane motif showed a comparable activity.

Keywords: 4-Hydroxycoumarin / 3-Acyl-4-hydroxycoumarins / 2H-Indene-1,3-dione / Antibacterial

Received: May 4, 2005; accepted: October 18, 2005

DOI 10.1002/ardp.200500127

Introduction

Many coumarin derivatives are biologically active [2, 3]. Much research has been focused on the inhibition of bacterial growth by naturally occurring coumarins (xanthotoxin, herniarin, umbelliferone, and scopoletin) and on the antifungal activity of umbelliferone, scopoletin, and coumarin itself [4–7]. Some coumarin derivatives (novobiocin and analogs) have proven very active as antibiotics [8, 9]. Among synthetic derivatives, several antibacterial 3-acyl- [10–14] and 3-carbamoyl-4-hydroxycoumarins [15, 16] have been described. The simplest one that proved moderately active, 3-acetyl-4-hydroxycoumarin [12], exists in its crystalline form, as shown in a X-ray diffraction study, predominantly as tautomer A (Scheme 1) with an intramolecular H-bond [17, 18].

In a search for more active compounds, we prepared a series of 3-acyl-4-hydroxycoumarins, **1a–d** and tested

their activity on *Staphylococcus aureus* ATCC 6538, *Propionibacterium acnes* ATCC 11827, and *Staphylococcus epidermidis* ATCC 12228. Results shed light on the relative importance for antibacterial action of the 4-hydroxycoumarin nucleus, the tricarbonylmethane motif and the side-chain length.

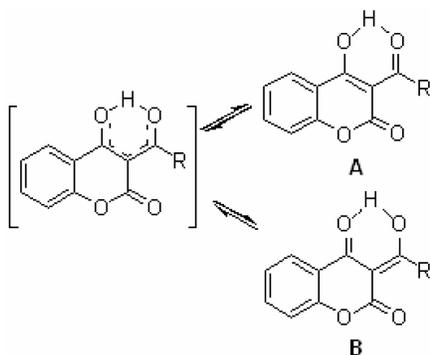
Results and discussion**Chemistry**

We began by reacting under sonochemical conditions [19] 4-hydroxycoumarin with several long-chain acyl chlorides (Scheme 1) to prepare a new series of 3-acyl-4-hydroxycoumarins (Scheme 2). The same reaction, when carried out on 4-hydroxyquinolin-2-(1H)-one, afforded the 4-O-acyl derivative (2-oxo-1,2-dihydroquinolin-4-yl-undec-10-enoate, **2a**) exclusively. In microbiological tests carried out on **2a** and on a few previously synthesized 3-alkyl-4-hydroxycoumarins and 4-alkyloxycoumarins [20] none of these compounds inhibited bacterial growth to an appreciable extent. We therefore concluded that anti-

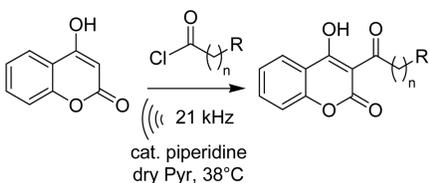
Correspondence: Giancarlo Cravotto, Dipartimento di Scienza e Tecnologia del Farmaco, Università di Torino, Via Pietro Giuria 9, 10125 Torino, Italy.

E-mail: giancarlo.cravotto@unito.it**Fax:** +39 011 670-7687

* See Reference [1].



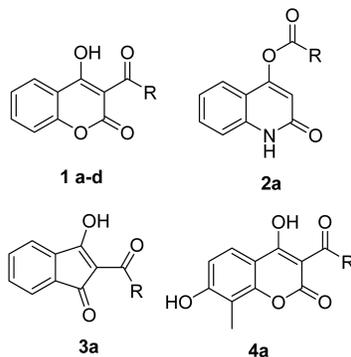
Scheme 1. Tautomers of 3-acyl-4-hydroxycoumarin.



Scheme 2. General procedure for the preparation of 3-acyl-4-hydroxycoumarins.

bacterial activity required the presence of a tricarbonyl-methane motif associated with a lipophilic side chain (C_{10} or longer). Provided these crucial features were presumed that replacing the 4-hydroxycoumarin with the 2*H*-indene-1,3-dione nucleus **3a** had no effect on the antimicrobial activity. We prepared a large number of acyl derivatives of which only the most active are described below (**1a–d**), namely those bearing an undec-10-enoyl (**1a**) [21], undec-10-ynoyl (**1b**), palmitoyl (**1c**), or (*Z*)-octadec-9-enoyl (**1d**) chain (Fig. 1).

We also prepared 4,7-dihydroxy-8-methyl-3-undec-10-enoyl-2*H*-chromen-2-one **4a** starting from 2,4-dihydroxy-3-methylacetophenone [22]. This was selectively protected (Scheme 3) in position 4 with 2-methoxyethoxymethyl chloride (MEM chloride) to yield an ether that was stable under the harsh conditions (NaH in toluene under reflux) required for cyclization with diethyl carbonate. After acylation with undec-10-enoyl chloride, the



- a** $R = -(CH_2)_8-CH=CH_2$
b $R = -(CH_2)_8-C\equiv CH$
c $R = -(CH_2)_{14}-CH_3$
d $R = -(CH_2)_7-CH=CH-(CH_2)_7-CH_3$

Figure 1. List of compounds synthesized and tested.

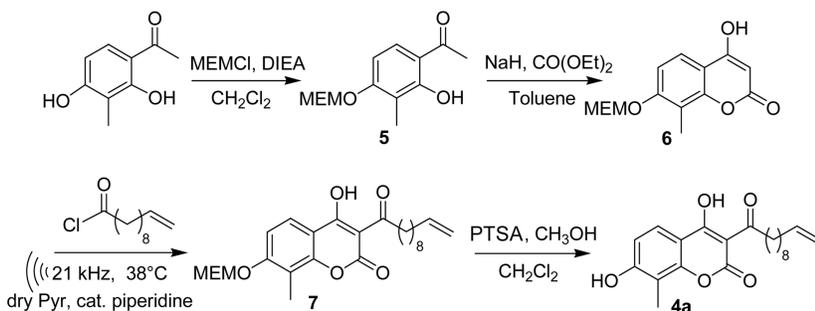
MEM ether was removed under acidic conditions with PTSA in $CH_2Cl_2/MeOH$.

Antimicrobial activity

As shown in Table 1, most active against *Staphylococcus aureus* ATCC 6538 were coumarins **1a**, **1b**, and the 1*H*-indene-1-one derivative **3a**; all of them bearing a terminal unsaturated bond on a C_{11} side chain. Acute toxicities of compounds **1a**, **1b**, and **3a** were very low (all mice survived a single dose of 2 g/kg without any marked symptoms of intolerance). The minimum inhibitory concentrations (MIC) of all compounds tested fell between 3.6 and 25.6 $\mu g/mL$ (Table 1). The three most active compounds, **1a**, **1b**, and **3a**, were tested on *Propionibacterium acnes* ATCC 11827 and *Staphylococcus epidermidis* ATCC 12228 (Table 2), confirming that the activities of **1a** and **3a** were close to each other, while **1b** was less active.

Table 1. Minimum inhibitory concentration (MIC) (mg/mL) on *Staphylococcus aureus* ATCC 6538.

| Compound | 1a | 1b | 1c | 1d | 3a | 4a |
|-------------|-----|-----|------|------|-----|------|
| MIC [mg/mL] | 3.6 | 6.2 | 25.0 | 25.6 | 6.8 | 22.6 |



Scheme 3. Synthesis of 4,7-dihydroxy-8-methyl-3-undec-10-enoyl-2*H*-chromen-2-one **4a**.

Table 2. Antimicrobial activity (MIC) of compounds **1a**, **1b**.

| Organism | MIC [$\mu\text{g/mL}$] | |
|--|--------------------------|-----------|
| | 1a | 1b |
| <i>Propionibacterium acnes</i> ATCC 11827 (1.0×10^3 c.f.u./mL) | 0.6 | 0.6 |
| <i>Staphylococcus epidermidis</i> ATCC 12228 (1.6×10^3 c.f.u./mL) | 1.2 | 1.8 |
| <i>Staphylococcus aureus</i> ATCC 6538 (1.5×10^3 c.f.u./mL) | 3.6 | 6.2 |

Conclusion

We obtained in good yields a number of new 3-acyl derivatives of 4-hydroxycoumarin **1a–d** and 4,7-dihydroxy-8-methylcoumarin **4a** that we found to be endowed with a marked antibacterial activity. This required the presence of a tricarbonylmethane motif associated with a lipophilic side chain (C_{10} or longer, best if bearing a terminal ethylene bond). Provided these crucial features were preserved, replacing 4-hydroxycoumarin with 2*H*-indene-1,3-dione **3a** hardly affected the antibacterial activity.

Financial support from Italian MIUR (COFIN 2004; prot. 2004037895) is gratefully acknowledged.

The study was partially supported by Medestea srl and we are grateful to Dr. Antonio Soleti for assistance in the biological evaluation.

Experimental

Chemistry

General methods

Anhydrous conditions were achieved (when indicated) by flame-drying flasks and other equipment. Reactions were monitored by TLC on Alugram Sil – Macherey-Nagel F254 (0.25 mm) plates (Macherey-Nagel, Düren, Germany), which were visualized by UV inspection and stained with a 5% KMnO_4 solution or by heating after a spray of 5% H_2SO_4 in ethanol. Macherey-Nagel silica gel was used for column chromatography. A Waters microPorasil column 7.8–300 (Waters Milford, MA, USA) was used for semi-preparative HPLC, a Gilson 133 refractive index refractometer (Gilson SA, Villiers-le-Bel, France) serving for peak detection. Melting points were obtained on a Büchi SMP-20 apparatus (Büchi Labortechnik, Flawil, Switzerland) and are uncorrected. $^1\text{H-NMR}$ (300 MHz) spectra were recorded on a Bruker 300 Advance spectrometer (Bruker BioSpin GmbH, Rheinstetten, Germany) at 25°C. CIMS were performed on a Finnigan-MAT TSQ70 (Thermo Electron Corporation, Bremen, Germany) with isobutane as reactant gas. Unless otherwise noted, commercially available reagents and solvents were used without further purification. Some synthetic steps were carried out under high-intensity ultrasound using a new sonochemical reactor developed in the authors' laboratory [22].

General procedure for the preparation of acyl chlorides

In a 50 mL two-necked, round-bottomed flask equipped with a magnetic stirrer, a dropping funnel and a condenser, long chain carboxylic acid (45 mmol) and anhydrous CH_2Cl_2 (0.8 mL/mmol) were mixed. SOCl_2 (1.5 equivalent mol) was added dropwise to the stirred mixture, which was subsequently heated under reflux for 10 h. The product was recovered by evaporating the solvent and the excess of SOCl_2 under vacuum.

General procedure for the acylation

The reaction was carried out under nitrogen atmosphere in the above mentioned sonochemical reactor that was thermostated by four Peltier modules. To the PTFE reaction vessel, 4-hydroxycoumarin, 4,7-dihydroxy-8-methylcoumarin or 2*H*-indene-1,3-dione, a catalytic amount of piperidine, and anhydrous pyridine (1 mL/mmol) were added. After the solution was cooled to 0°C, acyl chloride (1.6 equivalent mol) was slowly added through a septum. The mixture was sonicated (21 kHz, 40 W/cm²) for about 1.5 h under nitrogen at 38°C, and monitored by TLC. Work-up: the reacted mixture was diluted with EtOAc and washed with 5% HCl and NaHCO_3 , dried over Na_2SO_4 , filtered, and evaporated to dryness.

4-Hydroxy-3-undec-10-ynoyl-2*H*-chromen-2-one **1b**

Compound **1b** was obtained as white powder; mp. 109°C. IR (KBr) ν cm^{-1} 1724, 1618, 1553, 760; $^1\text{H-NMR}$ (CDCl_3 , 300 MHz): δ 8.07 (1H, dd, $J = 7.9$ Hz, 1.5 Hz, H-5), 7.70 (1H, td, $J = 7.9$ Hz, 1.5 Hz, H-7), 7.37–7.27 (2H, m, H-8 and H-6), 3.20 (2H, t, $J = 7.2$ Hz, 2'- CH_2), 2.13 (2H, td, $J = 6.9$ Hz, 2.2 Hz, 9'- CH_2), 1.95–1.30 (13H, m, aliph. chain). CIMS: 327 $[\text{M}+\text{H}]^+$. $R_f = 0.45$ (hexane/EtOAc (4:1)).

4-Hydroxy-3-palmitoyl-2*H*-chromen-2-one **1c** [11]

Compound **1c** was obtained as white powder; mp. 103–104°C. IR (KBr) ν cm^{-1} 1716, 1606, 1472, 1229, 899; $^1\text{H-NMR}$ (CDCl_3 , 300 MHz): δ 8.08 (1H, dd, $J = 7.6$ Hz, 1.5 Hz, H-5), 7.69 (1H, td, $J = 7.6$ Hz, 1.5 Hz, H-7), 7.37–7.26 (2H, m, H-8 and H-6), 3.20 (2H, t, $J = 7.3$ Hz, 2'- CH_2), 1.69 (2H, quintet, $J = 7.1$ Hz, 3'- CH_2), 1.42–1.16 (24H, m, aliph. chain), 0.88 (3H, t, $J = 6.9$ Hz, 16'- CH_3). CIMS: 401 $[\text{M}+\text{H}]^+$. $R_f = 0.6$ (hexane/EtOAc (4:1)).

4-Hydroxy-3-[(9*Z*)-octadec-9-enoyl]-2*H*-chromen-2-one **1d**

Compound **1d** was obtained as white powder; mp. 196–197°C. IR (KBr) ν cm^{-1} 1713, 1607, 1549, 1468, 768; $^1\text{H-NMR}$ (CDCl_3 , 300 MHz): δ 8.0 (1H, dd, $J = 7.8$ Hz, 1.5 Hz, H-5), 7.70 (1H, td, $J = 7.8$ Hz, 1.5 Hz, H-7), 7.27–7.37 (2H, m, H-8 and H-6), 5.35 (2H, m, H-9' and H-10'), 3.20 (2H, t, $J = 7.2$ Hz, 2'- CH_2), 2.01 (4H, m, 8'- CH_2 and 11'- CH_2), 1.73 (2H, quintet, $J = 6.7$ Hz, 3'- CH_2), 1.51–1.13 (20H, m, aliph. chain), 0.88 (3H, t, $J = 6.9$ Hz, 18'- CH_3). CIMS: 427 $[\text{M}+\text{H}]^+$. $R_f = 0.65$ (hexane/EtOAc (4:1)).

3-Hydroxy-2-undec-10-enoyl-2*H*-indene-1,3-dione **3a**

Compound **3a** was obtained as yellow oil. IR (KBr) ν cm^{-1} 2359, 1709, 1655, 1610, 883; $^1\text{H-NMR}$ (CDCl_3 , 300 MHz): δ 7.86–7.82 (2H, m, H-7, H-4), 7.76–7.67 (2H, m, H-5, H-6), 5.87–5.74 (1H, m, H-10'), 5.02–4.91 (2H, m, H-11'), 2.97 (2H, t, $J = 7.4$ Hz, 2'- CH_2), 2.12 (2H, m, 9'- CH_2), 1.62 (2H, quintet, 3'- CH_2), 1.48–1.11 (10H, m, aliph. chain). CIMS: 313 $[\text{M}+\text{H}]^+$. $R_f = 0.6$ (hexane/EtOAc (4:1)).

1-{2-Hydroxy-4-[(2-methoxyethoxy)methoxy]-3-methylphenyl}ethanone **5**

Compound **5** was obtained as white powder; mp. 57–59°C. IR (KBr) ν cm^{-1} 1634, 1499, 1422, 1381, 1333, 1260, 1073, 1101; $^1\text{H-NMR}$

NMR (CDCl₃, 300 MHz): δ 12.81 (1H, s, H-2), 7.59 (1H, d, J = 9.00 Hz, H-6), 6.72 (1H, d, J = 9.0 Hz, H-5), 5.37 (2H, s, H-1'), 3.85 (2H, t, J = 4.5 Hz, 3'-CH₂), 3.57 (2H, J = 4.5 Hz, 4'-CH₂), 3.39 (3H, s, 6'-OCH₃), 2.59 (3H, s, 2''-CH₃), 2.13 (3H, s, 3-CH₃). CIMS: 255 [M+H]⁺. R_f = 0.65 (hexane/EtOAc (3:2)).

4-Hydroxy-7-[(2-methoxyethoxy)methoxy]-8-methyl-2H-chromen-2-one **6**

To a magnetically stirred suspension of 0.355 g of NaH (8.12 mmol, 2 equivalent/mol) in 15 mL anhydrous toluene 1.03 g (4.06 mmol, 1 equivalent/mol) of compound **5** were added, followed by slowly dropping in 1.23 mL (10.16 mmol, 2.5 equivalent/mol) of diethylcarbonate. The mixture was refluxed for 12 h and the reaction was monitored by TLC, eluent CHCl₃/MeOH (9:1). Work-up: the reacted mixture was transferred to a separatory funnel and extracted with H₂O. When the aqueous phase was acidified with 2N H₂SO₄, a voluminous precipitate was obtained. The solid was collected on a Büchner funnel, washed with cold H₂O, and dried by heating at a temperature no higher than 60°C, yielding 0.76 g of **6** (yield 67%).

Compound **6**: white powder, mp. 132–134°C. IR (KBr) ν cm⁻¹ 1709, 1608, 1555, 1466, 1252, 1196, 972, 762; ¹H-NMR (CDCl₃, 300 MHz): δ 10.78 (1H, brs, 4-OH), 7.62 (1H, d, J = 8.8 Hz, H-5), 7.09 (1H, d, J = 8.8 Hz, H-6), 5.76 (1H, s, H-3), 5.40 (2H, s, 1'-CH₂), 3.90 (2H, t, J = 4.4 Hz, 3'-CH₂), 3.64 (3H, t, J = 4.4 Hz, 4'-CH₂), 3.42 (3H, s, 6'-OCH₃), 2.28 (3H, s, 8-CH₃). CIMS: 281 [M+H]⁺. R_f = 0.71 (CHCl₃/MeOH (9:1)).

4-Hydroxy-7-[(2-methoxyethoxy)methoxy]-8-methyl-3-undec-10-enoyl-2H-chromen-2-one **7**

Compound **7** was obtained as white powder; mp. 65–67°C. IR (KBr) ν cm⁻¹ 1725, 1613, 1559, 1458, 1248, 1078, 988; ¹H-NMR (CDCl₃, 300 MHz): δ 7.89 (1H, d, J = 8.9 Hz, H-5), 7.15 (1H, d, J = 8.9 Hz, H-6), 5.85 (1H, m, H-10''), 5.43 (2H, s, 1'-CH₂), 5.03–4.92 (2H, m, 11''-CH₂), 3.86 (2H, t, J = 4.35 Hz, 3'-CH₂), 3.57 (2H, t, J = 4.35 Hz, 4'-CH₂), 3.38 (3H, s, 6'-CH₃), 3.18 (2H, t, J = 7.3 Hz, 2''-CH₂), 2.30 (3H, s, 8-CH₃), 2.12 (2H, m, 9''-CH₂), 1.75 (2H, quintet, J = 7.0 Hz, 3''-CH₂), 1.57–1.20 (10H, m, aliph. chain). CIMS: 447 [M+H]⁺. R_f = 0.57 (hexane/EtOAc (3:2)).

4,7-Dihydroxy-8-methyl-3-undec-10-enoyl-2H-chromen-2-one **4a**

Compound **4a** was obtained as white powder; mp. 161–163°C. IR (KBr) ν cm⁻¹ 3480, 2922, 1688, 1618, 1568, 1090; ¹H-NMR (CDCl₃, 300 MHz): δ 9.8 (2H, brs, 2 × OH), 7.83 (1H, d, J = 8.3 Hz, H-5), 6.81 (1H, d, J = 8.3 Hz, H-6), 5.80 (1H, m, H-10'), 5.02–4.91 (2H, m, 11'-CH₂), 3.17 (2H, t, J = 7.3 Hz, 2'-CH₂), 2.31 (3H, s, 8-CH₃), 2.04 (2H, m, 9'-CH₂), 1.72 (2H, quintet, J = 7.1 Hz, 3'-CH₂) 1.57–1.27 (10H, m, aliph. chain). CIMS: 359 [M+H]⁺. R_f = 0.64 (hexane/EtOAc (3:2)).

Antimicrobial activity

Compounds to be tested were dissolved in acetone (10 mg/mL) and diluted with warm culture medium. Mueller-Hinton Broth containing 1% Tween 20 was used for tests with *Staphylococcus aureus* and *Staphylococcus epidermidis*; incubations were carried out aerobically at 36 ± 1°C. Actinomyces Broth containing 1% Tween 20 was used for *Propionibacterium acnes* that was incubated anaerobically at 36 ± 1°C. Duplicate blanks were incubated alongside culture aliquots containing scalar dilution of each tri-

carbonyl derivative. After 1–4 days of incubation, bacterial growth was estimated by turbidimetry or by plating on agar medium.

References

- [1] Part XIV in the series: The Chemistry of Coumarin Derivatives. For Part XIII, see: G. Cravotto, G. M. Nano, G. Palmisano, S. Tagliapietra, *Synthesis* **2003**, 8, 1286–1291.
- [2] R. D. H. Murray, J. Mendez, S. A. Brown, *The Natural Coumarins*, Wiley, Chichester, UK, **1982**.
- [3] J. R. S. Hoult, M. Paya, *Gen. Pharmacol.* **1996**, 27, 713–722.
- [4] L. Jurd, J. Corse, A. D. King, H. Bayne, K. Mihara, *Phytochem.* **1971**, 10, 2971–2974.
- [5] L. Jurd, A. D. King, K. Mihara, *Phytochem.* **1971**, 10, 2965–2970.
- [6] M. C. Recio, J. L. Rios, A. Villar, *Phytother. Res.* **1989**, 3, 117–125.
- [7] T. Ojala, S. Remes, P. Haansuu, H. Vuorela, R. Hiltunen, K. Haahtela, P. Vuorela, *J. Ethnopharm.* **2000**, 73, 299–305.
- [8] J. W. Hinman, H. Hoeksema, E. L. Caron, W. G. Jackson, *J. Am. Chem. Soc.* **1956**, 78, 1072–1074.
- [9] P. Laurin, M. Klich, C. Dupuis-Hamelin, P. Mauvais, P. Lassaigne, A. Bonnefoy, B. Musicki, *Bioorg. Med. Chem. Lett.* **1999**, 9, 2079–2084.
- [10] B. Musicki, A. M. Periers, P. Laurin, D. Ferroud, Y. Benedetti, S. Lachaud, F. Chatreaux, J.-L. Haesslein, A. Iltis, C. Pierre, *Bioorg. Med. Chem. Lett.* **2000**, 10, 1695–1699.
- [11] C. Ukita, S. Nojima, M. Matsumoto, *J. Am. Chem. Soc.* **1950**, 72, 5143–5144.
- [12] C. Ukita, K. Arakawa, *Pharm. Bull.* **1953**, 1, 255–260.
- [13] K. Arakawa, *Pharm. Bull.* **1953**, 1, 331–334.
- [14] D. Završnik, F. Basic, F. Becic, E. Becic, S. Jazic, *Period. Biologorum* **2003**, 105, 137–139.
- [15] Merck & Co., Inc., GB 856816, 1960, [Chem. Abstr. **1962**, 56, 2361].
- [16] B. Yang, J. Sutcliffe, C. J. Dutton, (Pfizer Inc., USA) US 5985912, 1999, [Chem. Abstr. **1999**, 131, 322536].
- [17] V. F. Traven, O. B. Safronova, L. I. Vorob'eva, T. A. Chibisova, I. N. Senchenya, *Russ. J. Gen. Chem.* **2000**, 70, 793–797.
- [18] K. A. Lyssenko, M. Yu. Antipin, *Russ. Chem. Bull. Int. Ed.* **2001**, 3, 418–431.
- [19] G. Cravotto, P. Cintas, *Chem. Soc. Rev.* **2005**, 34, 1–17.
- [20] G. Cravotto, G. M. Nano, G. Palmisano, S. Tagliapietra, *Heterocycles* **2003**, 60, 1351–1358.
- [21] G. Cravotto, G. Balliano, S. Tagliapietra, S. Oliaro-Bosso, G. M. Nano, *Chem. Pharm. Bull.* **2004**, 52, 1171–1174.
- [22] P. Laurin, D. Ferroud, M. Klich, C. Dupuis-Hamelin, P. Mauvais, P. Lassaigne, A. Bonnefoy, B. Musicki, *Bioorg. Med. Chem. Lett.* **1999**, 9, 2079–2084.
- [23] G. Cravotto, G. Omiccioli, L. Stevanato, *Ultrason. Sonochem.* **2005**, 12, 213–217.