

TETRAHEDRON

# Stereoselective Reductive Amination of β-Keto Esters Derived from Dipeptides. Stereochemical and Mechanistic Studies on the Formation of 5-Carboxymethyl-2-Oxopiperazine Derivatives

Rosario Patiño-Molina, Rosario Herranz, Mª Teresa García-López, and Rosario González-Muñiz\*

Instituto de Química Médica (CSIC), Juan de la Cierva 3, 28006 Madrid, Spain.

Received 9 August 1999; revised 4 October 1999; accepted 21 October 1999

Abstract. The stereoselective generation of 3,5-disubstituted and 3,5,6-trisubstituted 2-oxopiperazine derivatives can be accomplished by intramolecular reductive amination of  $\beta$ -keto esters derived from Z-Xaa-Gly-OH and Z-Xaa-Yaa-OH dipeptides, respectively. Differences in the stereoselectivity between the use of NaBH<sub>3</sub>CN and hydrogen as reducing agents are due to the reduction of different intermediates, as deduced from experiments of isotopic labelling with deuterium. © 1999 Elsevier Science Ltd. All rights reserved.

Keywords: Peptide mimetics, piperazinones, β-keto esters

The growing use of small organic molecules as non-peptide scaffolds in the search for peptidomimetics has created a demand for new methodology for the synthesis of these molecules.<sup>1-3</sup> Ideally, approaches for making conformationally constrained scaffolds should overcome two important challenges: the appendage of amino acid side chains onto the heterocycle and the stereocontrolled generation of the new chiral centers.<sup>4</sup> Among the variety of lactams which have been used successfully as scaffolds,<sup>5-8</sup> chiral piperidones, pyrrolidinones and piperazinones have emerged as preferred structures for the development of low molecular receptor ligands.<sup>8</sup> Moreover, these templates and structurally related bicyclic analogues have been shown to be effective structural tools for probing the active conformation of bioactive peptides and enzyme inhibitors.<sup>9,10</sup>

Our current interest in templates onto which pharmacologically relevant groups can be appended, led us to describe the preparation of 3,5- and 3,6-disubstituted 2-oxopiperazines from cyanomethyleneamino and methyleneamino pseudopeptides, respectively.<sup>11</sup> Due to the known possibility of obtaining high receptor binding affinity with three conveniently oriented binding groups,<sup>12</sup> we have now focused our attention on the 2-oxopiperazine derivatives 1 able to carry three amino acid side chains. In this sense,  $\beta$ -keto esters derived from dipeptides were envisaged as appropriate precursors to the corresponding 3,5,6-trisubstituted derivatives. Thus, the keto ester moiety could serve the dual purpose of allowing intramolecular cyclization, via reductive amination, and providing the Asp side chain at C-5 position of the heterocyclic ring. In addition to that, the versatility of the carboxylate group could supply opportunities for diversification.



0040-4020/99/\$ - see front matter @ 1999 Elsevier Science Ltd. All rights reserved. PII: S0040-4020(99)00957-6 To explore the suitability of the proposed synthetic route, we first investigated the synthesis of 3,5disubstituted piperazines (1,  $R^2 = H$ ), derived from Z-Xaa-Gly-OH. The stereochemical control found by application of two different reducing agents will be discussed and an explanation will be given in terms of reaction mechanism. The preparation of a 3,5,6-trisubstituted piperazine (1,  $R^2 \neq H$ ) was also undertaken.

### **RESULTS AND DISCUSSION**

According to the method reported for the preparation of  $\beta$ -keto esters derived from amino acids,<sup>13</sup> starting compounds 2-5 were synthesized by reaction of the corresponding dipeptide with carbonyldiimidazole followed by the treatment of the generated imidazolide with monoethyl or monomethyl malonate magnesium salt. The desired 3,5-disubstituted piperazines 6-9 were formed in high yield when the corresponding  $\beta$ -ketoesters 2-5 were hydrogenated at 45°C and 45 psi of pressure, using Pd-C as catalyst (Scheme 1, Table 1). Using this procedure, the removal of the Z protecting group and the reductive amination take place in a one-pot reaction. Although in slightly lower yield (Table 1), the piperazine ring is also formed in two steps involving removal of Z group, by catalytic hydrogenation, and reduction of the resulting intermediate(s) with NaBH<sub>3</sub>CN in the presence of ZnCl<sub>2</sub>.

The absolute configuration at C-5 was assigned by means of NOE experiments. Thus, isomers with R configuration at this position showed a weak (2-3%), but significant, NOE between H-3 and H-5, indicating that these hydrogens are located on the same side of the heterocyclic ring. Similarly, the observed NOE (3-4%) between H-5 and 3-CH<sub>3</sub> or 3-CH<sub>2</sub> in the 5S diastereoisomers revealed a *cis*-relationship between these groups.



Concerning the stereochemical course of the reaction, it can be noted that diastereoisomers with R configuration at C-5 were always obtained as major products. This result can be rationalized by reduction of any of the reaction intermediates (**A**, **B** or **C**) by attack on the less hindered side of the molecule and,

therefore, on the opposite side to the R<sup>1</sup> substituent. Accordingly, better stereoselectivities were found for the Phe derivatives 8 and 9 (Table 1, entries 5 to 7), for which R<sup>1</sup> is a benzyl group, than for the Ala analogue 7 (R<sup>1</sup> = Me, entries 3 and 4). Although the conjugated enamine B was the only intermediate that can be isolated and characterized, the participation of imine A and enamine C in the reductive amination process could not be ruled out. In fact, reduction with NaBH<sub>3</sub>CN/ZnCl<sub>2</sub> led to higher diastereoisomeric excesses than catalytic hydrogenation (Table 1), indicating that the reduction proceeds either through different intermediates in the two methods or that the extent of reduction of each intermediate is different in both reactions.

Entry	Starting Compd.	<b>R</b> <sup>1</sup>	R <sup>2</sup>	Method <sup>a</sup>	Solvent	Final Compd.	Yield (%)	<b>a/b</b> ratio
1	2	н	Et	А	EtOH	6	96	_
2	2	н	Et	В	EtOH	6	20 <sup>b</sup>	_
3	3	Me	Et	А	EtOH	7ab	89	1.7:1
4	3	Me	Et	в	EtOH	7ab	80	3.9:1
5	4	Bn	Et	А	EtOH	8ab	91	3.2:1
6	4	Bn	Et	В	EtOH	8ab	77	5.3:1
7	5	Bn	Me	А	MeOH	9ab	97	3.5:1
8	5	Bn	Me	Α	MeOD	9ab	90	3.5:1
9	5	Bn	Me	В	MeOD	9ab	75	5.6:1
10	5	Bn	Me	Bc	MeOH	9ab	64	5.7:1

Table 1.– 3,5–Disubstituted-2-oxopiperazines from  $\beta$ -Ketoesters Derived from Z-Xaa-Gly-OH Dipeptides

<sup>a</sup> Method A: H<sub>2</sub>/Pd-C, 45 psi, 45°C, 48-72 h; Method B: a)H<sub>2</sub>/Pd-C, 15 psi, 25°C, 2 h; b) NaBH<sub>3</sub>CN/ZnCl<sub>2</sub>, 25°C, 3 h. <sup>b</sup> This low yield is due to the difficulties found for the extraction of compound 6 from H<sub>2</sub>O. <sup>c</sup> Reaction with NaBD<sub>3</sub>CN

In order to clarify which intermediate(s) is(are) involved in each reducing method, the formation of compounds 9 was performed in MeOD following methods A and B (Table 1, entries 8 and 9).<sup>14</sup> First of all, intermediate **B** was generated by hydrogenolysis of the Z group under mild conditions (MeOD, 15 psi, r.t., 2 h). Then, this intermediate was stirred in MeOD at room temperature for 24 h, to facilitate the maximum incorporation of deuterium into the molecule (measured by the decrease in the corresponding signal in the  ${}^{1}H$ NMR spectrum). The low incorporation of this isotope at C-6 and the high isotopic labelling at the 5-CH (Figure 1), indicated that, under these conditions, imine A and enamine B are predominant in the equilibrium. According to this, the isotopic labelling at the 6-position of compound 9a and 9b, obtained by method B (NaBH<sub>3</sub>CN, r.t., 3 h), is almost insignificant. In this experiment, the predominant reduction of imine A was deduced from the almost complete deuteration of the 5-CH2 group, and afterwards confirmed by a similar reaction using NaBD<sub>3</sub>CN (Table 1, entry 10), in which compounds 9a and 9b showed the labelling exclusively at C-5 (Figure 1). When the hydrogenation reaction (45 psi, 45°C, 48 h) was applied to deuterated enamine **B**, the compounds **9a** and **9b** obtained showed a high incorporation of deuterium at both C-6 and 5-CH<sub>2</sub> positions. This result indicates that, as expected, the B = A = C equilibrium is favoured by the higher temperature and prolonged reaction time used in method A. However, the fact that the isotopic labelling at the 5-CH<sub>2</sub> group was lower for compounds 9 coming from the hydrogenation reaction (method A) than for those

obtained from method B, could only be explained if an approximately 20% of enamine intermediate B is reduced in the hydrogenation. As the incorporation of deuterium at the 5-CH<sub>2</sub> position in **9a** and **9b** was the same, it seems that, within the experimental error, reduction of enamine B by method A is not a selective process. On the other hand, the different isotopic labelling at C-6 position in compounds **9a** and **9b**, obtained from method A, could indicate that hydrogenation of enamine C preferentially takes place by the lower face of the molecule. From the above results, the partial hydrogenation of enamines B and C could account for the lower diastereoselectivity found in method A when compared to the reduction with NaBH<sub>3</sub>CN, for which we have demonstrated that the reductive amination exclusively take place through the imine intermediate A. Under the NaBH<sub>3</sub>CN/ZnCl<sub>2</sub> reduction conditions, a "chelation mechanism", involving the formation of a sixmembered cyclic complex through interactions between the ZnCl<sub>2</sub> catalyst and the carbonyl oxygen of the ester and the imine nitrogen, could contribute to the stabilization of imine A.



Figure 1.- Percentages of incorporation of deuterium (method B in bold)

To explore the preparation of 3,5,6-trisubstituted 2-oxopiperazines,  $\beta$ -ketoester 10 was prepared from Z-Phe-Ala-OH. This keto ester was obtained as an inseparable 10:1 mixture of L-L and L-D diastereoisomers due to the partial epimerization of the *C*-terminal residue during activation of the dipeptide with carbonyldiimidazole.<sup>15</sup> When this mixture was allowed to react under the conditions used in Method B, 2-oxopiperazines 11a, (35%), 11b (2%) and 11c (7%) were isolated. Traces of (3S,5S,6R) diastereoisomer 11d (= 1%) were also detected in the <sup>1</sup>H NMR spectrum of the crude reaction products. During the reductive amination of the  $\beta$ -keto ester 10 of L-L configuration the hydrogen preferentially enters on the opposite side to the R<sup>1</sup> and R<sup>2</sup> substituents, giving 11a as major compound. As expected, the 11a/11b ratio (18:1, R<sup>2</sup> = Me) is higher than that obtained for compounds 8 (a/b, 5.3:1, R<sup>2</sup> = H).



The absolute configurations at C-5 and C-6 in 11a, 11b and 11c were assigned by means of the coupling constants values and NOE experiments. Thus, The  $J_{5,6}$  value of 4 Hz for 11a indicated a *cis*-relationship between H-5 and H-6, while  $J_{5,6}$  values of 7 and 9.5 Hz for 11b and 11c, respectively, agreed with a *trans* disposition of these hydrogens. Concerning NOE experiments, isomers 11a and 11c show a weak NOE between H-3 and H-5 indicating that these atoms are located in the same face of the heterocyclic ring. Moreover, in the case of isomer 11c the weak NOE between H-5 and the 6-CH<sub>3</sub> group allowed us to propose an (3S,5R,6R) configuration for this isomer. On the other hand, irradiation of H-5 of compound 11b produces an enhancement in the signals due to the 3-CH<sub>2</sub> and 6-CH<sub>3</sub> groups, in agreement with a (3S,5S,6S) configuration.

Considering that the reductive amination of both L-L and L-D diastereoisomers of 10 proceeds to the same extent, the relative percentage of 11c, when compared to that obtained for 11a + 11b, is higher than that expected for the existence of 9% of L-D 10. Therefore, it seems that in the case of the 6-substituted piperazines, a certain amount of epimerization at C-6 occurred, probably due to the fact that the imine A-enamine C equilibrium is favoured by the presence of a substituent in position  $\alpha$  to the imine carbon.

In summary, we have demonstrated that the reductive amination of  $\beta$ -keto esters derived from dipeptides is a flexible method for the preparation of 2-oxopiperazines bearing amino acid side chains at C-3, C-5 and C-6 positions. The stereochemical course of this reaction was found to be dependent on the starting dipeptide derivative and on the reducing agent. As deduced from experiments of isotopic labelling with deuterium, the exclusive reduction of imine intermediates could account for the higher selectivities found for the reactions carried out with NaBH<sub>3</sub>CN. According to the synthetic approach described in this paper, variable amino acid side chains could be incorporated on the heterocyclic template in different spatial dispositions by starting from different dipeptide derivatives.

#### EXPERIMENTAL SECTION

<sup>1</sup>H NMR spectra were recorded with a Varian Unity 300 or a Varian Unity 500 spectrometers operating at 300 and 500 MHz, respectively, using TMS as internal standard. <sup>13</sup>C NMR spectra were recorded on the same instruments (75 MHz and 125 MHz); <sup>13</sup>C NMR assignments were made by means of heteronuclear H-C correlations (HMQC, HMBC). Elemental analyses were obtained on a CHN-O-RAPID instrument. Analytical TLC was performed on aluminium sheets coated with a 0.2 mm layer of silica gel 60 F254 (Merck). Silica gel 60 (230-400 mesh, Merck) was used for column chromatography. Analytical HPLC was performed on a Waters Nova-pak C<sub>18</sub> (3.9 x 150 mm, 4  $\mu$ m) column, with a flow rate of 1 mL/min, using a tuneable UV detector set at 214 nm. Mixtures of MeCN (solvent A) and 0.05% TFA in H<sub>2</sub>O (solvent B) were used as mobile phase. Dipeptide derivatives were purchased from Bachem. Monoethyl and momomethyl malonate magnesium salts were prepared as described.<sup>16</sup>

## Synthesis of $\beta$ -keto esters derived from dipeptides

General procedure.- A solution of the corresponding Z-dipeptide (5.4 mmol) in dry THF (15 mL) was treated with carbonyldiimidazole (0.96 g, 5.9 mmol) and stirred at r.t. for 1 h. Then, the corresponding monoalkyl malonate magnesium salt (5.9 mmol) was added. After stirring for 18 h at r.t., the solvent was

evaporated and the resulting residue was treated with 1N HCl (5 mL) and extracted twice with EtOAc (25 mL). The organic layer was washed with 10% NaHCO<sub>3</sub> and brine, dried over Na<sub>2</sub>SO<sub>4</sub> and evaporated, leaving a residue which was purified on a silica gel column as specified in each case.

**Ethyl 4-[N-(benzyloxycarbonyl)glycyl]amino-3-oxobutanoate** (2).– Yield: 80%. Eluent: EtOAc/hexane (2:1). White solid: mp 200-201°C. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 1.25 (t, 3H, CH<sub>3</sub> OEt, J = 7.1), 3.46 (s, 2H, H-2), 3.89 (m, 2H, α-Gly), 4.17 (m, 4H, H-4 and CH<sub>2</sub> OEt), 5.09 (CH<sub>2</sub> Z), 5.82 (brs, 1H, α-NH Gly), 7.03 (brs, 1H, 4-NH), 7.25 (m, 5H, C<sub>6</sub>H<sub>5</sub> Z). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): 13.94 (CH<sub>3</sub> Et), 44.17 (C-2), 46.48 (α-Gly), 49.14 (C-4), 61.67 (CH<sub>2</sub> OEt), 67.08 (CH<sub>2</sub> Z), 127.97, 128.14 and 128.45 (CH Ar), 136.01 (C Ar), 156.62, 166.57 and 169.54 (CO), 198.21 (C-3). Anal. Calcd for C<sub>16</sub>H<sub>20</sub>N<sub>2</sub>O<sub>6</sub>: C 57.14, H 5.99, N 8.33. Found: C 57.02, H 6.11, N 8.17.

**Ethyl 4-[N-(benzyloxycarbonyl)-L-alanyl]amino-3-oxobutanoate** (3).– Yield: 65%. Eluent: EtOAc/hexane (1:1). White solid: mp 87-89°C. <sup>1</sup>H NMR 300 MHz, CDCl<sub>3</sub>): δ 1.26 (t, 3H, CH<sub>3</sub> OEt, J = 7.1), 1.37 (d, 3H, α-CH<sub>3</sub> Ala, J = 7.1), 3.47 (s, 2H, H-2), 4.17 (m, 4H, H-4 and CH<sub>2</sub> OEt), 4.32 (m, 1H, α-Ala), 5.08 (m, 2H, CH<sub>2</sub> Z), 5.63 (d, 1H, α-NH Ala, J = 7.4), 7.01 (brs, 1H, 4-NH), 7.29 (m, 5H, C<sub>6</sub>H<sub>5</sub> Z). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): 13.98 (CH<sub>3</sub> Et), 18.50 (α-CH<sub>3</sub> Ala), 46.55 (C-2), 49.24 (C-4), 50.39 (α-Ala), 61.69 (CH<sub>2</sub> OEt), 66.98 (CH<sub>2</sub> Z), 127.99, 128.14 and 128.48 (CH Ar), 136.05 (C Ar), 155.95, 166.57 and 172.37 (CO), 198.13 (C-3). Anal. Calcd for C<sub>17</sub>H<sub>22</sub>N<sub>2</sub>O<sub>6</sub>: C 58.28, H 6.33, N 8.00. Found: C 57.98, H 6.01, N 7.85.

**Ethyl 4-[N-(benzyloxycarbonyl)-L-phenylalanyl]amino-3-oxobutanoate** (4).– Yield: 76%. Eluent: EtOAc/hexane (1:2). White solid: mp 102-104°C. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 1.27 (t, 3H, CH<sub>3</sub> OEt, J = 7.2), 3.09 (m, 2H, β-CH<sub>2</sub> Phe), 3.41 (s, 2H, H-2), 4.17 (m, 4H, H-4 and CH<sub>2</sub> OEt), 4.24 (m, 1H, α-Phe), 5.05 (m, 2H, CH<sub>2</sub> Z), 5.51 (d, 1H, α-NH Phe, J = 8.0), 6.77 (brs, 1H, 4-NH), 7.16-7.36 (m, 10H, C<sub>6</sub>H<sub>5</sub> Phe and Z). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): 14.01 (CH<sub>3</sub> Et), 38.35 (β-CH<sub>2</sub> Phe), 46.53 (C-2), 49.24 (C-4), 56.01 (α-CH Phe), 61.71 (CH<sub>2</sub> OEt), 67.03 (CH<sub>2</sub> Z), 127.02, 127.95, 128.14, 128.48, 128.64 and 129.19 (CH Ar), 136.01 and 136.19 (C Ar), 155.97, 166.49 and 171.27 (CO), 197.77 (C-3). Anal. Calcd for C<sub>23</sub>H<sub>26</sub>N<sub>2</sub>O<sub>6</sub>: C 64.78, H 6.14, N 6.57. Found: C 64.55, H 6.20, N 6.39.

Methyl 4-[N-(benzyloxycarbonyl)-L-phenylalanyl]amino-3-oxobutanoate (5).– Yield: 63%. Eluent: EtOAc/hexane (1:2). White solid: mp 101-103°C. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 3.09 (m, 2H, β-CH<sub>2</sub> Phe), 3.44 (s, 2H, H-2), 3.73 (s, 3H, OMe), 4.17 (m, 4H, H-4), 4.49 (m, 1H, α-Phe), 5.06 (m, 2H, CH<sub>2</sub> Z), 5.37 (d, 1H, α-NH Phe, J = 7.2), 6.62 (brs, 1H, 4-NH), 7.16-7.37 (m, 10H, C<sub>6</sub>H<sub>5</sub> Phe and Z). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): 38.32 (β-CH<sub>2</sub> Phe), 46.21 (C-2), 49.26 (C-4), 52.54 (OMe), 55.98 (α-CH Phe), 67.02 (CH<sub>2</sub> Z), 127.00, 127.92, 128.13, 128.46, 128.62 and 129.17 (CH Ar), 135.99 and 136.19 (C Ar), 155.96, 166.90 and 171.32 (CO), 197.68 (C-3). Anal. Calcd for C<sub>22</sub>H<sub>24</sub>N<sub>2</sub>O<sub>6</sub>: C 64.07, H 5.86, N 6.79. Found: C 64.23, H 6.07, N 6.52.

Ethyl 4(*R*,*S*)-[N-(benzyloxycarbonyl)-L-phenylalanyl]amino-3-oxopentanoate (10).– Yield: 64% (mixture of 4*S* and 4*R* diastereoisomersin 10:1 ratio). Eluent: EtOAc/hexane (1:2). 4*S* isomer: White solid: mp 129-131°C. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  1.27 (t, 3H, CH<sub>3</sub> OEt, J = 7.2), 1.28 (d, 3H, 5-H, J= 7.1), 3.08 (m, 2H,  $\beta$ -CH<sub>2</sub> Phe), 3.41 (s, 2H, H-2), 4.17 (q, 2H, CH<sub>2</sub> Et, J = 7.2), 4. 45 (m 1H, H-4), 4.58 (m, 1H,  $\alpha$ -Phe), 5.07 (s, 2H, CH<sub>2</sub> Z), 5.34 (d, 1H,  $\alpha$ -NH Phe, J = 7.6), 6.65 (d, 1H, 4-NH, J = 7.5), 7.16-7.38 (m, 10H, C<sub>6</sub>H<sub>5</sub> Phe and Z). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): 14.06 (CH<sub>3</sub> Et), 16.67 (C-5), 38.34 (β-CH<sub>2</sub> Phe), 45.71 (C-2), 54.25 (C-4), 56.12 (α-CH Phe), 61.62 (CH<sub>2</sub> OEt), 67.15 (CH<sub>2</sub> Z), 127.19, 128.06, 128.25, 128.54, 128.76 and 129.26 (CH Ar), 135.99 (C Ar), 155.83, 166.67 and 170.56 (CO), 201.15 (C-3). Anal. Calcd for

C<sub>24</sub>H<sub>28</sub>N<sub>2</sub>O<sub>6</sub>: C 65.44, H 6.41, N 6.36. Found: C 64.30, H 6.27, N 6.05. *4R* isomer: White foam. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 1.17 (d, 3H, 5-H, J = 7.1), 1.25 (t, 3H, CH<sub>3</sub> OEt, J = 7.1), 3.06 (m, 2H, β-CH<sub>2</sub> Phe), 3.47 (m, 2H, H-2), 4.16 (q, 2H, CH<sub>2</sub> Et, J = 7.1), 4. 43 (m 1H, H-4), 4.58 (m, 1H, α-Phe), 5.08 (s, 2H, CH<sub>2</sub> Z), 5.42 (d, 1H, α-NH Phe, J = 7.3), 6.44 (d, 1H, 4-NH, J = 7.1), 7.16-7.35 (m, 10H, C<sub>6</sub>H<sub>5</sub> Phe and Z).

## General procedures for the synthesis of 2-oxopiperazine derivatives

Method A.- A solution of the corresponding  $\beta$ -ketoester (2 mmol) in MeOH or MeOD (50 mL) was hydrogenated at 45°C and 45 psi of pressure for 2-3 days, using 10% Pd-C as catalyst. After filtration of the catalyst, the solvent was evaporated and the resulting residue was purified on a silica gel column, as specified. Method B.- A solution of the corresponding  $\beta$ -ketoester (2 mmol) in MeOH or MeOD (50 mL) was hydrogenated at r.t. and 15 psi of pressure for 2 h, using 10% Pd-C as catalyst. The catalyst was filtered and ZnCl<sub>2</sub> (0.14 g, 1 mmol) and NaBH<sub>3</sub>CN or NaBD<sub>3</sub>CN (0.38 g, 6 mmol) were added to the filtrate. After stirring for 3 h at r.t., the solvent was evaporated to dryness. The residue was extracted with EtOAc (50 mL) and washed with H<sub>2</sub>O. The organic layer was dried over Na<sub>2</sub>SO<sub>4</sub> and, after evaporation, the residue was purified on a silica gel column as specified in each case.

**5(R,S)-(Ethoxycarbonyl)methyl-2-oxopiperazine** (6).– Yield: 96% (from 2, method A) and 20% (from 2, method B). Eluent: CH<sub>2</sub>Cl<sub>2</sub>/MeOH (30:1). Syrup. Anal. Calcd for C<sub>8</sub>H<sub>14</sub>N<sub>2</sub>O<sub>3</sub>: C 51.60 H 7.58, N 15.04. Found: C 51.58, H 7.69, N 14.82.

(3S,5R) 5-(Ethoxycarbonyl)methyl-3-methyl-2-oxopiperazine (7a).– Yield: 56% (from 3, method A) and 64% (from 3, method B). Eluent: CH<sub>2</sub>Cl<sub>2</sub>/MeOH (30:1). Syrup. Anal. Calcd for C<sub>9</sub>H<sub>16</sub>N<sub>2</sub>O<sub>3</sub>: C 53.99 H 8.05, N 13.99. Found: C 54.13, H 7.85, N 14.06.

(3S,5S) 5-(Ethoxycarbonyl)methyl-3-methyl-2-oxopiperazine (7b). – Yield: 33% (from 3, method A) and 16% (from 3, method B). Eluent: CH<sub>2</sub>Cl<sub>2</sub>/MeOH (30:1). Syrup. Anal. Calcd for C<sub>9</sub>H<sub>16</sub>N<sub>2</sub>O<sub>3</sub>: C 53.99 H 8.05, N 13.99. Found: C 53.77, H 7.91, N 13.64.

(3S,5R) 3-Benzyl-5-(ethoxycarbonyl)methyl-2-oxopiperazine (8a).– Yield: 69% (from 4, method A) and 65% (from 4, method B). Eluent: acetone/hexane (2:1). Syrup. HPLC:  $t_R$  10.24 min (mobile phase A/B 10:90). Anal. Calcd for  $C_{15}H_{20}N_2O_3$ : C 65.20 H 7.29, N 10.14. Found: C 65.08, H 6.95, N 10.00.

(3S,5S) 3-Benzyl-5-(ethoxycarbonyl)methyl-2-oxopiperazine (8b).– Yield: 22% (from 4, method A) and 12% (from 4, method B). Eluent: acetone/hexane (2:1). Syrup. HPLC:  $t_R$  14.50 min (mobile phase A/B 10:90). Anal. Calcd for  $C_{15}H_{20}N_2O_3$ : C 65.20 H 7.29, N 10.14. Found: C 65.11, H 7.25, N 10.03.

(3S,5R) 3-Benzyl-5-(methoxycarbonyl)methyl-2-oxopiperazine (9a).– Yield: 76% (from 5, method A) and 54% (from 5, method B). Eluent: CH<sub>2</sub>Cl<sub>2</sub>/MeOH (30:1). Syrup. HPLC: t<sub>R</sub> 6.94 min (mobile phase A/B 8:92). Anal. Calcd for  $C_{14}H_{18}N_2O_3$ : C 64.11, H 6.92, N 10.68 Found: C 64.22, H 6.59, N 10.38.

(35,55) 3-Benzyl-5-(methoxycarbonyl)methyl-2-oxopiperazine (9b).– Yield: 21% (from 5, method A) and 11% (from 5, method B). Eluent: CH<sub>2</sub>Cl<sub>2</sub>/MeOH (30:1). Syrup. HPLC:  $t_R$  9.87 min (mobile phase A/B 8:92). Anal. Calcd for C<sub>14</sub>H<sub>18</sub>N<sub>2</sub>O<sub>3</sub>: C 64.11, H 6.92, N 10.68 Found: C 63.94, H 7.15, N 10.55.

(1*S*,3*S*,5*R*) 3-Benzyl-5-(ethoxycarbonyl)methyl-1-methyl-2-oxopiperazine (11a).– Yield: 35% (from 10, method B). Eluent: CH<sub>2</sub>Cl<sub>2</sub>/MeOH (30:1). Syrup. HPLC:  $t_R$  6.26 min (mobile phase A/B 15:85). Anal. Calcd for C<sub>16</sub>H<sub>22</sub>N<sub>2</sub>O<sub>3</sub>: C 66.19, H 7.64, N 9.65 Found: C 66.06, H 7.71, N 9.28.

(15,35,55) 3-Benzyl-5-(ethoxycarbonyl)methyl-1-methyl-2-oxopiperazine (11b).– Yield: 2% (from 10, method B). Eluent: CH<sub>2</sub>Cl<sub>2</sub>/MeOH (30:1). Syrup. HPLC:  $t_R$  9.83 min (mobile phase A/B 15:85). Anal. Calcd for C<sub>16</sub> H<sub>22</sub>N<sub>2</sub>O<sub>3</sub>: C 66.19, H 7.64, N 9.65 Found: C 65.96, H 7.44, N 9.37.

(1*R*,3*S*,5*R*) 3-Benzyl-5-(ethoxycarbonyl)methyl-1-methyl-2-oxopiperazine (11c).–Yield: 7% (from 10, method B). Eluent: CH<sub>2</sub>Cl<sub>2</sub>/MeOH (30:1). Syrup. HPLC:  $t_R$  4.93 min (mobile phase A/B 15:85). Anal. Calcd for C<sub>16</sub>H<sub>22</sub>N<sub>2</sub>O<sub>3</sub>: C 66.19, H 7.64, N 9.65 Found: C 66.13, H 7.59, N 9.55.

Compd.	1–H	3-Н	5-H	6-H	5-CH2	$\mathbb{R}^1$	$\mathbb{R}^2$	<b>R</b> <sup>3</sup>
6	7.28 (br s)	3.58 (s)	3.36 (m)	3.36 (m) 3.18 (m)	2.50 (m)	_	-	$\begin{array}{ c c c } 4.19 (q) \\ 1.30 (t) \\ (1 = 7 1) \end{array}$
7a	7.44 (br s)	3.39 (m)	3.22 (m)	3.11 (m) 2.96 (m)	2.92 (m)	1.17 (d) (5.8)	-	$\begin{array}{c} (3 = 7.12) \\ 3.98 (q) \\ 1.09 (t) \\ (J = 7.2) \end{array}$
7Ъ	7.26 (br s)	3.59 (m)	3.51 (m)	3.36 (m) 3.09 (m)	2.46 (m)	1.38 (d) (6.7)	-	$\begin{array}{c} 4.12 (q) \\ 1.23 (t) \\ (I = 7.1) \end{array}$
8a	7.48 (br s)	3.68 (dd) (9.7, 3.1)	3.24 (m)	3.24 (m) 3.03 (m)	2.35 (m)	3.43 (dd) (13.6, 3.1) 2.79 (dd) (13.6, 9.7)	-	$\begin{array}{c} 4.01 \text{ (q)} \\ 1.10 \text{ (t)} \\ \text{(J = 7.1)} \end{array}$
8b	7.01 (br s)	3.72 (dd) (10.6, 3.2)	3.55 (m)	3.27 (m) 3.12 (m)	2.41 (m)	3.24 (dd) (13.5, 3.2) 2.95 (dd) (13.5, 10.6)	-	4.08 (q) 1.19 (t) (J = 7.1)
9a	7.19 (br s)	3.66 (dd) (9.8, 3.3)	3.25 (m)	3.25 (m) 3.03 (m)	2.36 (m)	3.42 (dd) (13.8, 3.3) 2.80 (dd) (13.8, 9.8)	_	3.55 (s)
9Ъ	6.85 (br s)	3.72 (dd) (10.6, 3.1)	3.57 (m)	3.37 (m) 3.12 (m)	2.42(m)	3.263 (dd) (13.8, 3.1) 2.97 (dd) (13.8, 10.6)		3.63 (s)
11a <sup>a</sup>	6.70 (br s)	3.70 (dd) (8.4, 3.3)	3.43 (m)	3.43 (m) (J <sub>5.6</sub> =4.0)	2.28 (m)	3.30 (dd) (13.7, 3.3) 2.94 (dd) (13.7, 8.4)	0.95 (d) (5.7)	4.04 (q) 1.13 (t) (J = 7.2)
11b <sup>a</sup>	5.93 (br s)	3.71 (dd) (9.9, 3.3)	3.14 (m)	3.32 (m) (J <sub>5,6</sub> 7.0)	2.55 (dd) (15.4, 4.1) 2.28 (dd) (15.4, 8.8)	$\begin{array}{c} (13.7, 3.3) \\ 3.22 (dd) \\ (13.7, 3.3) \\ 2.99 (dd) \\ (13.7, 9.9) \end{array}$	1.14 (d) (6.4)	4.08 (q) 1.19 (t) (J = 7.2)
11c <sup>a</sup>	5.91 (br s)	3.67 (dd) (9.5, 3.3)	2.83 (m)	3.46 (m) (J <sub>5,6</sub> 9.5)	2.53 (dd) (15.7, 2.2) 2.28 (dd) (15.7, 9.9)	3.46 (dd) (13.8, 3.3) 2.83 (m)	1.12 (d) (5.2)	4.01 (q) 1.09 (t) (J = 7.1)

Table 2.- Significant <sup>1</sup>H-NMR Chemical Shifts (δ, ppm) and Coupling Constants (Hz) of Piperazine Derivatives (300 MHz, CDCl<sub>3</sub>)

<sup>a</sup> Registered at 500 MHz.

Compd.	C-2	C-3	C-5	C-6	5-CH2	R <sup>1a</sup>	R <sup>2</sup>	R
6	171.08	48.91	48.70	46.85	37.38		-	14.05 60.83
7 <b>a</b>	172.74	54.05	48.88	47.00	37.36	17.63	-	13.80 60.54
7b	173.46	51.80	44.36	47.06	37.12	18.78		14.06 60.71
8a	171.34	59.88	49.32	47.39	37.89	37.94		13.97 60.78
8b	171.95	58.07	44.37	47.10	37.40	37.88		14.07 60.80
9a	171.44	59.66	49.13	48.18	37.52	37.73	-	51.71
9b	171.88	58.13	44.37	47.15	37.24	37.86	~	51.84
11a <sup>b</sup>	171.24	59.97	50.46	52.09	37.84	36.98	16.36	14.02 60.75
11b <sup>b</sup>	171.49	57.76	52.87	51.23	36.97	37.94	20.34	14.21 60.84
11c <sup>b</sup>	170.97	59.87	53.35	56.42	37.72	36.83	19.33	13.99 60.85

Table 3.- Significant <sup>13</sup>C-NMR Chemical Shifts of Piperazine Derivatives (75 MHz, CDCl<sub>3</sub>)

<sup>a</sup> CH<sub>3</sub> and CH<sub>2</sub> groups in Ala and Phe derivatives, respectively. <sup>b</sup> Registered at 125 MHz.

ACKNOWLEDGEMENTS. We thank the Comisión Interministerial de Ciencia y Tecnología (SAF 97 0030), Fundación La Caixa (97/022) and Comunidad de Madrid (08.5/0006/1998) for financial support.

#### REFERENCES AND NOTES

- 1. For reviews on peptidomimetics see: a) Rees, D.C. Current Med. Chem. 1994, 1, 145. b) Giannis, A.; Rübsam, F. Adv. Drug Res. 1997, 29, 1.
- a) Hirschmann, R.; Sprengeler, P. A.; Kawasaki, T.; Leahy, J. W.; Shakespeare, W. C.; Smith III, A. B. J. Am. Chem. Soc. 1992, 114, 9699. b)Hirschmann, R.; Nicolau, K. C.; Pietranico, S.; Leahy, E. M.; Salvino, J.; Arison, B.; Cichy, M. A.; Spoors, P. G.; Shakespeare, W. C.; Sprengeler, P. A.; Hamley, P.; Smith III, A. B.; Reisine, T.; Raynor, K.; Maechler, L.; Donaldson, C.; Vale, W.; Freidinger, R. M.; Cascieri, M. R.; Strader C. D. J. Am. Chem. Soc. 1993, 115, 12550.
- Olson, G. L.; Bolin, D. R.; Bonner, M. R.; Bös, M.; Cook, C. M.; Fry, D. C.; Graves, B. J.; Hatada, M.; Hill, D. E.; Kahn, M.; Madison, V. S.; Rusiecki, V. K.; Sarabu, R.; Sepinwall, J.; Vincent, G. P.; Voss, M. E. J. Med. Chem. 1993, 36, 3039.
- a) Zydowsky, T.M.; Dellaria, J.F.; Nellans, H.N. J. Org. Chem. 1988, 53, 5607. b) Wolf, J.-P.; Rapoport, H. J. Org. Chem. 1989, 54, 3164.
- For reviews on peptide conformation mimetics see: a) Holzemann, G. Kontakte (Darmstadt) 1991, 1, 3, and 2, 55. b) M. Kahn, Synlett 1993, 821. c) R. M. J. Liskamp, Recl. Trav. Chim. Pays-Bas 1994, 113, 1.
   d) Hanessian, S.; McNaughton-Smith, G.; Lombart, H.-G.; Lubell, W.D. Tetrahedron 1997, 53, 12789.

- a) Sato, K.; Nagai, U. J. Chem. Soc., Perkin Trans. 1 1986, 1231 b) Genin, M. J.; Gleason, W. B.; Johnson, R. L. J. Org. Chem. 1993, 58, 860 c) Lombart, H.-G.; Lubell, W. D. J. Org. Chem., 1994, 59, 6147 d) Hanessian, S.; Ronan, B.; Laoui, A. Bioorg. Med. Chem. Lett. 1994, 4, 1397 e) Hanessian, S.; McNaughton-Smith, G. Bioorg. Med. Chem. Lett. 1996, 6, 1567 f) Slomczynskaya, U.; Chalmer, D. K.; Cornille, F.; Smythe, M. L.; Beusen, D. D.; Moeller, K. D.; Marshall, G. R. J. Org. Chem. 1996, 61, 1198 g) Nadin, A.; Derrer, S.; McGeary, R.P.; Goodman, J.M.; Raithby, P.R.; Holmes, A.B. J. Am. Chem. Soc. 1995, 117, 9768.
- a) Martín-Martínez, M.; García-López, M. T.; Herranz, R.; González-Muñiz, R. *Tetrahedron* 1995, 51, 10361.
   b) Andreu, D.; Ruiz, S.; Carreño, C.; Alsina, J.; Albericio, F.; Jiménez, M.A.; De la Figuera, N.; Herranz, R.; García-López, M.T.; González-Muñiz, R. J. Am. Chem. Soc. 1997, 119, 10579.
- a) Yu, K.-L.; Rajakumar, G.; Srivastava, L.K.; Mishra, R.K.; Jonhson, R.L. J. Med. Chem. 1988, 31, 1430. b) Shiosaki, K.; Craig, R.; Lin, C.W.; Barrett, R.W.; Miller, T.; Witte, D.; Wolfram, C.A.W.; Nadzan, A.M. Peptides, Chemistry and Biology (Ed.: J. Rivier), Escom, Leiden, 1990, p. 978. c) Logan, M.E.; Goswami, R.; Tomczuk, B.E.; Venepalli, B.R. Annu. Rep. Med. Chem. 1991, 26, 43. d) Flynn, D.L.; Villamil, C.I.; Becker, D.P.; Gullikson, G.W.; Moummi, C.; Yang, D.-C. Bioorg. Med. Chem. Lett. 1992, 2, 1251 e) Howbert, J.J.; Lobb, K.L.; Britton, T.C.; Mason, N.R.; Bruns, Bioorg. Med. Chem. Lett. 1993, 3, 875 f) Austel, V.; Himmelsbach, F.; Müller, T. Drugs Fut. 1994, 19, 757. g) Batt, A.R.; Kendrick, D.A.; Mathews, E.; Rooker, D.P.; Ryder, H.; Semple, G., Szelke, M. Bioorg. Med. Chem. Lett. 1994, 4, 867.
- a) Fredinger, R. M.; Veber, D. F.; Perlow, D. S.; Broods, J. R.; Saperstein, R. Science 1980, 210, 656 b) Aebi, J.D.; Guillaume, D.; Dunlap, B.E.; Rich, D.H. J. Med. Chem. 1988, 31, 1805.
- a) Thaisrivongs, S.; Pals, D.T.; Turner, S.R.; Kroll, L.T. J. Med. Chem. 1988, 31, 1369. b) Shuman, R.T.; Rothenberger, R.B.; Campbell, C.S.; Smith, G.F.; Gifford-Moore, D.S.; Gesellchen, P.D. J. Med. Chem. 1993, 36, 314.
- a) M.L. Suárez-Gea, M.T. García-López, R. González-Muñiz, S. Herrero y R. Herranz. Tetrahedron Lett., 1996, 37, 2083-2084. b) Bravo, A.; Gómez-Monterrey, I.; González-Muñiz, R.; García-López, M.T. J. Chem. Soc. Perkin Trans. 1 1991, 3117.
- 12. Farmer, P.S.; Ariëns, E.J. Trends Pharmacol. Sci. 1982, 3, 362.
- a) Brooks, D.W.; Lu, L.D.-L.; Masamune, S. Angew. Chem. Int. Ed. Engl. 1979, 18, 72-74. b) Aubry, N.; Plante, R.; Déziel, R. Tetrahedron Lett. 1990, 31, 6311-6312.
- a) Domínguez, M.J.; García-López, M.T.; Herranz, R.; Martín-Martínez, M.; González-Muñiz, R. J. Chem. Soc., Perkin Trans. 1, 1995, 2839-2843.b) Martín-Martínez M., Bartolomé-Nebreda J.M., Gómez-Monterrey I., González-Muñiz R., García-López M.T., Ballaz S., Barber A., Fortuño A., Del Río J., Herranz R., J. Med. Chem. 1997, 40, 3402-3407.
- 15. Paul, R.; Anderson, G.W. J. Am. Chem. Soc. 1960, 82, 4596-4600.
- 16. Brooks, D.W.; Lu, L.D.-L.; Masamune, S. Angew. Chem. Int. Ed. Engl. 1979, 18, 72-74.