

Preliminary communication

Synthesis and immunoadjuvant activities of novel *N*-acylmuramoyl dipeptides related to the lipid A constituent of the bacterial lipopolysaccharide*

MAKOTO KISO, YASUHIKO GOH, EIJI TANAHASHI, AKIRA HASEGAWA,
Department of Agricultural Chemistry, Gifu University, Kakamigahara, Gifu 504 (Japan)

HIROYUKI OKUMURA, and ICHIRO AZUMA

Institute of Immunological Science, Hokkaido University, Sapporo 060 (Japan)

(Received December 11th, 1980; accepted for publication, January 21st, 1981)

Bacterial cell-wall components have a variety of interesting immunomodulating activities. In recent years, much progress² has been made regarding the synthetic peptidoglycan derivatives as strong immunostimulants.

In our continuing effort³ to elucidate the relationships between the structure of the carbohydrate moiety and the biological activity of muramoyl dipeptide (MDP)**, which is the minimal, adjuvant-active structure of bacterial, cell-wall peptidoglycan⁴, we have demonstrated that not only is the sugar moiety essential for adjuvant activity, but also that the combining site of the lactoyl-dipeptide moiety and its configuration are critically important^{3e}. However, the sugar skeleton (2-amino-2-deoxy-D-glucose) was exchangeable with 2-amino-2-deoxy-D-mannose^{3a} or -D-galactose^{3e}, while retaining the activity. In addition to these findings, it has been shown that the 6-hydroxyl group can be replaced by an amino or acylamino group^{3c,d}, but that the activity is decreased dramatically by deoxygenation^{1,3c,d}, deamination^{3b}, or halogenation¹. Moreover, the 2-acetamido group could be replaced by a free amino^{3b}, a methylamino⁵, or a hydroxyl^{3b} group without decreasing the activity.

We have recently developed⁶ a new synthetic route to analogs of the lipid A component of the bacterial lipopolysaccharide (LPS), in order to elucidate the relationships between chemical structure and biological activity. The preliminary investigation⁷ revealed that 2-deoxy-2-(D-3-hydroxytetradecanoylamino)-D-glucose and one of its β -(1 \rightarrow 6)-linked disaccharide derivatives had antitumor, as well as *Limulus*-lysate gelation, activities. In view of the fact that the synthetic, lipophilic MDP derivatives showed potent antitumor and anti-infection activities² that were not found for MDP itself, our

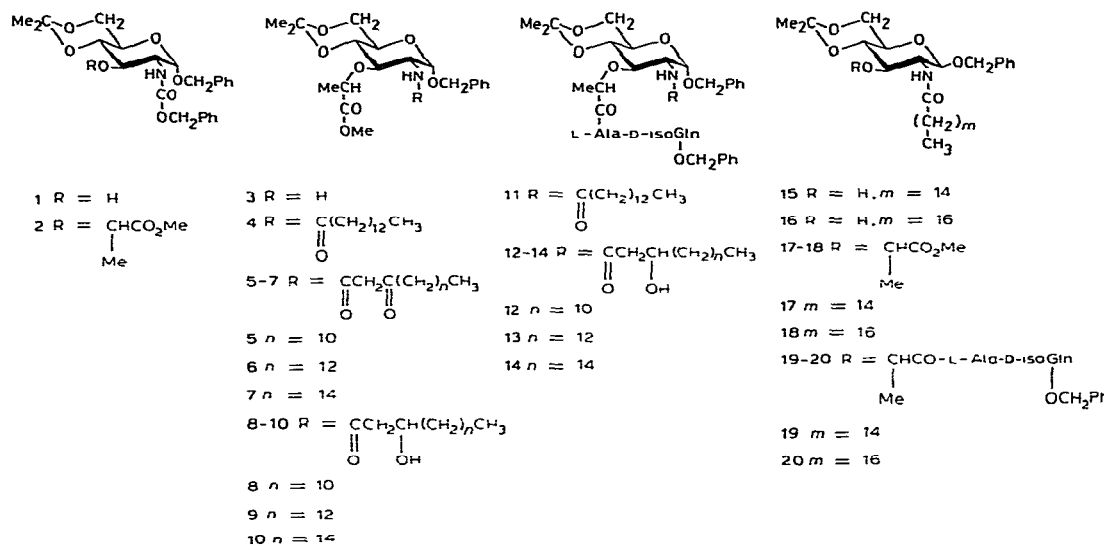
*Studies on Immunoadjuvant Active Compounds, Part XII. For Part XI, see ref. 1.

**A summary of this study, involving the biological investigation of the synthetic, repeating disaccharide–dipeptide units^{3f}, was presented at the Xth International Symposium on Carbohydrate Chemistry, Sydney, Australia, in July 1980.

interest has been directed to the synthesis of new, lipophilic, MDP analogs related to the lipid A constituent of LPS. We now describe the synthesis of novel *N*-acylmuramoyl dipeptides, and their immunoadjuvant activities on the induction of delayed-type hypersensitivity in guinea-pigs.

Benzyl 2-(benzyloxycarbonylamino)-2-deoxy-4,6-*O*-isopropylidene- α -D-glucopyranoside (**1**), a syrup, $[\alpha]_D^{25} +99.8^\circ$ (*c* 0.5, chloroform), was prepared by (benzyloxy-carbonyl)ation of benzyl 2-amino-2-deoxy-4,6-*O*-isopropylidene- α -D-glucopyranoside, m.p. 144–148° (dec.), $[\alpha]_D^{25} +123.2^\circ$ (*c* 1.11, chloroform), which can be obtained almost quantitatively by treatment of the corresponding 2-acetamido derivative^{3a} with hydrazine hydrate under reflux. Compound **1** can also be synthesized by direct acetalation of benzyl 2-(benzyloxycarbonylamino)-2-deoxy- α -D-glucopyranoside⁸ with 2,2-dimethoxypropane in dry 1,4-dioxane containing a trace of *p*-toluenesulfonic acid⁹.

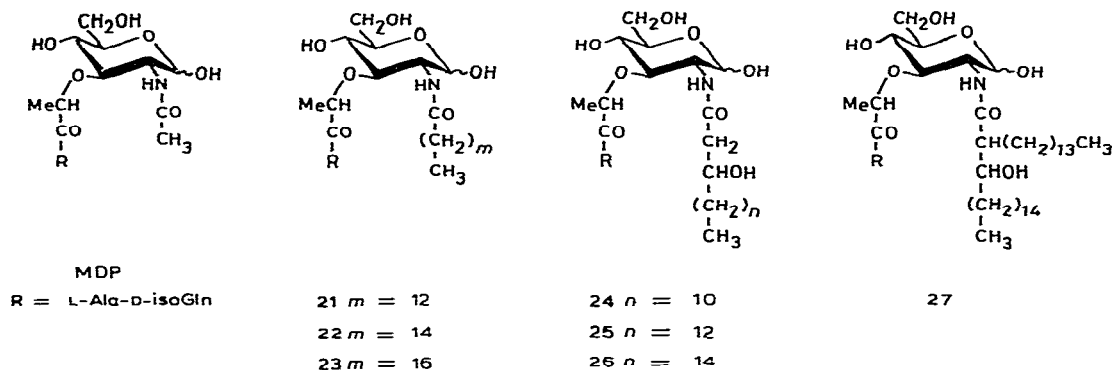
For the synthesis of the *N*-hexadecanoyl or *N*-octadecanoyl derivative, we employed the corresponding benzyl 2-(acylamino)-2-deoxy-4,6-*O*-isopropylidene- β -D-glucopyranoside as the starting material: **15** (*m* = 14), m.p. 114–117°, $[\alpha]_D^{25} -67.4^\circ$ (*c* 0.5, chloroform); and **16** (*m* = 16), m.p. 118°, $[\alpha]_D^{25} -65.5^\circ$ (*c* 1, chloroform), which were prepared stepwise by *O*-deacetylation of the benzyl 3,4,6-tri-*O*-acetyl-2-(acylamino)-2-deoxy- β -D-glucopyranosides^{6a}, and subsequent 4,6-*O*-isopropylidenation as just described.



The isopropylidene derivatives **1**, **15**, and **16** were condensed with L-2-chloropropionic acid in the presence of sodium hydride, to give 3-*O*-(D-1-carboxyethyl) derivatives, which were converted into the methyl esters by treatment with diazomethane in ether solution: **2**, m.p. 136–137°, $[\alpha]_D^{25} +83.2^\circ$ (*c* 0.202, chloroform); **17**, m.p. 108–110°, $[\alpha]_D^{25} -28.8^\circ$ (*c* 1.5, chloroform); and **18**, m.p. 96–97°, $[\alpha]_D^{25} -31.2^\circ$ (*c* 0.5, chloroform). The benzyloxycarbonyl group of **2** was removed by brief hydrogenation

(30–40 min) in methanol in the presence of 10% palladium–carbon catalyst, to afford 3, which was converted, by treatment with *N*-(tetradecanoyloxy)succinimide or *N*-(3-oxo-tetra-, -hexa-, or -octa-decanoyloxy)succinimide, into the corresponding *N*-fatty acyl derivatives: 4 (80%), m.p. 45–47°, $[\alpha]_D +76.4^\circ$ (*c* 0.2, chloroform); and 5–7 (55–70%) as amorphous materials. Compounds 5–7 were dissolved in methanol, and treated with sodium borohydride at 0°, to give 3-hydroxy-fatty acyl derivatives 8–10 in quantitative yields. After de-esterification of 4 and 8–10 with 0.1M aqueous potassium hydroxide in 1,4-dioxane, the acids formed were coupled with the benzyl ester of L-alanyl-D-isoglutamine, using dicyclohexylcarbodiimide and *N*-hydroxysuccinimide as activating agents, to yield the protected *N*-acylmuramoyl dipeptides as amorphous materials: 11, $[\alpha]_D +60.9^\circ$ (*c* 0.33, chloroform); 12 (*n* = 10), $[\alpha]_D +50.6^\circ$ (*c* 1.1, chloroform); 13 (*n* = 12), $[\alpha]_D +54.5^\circ$ (*c* 0.59, chloroform); and 14 (*n* = 14), $[\alpha]_D +45.0^\circ$ (*c* 0.69, chloroform). Similar treatment of 17 and 18 gave 19 (*m* = 14), $[\alpha]_D -7.62^\circ$ (*c* 1.94, chloroform) and 20 (*m* = 16), $[\alpha]_D -8.3^\circ$ (*c* 0.7, chloroform), respectively.

Hydrolytic removal of the 4,6-*O*-isopropylidene group from 11, 12–14, 19, and 20 with 70% acetic acid, and subsequent hydrogenation in the presence of 10% palladium–carbon catalyst, gave the desired (*N*-acylmuramoyl)-L-alanyl-D-isoglutamines as amorphous materials after lyophilization: 21 (*m* = 12), $[\alpha]_D +12.2^\circ$ (*c* 0.34, methanol); 22 (*m* = 14), $[\alpha]_D +13.0^\circ$ (*c* 0.2, water); 23 (*m* = 16), $[\alpha]_D +15.3^\circ$ (*c* 0.4, water); 24 (*n* = 10), $[\alpha]_D +19.7^\circ$ (*c* 0.46, methanol); 25 (*n* = 12), $[\alpha]_D +23.7^\circ$ (*c* 0.95, methanol); and 26 (*n* = 14), $[\alpha]_D +6.8^\circ$ (*c* 0.205, methanol). In order to clarify the effect of the



alkyl-chain length of the *N*-acyl moiety, we also prepared 2-deoxy-2-(3-hydroxy-2-tetradecyl-octadecanoylamino)-3-*O*-(D-2-propionyl-L-alanyl-D-isoglutamine)-D-glucopyranose (27), amorphous, $[\alpha]_D +11.3^\circ$ (*c* 0.2, methanol, equil.), starting from the 3-hydroxy-2-tetradecyl-octadecanoyl¹⁰ derivative of compound 3.

The immunoadjuvant activities of 21–27 on the induction of delayed-type hypersensitivity to azobenzenearsonate-*N*-acetyl-L-tyrosine (ABA-*N*-acetyl-L-tyrosine)¹¹ in guinea-pigs were examined. As shown in Table I, all of the synthetic MDP analogs exhibited potent activity. Compound 21 showed not only induration, but also a clear necrotic reaction.

TABLE I

ADJUVANT ACTIVITY OF NOVEL *N*-ACYLMURAMOYL DIPEPTIDES (21–27) ON THE INDUCTION OF DELAYED-TYPE HYPERSENSITIVITY TO ABA-*N*-ACETYL-L-TYROSINE IN GUINEA-PIGS

Compound ^a	Skin reaction ^b with ABA-BSA (50 µg) (diam. in mm ±SE) ^c at	
	24 h	48 h
21	21.0 ±1.2	19.4 ±0.8
22	20.8 ±0.4	18.3 ±0.4
23	21.0 ±0.8	16.5 ±0.6
24	21.1 ±1.1	21.1 ±0.8
25	20.5 ±0.4	17.8 ±0.6
26	20.1 ±1.0	19.5 ±1.1
27	21.5 ±1.0	18.0 ±0.7
<i>N</i> -Acetylmuramoyl-L-alanyl-D-isoglutamine (MDP)	20.3 ±0.8	19.8 ±0.9
Control ^d	0	0

^a At a dose of 100 µg. ^b Four animals. ^c The data indicate the average diameter ± standard-error (SE) of the skin reaction (induration); ABA-BSA, azobenzenearsonate-*N*-acetyl-L-tyrosine-bovine serum albumin. ^d ABA-*N*-acetyl-L-tyrosine in Freund's incomplete adjuvant (FIA), or FIA alone.

REFERENCES

- 1 A. Hasegawa, E. Tanahashi, Y. Goh, M. Kiso, and I. Azuma, *Carbohydr. Res.*, 92 (1981) in press.
- 2 E. Lederer, *J. Med. Chem.*, 23 (1980) 819–825.
- 3 (a) A. Hasegawa, Y. Kaneda, M. Amano, M. Kiso, and I. Azuma, *Agric. Biol. Chem.*, 42 (1978) 2187–2189; (b) M. Kiso, Y. Kaneda, H. Okumura, A. Hasegawa, I. Azuma, and Y. Yamamura, *Carbohydr. Res.*, 79 (1980) C17–C19; (c) A. Hasegawa, H. Okumura, M. Kiso, I. Azuma, and Y. Yamamura, *ibid.*, 79 (1980) C20–C23; (d) A. Hasegawa, H. Okumura, M. Kiso, I. Azuma, and Y. Yamamura, *Agric. Biol. Chem.*, 44 (1980) 1301–1308, 1309–1313; (e) M. Kiso, Y. Kaneda, Y. Goh, A. Hasegawa, and I. Azuma, *ibid.*, 44 (1980) 1971–1973; (f) M. Kiso, Y. Kaneda, R. Shimizu, and A. Hasegawa, *Carbohydr. Res.*, 83 (1980) C8–C11.
- 4 (a) F. Ellouz, A. Adam, R. Ciorbaru, and E. Lederer, *Biochem. Biophys. Res. Commun.*, 59 (1974) 1317–1325; (b) C. Merser, P. Sinaÿ, and A. Adam, *ibid.*, 66 (1975) 1316–1322; (c) S. Kotani, Y. Watanabe, F. Kinoshita, T. Shimono, I. Morisaki, T. Shiba, S. Kusumoto, Y. Tarumi, and K. Ikenaka, *Biken J.*, 18 (1975) 105–111.
- 5 M. Kiso, Y. Kaneda, K. Nishihori, A. Hasegawa, H. Okumura, I. Azuma, and Y. Yamamura, unpublished results.
- 6 (a) M. Kiso, H. Nishiguchi, and A. Hasegawa, *Carbohydr. Res.*, 81 (1980) C13–C15; (b) M. Kiso, H. Nishiguchi, S. Murase, and A. Hasegawa, *ibid.*, 88 (1981) C5–C9; (c) M. Kiso, H. Nishiguchi, K. Nishihori, A. Hasegawa, and I. Miura, *ibid.*, 88 (1981) C10–C13; (d) M. Kiso, K. Nishihori, and A. Hasegawa, *Agric. Biol. Chem.*, 45 (1981) 545–548.
- 7 M. Kiso, H. Nishiguchi, A. Hasegawa, H. Okumura, and I. Azuma, *Agric. Biol. Chem.*, in press.
- 8 K. Heyns and H. Paulsen, *Chem. Ber.*, 88 (1951) 188–195.
- 9 A. Hasegawa and M. Kiso, *Carbohydr. Res.*, 79 (1980) 265–270; see also, many references cited therein.
- 10 S. Kusumoto, M. Inage, T. Shiba, I. Azuma, and Y. Yamamura, *Tetrahedron Lett.*, (1978) 4899–4902.
- 11 (a) I. Azuma, K. Kamisango, I. Saiki, Y. Tanio, S. Kobayashi, and Y. Yamamura, *Infect. Immun.*, 29 (1980) 1193–1196; (b) I. Azuma, H. Okumura, I. Saiki, Y. Tanio, M. Kiso, A. Hasegawa, and Y. Yamamura, *ibid.*, in press.