



Chemistry A European Journal

 **Chemistry
Europe**
European Chemical
Societies Publishing

Accepted Article

Title: A Buchwald-Hartwig Protocol to Enable Rapid Linker Exploration of Cereblon E3-Ligase PROTACs

Authors: Thomas G Hayhow, Rachel E. A. Borrow, Coura R. Diène, Gary Fairley, Charlene Fallan, Shaun M. Fillery, James S. Scott, and David W. Watson

This manuscript has been accepted after peer review and appears as an Accepted Article online prior to editing, proofing, and formal publication of the final Version of Record (VoR). This work is currently citable by using the Digital Object Identifier (DOI) given below. The VoR will be published online in Early View as soon as possible and may be different to this Accepted Article as a result of editing. Readers should obtain the VoR from the journal website shown below when it is published to ensure accuracy of information. The authors are responsible for the content of this Accepted Article.

To be cited as: *Chem. Eur. J.* 10.1002/chem.202003137

Link to VoR: <https://doi.org/10.1002/chem.202003137>

WILEY-VCH

A Buchwald-Hartwig Protocol to Enable Rapid Linker Exploration of Cereblon E3-Ligase PROTACs

Thomas G. Hayhow,^{*a} Rachel E. A. Borrows,^a Coura R. Diène,^a Gary Fairley,^b Charlene Fallan,^a Shaun M. Fillery,^a James S. Scott,^a and David W. Watson^a

[a] T. G. Hayhow, R. E. A. Borrows, C. R. Diène, C. Fallan, S. M. Fillery, J. S. Scott, D. W. Watson
Medicinal Chemistry, Oncology R&D, Research and Early Development
AstraZeneca
Cambridge Science Park, Unit 310 Darwin Building, Cambridge, CB4 0WG, United Kingdom
E-mail: thomas.hayhow@astrazeneca.com

[b] G. Fairley
Medicinal Chemistry, Oncology R&D, Research and Early Development
AstraZeneca
Macclesfield Campus, Etherow Building, Silk Road Industrial Estate, Macclesfield, SK10 2NA, United Kingdom

Supporting information for this article is given via a link at the end of the document.

Abstract: A palladium-catalysed Buchwald-Hartwig amination for lenalidomide-derived aryl bromides was optimised using high throughput experimentation (HTE). The substrate scope of the optimised conditions was evaluated for a range of alkyl- and arylamines and functionalised aryl bromides. The methodology allows access to new cereblon-based bifunctional proteolysis targeting chimeras with a reduced step count and improved yields.

Introduction

Immunomodulatory imide drugs (IMiDs) such as thalidomide (**1**) have been used since the late 1950s, including the treatment of nausea in pregnant women.^[1] While thalidomide was removed from the market in 1961 due to teratogenicity,^[2] it was subsequently found to be a useful treatment for leprosy.^[3] Interest in the immuno-modulatory properties of **1**^[4] resulted in further clinical studies and it was shown to be effective in the treatment of multiple myeloma (MM) in 1999 and approved for use in MM by the US Food and Drug Administration in 2006.^[2, 5] This success led to the development of more potent analogues such as lenalidomide (**2**) and pomalidomide (**3**), that were approved for MM in 2005 and 2013 respectively.^[6]

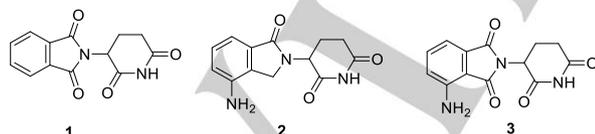


Figure 1. Structures of IMiD drugs thalidomide (**1**), lenalidomide (**2**) and pomalidomide (**3**).

The biological targets of the IMiD drugs were initially unknown. One protein that thalidomide binds to is cereblon (CRBN), which is believed to be responsible for the teratogenicity of the drug;^[2] at least partially driven by the degradation of SALL4^[7] and p63.^[8] CRBN associates with DBB1-CUL4A protein complex, an E3 ubiquitin ligase.^[9] One of the many functions of ubiquitin ligases

is the labelling of proteins for degradation by the 26S proteasome through linear ubiquitination of K48.^[10] In the last decade there has been considerable interest in hijacking the ubiquitination function of E3 ligases to target specific proteins for degradation. One widely used approach involves a heterobifunctional molecule joining an E3 ligase binder, such as CRBN, to another protein-of-interest (POI) binder, via a chemical linker. In many cases degradation of the POI is observed. These compounds are known as proteolysis targeting chimeras (PROTACs) and have used a number of different E3 ligases such as MDM2,^[11] Von Hippel-Lindau (VHL)^[12] and CRBN.^[13] CRBN based PROTACs are of considerable interest since IMiDs have the smallest molecular weight and the fewest hydrogen-bond donors (HBD) of the E3 ligase binders disclosed, which may improve the oral exposure of these new modalities.^[14]

CRBN-based PROTACs have been demonstrated to degrade a range of targets such as Brd4, for example dBET1 (**4**)^[13] and ARV-825 (**5**),^[15] in addition to multiple kinases with TL12-186 (**6**),^[16] Bruton's tyrosine kinase with P13I (**7**),^[17] and RIPK2 with PROTAC **3** (**8**) (Figure 2).^[18]

FULL PAPER

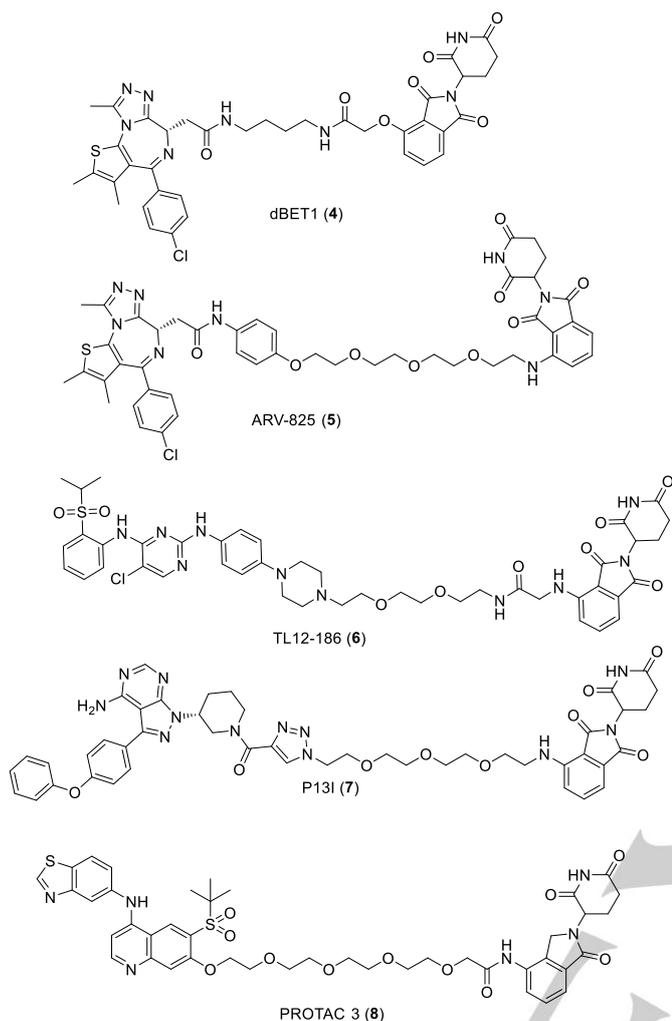


Figure 2. Structures of CRBN based PROTACs.

Published imide-based PROTACs include examples with linkers attached at either the 4- or 5-position of the phthalimide ring system. The compounds exemplified in Figure 2 have the linker attached at the 4-position of the IMiD. Within the patent literature there are an increasing number of examples with attachment at the 5-position of the IMiDs. Both C4 Therapeutics and Arvinas have attached the linkers of their PROTACs via the 5-position on both pomalidomide-based (**9**)^[19] and lenalidomide-based CRBN binders (**10**^[20] and **11**^[21]) (Figure 3).

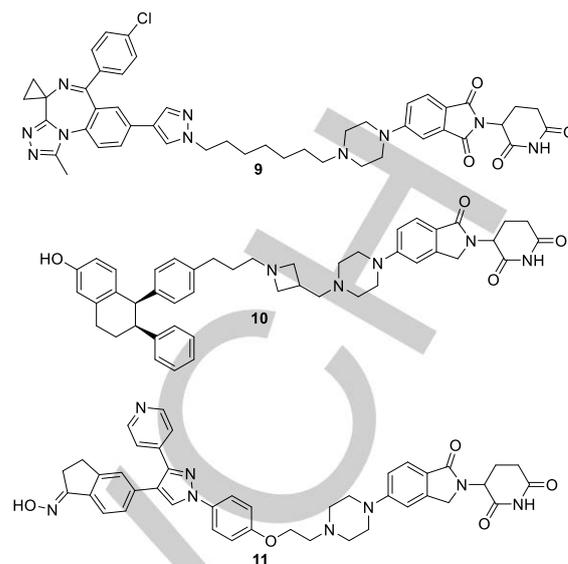
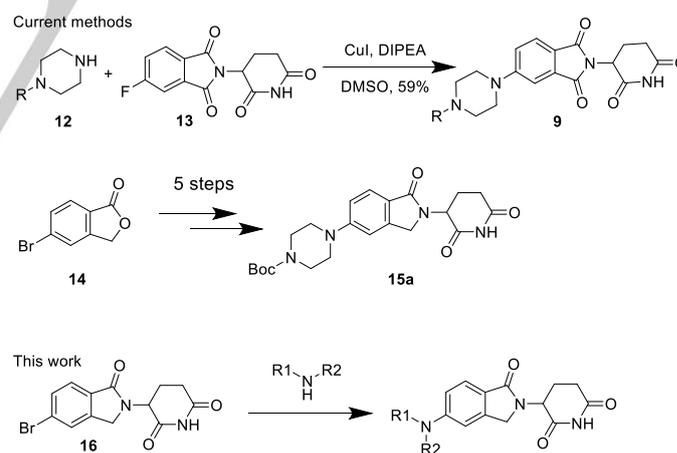


Figure 3. Structures of N-linked 5-substituted CRBN PROTACs.

The pomalidomide-based PROTACs, such as **9**, can be accessed in one step from 2-(2,6-dioxo-3-piperidyl)-5-fluoro-isoindoline-1,3-dione (**13**) using S_NAr methodology. Access to related lenalidomide-based PROTACs is less straightforward as nucleophilic aromatic substitution is no longer viable to introduce amines due to the reduced electrophilicity of the bicyclic ring system. Current methods involve a 5-step synthesis to key intermediate **15a**, with Boc-piperazine introduced in the first step and these lengthy syntheses hamper SAR exploration (Scheme 1).



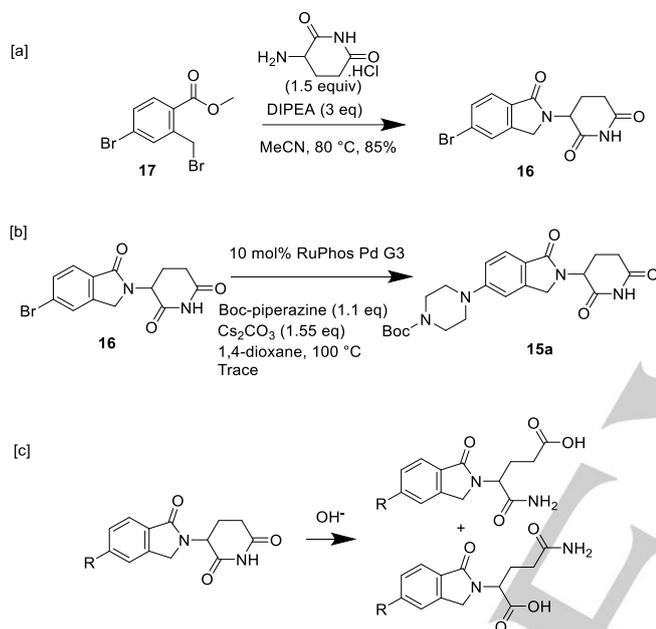
Scheme 1. Methods to synthesise CRBN binders.

To enable our medicinal chemistry exploration of lenalidomide based CRBN PROTACs, we aimed to develop a concise and robust synthetic route that would provide rapid access to a diverse set of PROTAC linkers for SAR development. Our strategy was to install the amine fragment using a Buchwald-Hartwig amination and crucially perform this key coupling as the last step of the sequence, rather than the first, to facilitate exploration of structural diversity.

FULL PAPER

Results and Discussion

The key Buchwald-Hartwig precursor, bromide **16**, was prepared in a single step from commercially available starting material **17** (Scheme 2a). With this in hand, palladium-catalysed Buchwald-Hartwig coupling conditions were examined on **16** using Boc-piperazine (**18**), cesium carbonate and RuPhos Pd G3 but disappointingly only trace amounts of desired product were obtained (Scheme 2b). Accurate analysis of the reaction profiles by LCMS required the use of an acidic diluent, 5% acetic acid in acetonitrile. Dissolution of the analytical samples in methanol caused significant impurities with masses corresponding to hydrolysis of the glutarimide ring, presumably from trace amounts of aqueous base (Scheme 2c).



Scheme 2. [a] Synthesis of key bromide starting material **16** [b] Trial Buchwald-Hartwig conditions [c] Hydrolysis of glutarimide by aqueous base.

High throughput experimentation (HTE) was used for the rapid identification of improved reaction conditions, using minimal quantities of intermediates. A screen was run to compare the original RuPhos Pd G3 catalysed conditions with variation of catalyst and ligand, base, and solvent (Table 1, full screen data available in SI, Tables S1 and S2). For the HTE optimisation, conversion to **15a** (Conv) was determined by comparison of the peak areas of **15a** and **16** where 100 indicates complete consumption of **16** (Equation S1 in SI). The product to internal standard ratio (P/IS) was used to determine the degree of conversion to **15a**. P/IS was calculated by the peak area of **15a** being divided by the peak area of the internal standard, *m*-terphenyl, and normalised to 1 for the highest ratio per screen. In order to determine the success of a reaction both conversion of **16** and P/IS were considered. In examples where conversion was high and P/IS was poor, the major by-products observed were the hydrolysed glutarimides of both the starting bromide and the aminated product (Scheme 2c) in addition to unidentified minor impurities.

Table 1. Investigation of catalysts, solvents and bases

| Entry [a] | Pd Catalyst Ligand | / | Base ^[b] | Solvent [c] | Conv (%) | P/IS 17 h |
|--------------|----------------------------------|---|------------------------------------|----------------|-------------------|--------------|
| 1 | RuPhos Pd G3 | | Cs ₂ CO ₃ | 1,4-dioxane | 22 ^[d] | N/A |
| 2 | RuPhos Pd G3 | | Cs ₂ CO ₃ | <i>t</i> -AmOH | 100 | 0.64 |
| 3 | RuPhos Pd G3 | | Cs ₂ CO ₃ | DMF | 62 | 0.61 |
| 4 | BrettPhos Pd G3 | | Cs ₂ CO ₃ | DMF | 0 | 0.00 |
| 5 | DavePhos Pd G3 | | Cs ₂ CO ₃ | DMF | 20 | 0.18 |
| 6 | Pd(OAc) ₂ XantPhos | / | Cs ₂ CO ₃ | DMF | 9 | 0.03 |
| 7 | Pd-PEPPSI-IPent | | Cs ₂ CO ₃ | DMF | 100 | 1.00 |
| 8 | RuPhos Pd G3 | | K ₃ PO ₄ | 1,4-dioxane | 0 ^[d] | N/A |
| 9 | RuPhos Pd G3 | | NaO ^t Bu ^[e] | 1,4-dioxane | 100 | 0.45 |
| 10 | RuPhos Pd G3 | | LiHMDS ^[e] | 1,4-dioxane | 74 | 0.41 |

[a] ArBr (1 eq), *N*-Boc-piperazine (1.3 eq), cat / L (10 mol%), 80 °C, glove box [b] 3 eq unless otherwise stated, [c] 30 volumes, [d] 21 hours [e] 2 eq.

The initial reaction conditions afforded partial conversion to product after 21 hours (Entry 1). Switching to more polar solvents, *tert*-amyl alcohol and DMF, enabled more rapid consumption of bromide **16** than in 1,4-dioxane (Entries 2 and 3). Solubility studies revealed that **16** is fully soluble in DMF and sparingly soluble in *tert*-amyl alcohol at 80 °C in 30 volumes of solvent (SI Table S4). Solubility was likely not a contributing factor to the rate of reaction. Entries 3–7 compared a variety of catalysts and while most saw some conversion, BrettPhos Pd G3 was unable to catalyse the desired reaction (Entry 4). DavePhos Pd G3 (Entry 5) and Pd(OAc)₂ / Xantphos (Entry 6) showed less conversion after 17 hours than RuPhos Pd G3. The standout result used Pd-PEPPSI-IPent an *N*-heterocyclic carbene containing ligand,^[22] which showed complete conversion after 17 hours and with the greatest P/IS (Entry 7). Using K₃PO₄ as a base gave no desired product (Entry 8). The use of stronger bases gave high levels of conversion compared to Cs₂CO₃, although modest P/IS (Entries 9 and 10).

Table 2. Optimisation with Pd-PEPPSI-IPent

| Entry [a] | Base ^[b] | Solvent [c] | Conv 3 h (%) | P/IS 3 h | Conv 21 h (%) | P/IS 21 h |
|------------------|------------------------------------|--------------------|-----------------|-------------|------------------|--------------|
| 1 | Cs ₂ CO ₃ | DMF | 34 | 0.72 | 100 | 0.85 |
| 2 ^[d] | Cs ₂ CO ₃ | DMF | 31 | 0.70 | 100 | 0.96 |
| 3 ^[e] | Cs ₂ CO ₃ | DMF | 30 | 0.67 | 100 | 0.93 |
| 4 | NaO ^t Bu ^[f] | DMF | 100 | 0.65 | 100 | 0.63 |
| 5 | Cs ₂ CO ₃ | DMF ^[g] | 89 | 0.96 | 100 | 0.57 |
| 6 | Cs ₂ CO ₃ | DMF ^[h] | 93 | 0.98 | 95 | 1.00 |

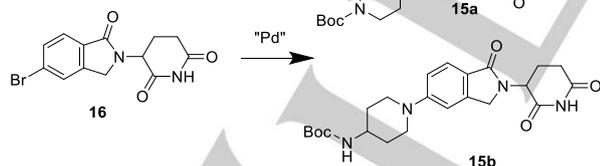
FULL PAPER

| | | | | | | |
|----|---------------------------------|--------------------|-----|------|-----|------|
| 7 | Cs ₂ CO ₃ | 2-MeTHF | 74 | 0.91 | 89 | 0.74 |
| 8 | Cs ₂ CO ₃ | 1,4-dioxane | 73 | 1.00 | 81 | 0.63 |
| 9 | Cs ₂ CO ₃ | CH ₃ CN | 13 | 0.37 | 100 | 0.48 |
| 10 | Cs ₂ CO ₃ | ^t AmOH | 100 | 0.93 | 100 | 0.41 |

[a] ArBr (1 eq), *N*-Boc-piperazine (1.3 eq), Pd-PEPPSI-IPent (10 mol%), 80 °C, glove box [b] 3 eq unless otherwise stated [c] 30 vols unless otherwise stated, [d] 2 mol% cat, [e] 5 mol% cat, [f] 2 eq NaO^tBu, [g] 20 vols, [h] 10 vols.

Given the excellent conversion and P/IS using Pd-PEPPSI-IPent, conditions were further optimised using this catalyst (Table 2, selected examples, full screen data available in SI Table S3). Reducing the level of catalyst loading from 10 mol% to 5 mol% or 2 mol% had no significant effect on reaction progression (Entries 1, 2 and 3). Use of strong base NaO^tBu showed complete consumption of starting material within 3 hours, although at a reduced P/IS, suggesting side reactions also occurring (Entry 4). Running the reaction at higher concentration (Entries 5 and 6) increased conversion compared to 30 volumes at 3 hours (Entry 1). A variety of solvents afforded good consumption of starting material, although P/IS ratios were varied, especially on prolonged heating (Entries 7 – 10).

On scaling up the conditions optimised by HTE and performing the reaction outside of a glovebox, it was found that the temperature needed to be increased to >90 °C for efficient cross-coupling to take place. Additionally, using DMF as a solvent made extraction during aqueous work ups more difficult compared with 1,4-dioxane. Therefore, we sought to further optimise the amination with a focus on nucleophile diversity by assessing a range of similar, highly sterically hindered PEPPSI precatalysts (Scheme 4). Pd-PEPPSI-IHept,^[23] Pd-PEPPSI-IPent^{Cl},^[24] Pd-PEPPSI-IHept^{Cl},^[25] and (DiMeIHept^{Cl})Pd(cinnamyl)Cl^[26] with both Boc-piperazine (**18**) and (*N*-Boc-4-amino)piperidine (**19**) were evaluated (Table 3).



Scheme 3. Reaction with Boc-piperazine (**18**) and (*N*-Boc-4-amino)piperidine (**19**).

Table 3. Variation of PEPPSI catalyst and substrates

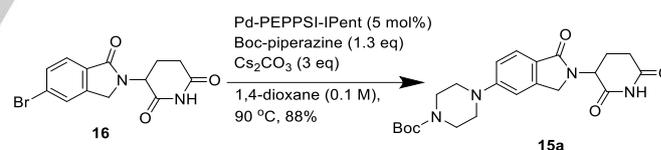
| Entry ^[a] | Pd-PEPPSI Catalyst | Substrate | Product | Yield (%) ^[b] |
|----------------------|--------------------|-----------|------------|--------------------------|
| 1 | IPent | 18 | 15a | 44 |
| 2 | IPent | 19 | 15b | 51 |

| | | | | |
|----|-------------------------|-----------|------------|----|
| 3 | IHept | 18 | 15a | 42 |
| 4 | IHept | 19 | 15b | 33 |
| 5 | IPent ^{Cl} | 18 | 15a | 63 |
| 6 | IPent ^{Cl} | 19 | 15b | 67 |
| 7 | IHept ^{Cl} | 18 | 15a | 70 |
| 8 | IHept ^{Cl} | 19 | 15b | 76 |
| 9 | IHept ^{Cl[e]} | 18 | 15a | 29 |
| 10 | DiMeIHept ^{Cl} | 18 | 15a | 38 |
| 11 | DiMeIHept ^{Cl} | 19 | 15b | 35 |

[a] ArBr (1 eq), amine (1.3 eq), catalyst (5 mol%), cesium carbonate (3 eq) 1,4-dioxane (0.1 M), 100 °C, [b] Isolated yield, [c] 2.5 mol% catalyst.

These small scale reactions demonstrated the clear superiority of the more electron-deficient PEPPSI^{Cl} catalysts compared to the ligands without the Cl on the backbone with both amines (Entries 1 – 4 vs 5 – 8). We attribute the lower conversions using DiMeIHept^{Cl} (Entries 10 and 11) compared to the IPent^{Cl} and IHept^{Cl} to the increased and less flexible steric bulk of the ligand. On the basis of achieving higher isolated yields with Pd-PEPPSI-IHept^{Cl}, this catalyst was chosen for further investigation. Attempts were made to lower the catalyst loading from 5 mol%, and while desired products were observed using these conditions, they did not enable full consumption of the starting material (Entry 9). Therefore, we identified our optimised conditions as 5 mol% Pd-PEPPSI-IHept^{Cl}, 3 equivalents of dried cesium carbonate in degassed dioxane at 0.1 M with 1.3 equivalents of amine (Entries 7 and 8).

To demonstrate the utility of the Buchwald-Hartwig approach to produce large-scale quantities of intermediates, the coupling was performed on 19 g of bromide **16**, affording **15a** in an excellent 88% isolated yield (Scheme 4).^[27]



Scheme 4. Large scale preparation of **15a**.

With our newly developed conditions in hand, we were keen to expand the substrate scope of the amination beyond **15a** and **15b**. Using the optimised Pd-PEPPSI-IHept^{Cl} conditions a wide variety of amines were found to be competent in the cross-coupling with bromide **16**. Complete consumption of the starting material usually allowed isolation of the clean products by trituration after 3 hours (Figure 4).

FULL PAPER

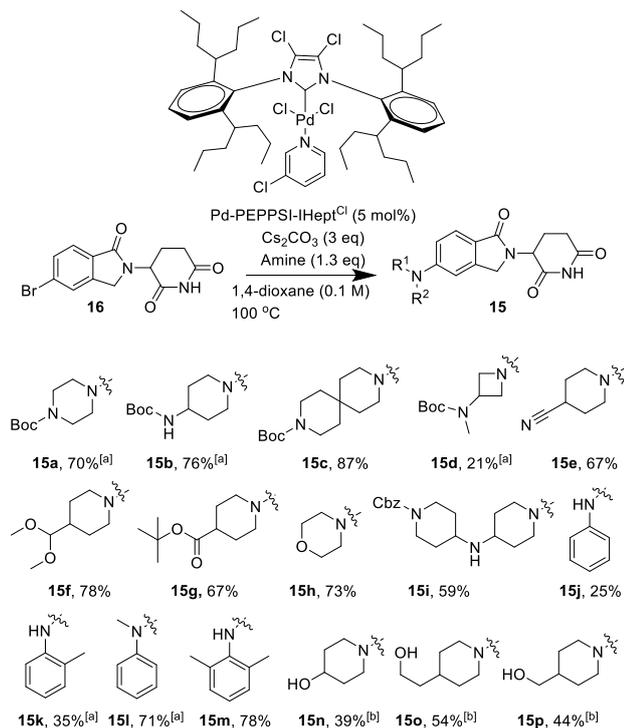


Figure 4. Substrates allowing full conversion, isolated by trituration unless otherwise stated. [a] Isolated by column chromatography, [b] 3 eq of amine.

Beyond Boc-protected amines (**15a-d**), amines bearing nitrile (**15e**), acetal (**15f**), ester (**15g**) and ether (**15h**) substituents were excellent coupling partners in this transformation. A di-basic substrate formed desired product (**15i**) coupling via the less sterically hindered nitrogen. In addition, anilines, including primary anilines and *ortho*-substituted anilines (**15j-m**) resulted in full conversion of bromide **16**. Alcohol containing amines proved more challenging. However, increasing the amine loading from 1.3 to 3 equivalents allowed for full conversion to the alcohol containing desired products (**15n-p**).

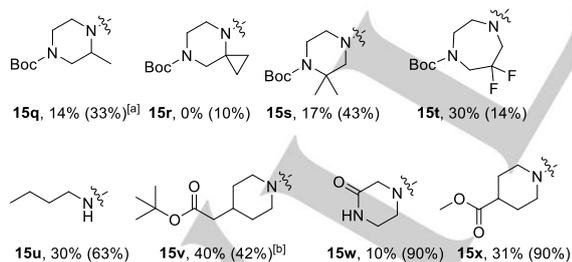


Figure 5. Substrates showing partial conversion. First value is isolated yield, value in parentheses is LCMS conversion [a] 3 eq of amine, Pd-PEPPSI-IHeptCl (5 mol%), [b] Pd-PEPPSI-IHeptCl (10 mol%).

More sterically hindered amines also proved challenging to couple. The alpha-methylpiperazine **15q** underwent poor conversion using Pd-PEPPSI-IHeptCl and was eventually made in sufficient quantities for isolation by increasing the amine to 3 equivalents and using Pd-PEPPSI-IHeptCl, consistent with our hypothesis that the increased conversion of the bulky amine is facilitated by using a less sterically hindered catalyst. Bulky

substitution around the piperazine ring greatly reduced the conversion of bromide **16** to the desired products (**15r** and **15s**). Boc-3,3-difluorohomopiperazine underwent limited coupling to **15t**, presumably affected by the reduced nucleophilicity of the amine. Butylamine allowed partial conversion to **15u**. Unexpectedly, the *tert*-butyl ester gave only 42% conversion to **15v**, significantly lower than the other ester examples **15g** and **15x**. The coupling with piperazinone demonstrated a single product **15w**, coupling *via* the amine moiety rather than the amide. The conversion of **16** to **15w** was efficient, if not complete; isolation of the final product was difficult due to the hydrophilicity of the compound. Similarly, the final yield of **15x** was lower than expected given the conversion from **16**.

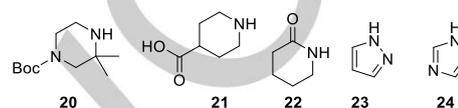
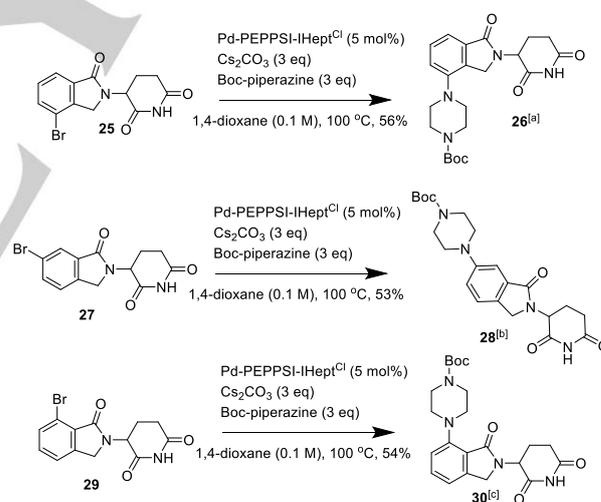


Figure 6. Substrates that gave no desired product.

Not all substrates showed conversion or gave desired products. Gem-dimethyl piperazine (**20**) was too sterically hindered to undergo reaction. Similarly, Buchwald-Hartwig couplings with piperidine-4-carboxylic acid (**21**), piperidinone (**22**), pyrazole (**23**) and imidazole (**24**) showed no evidence for the desired products.



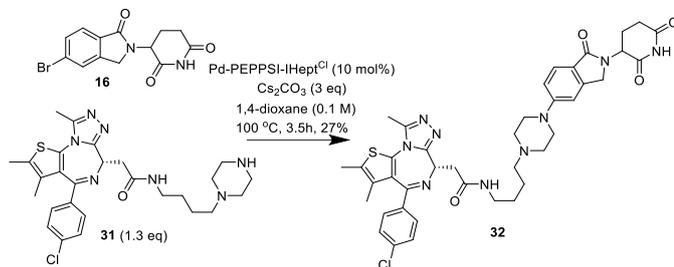
Scheme 5. Reaction of other bromoisindolinones. [a] 16 hours, [b] 5 hours, [c] recharged with catalyst after 24 hours and full conversion 2 hours subsequently.

Next, we investigated the Pd-catalyzed coupling of Boc-piperazine to other bromoisindolinones (Scheme 5). These gave desired products in moderate yields using our standard conditions and 3 equivalents of the amine, allowing exploration of vectors from all of the aromatic positions of the isoindolinone (**26**, **28** and **30**).

As a final test of the reaction conditions, we sought to make a potential PROTAC by introducing the isoindolinone E3-binder as a final step. A complex functionalised amine linker attached to JQ1(+) **31** was prepared. Using our standard conditions, coupling of **31** with bromoisindolinone **16** was undertaken in 3.5

FULL PAPER

hours to provide a complete PROTAC-like molecule **32**.^[29] This, to the best of our knowledge, is the first example of introducing a lenalidomide-like E3 ligase binder as a final step using a palladium-catalysed Buchwald-Hartwig amination protocol (Scheme 6).



Scheme 6. Buchwald-Hartwig coupling as a final step to synthesise a complete PROTAC.

Conclusion

We have reported a general method for the palladium-catalysed Buchwald-Hartwig coupling of amines to 3-(5-bromo-1-oxoisindolin-2-yl)piperidine-2,6-dione (**16**) with a wide amine substrate scope. Additionally, we have demonstrated the ability to substitute all positions around the aromatic ring of the isindolinone. Using these reaction conditions, we introduced for the first time a lenalidomide-based CRBN-binding moiety as a final step in the preparation of a potential PROTAC. We believe this methodology has utility in allowing rapid access to increasingly varied linkers to help identify PROTACs for *in-vitro* and *in-vivo* degradation of proteins through late stage diversification.

Experimental Section

Exemplar synthesis of *tert*-butyl (1-(2-(2,6-dioxopiperidin-3-yl)-1-oxoisindolin-5-yl)piperidin-4-yl)carbamate (**15b**):

Keywords: Amination • Cereblon • Drug Discovery • PEPPSI • PROTAC

...

Pd-PEPPSI-IHeptCl (0.030 g, 0.030 mmol) was added to 3-(5-bromo-1-oxoisindolin-2-yl)piperidine-2,6-dione (**16**) (0.2 g, 0.62 mmol), cesium carbonate (0.605 g, 1.86 mmol) and 4-(*N*-Boc-amino)piperidine (**19**) (0.161 g, 0.80 mmol) in 1,4-dioxane (6 mL) at 20 °C under nitrogen. The resulting mixture was vacuum degassed, backfilling with nitrogen and stirred at 100 °C for 3 hours. The reaction mixture was diluted with DCM (15 mL) and washed sequentially with 5% AcOH in water (10 mL) and saturated brine (10 mL). The organic layer was dried with MgSO₄, filtered and evaporated to afford crude product. The crude product was purified by flash silica chromatography, elution gradient 0 to 5% MeOH in DCM. Pure fractions were evaporated to dryness to afford the title compound (0.208 g, 76%) as a beige powder. ¹H NMR (500 MHz, DMSO-d₆): δ= 10.93 (s, 1H), 7.49 (d, *J*=8.5 Hz, 1H), 7.09 – 6.98 (m, 2H), 6.84 (d, *J*=7.5 Hz, 1H), 5.04 (dd, *J*=13.3, 5.1 Hz, 1H), 4.31 (d, *J*=16.9 Hz, 1H), 4.19 (d, *J*=16.9 Hz, 1H), 3.82 (d, *J*=13.1 Hz, 2H), 3.53 – 3.41 (m, 1H), 2.90 (t, *J*=12.7 Hz, 3H), 2.58 (dd, *J*=14.4, 3.1 Hz, 1H), 2.41 – 2.30 (m, 1H), 1.99 – 1.92 (m, 1H), 1.79 (d, *J*=10.4 Hz, 2H), 1.47 – 1.35 (m, 11H); ¹³C NMR (126 MHz, DMSO-d₆): δ=172.9, 171.3, 168.3, 154.9, 153.4, 144.1, 123.8, 120.9, 114.8, 108.4, 77.5, 51.4, 47.4, 46.9, 46.9, 31.3, 31.0, 28.3, 22.6; IR (ATR): 3361 (br), 3213 (br), 1676 (s), 1662 (s), 1614 (m), 1318 (m), 1225 (m), 770 (w) cm⁻¹; HR-MS(ESI): *m/z* calculated for C₂₃H₃₀N₄O₅ [M+H]⁺:443.2289, found: 443.2279.

Full experimental details can be found in the Supporting Information.

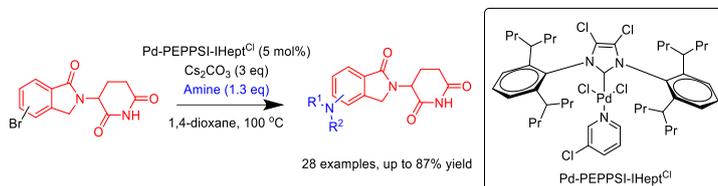
Acknowledgements

The authors are thankful for the advice of Prof Michael G. Organ on the storage and handling of PEPPSI catalysts. We express our gratitude to the analytical and structural chemistry group within AstraZeneca Oncology Chemistry, in particular to Paul Davey for the high resolution mass spectrometry, Eva Lenz for assistance with NMR spectrometry, and Daniel Rhodes and Joanna Raubo for the optical rotations.

Conflict of interest

The authors declare no conflict of interest.

Entry for the Table of Contents



PROTAC assembly: A method to introduce diverse amines to lenalidomide-like scaffolds is described. The products from the Buchwald-Hartwig cross couplings could be used to access a range of PROTACs in a short number of subsequent steps. The coupling is effective from milligram to multigram scale.

- [1] J. H. Kim, A. R. Scialli, *Toxicol. Sci.* **2011**, *122*, 1-6.
- [2] T. Ito, H. Ando, H. J. C. Handa, M. L. *Cell. Mol. Life Sci.*, **2011**, *68*, 1569-1579.
- [3] J. Sheskin, *Clin. Pharmacol. Ther.* **1965**, *6*, 303-306.
- [4] a) O. Gutierrez-Rodriguez, *Arthritis Rheum.* **1984**, *27*, 1118-1121; b) M. H. Hamza, *Clin. Rheumatol.* **1986**, *5*, 365-371; c) D. M. McCarthy, E. J. Kanfer, A. J. Barrett, *Biomed. & Pharmacother.* **1989**, *43*, 693-697.
- [5] S. Singhal, J. Mehta, R. Desikan, D. Ayers, P. Roberson, P. Eddlemon, N. Munshi, E. Anaisie, C. Wilson, M. Dhodapkar, J. Zeddis, B. Barlogie, *N. Engl. J. Med.* **1999**, *341*, 1565-1571.
- [6] a) X. Armoiry, G. Aulagner, T. Facon, *J. Clin. Pharm. Ther.* **2008**, *33*, 219-226; b) R. Rios-Tamayo, A. Martin-Garcia, C. Alarcon-Payer, D. Sanchez-Rodriguez, A. de la Guardia, C. G. Garcia Collado, A. Jimenez Morales, M. Jurado Chacon, J. Cabeza Barrera, *Drug Des. Dev. Ther.* **2017**, *11*, 2399-2408.
- [7] K. A. Donovan, J. An, R. P. Nowak, J. C. Yuan, E. C. Fink, B. C. Berry, B. L. Ebert, E. S. Fischer, *eLife* **2018**, *7*.
- [8] T. Asatsuma-Okumura, H. Ando, M. De Simone, J. Yamamoto, T. Sato, N. Shimizu, K. Asakawa, Y. Yamaguchi, T. Ito, L. Guerrini, H. Handa, *Nat. Chem. Biol.* **2019**, *15*, 1077-1084.
- [9] S. Angers, T. Li, X. Yi, M. J. MacCoss, R. T. Moon, N. Zheng, *Nature* **2006**, *443*, 590-593.
- [10] J. S. Thrower, L. Hoffman, M. Rechsteiner, C. M. Pickart *EMBO J.* **2000**, *19*, 94-102.
- [11] A. R. Schneekloth, M. Puchault, H. S. Tae, C. M. Crews, *Bioorg. Med. Chem. Lett.* **2008**, *18*, 5904-5908.
- [12] D. P. Bondeson, A. Mares, I. E. Smith, E. Ko, S. Campos, A. H. Miah, K. E. Mulholland, N. Routly, D. L. Buckley, J. L. Gustafson, N. Zinn, P. Grandi, S. Shimamura, G. Bergamini, M. Faeth-Savitski, M. Bantscheff, C. Cox, D. A. Gordon, R. R. Willard, J. J. Flanagan, L. N. Casillas, B. J. Votta, W. den Besten, K. Famm, L. Kruidenier, P. S. Carter, J. D. Harling, I. Churcher, C. M. Crews, *Nat. Chem. Biol.* **2015**, *11*, 611-617.
- [13] G. E. Winter, D. L. Buckley, J. Paulk, J. M. Roberts, A. Souza, S. Dhe-Paganon, J. E. Bradner, *Science* **2015**, *348*, 1376-1381.
- [14] a) S. D. Edmondson, B. Yang, C. Fallan, *Bioorg. Med. Chem. Lett.* **2019**, *29*, 1555-1564; b) H. J. Maple, N. Clayden, A. Baron, C. Stacey, R. Felix, *Med. Chem. Commun.* **2019**, *10*, 1755-1764.
- [15] J. Lu, Y. Qian, M. Altieri, H. Dong, J. Wang, K. Raina, J. Hines, J. D. Winkler, A. P. Crew, K. Coleman, C. M. Crews, *Chem. Biol.* **2015**, *22*, 755-763.
- [16] H. T. Huang, D. Dobrovolsky, J. Paulk, G. Yang, E. L. Weisberg, Z. M. Doctor, D. L. Buckley, J. H. Cho, E. Ko, J. Jang, K. Shi, H. G. Choi, J. D. Griffin, Y. Li, S. P. Treon, E. S. Fischer, J. E. Bradner, L. Tan, N. S. Gray, *Cell Chem. Biol.* **2018**, *25*, 88-99.
- [17] Y. Sun, X. Zhao, N. Ding, H. Gao, Y. Wu, Y. Yang, M. Zhao, J. Hwang, Y. Song, W. Liu, Y. Rao, *Cell Res.* **2018**, *28*, 779-781.
- [18] A. Mares, A. H. Miah, I. E. D. Smith, M. Rackham, A. R. Thawani, J. Cryan, P. A. Haile, B. J. Votta, A. M. Beal, C. Capriotti, M. A. Reilly, D. T. Fisher, N. Zinn, M. Bantscheff, T. T. MacDonald, A. Vossenkamper, P. Dace, I. Churcher, A. B. Benowitz, G. Watt, J. Denyer, P. Scott-Stevens, J. D. Harling, *Commun. Biol.* **2020**, *3*, 140.
- [19] A. J. Phillips, G. G. Nasveschuk, J. A. Henderson, Y. Liang, C. Chen, M. Duplessis, M. He, K. Lazarski, WO2017197051 A1 2017, 2017.
- [20] A. P. Crew, Y. Qian, H. Dong, J. Wang, K. R. Hornberger, C. M. Crews, WO2018102725 A1 2018, 2018.
- [21] A. P. Crew, K. R. Hornberger, J. Wang, H. Dong, Y. Qian, C. M. Crews, S. Jaime-Figueroa WO2018119448 A1 2018, 2018.
- [22] a) M. G. Organ, S. Calimsiz, M. Sayah, K. H. Hoi, A. J. Lough, *Angew. Chem. Int. Ed. Engl.* **2009**, *48*, 2383-2387; b) S. Shi, M. Szostak, *Chem. Commun.* **2017**, *53*, 10584-10587; c) P. Lei, G. Meng, Y. Ling, J. An, M. Szostak, *J. Org. Chem.* **2017**, *82*, 6638-6646; d) Y. Zhang, V. César, G. Storch, N. Lugan, G. Lavigne, *Angew. Chem. Int. Ed. Engl.* **2014**, *53*, 6482-6486; e) S. Meiries, K. Speck, D. B. Cordes, A. M. Z. Slawin, S. P. Nolan, *Organometallics* **2013**, *32*, 330-339; f) A. Chartoire, X. Frogneux, A. Boreux, A. M. Z. Slawin, S. P. Nolan, *Organometallics* **2012**, *31*, 6947-6951.
- [23] S. Meiries, G. Le Duc, A. Chartoire, A. Collado, K. Speck, K. S. Athukorala Arachchige, A. M. Slawin, S. P. Nolan, *Chem. Eur. J.* **2013**, *19*, 17358-17368.
- [24] K. H. Hoi, J. A. Coggan, M. G. Organ, *Chem. Eur. J.* **2013**, *19*, 843-845.
- [25] B. Atwater, N. Chandrasoma, D. Mitchell, M. J. Rodriguez, M. G. Organ, *Chem. Eur. J.* **2016**, *22*, 14531-14534.
- [26] S. Sharif, J. Day, H. N. Hunter, Y. Lu, D. Mitchell, M. J. Rodriguez, M. G. Organ, *J. Am. Chem. Soc.* **2017**, *139*, 18436-18439.
- [27] Pd-PEPPSI-IPent was used instead of Pd-PEPPSI-IHeptCl due to catalyst availability. The authors found during preparatory scale-up work that raising the reaction temperature from 80 °C to 90 °C and isolating the product via trituration rather than column chromatography improved the yield over small scale reactions (e.g. Table 3, Entry 1).

FULL PAPER

- [28] P. Filippakopoulos, J. Qi, S. Picaud, Y. Shen, W. B. Smith, O. Fedorov, E. M. Morse, T. Keates, T. T. Hickman, I. Felletar, M. Philpott, S. Munro, M. R. McKeown, Y. Wang, A. L. Christie, N. West, M. J. Cameron, B. Schwartz, T. D. Heightman, N. La Thangue, C. A. French, O. Wiest, A. L. Kung, S. Knapp, J. E. Bradner, *Nature* **2010**, *468*, 1067-1073.
- [29] The experiment was terminated prior to complete consumption of **16**, as analysis of the reaction mixture by LCMS revealed the formation of undesired by-products.

WILEY-VCH

Accepted Manuscript