Article

First Synthesis and Structural Elucidation of (-)-Presphaerene¹

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The first total synthesis of (–)-presphaerene (**1**) was achieved from (*R*)-glyceraldehyde **9** in 19 steps, demonstrating the novel "folding and allylic strain-controlled" intramolecular ester enolate $S_N 2'$ alkylation strategy could be extended to the stereoselective synthesis of cyclopentanoid natural products. The present study also established the relative and absolute stereochemistry of **1**, and the absolute structures of co-occurring sphaeroanes from the red alga *Sphaerococcus coronopifolius*.

Introduction

The sphaeroanes with a skeleton of 2,7-cycloneodolabellane such as (–)-presphaerene (1), (+)-presphaerol (2), (+)-isosphaerodiene 1 (3), and (+)-isosphaerodiene 2 (4) were isolated from the red alga *Sphaerococcus coronopifolius* by Fattorusso and co-workers (Figure 1).² The structures of these natural products were determined by a combination of chemical correlation, NMR spectroscopy, and X-ray crystallography except for their absolute configurations and the C-7 stereochemistry of 1. In detail, the relative configuration of **3** was resolved by X-ray crystallography. The chemical transformation of **2** into a mixture of **3** and **4** by treatment with acetyl chloride in refluxing xylene, coupled with NMR spectroscopy, led to the tentative assignment of **2**. Aromatization of **2** with SeO₂ also produced **1**.

Previously, we reported the application of the novel and highly stereoselective "folding and allylic strain-controlled" intramolecular enolate S_N2' alkylation methodology to the synthesis of cyclohexanoid natural products.³ Encouraged by the report that some of the sphaeroanes are biologically active,⁴ we set out to examine the feasibility of the internal S_N2' alkylation for the construction of highly functionalized cyclopentane systems with a particular emphasis on the stereochemical outcome. Described herein is the first total synthesis and complete structural elucidation of **1** utilizing a folding and allylic



(+)-isosphaerodiene 1 (3) (+)-isosphaerodiene 2 (4)

FIGURE 1. Structures of 1-4.

strain-controlled $^{\scriptscriptstyle 5}$ intramolecular ester enolate $S_N 2'$ alkylation and an intramolecular Friedel–Crafts acylation as key steps.

Results and Discussion

Our retrosynthetic analysis for **1** (Scheme 1) called for the intermediacy of tricyclic ketone **5** and highly functionalized cyclopentanecarboxylate **7** as key intermediates, which could be constructed by intramolecular

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⁽¹⁾ Taken in part from the doctoral thesis of J. Lee, Seoul National University, 2001.

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SCHEME 1. **Retrosynthetic Analysis of 1**



Friedel-Crafts acylation and S_N2' alkylation from acyclic precursors 6 and 8, respectively.

The requisite intramolecular S_N2' alkylation substrate **8** was prepared from readily available (*R*)-glyceraldehyde **9**⁶ by a straightforward manner as depicted in Scheme 2. (R)-Glyceraldehyde 9 was transformed to an easily separable mixture of Z-olefin 11 and E-olefin 12 (11:1) by a Wittig reaction with *m*-tolylphosphonium bromide 10^7 in a total 90% yield. Z-Olefin 11 could be cleanly isomerized to *E*-olefin 12 by treatment with thiophenol in the presence of AIBN in refluxing benzene according to the protocol described by Schwarz⁸ in 76% yield. It is worthwhile to mention the stereoselective access to both Z- and E-olefin is an important factor for the determination of the absolute stereochemistry of 1 in our synthetic scheme. Removal of the acetonide group of olefin **12**, followed by a selective monobenzovlation⁹ of the resulting diol 13 (>95% ee),¹⁰ furnished monobenzoate 14 in 88% yield for the two steps. Subjection of allylic alcohol 14 to Johnson ortho ester Claisen rearrangement conditions¹¹ and subsequent deprotection of the benzoyl protecting group of the resulting diester 15 by a transesterification led to the formation of hydroxy ester 16 in 85% overall yield. Catalytic hydrogenation of olefin 16 using PtO₂ and PCC oxidation of the resulting alcohol 17 produced the corresponding aldehyde 18 in 74% yield for the two steps. The aldehyde 18 was elaborated to the cyclization precursor 8 by a convenient three-step se-

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^a Reagents and conditions: (i) t-BuOK, THF, -78 to -30 °C, 90%; (ii) PhSH, AIBN, benzene, reflux, 3 h, 76%; (iii) 60% aqueous AcOH, rt, 6 h, 97%; (iv) BzCl, Et₃N, CH₂Cl₂, -40 °C, 4 h, 91%; (v) CH₃CH₂CH(OEt) ₃, phenol, toluene, reflux, 20 h, 87%; (vi) NaOEt, EtOH, rt, 7 h, 98%; (vii) PtO2, H2, EtOH, rt, 7 h, 88%; (viii) PCC, NaOAc, CH₂Cl₂, 0 °C, 2 h, 84%; (ix) Ph₃P=C(CH₃)CHO, toluene, reflux, 8 h; (x) NaBH₄, EtOH, 0 °C, 0.5 h, 78% for two steps; (xi) CBr₄, Ph₃P, CH₂Cl₂, 0 °C, 2 h, 89%.

quence. Wittig reaction of aldehyde 18 with 2-(triphenylphosphoranylidene)propionaldehyde, NaBH4 reduction of the corresponding α,β -unsaturated aldehyde **19**, and finally bromination of the resulting allylic alcohol 20 by the protocol described by Hooz¹² afforded the desired allylic bromide 8 in 69% overall yield over the three steps.

Allylic bromide **8** underwent a smooth $S_N 2'$ cyclization upon treatment with LHMDS in THF for 22 h at room temperature to afford a highly functionalized cyclopentanecarboxylate, 7, as a major component along with diastereomeric 7-iso and 7-cis in a 9.9:3.3:1 ratio (capillary GLC analysis) in a total 86% yield. These stereoisomers could be separated by preparative HPLC, and the relative stereochemistry of the cyclized products was established by NOE experiments as shown in Scheme 3. In addition, the second major isomer, 7-iso, could be converted to the desired isomer 7 by a straightforward three-step sequence (ozonolysis, epimerization, Wittig reaction) as described in Scheme 3, which reinforces our structural assignment.

The observed stereoselectivity can best be rationalized by considering that the reaction proceeds via chairlike "double H-eclipsed" transition-state geometry 21 where the nucleophilic ester enolate moiety and electrophilic allylic bromide assume a "H-eclipsed" conformation with

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SCHEME 3



SCHEME 4^a



^a Reagents and conditions: (i) DIBALH, toluene, -78 to -30 °C, 93%; (ii) PCC, NaOAc, 0 °C, 2.5 h, 77%; (iii) TMSCH₂CO₂Bn, LDA, THF, -25 °C, 2 h; (iv) PtO₂, H₂, EtOAc, rt, overnight, 92% for two steps; (v) PPA, 90 °C, overnight, 89%.

the bulky *m*-tolyl appendage in an equatorial position, affording a cyclopentanecarboxylate with three contiguous stereogenic centers.³

Treatment of the desired cyclopentanecarboxylate **7** with DIBALH led to the formation of primary alcohol **23** (Scheme 4), which was further oxidized with PCC to the corresponding aldehyde **24** (72% overall yield). Peterson olefination¹³ of aldehyde **24** with benzyl (trimethylsilyl)-acetate¹⁴ yielded α,β -unsaturated benzyl ester **25**, and then simultaneous catalytic hydrogenation of olefin and hydrogenolysis of benzyl ester with PtO₂ afforded carboxylic acid **6** in 92% overall yield. It is quite interesting to note that the Peterson olefination yielded *E*-olefinic ester in an exclusive manner. Intramolecular Friedel–Crafts acylation^{13a,15} of carboxylic acid **6** with PPA gave the desired seven-membered tricyclic ketone **5** in high yield (89%).

To establish unambiguously the configuration of the secondary methyl group at C-7, tricyclic ketone **5** was first treated with methyllithium and SOCl₂ to provide a 5:1 mixture of *endo*-olefin **26** and *exo*-olefin **27** as depicted in Scheme 5 .¹⁶ Examination of the most stable conforma-

tion of endo-olefin 26, which was generated by a systematic Monte Carlo conformational search and subsequent energy minimization using MM2 calculation, revealed the β -face of the olefin is distinctively less hindered as shown in Figure 2.17 As we expected, catalytic hydrogenation of endo-olefin 26 with 10% Pd/C produced exclusively a quantitative yield of the 7- α -methyl isomer, which turned out to be 7-*epi*-presphaerene ($\mathbf{1}'$). However, use of PtO₂ as catalyst instead of Pd/C for the hydrogenation of endoolefin **26** yielded a small amount (10%) of **1**, the desired 7- β -methyl isomer, in addition to **1**' as a major product (90%). These two isomeric compounds could be separated by column chromatography on AgNO₃-impregnated silica gel, and their structure was fully determined by the analyses of their NOE, DEPT, 1H-1H COSY, and ¹H⁻¹³C COSY spectra. Moreover, the spectrum of the minor 7- β -methyl isomer was in agreement with the spectral data of natural 1 reported in the literature.^{2c,e}

Since it was not so obvious from the inspection of the most stable conformation of *exo*-olefin **27** (Figure 2) which side of the *exo*-methylene double bond is more hindered,¹⁷ we hoped that we had a better chance of obtaining the desired 7- β -methyl isomer from *exo*-olefin **27** as a major product. The catalytic hydrogenation of *exo*-olefin **27**, also prepared from tricyclic ketone **5** by a Wittig reaction in high yield, with 10% Pd/C produced only **1**' in almost quantitative yield. We reasoned that the reduction with Pd catalyst might proceed through *endo*-olefin **26** via in situ isomerization.¹⁸

Taking advantage of these experimental findings, *exo*olefin **27** was hydrogenated in ethanol using PtO_2 as

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(-)-presphaerene (1) 7-epi-presphaerene (1')

Catalyst	Substrate	Solvent	Products (1 : 1')
10% Pd/C PtO ₂	26 26	EtOH EtOH	0 : 100 10 : 90
10% Pd/C	27	EtOH	0:100
PtO ₂	27	<i>n</i> -BuOH	75: 25
PtO ₂	27	<i>n</i> -PrOH	75: 25
PtO ₂	27	<i>i-</i> PrOH	71: 29
PtO ₂	27	EtOH	75: 25
PtO ₂	27	MeOH	57: 43
PtO ₂	27	EtOAC	60: 40
PtO ₂	27	hexane	57: 43
Pt/C	27	EtOH	50: 50
Pt/Al ₂ O ₃	27	EtOH	50: 50

catalyst, which is known to be less prone to isomerization,¹⁸ to give rise to a 3:1 mixture of **1** and **1'**, to our delight (Scheme 5). Use of other solvents such as MeOH, EtOAc, and *n*-hexane exhibited somewhat decreased stereoselectivity than ethanol. It is interesting to note that use of Pt/C or Pt/Al₂O₃ for the hydrogenation of *exo*olefin **27** afforded a 1:1 mixture of **1** and **1'**.

In summary, the first total synthesis and structure determination of 1 have been accomplished from (R)glyceraldehyde 9 employing a folding and doubly allylic strain-controlled intramolecular ester enolate S_N2' alkylation and an intramolecular Friedel-Crafts acylation as key steps. The present synthesis also established the absolute structures of co-occurring sphaeroanes from the red alga *S. coronopifolius* such as 2-4 as a result. More importantly, we have demonstrated that the internal $S_N 2'$ methodology is a viable method for the stereoselective construction of highly functionalized cyclopentanoid natural products, though the observed stereoselectivity seems to be slightly inferior to that of the corresponding sixmembered cases. During the course of the study we introduced benzyl (trimethylsilyl)acetate¹⁴ for the purpose of introducing a two-carbon acetic acid unit to a very hindered position and observed a very interesting interplay of catalyst and molecular geometry in the catalytic



FIGURE 2. The most stable conformations of *endo*-olefin **26** and *exo*-olefin **27** by a systematic Monte Carlo conformational search and subsequent energy minimization using MM2 calculation.¹⁷

hydrogenation. Currently, efforts are being made to apply this internal $S_N 2'$ alkylation strategy to the syntheses of various biologically active natural products in our laboratories.

Experimental Section

(Z/E,4S)-2,2-Dimethyl-4-(2-m-tolylvinyl)[1,3]dioxolane (11 and 12). To a mixture of *m*-tolylphosphonium bromide 10 (1374 mg, 3.07 mmol) and anhydrous THF (5.1 mL) was added a 1.0 M solution of t-BuOK (2.76 mL) in THF at 0 °C. After 1 h at ambient temperature, the mixture was cooled to -78 °C, and a solution of (*R*)-glyceraldehyde **9** (180.0 mg, 1.38 mmol) in THF (1.5 mL) was added. The mixture was stirred and slowly warmed to -30 °C over 3 h. The reaction mixture was quenched with a few drops of saturated aqueous NH₄Cl solution and filtered through a pad of silica gel. The filtrate was concentrated in vacuo, and the resulting residue was purified by column chromatography on silica gel (hexane/ EtOAc, 15:1) to afford Z-olefin 11 (247.0 mg) and E-olefin 12 (22.5 mg) in a total 90% yield. Data for Z-olefin 11: ¹H NMR (500 MHz, CDCl₃) δ 7.23 (t, J = 7.5 Hz, 1H), 7.09 (d, J = 7.5Hz, 1H), 7.07 (s, 1H), 7.06 (d, J = 7.8 Hz, 1H), 6.68 (d, J =11.6 Hz, 1H), 5.68 (dd, J = 11.5, 8.9 Hz, 1H), 4.91 (dddd, J =8.8, 7.6, 6.3, 1.2 Hz, 1H), 4.14 (dd, J = 8.1, 6.1 Hz), 3.66 (t, J = 7.9 Hz, 1H), 2.35 (s, 3H), 1.47 (s, 3H), 1.38 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) & 137.9, 136.1, 134.1, 129.4, 128.9, 128.2, 128.1, 125.7, 109.3, 72.4, 69.7, 26.8, 25.9, 21.4; IR (neat) 1604, 1454, 1060 cm⁻¹; $[\alpha]^{20}_{D} = -37.3$ (*c* 3.04, CHCl₃); HRMS (EI) m/z calcd for C₁₄H₁₈O₂ (M⁺) 218.1307, found 218.1313. Data for *E*-olefin **12**: ¹H NMR (500 MHz, CDCl₃) δ 7.20-7.18 (m, 3H), 7.06 (br d, J = 6.6 Hz, 1H), 6.63 (dd, J = 15.8, 0.6 Hz, 1H), 6.14 (dd, J = 15.8, 7.6 Hz, 1H), 4.67 (dddd, J = 8.5, 7.6, 6.4, 1.1 Hz, 1H), 4.15 (dd, J = 8.1, 6.2 Hz, 1H), 3.67 (t, J = 8.0 Hz, 1H), 2.33 (s, 3H), 1.47 (d, J = 0.5 Hz, 3H), 1.43 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 138.1, 136.2, 133.4, 128.7, 128.4, 127.3, 126.5, 123.8, 109.4, 77.2, 69.5, 26.7, 25.9, 21.3; IR (neat) 1604, 1454, 1059 cm⁻¹; $[\alpha]^{20}_{D} = +58.5$ (*c* 2.02, CHCl₃); HRMS (EI) m/z calcd for C₁₄H₁₈O₂ (M⁺) 218.1307, found 218.1305.

E-Olefin 12 from Z-Olefin 11. To a solution of Z-olefin **11** (2094 mg, 9.59 mmol) in benzene (9.6 mL) were added thiophenol (0.49 mL, 4.77 mmol) and AIBN (236 mg, 1.44 mmol). The reaction mixture was refluxed for 3 h. Removal of the solvent and column chromatography of the resulting residue on silica gel (hexane/EtOAc, 30:1) gave *E*-olefin **12** (1582 mg) in 76% yield.

(2.5)-4-*m*-Tolylbut-3-ene-1,2-diol (13). A mixture of *E*olefin 12 (645.7 mg, 2.96 mmol) and 60% aqueous acetic acid (5.9 mL) was stirred for 6 h at room temperature. After neutralization with saturated aqueous NaOH solution, the mixture was extracted with EtOAc (30 mL × 5). The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, and concentrated in vacuo. The residue was purified by column chromatography on silica gel (hexane/EtOAc/MeOH, 8:16:1) to give diol 13 (510.0 mg) in 97% yield: ¹H NMR (500 MHz, CDCl₃) δ 7.22–7.17 (m, 3H), 7.07 (d, *J* = 7.1 Hz, 1H),

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6.65 (d, J = 16.0 Hz, 1H), 6.18 (dd, J = 16.0, 6.4 Hz, 1H), 4.41 (dt, J = 6.5, 3.5 Hz, 1H), 3.74 (dd, J = 11.2, 3.5 Hz, 1H), 3.59 (dd, J = 11.2, 7.4 Hz, 1H), 2.45 (br s, 1H), 2.34 (s, 3H), 2.23 (br s, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 137.9, 136.2, 131.7, 128.4, 128.3, 127.4, 127.2, 123.5, 73.2, 66.4, 21.2; IR (neat) 3389, 1074 cm⁻¹; [α]²⁰_D = +29.5 (*c* 1.91, CHCl₃); HRMS (EI) *m*/*z* calcd for C₁₁H₁₄O₂ (M⁺) 178.0994, found 178.0993.

Benzoic Acid (E,2S)-2-Hydroxy-4-m-tolylbut-3-enyl Ester (14). To a solution of diol 13 (245.0 mg, 1.38 mmol) in CH₂Cl₂ (6.9 mL) were added Et₃N (0.57 mL, 4.09 mmol) and benzoyl chloride (0.24 mL, 2.07 mmol) at $-78\ {\rm °C}.$ The mixture was stirred for 4 h at -40 °C and cooled to -78 °C. A 3% aqueous HCl solution (1.0 mL) was added to the mixture at -78 °C with vigorous agitation. After 1 h, the mixture was poured into brine and extracted with EtOAc (30 mL \times 4). The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, and concentrated at reduced pressure. The resulting residue was purified by column chromatography on silica gel (hexane/EtOAc, 2:1) to produce benzoate 14 (352.5 mg) in 91% yield: ¹H NMR (500 MHz, CDCl₃) δ 8.07 (d, J =8.1 Hz, 1H), 8.06 (d, J = 8.6 Hz, 1H), 7.59-7.55 (m, 1H), 7.45 (dd, J = 8.1, 7.5 Hz, 2H), 7.23-7.19 (m, 3H), 7.08 (dd, J = 6.9, 1.3 Hz, 1H), 6.74 (dd, J = 16.0, 1.2 Hz, 1H), 6.25 (dd, J = 15.9, 6.2 Hz, 1H), 4.72-4.68 (m, 1H), 4.50 (dd, J = 11.4, 3.7 Hz, 1H), 4.37 (dd, J = 11.4, 7.4 Hz, 1H), 2.34 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 166.7, 138.2, 136.2, 133.2, 132.6, 129.8, 129.7, 128.8, 128.5, 128.4, 127.3, 126.9, 123.8, 71.1, 68.5, 21.3; IR (neat) 3443, 1714, 1275 cm⁻¹; $[\alpha]^{20}_{D} = +3.4$ (*c* 0.79, CHCl₃); HRMS (EI) m/z calcd for C18H18O3 (M+) 282.1256, found 282.1247.

Benzoic Acid (E,4S)-5-Ethoxycarbonyl-4-m-tolylhex-2envl Ester (15). To a solution of benzoate 14 (352.5 mg, 1.25 mmol) in toluene (6.2 mL) were added triethylorthopropionate (2.5 mL, 12.4 mmol) and phenol (12 mg, 0.13 mmol) at room temperature. The reaction mixture was refluxed for 20 h and cooled to room temperature. The mixture was concentrated in vacuo, and the resulting residue was purified by column chromatography on silica gel (hexane/EtOAc, 15:1) to afford allylic benzoate 15 (399.4 mg) in 87% yield: ¹H NMR (500 MHz, CDCl₃) & 8.04-8.00 (m, 2H) 7.57-7.53 (m, 1H), [7.43 (t, J = 7.7 Hz) and 7.42 (t, J = 7.6 Hz), 2 H], [7.21 (t, J = 7.5 Hz) and 7.17 (t, J = 7.8 Hz), 1H], [7.04 (d, J = 7.2 Hz) and 7.01-6.98 (m), 3H], [6.03 (dd, J = 15.4, 8.6 Hz) and 5.96 (dd, J =15.3, 9.3 Hz), 1H], [5.79 (td, J = 15.3, 6.2 Hz) and 5.72 (td, J = 15.3, 6.3 Hz), 1H], [4.82-4.74 (m) and 4.73 (dd, J = 6.3, 1.0 m)Hz), 2H], [4.10 (q, J = 7.1 Hz) and 3.94–3.85 (m), 2H], [3.51 (t, J = 9.6 Hz) and 3.44 (t, J = 9.6 Hz), 1H], 2.84–2.77 (m, 1H), [2.34 (s) and 2.31 (s), 3H], [1.21 (t, J = 6.4 Hz) and 0.98 (t, J = 7.0 Hz), 3H], [1.22 (d, J = 5.8 Hz) and 0.97 (d, J = 7.0Hz), 3H]; ¹³C NMR (75 MHz, CDCl₃) δ 175.5, 175.0, 166.2, 141.7, 140.8, 138.3, 137.9, 136.7, 135.7, 132.9, 132.8, 130.2, 129.52, 129.50, 128.7, 128.6, 128.5, 128.29, 128.26, 128.23, 127.6, 127.4, 126.1, 125.0, 124.9, 124.6, 65.09, 65.07, 60.3, 60.0, 52.4, 52.3, 45.3, 44.9, 21.38, 21.35, 15.8, 15.6, 14.2, 13.8; IR (neat) 1725, 1271 cm⁻¹; HRMS (EI) *m*/*z* calcd for C₂₃H₂₆O₄ (M⁺) 366.1831, found 366.1845.

(E,3S)-6-Hydroxy-2-methyl-3-m-tolylhex-4-enoic Acid Ethyl Ester (16). Sodium ethoxide (1.82 mL, 1.0 M solution in ethanol) was added to a solution of allylic benzoate 15 (222 mg, 0.61 mmol) in ethanol (1.2 mL). The mixture was stirred for 7 h at room temperature under an argon atmosphere and neutralized with saturated aqueous NH₄Cl solution. The solvent was removed at reduced pressure, and the resulting residue was dissolved in EtOAc and washed with brine. The organic layer was dried over anhydrous Na₂SO₄ and concentrated in vacuo. The residue was purified by column chromatography on silica gel (hexane/EtOAc, 4:1) to afford allylic alcohol 16 (156 mg) in 98% yield: 1H NMR (500 MHz, CDCl₃) δ [7.20 (t, J = 7.5 Hz) and 7.16 (t, J = 7.8 Hz), 1H], [7.04 (d, J = 7.2 Hz and 7.00–6.97 (m), 3H], [5.87 (dd, J = 15.2, 8.6 Hz) and 5.81 (dd, J = 15.4, 9.1 Hz), 1H], [5.73 (td, J = 15.2, 5.3 Hz) and 5.66 (td, J = 15.3, 5.6 Hz), 1H], [4.18–4.11 (m) and 3.93–3.86 (m), 2H], [4.12 (t, J = 5.7 Hz) and 4.05 (t, J = 5.4 Hz, 2H], [3.46 (t, J = 9.5 Hz) and 3.40 (t, J = 9.7 Hz), 1H], 2.82–2.75 (m, 1H), [2.34 (s) and 2.31 (s), 3H], [1.34 (t, J = 6.0 Hz) and 1.28 (t, J = 6.1 Hz), 1H], [1.26 (t, J = 7.2 Hz) and 0.98 (t, J = 7.1 Hz), 3H], [1.21 (d, J = 6.9 Hz) and 0.96 (d, J = 6.9 Hz), 3H]; ¹³C NMR (75 MHz, CDCl₃) δ 175.7, 175.3, 142.1, 141.2, 138.2, 137.8, 133.3, 132.1, 131.3, 130.1, 128.7, 128.5, 128.4, 128.2, 127.4, 127.3, 124.9, 124.6, 63.13, 63.08, 60.3, 60.0, 52.3, 52.2, 45.4, 45.0, 21.4, 21.3, 15.8, 15.5, 14.2, 13.8; IR (neat) 3444, 1732 cm⁻¹; HRMS (EI) *m*/*z* calcd for C₁₆H₂₀O₂ (M⁺ – H₂O) 244.1463, found 244.1464.

(3S)-6-Hydroxy-2-methyl-3-m-tolylhexanoic Acid Ethyl Ester (17). To a solution of allylic alcohol 16 (264.0 mg, 1.01 mmol) in ethanol (2.0 mL) was added PtO₂ (5 mg) at room temperature. After 7 h at room temperature under a hydrogen atmosphere, the mixture was filtered through a pad of Celite. The filtrate was concentrated in vacuo, and the resulting residue was purified by column chromatography on silica gel (hexane/EtOAc, 5:1) to afford alcohol 17 (235.1 mg) in 88% yield: ¹H NMR (500 MHz, CDCl₃) δ [7.18 (t, J = 7.7 Hz) and 7.15 (t, J = 8.1 Hz), 1H], [7.02 (d, J = 7.4 Hz) and 6.99 (d, J= 7.6 Hz), 1H], 6.96–6.92 (m, 2H), [4.18 (q, J = 7.1 Hz) and 3.92-3.86 (m), 2H], [3.57 (t, J = 6.3 Hz) and 3.54 (t, J = 6.3Hz), 2H], [2.79 (ddd, J = 11.8, 8.4, 3.4 Hz) and 2.73 (dt, J = 10.4, 4.2 Hz), 1H], 2.68-2.60 (m, 1H), [2.33 (s) and 2.31 (s), 3H], [1.89-1.82 (m) and 1.68-1.59 (m), 2H], 1.39-1.25 (m, 3H), 1.29 (t, J = 7.1 Hz) and 0.99 (t, J = 7.1 Hz), 3H], [1.21 (d, J = 6.9 Hz) and 0.90 (d, J = 6.9 Hz), 3H]; ¹³C NMR (75 MHz, CDCl₃) δ 176.7, 175.6, 142.4, 141.9, 137.9, 137.5, 129.00, 128.95, 128.2, 128.0, 127.3, 127.1, 125.2, 125.1, 62.6, 62.4, 60.3, 59.9, 48.6, 48.3, 46.1, 46.0, 30.6, 30.5, 27.8, 21.4, 16.2, 14.8, 14.2, 13.8; IR (neat) 3437, 1731 cm⁻¹; HRMS (EI) m/z calcd for C₁₆H₂₄O₃ (M⁺) 264.1725, found 264.1725.

(3S)-2-Methyl-6-oxo-3-m-tolylhexanoic Acid Ethyl Ester (18). To a solution of alcohol 17 (236.7 mg, 0.90 mmol) in dry CH₂Cl₂ (9.0 mL) were added NaOAc (490 mg, 5.97 mmol), molecular sieves (4 Å, 400 mg), and PCC (429 mg, 1.99 mmol) at 0 °C. After 2 h at 0 °C, the mixture was diluted with ether. The mixture was stirred for 0.5 h and filtered through a pad of Florisil. The filtrate was concentrated at reduced pressure, and the resulting residue was purified by column chromatography on silica gel (hexane/EtOAc, 7:1) to afford aldehyde 18 (198.0 mg) in 84% yield: ¹H NMR (500 MHz, CDCl₃) δ [9.66 (t, J = 1.1 Hz) and 9.60 (t, J = 1.4 Hz), 1H], [7.19 (t, J = 7.5Hz) and 7.15 (t, J = 7.7 Hz), 1H], [7.04 (d, J = 7.5) and 7.01 (d, J = 7.5 Hz), 1H], 6.93-6.90 (m, 2H), [4.19 (q, J = 7.1 Hz) and 3.93-3.84 (m), 2H], 2.80-2.61 (m, 2H), [2.33 (s) and 2.31 (s), 3H], [2.28-2.13 (m) and 2.00-1.93 (m), 3H], 1.90-1.78 (m, 1H), [1.30 (t, J = 7.2 Hz) and 0.98 (t, J = 7.1 Hz), 3H], [1.25 (d, J = 6.9 Hz) and 0.91 (d, J = 6.8 Hz), 3H]; ¹³C NMR (75) MHz, CDCl₃) δ 176.1, 175.2, 141.4, 140.8, 138.2, 137.8, 128.9, 128.8, 128.4, 128.2, 127.6, 127.5, 125.2, 125.1, 60.4, 59.9, 48.3, 47.9, 46.0, 45.8, 42.0, 41.9, 26.8, 24.1, 21.4, 21.3, 16.3, 14.9, 14.1, 13.8; IR (neat) 2723, 1730 cm⁻¹; HRMS (EI) *m*/*z* calcd for C₁₆H₂₂O₃ (M⁺) 262.1569, found 262.1569.

(E,3S)-8-Hydroxy-2,7-dimethyl-3-m-tolyloct-6-enoic Acid Ethyl Ester (20). A mixture of 2-(triphenylphosphoranylidene)propionaldehyde (226 mg, 0.71 mmol), aldehyde 18 (155.2 mg, 0.59 mmol), and toluene (2.0 mL) was refluxed for 8 h. The mixture was filtered through a pad of silica gel, and the filtrate was concentrated to give crude enal 19. The crude product was dissolved in ethanol (2.0 mL), and NaBH₄ (45 mg, 1.19 mmol) was added at 0 °C. After 0.5 h at the same temperature, the mixture was quenched with saturated aqueous NH₄Cl, and ethanol was removed at reduced pressure. The resulting residue was dissolved in EtOAc and washed with brine. The organic layer was dried over anhydrous Na₂SO₄ and concentrated in vacuo. The resulting residue was purified by column chromatography on silica gel (hexane/EtOAc, 10:1) to afford allylic alcohol 20 (140.0 mg) in 78% yield from aldehyde 18: ¹H NMR (500 MHz, CDCl₃) δ [7.18 (t, J = 7.5 Hz) and 7.15 (t, J = 7.9 Hz), 1H], [7.02 (d, J = 7.5 Hz) and 6.99 (d, J = 7.9

Hz), 1H], 6.95–6.91 (m, 2H), [5.33 (t, J = 6.5 Hz) and 5.29 (t, J = 6.9 Hz), 1H], [4.17 (q, J = 6.9 Hz) and 3.95–3.86 (m), 4H], [2.78 (ddd, J = 11.3, 8.5, 2.9 Hz) and 2.72 (dt, J = 10.2, 4.3 Hz), 1H], 2.67–2.58 (m, 1H), [2.33 (s) and 2.31 (s), 3H], [1.87–1.78 (m) and 1.70–1.62 (m), 4H], [1.50 (s) and 1.48 (s), 3H], [1.29 (dt, J = 7.1, 0.5 Hz) and 0.99 (dt, J = 7.2, 0.5 Hz), 3H], [1.20 (d, J = 6.7 Hz) and 0.89 (d, J = 6.8 Hz), 3H]; ¹³C NMR (75 MHz, CDCl₃) δ 176.6, 175.6, 142.3, 141.8, 137.7, 137.3, 134.93, 134.91, 129.0, 128.1, 127.8, 127.1, 127.0, 125.4, 125.3, 125.2, 125.1, 68.5, 60.2, 59.8, 48.4, 48.2, 46.1, 45.9, 34.2, 31.4, 25.31, 25.26, 21.32, 21.27, 16.1, 14.7 14.1, 13.8, 13.4; IR (neat) 3443, 1731 cm⁻¹; HRMS (EI) *m*/*z* calcd for C₁₉H₂₈O₃ (M⁺) 286.1933, found 286.1937.

(E,3S)-8-Bromo-2,7-dimethyl-3-m-tolyloct-6-enoic Acid Ethyl Ester (8). To a solution of allylic alcohol 20 (105.0 mg, 0.34 mmol) in CH₂Cl₂ (3.4 mL) were added CBr₄ (229 mg, 0.69 mmol) and Ph₃P (136 mg, 0.52 mmol) at 0 °C. After 2 h, the mixture was diluted with hexane and filtered through a pad of silica gel. The filtrate was concentrated, and the resulting residue was purified by column chromatography (hexane/ EtOAc, 20:1) to give allylic bromide 8 (112.7 mg) in 89% yield: ¹H NMR (500 MHz, CDCl₃) δ [7.18 (t, J = 7.5 Hz) and 7.15 (t, J = 7.9 Hz), 1H], [7.02 (d, J = 7.5 Hz) and 6.99 (d, J= 8.0 Hz), 1H], 6.94–6.90 (m, 2H), [5.53 (t, J = 6.7 Hz) and 5.49 (t, J = 6.9 Hz), 1H], [4.18 (q, J = 7.1 Hz) and 3.94–3.86 (m), 4H], [2.76 (ddd, J = 11.7, 8.6, 3.0 Hz) and 2.71 (dt, J =9.8, 5.0 Hz), 1H], 2.66-2.57 (m, 1H), [2.33 (s) and 2.31 (s), 3H], 1.85-1.75 (m, 3H), 1.70-1.62 (m, 2H], [1.57 (s) and 1.55 (s), 3H], [1.29 (t, J = 7.1 Hz) and 1.00 (t, J = 7.1 Hz), 3H], [1.20 (d, J = 7.0 Hz) and 0.89 (d, J = 6.8 Hz), 3H]; ¹³C NMR (75 MHz, CDCl₃) δ 176.4, 175.5, 142.1, 141.6, 137.9, 137.5, 132.3, 132.2, 130.9, 129.16, 129.14, 128.3, 128.0, 127.3, 127.2, 125.3, 125.2, 60.3, 59.9, 48.5, 48.2, 46.11, 46.06, 41.8, 41.7, 33.7, 31.0, 26.1, 26.0, 21.44, 21.41, 16.3, 14.8, 14.5, 14.3, 13.9; IR (neat) 1730 cm $^{-1};~HRMS$ (EI) ${\it m/z}~calcd~for~C_{19}H_{28}BrO_2$ (MH $^+)$ 367.1273, found 367.1268.

Cyclization of Allylic Bromide 8. To a solution of allylic bromide 8 (62.0 mg, 0.17 mmol) in THF (17 mL) was added LHMDS (0.84 mL, 1.0 M solution in THF) at 0 °C. After 22 h at room temperature, the mixture was cooled to 0 °C and quenched with saturated aqueous NH4Cl solution. The solvent was removed at reduced pressure, and the residue was dissolved in EtOAc. The mixture was washed with brine, dried over anhydrous Na₂SO₄, and concentrated in vacuo. The resulting residue was purified by column chromatography on silica gel (hexane/EtOAc, 20:1) to give a mixture of 7, 7-iso, and 7-cis (9.9:3.3:1, by capillary GLC analysis) in a total 86% yield, and the isomers were separated by NP-preparative HPLC. Data for cylopentanecarboxylate 7: ¹H NMR (500 MHz, CDCl₃) δ 7.15 (t, J = 7.6 Hz, 1H), 7.00 (d, J = 7.6, 1H), 6.96 (s, 1H), 6.96 (d, J = 7.6 Hz), 1H], 4.83 (t, J = 1.6 Hz, 1H), 4.79 (d, J = 0.8 Hz, 1H), 4.16–4.04 (m, 2H), 3.96 (dd, J = 11.5, 6.8 Hz, 1H), 2.48 (t, J = 8.7 Hz, 1H), 2.31 (s, 3H), 2.11-2.05 (m, 1H), 2.04-1.96 (m, 1H), 1.96-1.90 (m, 2H), 1.77 (s, 3H), 1.25 (t, J = 7.2 Hz, 3H), 0.91 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 176.4, 145.3, 141.1, 137.4, 129.4, 127.8, 127.0, 125.6, 111.9, 60.4, 59.5, 55.6, 51.8, 30.0, 29.5, 23.3, 22.9, 21.5, 14.2; IR (neat) 1719 cm⁻¹; $[\alpha]^{20}_{D} = +46.5$ (*c* 0.43, CHCl₃); HRMS (EI) *m*/*z* calcd for C₁₉H₂₆O₂ (M⁺) 286.1933, found 286.1932. Data for 7-iso. ¹H NMR (500 MHz, CDCl₃) δ 7.14 (t, J = 7.9 Hz, 1H), 7.01 (d, J = 7.5 Hz, 1H), 6.93 (br s, 2H), 4.87 (q, J = 1.4 Hz, 1H), 4.77 (d, J = 0.5 Hz, 1H), 4.15 (q, J = 7.1 Hz, 2H), 3.73 (t, J = 10.1Hz, 1H), 3.27 (t, J = 9.8 Hz, 1H), 2.30 (s, 3H), 2.13–2.08 (m, 2H), 2.01–1.95 (m, 2H), 1.65 (s, 3H), 1.24 (t, J = 7.2 Hz, 3H), 0.72 (s, 3H); $^{13}\mathrm{C}$ NMR (75 MHz, CDCl_3) δ 177.5, 144.4, 139.6, 137.4, 129.2, 127.8, 127.4, 125.3, 111.9, 60.5, 55.6, 55.0, 54.9, 26.5, 25.6, 23.2, 21.5, 14.2, 11.3; IR (neat) 1726 cm⁻¹; $[\alpha]^{20}$ _D = -25.1 (c 0.25, CHCl₃); HRMS (EI) m/z calcd for C₁₉H₂₆O₂ (M⁺) 286.1933, found 286.1935. Data for 7-cis: 1H NMR (500 MHz, CDCl₃) δ 7.13 (t, J = 7.7 Hz, 1H), 7.00 (s, 1H), 6.99 (d, J = 7.4Hz, 2H), 4.86 (d, J = 0.9 Hz, 1H), 4.78 (s, 1H), 3.54 (dq, J =7.1, 1.6 Hz, 2H), 3.38 (dd, J = 12.5, 5.9 Hz, 1H), 2.93 (dd, J = 11.4, 7.0 Hz, 1H), 2.31 (s, 3H), 2.09 (dq, J = 11.9, 6.0 Hz, 1H), 2.03–1.98 (m, 1H), 1.96 (dq, J = 6.0, 1.2 Hz, 1H), 1.79 (dq, J = 12.2, 6.3 Hz, 1H), 1.63 (s, 3H), 1.18 (s, 3H), 0.84 (t, J = 7.2 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 176.0, 145.0, 141.1, 137.1, 129.2, 127.6, 127.3, 125.5, 111.3, 60.1, 59.1, 55.0, 52.8, 30.8, 29.1, 23.5, 21.4, 13.6; IR (neat) 1727 cm⁻¹; [α]²⁰_D = +15.5 (*c* 0.075, CHCl₃); HRMS (EI) *m*/*z* calcd for C₁₉H₂₆O₂ (M⁺) 286.1933, found 286.1930.

(1R,2S,5R)-2-Acetyl-1-methyl-5-m-tolylcyclopentanecarboxylic Acid Ethyl Ester (22). To a solution of cyclopentanecarboxylate 7 (11.0 mg, 0.038 mmol) in EtOAc (1 mL) were added a few drops of saturated ozone solution in EtOAc every 20 min at -78 °C until the starting material disappeared on TLC. After removal of excess ozone by a stream of nitrogen, Ph₃P (30 mg, 0.11 mmol) was added to the mixture. It was stirred overnight and filtered through a pad of silica gel. The filtrate was concentrated, and the resulting residue was purified by column chromatography on silica gel (hexane/ EtOAc, 10:1) to give ketone 22 (9.4 mg) in 85% yield: ¹H NMR (500 MHz, CDCl₃) δ 7.17 (t, J = 7.5 Hz, 1H), 7.03 (d, J = 7.6Hz, 1H), 7.00 (s, 1H), 6.99 (d, J = 7.5 Hz, 1H), 4.12 (dq, J = 7.1, 1.4 Hz, 2H), 3.68 (dd, J = 9.8, 6.7 Hz, 1H), 2.85 (t, J = 8.1Hz, 1H), 2.33 (s, 3H), 2.17 (s, 3H), 2.20-2.00 (m, 4H), 1.25 (t, J = 7.2 Hz, 3H), 1.00 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 208.8, 176.5, 140.3, 137.4, 129.8, 127.8, 127.3, 125.9, 63.4, 60.7, 54.7, 52.2, 30.0, 29.9, 27.8, 22.5, 21.5, 14.0; IR (neat) 1712, 1233 cm⁻¹; $[\alpha]^{20}_{D} = +56.4$ (*c* 0.70, CHCl₃); HRMS (EI) *m*/*z* calcd for C₁₈H₂₄O₃ (M⁺) 288.1725, found 288.1721.

(1R,2R,5R)-2-Acetyl-1-methyl-5-m-tolylcyclopentanecarboxylic Acid Ethyl Ester (22-iso). To a solution of cyclopentanecarboxylate 7-iso (24.6 mg, 0.086 mmol) in EtOAc (1 mL) were added a few drops of saturated ozone solution in EtOAc every 10 min for 2 h, and 0.5 mL of saturated ozone solution in EtOAc was added to the mixture every 1 h for 4 h at -78 °C. After removal of excess ozone by a stream of nitrogen, Ph₃P (68 mg, 0.26 mmol) was added to the mixture. The mixture was stirred overnight and filtered through a pad of silica gel. The filtrate was concentrated, and the resulting residue was purified by column chromatography on silica gel (hexane/EtOAc, 10:1) to give ketone 22-iso (14.0 mg) in 82% yield: ¹H NMR (500 MHz, CDCl₃) δ 7.16 (t, J = 7.8 Hz, 1H), 7.04 (d, J = 7.6 Hz, 1H), 6.76 (br s, 2H), 4.21 (q, J = 7.1 Hz, 2H), 3.71 (t, J = 9.5 Hz, 1H), 3.66 (dd, J = 12.1, 8.2 Hz, 1H), 2.38–2.29 (m, 1H), 2.31 (s, 3H), 2.17 (dq, J = 12.2, 6.3 Hz, 1H), 2.06 (s, 3H), 2.10–2.03 (m, 1H), 1.95 (dtd, J = 13.2, 9.5,6.3 Hz, 1H), 1.25 (t, J = 7.2 Hz, 3H), 0.78 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) & 207.5, 176.5, 138.2, 137.5, 129.2, 127.9, 127.8, 125.4, 61.1, 61.0, 55.3, 30.1, 26.7, 22.4, 21.4, 14.2, 12.8; IR (neat) 1712, 1268 cm⁻¹; $[\alpha]^{20}_{D} = -87.3$ (*c* 0.48, CHCl₃); HRMS (EI) m/z calcd for C₁₈H₂₄O₃ 288.1725, found 288.1725.

Epimerization of Ketone 22 to 22-*iso.* A mixture of ketone **22** (7.0 mg, 0.024 mmol) and DBU (0.5 mL) was stirred for 20 h at room temperature and filtered through a pad of silica gel to give a mixture of ketone **22** and epimer **22-***iso* (6.8 mg, 1:4.2 by 400 MHz ¹H NMR analysis) in a total 97% yield. The mixture was separated by column chromatography on silica gel (hexane/EtOAc, 10:1).

Epimerization of Ketone 22-*iso* **to 22.** A mixture of ketone **22-***iso* (5.5 mg, 0.019 mmol) and DBU (0.5 mL) was stirred for 36 h at room temperature and then filtered through a pad of silica gel to give a mixture of ketone **22-***iso* and epimer **22** (5.5 mg, 4.2:1 by 400 MHz ¹H NMR analysis). The mixture was separated by column chromatography on silica gel (hexane/EtOAc, 10:1).

Cyclopentanecarboxylate 7 from 22. A mixture of methyltriphenylphosphonium iodide (56 mg, 0.14 mmol) and *n*-BuLi (0.078 mL, 1.6 M solution in hexane) in THF (1.0 mL) was stirred for 30 min at -78 °C. To the mixture was added a solution of ketone **22** (8.1 mg, 0.028 mmol) in THF (0.5 mL) at -78 °C, and the mixture was stirred for 10 h at room temperature. The reaction mixture was quenched with a few drops of saturated aqueous NH₄Cl solution and filtered

through a pad of silica gel. The filtrate was concentrated in vacuo, and the resulting residue was purified by column chromatography on silica gel (hexane/EtOAc, 20:1) to afford cyclopentanecarboxylate 7 (6.1 mg) in 76% yield.

((1S,2R,5R)-2-Isopropenyl-1-methyl-5-m-tolylcyclopentyl)methanol (23). To a solution of cyclopentanecarboxylate 7 (13.0 mg, 0.045 mmol) in dry toluene (0.9 mL) was added DIBALH (0.27 mL, 1.0 M solution in hexane) at -78 °C. The reaction mixture was stirred and slowly warmed to -30 °C over 3 h. The mixture was cooled to -78 °C again, and methanol (0.5 mL) was added to the mixture. The mixture was stirred overnight at room temperature and filtered through a pad of Celite. The filtrate was concentrated at reduced pressure, and the resulting residue was purified by column chromatography on silica gel (hexane/EtOAc, 10:1) to afford alcohol 23 (10.2 mg) in 93% yield: ¹H NMR (500 MHz, CDCl₃) δ 7.17 (t, J = 7.8 Hz, 1H), 7.02–6.99 (m, 3H), 4.98 (t, J = 1.5Hz, 1H), 4.96 (s, 1H), 3.45 (br d, J = 4.1 Hz, 2H), 3.20 (dd, J = 10.9, 7.7 Hz, 1H), 2.37 (dd, J = 10.9, 6.9 Hz, 1H), 2.33 (s, 3H), 2.05-1.95 (m, 2H), 1.92 (s, 3H), 1.91-1.86 (m, 1H), 1.85-1.78 (m, 2H), 0.66 (s, 3H); 13 C NMR (75 MHz, CDCl₃) δ 148.4, 141.9, 137.4, 129.9, 127.8, 126.9, 126.0, 111.7, 68.4, 56.1, 50.4, 48.7, 29.7, 24.6, 23.0, 21.5; IR (neat) 1040 cm⁻¹; $[\alpha]^{20}_{D} = +15.2$ (c 0.35, CHCl₃); HRMS (EI) m/z calcd for C₁₇H₂₄O (M⁺) 244.1827, found 244.1828.

(1S,2R,5R)-2-Isopropenyl-1-methyl-5-m-tolylcyclopentanecarbaldehyde (24). To a solution of alcohol 23 (12.0 mg, 0.050 mmol) in dry CH₂Cl₂ (1 mL) were added NaOAc (26 mg, 0.32 mmol), molecular sieves (4 Å, 22 mg), and PCC (24 mg, 1.08 mmol) at 0 °C. After 2.5 h at the same temperature, the mixture was diluted with ether and filtered through a pad of Florisil. The filtrate was concentrated, and the resulting residue was purified by column chromatography on silica gel (hexane/EtOAc, 15:1) to afford aldehyde 24 (9.2 mg) in 77% yield: ¹H NMR (500 MHz, CDCl₃) δ 9.57 (s, 1H), 7.15 (t, J = 7.9 Hz, 1H), 7.00 (d, J = 7.5 Hz), 6.90 (d, J = 7.9 Hz, 1H), 6.89 (s, 1H), 4.92 (t, J = 1.4 Hz, 1H), 4.92 (d, J = 1.4 Hz, 1H), 3.65 (dd, J = 11.5, 6.7 Hz, 1H), 2.54 (dd, J = 11.5, 6.7 Hz, 1H), 2.31 (s, 3H), 2.16 (dtd, J = 12.3, 6.2, 1.7 Hz, 1H), 2.10 (dq, J = 11.7, 6.2 Hz, 1H), 1.97 (dtd, J = 12.7, 6.4, 1.7 Hz, 1H), 1.88 (dq, J = 11.9, 6.7 Hz, 1H), 1.72 (s, 3H), 0.82 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 205.6, 143.0, 140.2, 137.7, 129.4, 128.0, 127.2, 125.6, 113.1, 58.7, 57.3, 48.9, 30.0, 29.9, 23.4, 21.5, 19.8; IR (neat) 2710, 1721 cm⁻¹; $[\alpha]^{20}_{D} = +39.7$ (*c* 0.22, CHCl₃); HRMS (EI) m/z calcd for C17H22O (M⁺) 242.1671, found 242.1672.

3-((1S,2R,5S)-2-Isopropyl-1-methyl-5-m-tolylcyclopentyl)propionic Acid (6). To a solution of benzyl (trimethylsilyl)acetate (46 mg, 0.21 mmol) in THF (1.0 mL) was added LDA (0.21 mL, 0.5 M solution in THF) at -78 °C. After 0.5 h, a solution of aldehyde 24 (10.0 mg, 0.041 mmol) in THF (0.5 mL) was added to the mixture at the same temperature, and the mixture was stirred for 2 h at -25 °C. The mixture was poured into saturated aqueous NH₄Cl solution and extracted with EtOAc (20 mL \times 3). The combined organic layers were washed with brine, dried over anhydrous Na2SO4, and concentrated in vacuo to give crude benzyl ester 25. The crude product was dissolved in EtOAc (2.0 mL), and PtO₂ (1 mg) was added. The mixture was stirred overnight at ambient temperature under a hydrogen atmosphere. The mixture was filtered through a pad of Celite, and the filtrate was concentrated in vacuo. The resulting residue was purified by column chromatography on silica gel (hexane/EtOAc, 2:1) to afford carboxylic acid 6 (11.0 mg) in 92% overall yield: ¹H NMR (500 MHz, CDCl₃) δ 7.15 (t, J = 8.0 Hz, 1H), 6.99 (d, J = 7.4 Hz, 1H), 6.94 (d, J = 8.0 Hz, 1H), 6.94 (s, 1H), 2.91 (t, J = 7.5 Hz, 1H), 2.49-2.35 (m, 2H), 2.33 (s, 3H), 2.11-1.98 (m, 2H), 1.89-1.73 (m, 3H), 1.72–1.65 (m, 1H), 1.60–1.50 (m, 2H), 0.96 (d, J =6.7 Hz, 3H), 0.94 (d, J = 6.7 Hz, 3H), 0.66 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) & 180.7, 144.4, 137.2, 130.0, 127.7, 126.6, 126.1, 56.3, 53.9, 46.7, 30.9, 30.1, 29.9, 28.6, 27.8, 24.1, 23.5, 22.2, 21.5; IR (neat) 1708, 1297 cm⁻¹; $[\alpha]^{20}_{D} = +38.2$ (c 1.03, CHCl₃); HRMS (EI) m/z calcd for $C_{19}H_{28}O_2$ (M⁺) 288.2089, found 288.2087.

(3R,3aS,10bS)-3-Isopropyl-3a,9-dimethyl-2,3,3a,4,5,10bhexahydro-1H-benzo[e]azulen-6-one (5). A mixture of carboxylic acid **6** (25.3 mg, 0.089 mmol) and PPA (1 mL) was stirred overnight at 90 °C. Crushed ice was added, and the mixture was extracted with EtOAc (20 mL \times 3). The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, and concentrated in vacuo. The resulting residue was purified by column chromatography on silica gel (hexane/ EtOAc, 20:1) to give ketone 5 (21.0 mg) in 89% yield: ¹H NMR (500 MHz, CDCl₃) δ 7.66 (d, J = 7.7 Hz, 1H), 7.08 (d, J = 7.7Hz, 1H), 7.07 (s, 1H), 3.18 (dd, J = 12.4, 5.5 Hz, 1H), 2.83 (t, J = 6.4 Hz, 1H), 2.83 (t, J = 7.2 Hz, 1H), 2.38 (s, 3H), 2.14 (dt, J = 12.8, 6.4 Hz, 1H), 2.09 (dt, J = 10.9, 5.5 Hz, 1H), 1.98 (ddd, J = 11.4, 5.7, 5.7 Hz, 1H), 1.85–1.73 (m, 3H), 1.41–1.25 (m, 2H), 0.97 (d, J = 6.5 Hz, 3H), 0.92 (d, J = 6.6 Hz, 3H), 0.61 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 205.6, 142.2, 142.1, 137.2, 127.85, 127.82, 126.9, 59.8, 53.0, 43.7, 41.6, 33.7, 30.1, 29.4, 28.5, 27.7, 23.5, 22.2, 21.7; IR (neat) 1673 cm⁻¹; $[\alpha]^{20}_{D} =$ -140.9 (c 0.64, CHCl₃); HRMS (EI) m/z calcd for C₁₉H₂₆O (M⁺) 270.1984, found 270.1984.

(3R,3aS,10bS)-3-Isopropyl-3a,6,9-trimethyl-1,2,3,3a,4,-10b-hexahydrobenzo[e]azulene (26). To a solution of ketone 5 (8.0 mg, 0.030 mmol) in ether (1.0 mL) was added methyllithium (0.2 mL, 1.5 M solution in ether) at -78 °C, and the mixture was stirred for 1 h at the same temperature. SOCl₂ (0.04 mL, 0.54 mmol) was then added to the mixture. After 10 min at -78 °C, the mixture was stirred for 1 h at room temperature, poured into ice, and extracted with ether (15 mL \times 3). The combined organic layers were washed with brine, dried over anhydrous MgSO₄, and concentrated in vacuo. The residue was purified by column chromatography on silica gel (hexane) to afford a mixture of *endo*-olefin **26** and *exo*-olefin 27 (7.0 mg, 5.3:1 by 500 MHz ¹H NMR analysis) in a total 88% yield. The endo-olefin 26 and exo-olefin 27 were separated by column chromatography on AgNO3-impregnated silica gel (hexane). Data for endo-olefin 26: ¹H NMR (500 MHz, CDCl₃) δ 7.32 (d, J = 8.0 Hz, 1H), 7.01 (d, J = 8.1 Hz, 1H), 6.99 (s, 1H), 5.85–5.83 (m, 1H), 2.83 (t, J = 9.3 Hz, 1H), 2.57 (br d, J = 15.8 Hz, 1H), 2.35 (s, 3H), 2.20 (t, J = 1.5 Hz, 3H), 2.21-2.15 (m, 1H), 1.92-1.88 (m, 1H), 1.84-1.79 (m, 1H), 1.77-1.68 (m, 1H), 1.30–1.19 (m, 2H), 0.97 (d, J = 6.4 Hz, 3H), 0.92 (d, J = 6.6 Hz, 3H), 0.42 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 140.7, 135.8, 135.4, 130.5, 128.4, 126.6, 126.0, 125.9, 59.6, 53.6, 42.3, 41.2, 30.6, 28.3, 27.7, 27.2, 25.7, 23.6, 22.8, 21.3; IR (neat) 1574, 1496 cm⁻¹; $[\alpha]^{20}_{D} = -320.1$ (*c* 0.10, CHCl₃); HRMS (EI) *m*/*z* calcd for C₂₀H₂₈ (M⁺) 268.2191, found 268.2190.

Reduction of *endo*-**Olefin 26** with 10% Pd/C. A mixture of *endo*-olefin **26** (2.3 mg, 0.0086 mmol) and 10% Pd/C (2 mg) in ethanol (1 mL) was stirred for 1 h at 15 °C under a hydrogen atmosphere. The mixture was filtered over a pad of Celite to give only 7-*epi*-presphaerene (**1**) (2.3 mg) in quantitative yield: ¹H NMR (500 MHz, CDCl₃) δ 7.10 (d, J = 7.8 Hz, 1H), 6.97 (d, J = 7.8 Hz, 1H), 6.95 (s, 1H), 3.14 (dd, J = 12.6, 5.2 Hz, 1H), 3.05–2.98 (m, 1H), 2.36 (s, 3H), 1.93–1.82 (m, 4H), 1.82–1.72 (m, 2H), 1.65 (td, J = 14.0, 1.0 Hz, 1H), 1.47–1.30 (m, 3H), 1.34 (d, J = 7.0 Hz, 3 H), 0.92 (d, J = 6.5 Hz, 3H), 0.90 (d, J = 6.6 Hz, 3H), 0.40 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 142.4, 141.9, 134.4, 126.3, 126.0, 123.8, 59.2, 53.3, 40.3, 38.7, 35.6, 32.3, 30.0, 27.80, 27.75, 25.1, 24.2, 21.9, 21.6, 21.1; IR (neat) 1455 cm⁻¹; [α]²⁰_D = -91.8 (*c* 0.20, CHCl₃); HRMS (EI) *m*/*z* calcd for C₂₀H₃₀ (M⁺) 270.2348, found 270.2348.

Reduction of *endo*-Olefin 26 with PtO₂. A mixture of *endo*-olefin 26 (2.5 mg, 0.0093 mmol) and PtO₂ (1 mg) in ethanol (1 mL) was stirred for 1 h at 15 °C under a hydrogen atmosphere. The mixture was filtered over a pad of Celite to give a mixture of (–)-presphaerene (1) and 1' (2.5 mg, 1:9 by 400 MHz ¹H NMR analysis) in quantitative yield.

(3*R*,3a.*S*,10b.*S*)-3-Isopropyl-3a,9-dimethyl-6-methylene-1,2,3,3a,4,5,6,10b-octahydrobenzo[*e*]azulene (27). A mixture of methyltriphenylphosphonium iodide (112 mg, 0.28

mmol) and n-BuLi (0.15 mL, 1.6 M solution in hexane) in THF (2.0 mL) was stirred for 0.5 h at -78 °C. To the mixture was added ketone 5 (10.7 mg, 0.040 mmol) in THF (1.0 mL) at the same temperature. After 2 h at room temperature, the mixture was quenched with saturated aqueous NH4Cl solution and filtered through a pad of silica gel. The filtrate was concentrated in vacuo, and the resulting residue was purified by column chromatography on silica gel (hexane) to give exo-olefin 27 (9.5 mg) in 89% yield: ¹H NMR (500 MHz, CDCl₃) δ 7.03 (d, J = 7.5 Hz, 1H), 6.94 (d, J = 7.6 Hz, 1H), 6.91 (s, 1H), 5.11(dd, J = 2.2, 1.3 Hz, 1H), 4.86 (d, J = 2.5 Hz, 1H), 2.95 (dd, J= 12.7, 5.5 Hz, 1H), 2.45 (td, J = 13.2, 4.1 Hz, 1H), 2.35-2.29 (m, 1H), 2.33 (s, 3H), 1.95 (td, J = 12.8, 4.0 Hz, 1H), 1.90-1.81 (m, 3H), 1.80-1.71 (m, 2H), 1.42-1.37 (m, 1H), 1.33-1.24 (m, 1H), 0.94 (d, J = 6.4 Hz, 3H), 0.89 (d, J = 6.6 Hz, 3H), 0.62 (s, 3H); 13 C NMR (100 MHz, CDCl₃) δ 153.3, 141.4, 139.9, 136.3, 127.3, 126.1, 113.7, 59.3, 54.8, 40.8, 39.9, 32.4, 29.7, 27.9, 27.8, 24.7, 24.2, 24.8, 21.4; IR (neat) 1628, 1448 cm⁻¹; $[\alpha]^{20}_{D} = -165.3$ (*c* 0.46, CHCl₃); HRMS (EI) *m*/*z* calcd for C₂₀H₂₈ (M⁺) 268.2191, found 268.2193.

Reduction of *exo*-**Olefin 27 with 10% Pd/C.** A mixture of *exo*-olefin **27** (2.3 mg, 0.0086 mmol) and 10% Pd/C (2 mg) in ethanol (1.0 mL) was stirred for 1 h at 15 °C under a hydrogen atmosphere. The mixture was filtered over a pad of Celite to give **1**' (2.3 mg) in quantitative yield.

Reduction of *exo***-Olefin 27 with PtO₂.** A mixture of *exo*olefin **27** (9.5 mg, 0.035 mmol) and PtO₂ (1 mg) in ethanol (2.0 mL) was stirred for 3 h at 15 °C under a hydrogen atmosphere. The mixture was filtered through a pad of Celite to give a mixture of **1** and **1**' (9.5 mg, 3:1 by 500 MHz ¹H NMR analysis) in quantitative yield. **1** and **1**' were separated by column chromatography on AgNO₃-impregnated silica gel (hexane). Data for **1**: ¹H NMR (500 MHz, CDCl₃) δ 6.94 (d, J = 7.5 Hz, 1H), 6.92 (s, 1H), 6.87 (d, J = 7.4 Hz, 1H), 3.20 (dd, J = 12.2, 5.6 Hz, 1H), 3.09–3.04 (m, 1H), 2.28 (s, 3H), 2.03–1.93 (m, 2H), 1.90–1.77 (m, 4H), 1.74–1.66 (m, 2H), 1.44–1.33 (m, 2H), 1.31 (d, J = 7.6 Hz, 3H), 0.93 (d, J = 6.5 Hz, 3H), 0.90 (d, J = 6.6 Hz, 3H), 0.51 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 142.4, 140.9, 135.0, 129.5, 127.6, 126.2, 58.9, 53.3, 41.7, 41.6, 32.9, 28.6, 28.5, 28.2, 27.9, 24.2, 24.0, 21.3, 21.2, 18.3; IR (neat) 1455 cm⁻¹; [α]²⁰_D = -90.2 (*c* 0.40, CHCl₃); HRMS (EI) *m*/*z* calcd for C₂₀H₃₀ (M⁺) 270.2348, found 270.2347.

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Supporting Information Available: ¹H NMR comparison of natural and synthetic **1** and **1**', the most stable conformations of *endo*-olefin **26** and *exo*-olefin **27**, along with copies of the ¹H and ¹³C NMR spectra for **1**, **1**', **5–20**, and **22–27**, ¹H–¹H COSY, ¹H–¹³C COSY, and DEPT spectra for **1**, **1**', **5**, **7**, **7**-*iso*, and **7**-*cis*, and NOE spectra for **1**, **1**', **7**, **7**-*iso*, and **7**-*cis* (PDF). This material is available free of charge via the Internet at http://pubs.acs.org.

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