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The P₁ *N*-isopropyl motif bearing hydroxyethylene dipeptide isostere analogues of aliskiren are in vitro potent inhibitors of the human aspartyl protease renin

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ABSTRACT

Novel nonpeptide small molecule renin inhibitors bearing an *N*-isopropyl P_1 motif were designed based on initial lead structures **1** and aliskiren (**2**). (P_3 - P_1)-Benzamide derivatives such as **9a** and **34**, as well as the corresponding P_1 basic tertiary amine derivatives **10** and **35** were found to display low nanomolar inhibition against human renin in vitro.

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Hypertension is a major risk factor of high prevalence worldwide for cardiovascular diseases.¹ The pivotal functions of the renin-angiotensin-aldosterone system (RAAS) in cardiovascular (patho)physiology including the regulation of blood pressure, as well as electrolyte and body fluid homeostasis have been well established.² The RAAS has emerged as an important target for antihypertensive drug therapy, with angiotensin (Ang)-converting enzyme inhibitors (ACEi) and AT1-receptor blockers (ARBs) having proven effectiveness in preventing cardiovascular and renal events.³ Direct blockade of highly substrate-specific renin activity attenuates the formation of Ang I and Ang II, the principal mediator of the RAAS, even at elevated circulating plasma renin levels due to negative feedback mediated release from the kidney.^{4,5} The attractiveness of the aspartyl protease renin as an advantageous target^{6,7} with the potential to provide more effective suppression of the RAAS has stimulated vast efforts to develop small molecule, orally efficacious renin inhibitors.⁸

We have discovered recently several series of potent, highly soluble, and orally active non-peptide chemotype renin inhibitors, such as **1** and **2** (Fig. 1).⁹ These transition-state mimetic (TSM) inhibitors emerged from an unprecedented topological structure-based design approach incorporating a direct spacer link of the P_1 and P_3 moieties and concomitant removal of the P_2 - P_4 backbone



Figure 1. Renin inhibitors **1** and **2**, and 'simplification' strategies A and B by *N* versus chiral CH' replacements at the P₁ and/or P₁' positions.

of classical peptide-like renin inhibitors.^{10–13} Aliskiren (**2**) is a highly potent selective inhibitor of human renin demonstrating sustained blood pressure lowering efficacy in vivo upon once-daily oral dosing.¹² It is the first direct renin inhibitor to be marketed as a new therapy for hypertension,¹³ a hallmark of more than three decades of intense research efforts worldwide.

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Our strategy during initial lead optimization of **1** and related analogues was to reduce the chemical complexity of these novel chemotype renin inhibitors by 'simplifying' structural elements of the key TSM pharmacophore. Computational modeling based on the X-ray structure of inhibitor **1** bound to the active site of recombinant human renin (rh-renin)^{9,10} suggested replacement of the chirality center either at P_1 ' ('reversed carboxamides', approach A; Fig. 1),¹⁴ or at P_1 and introduction of a functional spacer group to tether the substituted phenyl moiety (approach B) to be viable concepts.

We report herein our results on establishing a new series of non-peptide renin inhibitors based on the *N*-isopropyl benzamide P_1-P_3 motif and the corresponding *N*-isopropyl-*N*-benzylamine analogues (approach B). A divergent chemistry strategy provided ready access to a variety of analogues, starting from advanced versatilely protected amine TSM intermediates. This tactic afforded the opportunity for broad SAR investigation addressing structural modifications of the P_3-P_1 pharmacophore as well as the P_3^{sp} side chain.

The stereoselective synthesis of the key intermediate (2R,4S,5S)-**8** bearing a P₁' Me at the hydroxyethylene TSM portion is depicted in Scheme 1. Starting from commercial OBn-protected N-BOC-serine 3, aldehyde 4 was obtained in >85% yield over 2 steps with >95% ee (by chiral HPLC of the corresponding alcohol, obtained via LiBH₄/THF reduction, on Chiralcel OJ with *n*-hexaneisopropanol 96:4). Adaptation of the chiral homo-enolate addition to amino acid derived aldehydes, as reported by DeCamp et al.,¹⁵ provided the hydroxyethylene moiety in an efficient and reliable manner on a multi-gram scale. Thus, metallation of 3-iodo-2(R)methyl-propionic acid methyl ester with Zn(Cu)/Cl₃Ti(OiPr) and subsequent reaction (4 equiv of the homo-enolate species in excess) with **4** furnished the γ -hydroxy esters 4S-**5a**/4R-**5b** (4:1 ratio) in 50–60% overall yield.¹⁶ Further transformation to the respective lactones under acidic conditions was followed by direct aminolysis with *n*-butylamine at 80 °C without detectable epimerization at



Scheme 1. Reagents and conditions: (a) MeI, K₂CO₃, DMF, 70 °C, 95%; (b) DIBAL, toluene, -78 °C, >90%; (c) 3-iodo-2(*R*)-methyl-propionic acid methyl ester, Zn(Cu), TiCl₄–Ti(OiPr)₄, *N,N*-dimethyl acetamide, toluene, -20 °C; (d) AcOH, toluene, reflux. (e) *n*-BuNH₂, 80 °C; (f) recrystallisation from *n*-hexane–CHCl₃, 40% (4 steps from **3**); (g) 2-methoxypropene, cat. TsOH, CH₂Cl₂, rt, 97%. (h) H₂, cat. Pd/C 10%, MeOH; (i) MsCl, Et₃N, CH₂Cl₂ o °C; (j) NaN₃, DMPU, 40 °C, 95% (3 steps); (k) H₂, cat. Pd/C 10%, MeOH; (i) acetone, NaBH₃CN, rt, >90% (2 steps); (m) 4-methoxy-3-(3-methoxypropoxy)-benzaldehyde, Ti(OiPr)₄, NaBH₃CN, MeOH; 40%; or amide coupling to 3,4-substituted benzoic acids using HBTU, Et₃N, MeCN; (n) cat. TsOH, MeOH, rt; (o) 4 N HCl, dioxane, -20 °C; followed by lyophilisation.

C2. The desired 4(*S*)-**6** (mp 105 °C)¹⁷ was isolated in high diastereomeric purity (>17:1 by RP-HPLC) after single crystallization from *n*hexane–CHCl₃ in 40% yield from aldehyde **4**.

Intermediate 6 was then subjected to N,O-acetal formation (97%), followed by hydrogenolysis of the benzylether, mesylate formation, and finally displacement with NaN₃ in DMPU to afford azide 7 in 83% (3 steps). Azide reduction and reductive amination with acetone/NaBH₃CN yielded the secondary amine 8 in good overall yield. Inhibitors **9a-i** bearing a *tertiary N*-isopropyl-benzamide group at the P₁ position, as listed in Table 1, were prepared via the activated esters from the corresponding benzoic acids¹⁸ using HBTU.¹⁹ Removal of the N,O-acetal protecting group with cat. TsOH in MeOH proceeded uneventfully and in good isolated yields (>80%). N-BOC deprotection with 4 N HCl-dioxane was conducted at -20 °C in order to avoid formation of the dihvdro-1.3imidazole **37** as a major side product (Fig. 2, vide infra) affording the inhibitors as their mono-HCl salts after lyophilisation. Inhibitor **10** (di-HCl salt) bearing an *N*-isopropyl substituted tertiary amine was obtained by reductive alkylation of 8 with 4-methoxy-3-(3methoxy-propoxy)-benzaldehyde²⁰ in the presence of a Lewis acid (NaBH₃CN, Ti(OiPr)₄; 40% yield), followed by two-step N,Odeprotection.

The sulfonamide analogue **14** was prepared according to Scheme 2. In brief, chlorosulfonylation of anisaldehyde **11** rendered **12** in high yield, which then was coupled to secondary amine **8** in the presence of DIEA as base. Baeyer–Villiger oxidation with *m*-chloro-perbenzoic acid and subsequent hydrolysis afforded intermediate **13**, which was O-alkylated to introduce the P_3^{sp} methoxypropoxy side chain. Final stepwise *N*,*O*-deprotection afforded **14** in good overall yield. **15** was obtained in a similar fashion via coupling of *p*-OMe-phenyl-sulfonylchloride to **8** (82% yield).

The synthesis of 3,4-disubstituted benzoic acids **18**, **20**, and **23–25** with a modified P_3 moiety was achieved according to standard protocols from readily available starting materials (Scheme 3).¹⁸

The synthesis of the O-silyl protected TSM intermediate (2S,4S,5S)-**33** with isopropyl at the P₁' position is described in Scheme 4. Thus, Evans alkylation of the lithium enolate derived from (S)-27 with (E)-4-bromo-but-2-envloxymethyl)-benzene²¹ gave 28 in 77% yield with >95% de. Bromolactonization of 28 provided trans-29 in high yield as the only isolated isomer. Displacement of the bromine in 29 under non-optimized reaction conditions afforded azide **30** $(>97\% \text{ ee})^{22}$ in 30% isolated yield together with the elimination side product **31** (60%). The relative stereochemistry of lactones 29 and 30 was determined by ¹H and ¹³C NMR as reported previously.²³ Azide **30** was then converted to the N-BOC protected lactone 32 by selective hydrogenation in the presence of (BOC)₂O followed by O-debenzylation, mesylation, and final azide substitution. Inhibitor 34 was prepared from 33 by the reaction sequence analogous to the route described for 9a in Scheme 1. Inhibitor **35** bearing a tertiary amine at P₁ was obtained as described in Scheme 4.

Structure–activity relationship (SAR) data based on IC_{50} s for inhibition of rh-renin, as well as of endogenous renin in human plasma, is summarized for a set of representative inhibitors in Table 1. We were gratified to find the in vitro potency of the P₁ *N*-isopropyl benzamide **9a** to be comparable to that of the corresponding P₁ carba-analogue **1** in the purified renin assay, albeit enzyme affinity of **9a** dropped fivefold in the plasma assay as compared to **1**. Extending previous SAR^{10,24} by modification of the P₃^{sp} side chain revealed that the phenol ether oxygen of **9a** is not important for binding interactions to purified renin (**9b** to **9d**). The ether link in **9a** was replaced by *CH*₂ to afford an in vitro highly potent inhibitor of rh-renin, **9b**, which however was 15-fold less active in the presence of plasma.²⁵ A similar potency loss in the plasma assay using angiotensinogen as the endogenous substrate has been observed in several other non-peptide

Table 1

In vitro enzyme inhibition data for human renin



| No. | Х | R ¹ | R ² | R ³ | IC ₅₀ (nM), purified ^a | IC ₅₀ (nM), plasma ^b |
|-----------------|-----------------|--|----------------|----------------|--|--|
| 1 | | | | | 1 | 1 |
| 2 | | | | | 0.6 | 0.6 |
| 9a | CO | O(CH ₂) ₃ OMe | OMe | Me | 2 | 5 |
| 9b | CO | (CH ₂) ₄ OMe | OMe | Me | 1 | 15 |
| 9c ^d | CO | CH=CH(CH ₂) ₂ OMe | OMe | Me | 1 | 4 |
| 9d | CO | CH ₂ NHCO ₂ Me | OMe | Me | 3 | 2 |
| 9e | CO | (CH ₂) ₂ NHCO ₂ Me | OMe | Me | 100 | nd ^c |
| 9f | CO | O(CH ₂) ₃ OMe | Me | Me | 8 | 13 |
| 9g | CO | O(CH ₂) ₃ OMe | Et | Me | 2 | 7 |
| 9h | CO | O(CH ₂) ₃ OMe | Br | Me | 8 | 21 |
| 9i | CO | O(CH ₂) ₃ OMe | ССН | Me | 6 | 14 |
| 34 | CO | O(CH ₂) ₃ OMe | Et | iPr | 1 | 10 |
| 14 | SO ₂ | O(CH ₂) ₃ OMe | OMe | Me | 300 | nd ^c |
| 15 | SO ₂ | Н | OMe | Me | >100,000 | nd ^c |
| 10 | CH ₂ | O(CH ₂) ₃ OMe | OMe | Me | 13 | 14 |
| 35 | | O(CH ₂) ₃ OMe | OMe | iPr | 7 | 7 |

^a Purified rh-renin assay at pH 7.4.¹¹

^b Inhibition of endogenous renin in plasma from human.¹¹

^c Not determined.

^d Stereoisomer ratio *Z*:*E* = 10:1. All compounds showed similar IC₅₀s against marmoset plasma renin, with high selectivity to bovine cathepsin D and porcine pepsin (IC₅₀ > 100 μ M). **9a**, **9g**, **9h**, **10** had IC₅₀ > 20 μ M for rat renin (for **35**, IC₅₀ = 1.4 μ M). Calculated log *P* for **9a**: 2.74; **9b**: 3.60; **9g**: 4.08; **34**: 5.00; **35**: 2.94.



Scheme 2. Reagents and conditions: (a) $HC(OEt)_3$, NH_4Cl , EtOH, reflux, 90%. (b) $CISO_3H$, 3 h at -5 °C, then 2 h at rt, 80%; (c) **8**, DIEA, CH_2Cl_2 , 0 °C, 16 h, 72%; (d) MCPBA, $CHCl_3$, 48 h, rt; then Et_3N , MeOH, rt, 2 d, 55%. (e) $MeO(CH_2)_3I$, K_2CO_3 , MeCN, reflux, 18 h, 82%. (f) CSA, MeOH, rt, 89%; (g) 4 N HCl-dioxane, 0 °C, 2 h, 82%.



Figure 2. Decomposition products of inhibitor 9a.

compound series upon minor structural modifications leading to a subtle increase in overall hydrophobicity of the inhibitors.^{10,11,26}

A methyl carbamate group tethered to the phenyl by a methylene spacer provided highly active **9d** with comparable enzyme affinities in both in vitro assays. Both the NH and the MeO oxygen of the carbamate were found in reasonable H-bonding distance to the Ser219 side chain OH according to modeling. In contrast, the homologous carbamate **9e** with a two-carbon tether was only weakly active against rh-renin.



Scheme 3. Reagents and conditions: (a) $(CHO)_n$, HCl, CHCl₃, rt, 86%; (b) NaN₃, KI, DMPU, 100 °C, 81%; (c) Ph₃P, THF; then H₂O, 91%; (d) MeOCOCl, DIEA, dioxane, 60%; (e) 1 N NaOH, MeOH; (f) NaCN, KI, MeCN, 100 °C, 76%; (g) H₂, RaNi, 79%; (h) MeOCOCl, CH₂Cl₂, 0 °C to rt, 4 h, 87%; (i) Br₂, AcOH, rt, 12 h, 15%; (j) H₂SO₄, MeOH, reflux, 18 h, 95%; (k) MeO(CH₂)₃I, K₂CO₃, reflux, 16 h, 98%; (l) trimethylsilylacetylene, cat. Cl₂Pd(PPh₃)₂, Cul, Et₃N, rt, 15 h, 100%; (m) 1 N KOH, MeOH, 97%; (n) H₂, cat. Pd/C 10%, EtOH, 77%.

Next, various small hydrophobic residues such as alkyl and halogen were investigated as replacements of the *para*-OMe group which interacts with the S₃ site of renin (Table 1, **9f**–**i**). The P₃ ethyl derivative **9g** was equipotent to **9a** in both assays, whereas a threeto four -fold increase in IC₅₀s was observed for **9f**, **9h**, and **9i**. Inhibitor **34** with a P₁' isopropyl substituted hydroxyethylene isostere showed similar in vitro activity as the P₁' Me analogue **9g**. The relative increase in IC₅₀ of **9g** in presence of plasma was surprisingly



Scheme 4. Reagents and conditions: (a) LiN(TMS)₂, THF, -78 °C, 77%; (b) NBS, 1:1 DME-H₂O, 0 °C to rt, 15 h, 74%; (c) NaN₃, DMPU, 40 °C, 18 h; **30** (30%) and **31** (60%); (d) H₂, cat. Pd/C 10%, BOC₂O, EtOAc, 82%; (e) H₂, cat. Pd/C 10%, (f) MsCl, Et₃N, CH₂Cl₂; (g) NaN₃, DMPU, 60 °C, 20 h, 95% (3 steps from **30**); (h) 1 M LiOH, 2:1 DME-H₂O, rt, 3 h; (i) TBDMSiCl, imidazole, DMF, >80% (2 steps); (j) HBTU, Et₃N, H₂NCH₂C(CH₃)₂CONH₂, DMF, 89%. (k) H₂, cat. Pd/C 10%, EtOAc; (l) acetone, NaBH₃CN, 4 Å MS, MeOH, 74%. (2 steps); (m) 4-methoxy-3-(3-methoxy-propoxy)-benzylbromide¹⁸, K₂CO₃, DMF, 50%; (n) Bu₄NF, THF, rt, 63%; (o) 4 N HCl-dioxane, rt, 90%.

more pronounced, in contrast to the SAR observed for the inhibitor **1** series.¹⁰

The sulfonamide P_3-P_1 spacer was explored with the aim to identify potent inhibitors with improved physicochemical properties including crystallinity and chemical stability in plasma (vide infra). Compound 14 with an IC_{50} of 0.3 μ M towards rh-renin was found to be greater than 100 times less potent compared to the carboxamide analogue 9a (Table 1). Compound 15 lacking the P₃^{sp} side chain did not show significant enzyme affinity up to 100 μ M. These results confirm previous findings,^{24,27} that binding interactions to the contiguous S_3-S_1 hydrophobic pocket of renin alone are not sufficient for strong enzyme affinity, and further suggest the methoxypropoxy group of inhibitor 14 to interact with the non-substrate P₃^{sp} binding site. Computational docking of **14** into the renin active site predicted a similar binding pose in an energy-minimized conformation as was observed by X-ray for inhibitor 1 complexed to rh-renin.^{9,10} The modeling pose suggested a Hbond of a sulfonyl oxygen to the flap Thr₇₇ NH, but also an unfavorable interaction of the sulfonyl with Gly₂₁₇. This could explain at least in part the marked drop in potency of 14 versus 9a.

Interestingly, the 3,4-dialkoxybenzyl substituted tertiary amine **10** showed high potency with similar affinities to recombinant and plasma renin (Table 1).²⁸ Compared to benzamide **9a**, compound **10** was only threefold less active in vitro in the physiologically more relevant plasma renin assay ($IC_{50}s$ of 14 vs 5 nM, respectively). The increased potency of **9a** over **10** was considered to be due to a H-bond interaction of the spacer carbonyl to Thr₇₇ of the flap domain in its closed conformation. Other contributing factors could be a potential intramolecular H-bond favoring a solution conformation of **9a** similar to its enzyme-bound conformation, or different desolvatation energies for **9a** and **10** required for binding into the solvent-shielded hydrophobic P_3-P_1 site of the catalytic cleft. To the best of our knowledge, **10** is the first reported low nanomolar inhibitor of renin, which incorporates a basic amine

at the P₁ position. This encouraging result prompted us, subsequent to the discovery of the highly potent development candidate **2** bearing an optimized P₂' carboxamide moiety¹¹, to explore its corresponding basic P₁ analogue **35**.²⁸ We envisaged that the P₁' isopropyl group of the dipeptide TSM, replacing the smaller Me group in **10**, would improve the in vitro enzyme inhibition, and in particular oral in vivo potency, as demonstrated previously for the lead series.¹¹ Indeed inhibitor **35** showed strong inhibition in both human renin in vitro assays with IC₅₀s of 7 nM. However, **35** was only equipotent to the P₁' Me analogue towards plasma renin, and more than 10-fold less active as compared to **2** (Table 1). The significant reduction in the potency of **35** limited its potential attractiveness for more extensive investigations.

We had established a primate in vivo pharmacology model in telemetered sodium-depleted marmosets (Callthrix jacchus) for non-invasive measurement of blood pressure (BP), heart rate, and motor activity in freely moving animals to study oral efficacy of renin inhibitors and other RAS blocking agents.²⁹ Inhibitor 9a, administered orally in saline solution at 3 mg/kg to normotensive marmosets pre-treated with furosemide (N = 5; 6 mg/kg/d for 1 week), caused a significant change in mean arterial pressure (MAP), compared to vehicle-treated animals, with a rapid onset of action and paralleled by partial inhibition of plasma renin activity (PRA). The maximal depressor effect Δ MAP of -18 mmHg observed 90 min post-dose was similar to ΔMAP_{max} of inhibitor 1 at the 3 mg/kg po dose in this model. However, and in marked contrast to the sustained BP lowering potency of **1** over up to 8 h,¹⁰ **9a** had only short-lasting effects, with complete recovery to baseline after 4 h post-dose. Total peak plasma exposure was found to be low (0.07 μ mol/L at 1.5 h post-dose). The inferior potency of 9a in vivo suggested a poor pharmacokinetics profile due to low oral absorption and/or rapid clearance in marmosets.

The chemical stability of inhibitors **9a** and **9g** in aqueous phosphate or Tris buffer solutions at pH 6.0, 7.4, and 8.0 was investigated. Both compounds showed significant decomposition up to 30% and 12%, respectively, after 24 h at 37 °C. A trans-acylation reaction to form the secondary amide **36** (Fig. 2) was found to be the major decomposition pathway under weakly acidic conditions, whereas in neutral or slightly basic solutions the formation of the dihydro-1,3-imidazole derivative **37**³⁰ was observed in addition. In DMSO at 100 °C, **9a** was converted exclusively into **36**³¹ within 1 h. **36** (rh-renin IC₅₀ = 0.67 μ M) was stable in aqueous buffers over a broad pH range. Additional studies are required to demonstrate whether the limited stability of **9a** in physiological buffer may at least partially contribute to the short duration of action in vivo in marmosets.

In summary, the synthesis, the chemical stability and in vitro SAR for inhibition of human renin has been explored for a novel series of 'symmetrized' hydroxyethylene dipeptide mimetics. From this work, the first low nanomolar renin inhibitors **10** and **35** with a basic tertiary amine at the P₁ position are described. The (P_3-P_1) -benzamide analogue **9a** showed potent but transient blood pressure lowering effects in Na-depleted marmosets. In subsequent studies as part of our continued renin inhibitor program, the *N*-isopropyl benzamide (P_3-P_1) -pharmacophore, as in **9a**, has been combined with a pyrrolidine-based center scaffold as TSM surrogate to afford highly potent candidate inhibitors with promising in vivo activity profiles.^{8,18} A full account of this more recent work will be disclosed in due course.

Acknowledgments

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- 16. Typical procedure for the preparation of 5a/5b: To a suspension of Zn(Cu) (43.3 g) in toluene (130 mL) was added a solution of 3-iodo-2(R)-methylpropionic acid methyl ester (92.5 g, 0.41 mol) in N,N-dimethyl acetamide (69 mL) and toluene (560 mL) at rt over 10 min. The suspension was stirred at rt for 1 h and then refluxed for 5 h. To a solution of Ti(OiPr)₄ (30.1 mL) in toluene (105 mL) and CH2Cl2 (530 mL) was added TiCl4 (33.3 mL) at <25 °C. The solution was stirred at rt for 15 min, followed by addition of the Znhomoenolate solution while keeping the temperature below -20 °C. Then, a solution of 4 (26.4 g, 0.09 mol) in CH2Cl2 (130 mL) was added below -35 °C, and the mixture was stirred at -20 °C for 2 days. Standard workup afforded a 4:1 mixture of **5a/5b** (27.7 g, 77%) as colorless oil.
- 17 Analytical data for 4(S)-6: TLC Rf 0.33 (hexane/EtOAc 1:1); mp 105 °C; ¹H NMR (400 MHz, DMSO- d_6): δ 7.65 (t, J = 8 Hz, 1H), 7.36–7.27 (m, 5H), 6.29 (d,

J = 12 Hz, 1H), 4.46 (s, 2H), 3.58-3.48 (m, 3H), 3.05-3.00 (m, 2H), 2.47-2.44 (m, 1H), 1.66–1.60 (m, 1H), 1.39 (s, 9H), 1.36–1.20 (m, 6H), 0.99 (d, J = 8 Hz, 3H), 0.85 (t, J = 8 Hz, 3H). HRMS [M+H]⁺ calcd 423.2854; found 423.2853.

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- Derivative 37, isolated as major side product from N-BOC-9a deprotection with 4 N HCl-dioxane at rt showed weak affinity to rh-renin (IC₅₀ = 1 μ M).
- The trans-acyl decomposition product 36 was synthesized according to the following reaction scheme:



Reaction conditions: (a) Cbz-Cl, Et₃N, CH₂Cl₂, 0 °C to rt, 18 h, (72%); (b) cat. p-TsOH, MeOH, 58%; (c) TFA, CH2Cl2, 0 °C to rt, 3 h, 75%; (d) 4-methoxy-3-(3methoxy-propoxy)-benzoic acid, HOBT, EDC, N-methylmorpholine, DMF, rt, 16 h, 78%; (e) H₂, cat. Pd/C, BOC₂O, MeOH, rt, 7 h, 89%. (f) 4 N HCl-dioxane, rt, 69%