REGULAR ARTICLE



Stannous chloride catalyzed synthesis of Schiff bases from hydroxybenzaldehydes and determination of their antioxidant activity by ABTS and DPPH assay

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Abstract. Phenolic compounds play a very important role in human life because of their antioxidant activity which can prevent harmful diseases caused by free radicals. In the present work, we have synthesized some Schiff bases by the reaction of different hydroxybenzaldehydes and primary aromatic diamines using Stannous Chloride $(SnCl_2 \cdot 2H_2O)$ as the catalyst. The products were characterized by FT-IR spectroscopy, GCMS and NMR spectroscopy. Furthermore, the antioxidant activity of the Schiff bases were determined by using DPPH assay and ABTS assay and the results were compared with a standard compound, trolox as well as with the parent aldehydes. The synthesized compounds were found to have better antioxidant activity than their corresponding parent aldehydes.

Keywords. Phenolic compounds; Schiff bases; antioxidant activity; ABTS assay; DPPH assay.

1. Introduction

In recent years, there is an escalation in research in the areas related to the prevention of diseases, especially the role of free radicals and antioxidants. The oxidant by-products of normal metabolism are either free radicals or molecular species capable of generating free radicals that are naturally produced in the body and play important roles in many normal cellular processes like cell signalling and homeostasis. 1 But abnormally high concentrations of free radicals produced by ionizing radiation and other environmental toxins such as cigarette smoke, some metals and highoxygen atmospheres can be hazardous to the body and play a significant role in the damage of various biological macromolecules like cellular DNA, proteins, cell membrane, etc., leading to the development of cancer and other health conditions, including Diabetes mellitus, Hypertension, Alzheimer disease, immune-system decline, brain dysfunction, and aging of the body. Here comes the need for special substances that are proficient in blocking the activity of free radicals and thus prevent the damage of cells—'antioxidants.' Antioxidants can inhibit the oxidative mechanism that leads to degenerative diseases as they can terminate the deleterious chain reactions by removing the free radical intermediates and also can inhibit other oxidation reactions. In this process, they themselves do get oxidized, so antioxidants are often termed as reducing agents. Moreover, antioxidants have been used to prevent food rancidity as they can slow down the oxidative degradation of polyunsaturated fatty acids and also, compounds with antioxidant activity are found to possess a lot of other significant activities such as anticancer, anti-cardiovascular, antiinflammatory, etc.^{3,4} Mainly, phenolic compounds are one of the most important classes of bioactive antioxidants present in human diet because of their ability to scavenge free radicals as they are able to donate hydrogen atom by breaking the O-H bond to a free radical, thereby preventing the propagation of chain at some stage in the oxidation process and finally inhibiting or retarding the entire process of oxidation.⁵

Among the most efficient antioxidant materials, Schiff bases are considered as one of the most

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important classes of compounds. They are the condensation products of primary amines with carbonyl compounds, i.e., aldehydes or ketones; are termed as Schiff bases as they were first reported by Schiff⁶ in 1864 and have substantial biological activity. In Schiff bases, the oxygen of the carbonyl group is replaced by nitrogen to form the C=N bond. The significant biological properties of Schiff bases are ascribable to the existence of the intra-molecular hydrogen bonds and the proton transfer equilibrium. In medicinal and pharmaceutical fields, Schiff bases have acquired very high significance due to a broad spectrum of biological activities like anti-inflammatory, ⁷ analgesic, ⁸ antimicrobial, ⁹ anticonvulsant, 10 antitubercular, 11 anticancer, 12 antioxidant, 13 anthelmintic, 14 etc. The nitrogen atom of the azomethine group interferes in normal cell processes by being involved in the formation of a hydrogen bond with the active centres of cell constituents. 15,16 Apart from their biological significance, Schiff bases are also used as catalysts, dyes, pigments, polymer stabilizers, intermediates in organic synthesis ¹⁷ and corrosion inhibitors. 18 Schiff bases are used as ligands for preparing metal complexes having a series of different structures or as intermediates for amino acid synthesis. ¹⁹ They commonly co-ordinate through the N-atom of the azomethine group or O-atom of the deprotonated phenolic group and behave as flexidentate ligands. The azomethine nitrogen of Schiff bases and other donor atoms like oxygen play a very important role in coordination chemistry. 20

In our study, we synthesized phenolic aldehyde based di-imines. To synthesize the di-imines, we used 1,2-Phenylenediamine; 1,3-phenylenediamine and 1,4-phenylenediamine as the primary amine substrates along with three different hydroxyl substituted aromatic aldehydes. We used a Lewis acid (SnCl₂ · 2H₂O) to catalyze the reactions and then we studied their antioxidant scavenging behaviour towards ABTS (2, 2'-azinobis-3-ethylbenzothiazoline-6-sulfonic acid) radical and DPPH (1,1-diphenyl-2-picrylhydrazyl) radical.

2. Experimental

2-hydroxybenzaldehyde was purchased from 'MERCK', 3-hydroxybenzaldehyde from 'Sigma Aldrich', 4-hydroxybenzaldehyde from SRL (Sisco Research Laboratory). 1,2-Phenylenediamine; 1,3-phenylenediamine and 1,4-phenylenediamine were purchased from TCI (Tokyo Chemical Industry Co., Ltd). SnCl₂ · 2H₂O was purchased from BDH. ABTS and DPPH were purchased from Sigma Aldrich. Methanol was of LR grade and purchased from 'Rankem' and dichloromethane was purchased from 'SRL' and both were distilled before use.

The infrared spectra of the Schiff bases were recorded within the range 400–4000 cm $^{-1}$ using Shimadzu FT-IR spectrophotometer, model: Prestige 21. 1 HNMR spectra of the Schiff bases were recorded with JEOL, 400 MHz and Bruker Avance III 500 MHz FT-NMR spectrophotometer using DMSO-d $_{6}$ as the solvent and TMS as the internal standard. The mass spectra of the compounds were recorded using Agilent GC-7820A/MS5975 analyzer in methanol. The ABTS and DPPH radical absorptions were recorded with UV–visible spectrophotometer, Hitachi model: U-3900H.

For the preparation of the Schiff bases, a dry round bottom flask (50 mL) was equipped with an efficient magnetic stirrer. 0.1 mmol of SnCl₂·2H₂O was slowly added to the mixture of 2 mmol of an aldehyde and 1 mmol of the primary amine dissolved in dichloromethane. The reaction mixture was allowed to stir at room temperature. The progress of the reaction was monitored by TLC. After the completion of the reaction, the solid products were separated from the reaction mixture by filtration and washed with dichloromethane and water. The desired Schiff bases were synthesized according to the reaction schemes showed in Table 1.

2.1 DPPH assay

The antioxidant activities of the synthesized imines were measured on the basis of the scavenging of the stable 1,1diphenyl-2-picrylhydrazyl (DPPH) free radical according to the method described by Brand-Williams et al. 21 with slight modification. The stock solution of DPPH free radical was prepared by dissolving 0.004 g in 10 mL of methanol in dark. For each different concentrations of imine (2, 4, 6, 8 and 10 mmol/L), blank solutions were prepared by adding 200 μL of DPPH free radical stock solution in 3 mL of methanol and their absorbance were measured at 517 nm. Then, in each blank solution, 100 µL of the imine with different concentration (2, 4, 6, 8 and 10 mmol/L) was added and kept in dark for 30 min. After 30 min their absorbance were measured at 517 nm. By plotting the values of percentage inhibition as the abscissa and the concentration as the ordinate, we calculated the IC₅₀ values for each of the samples. The IC₅₀ value represents the concentration of an antioxidant or inhibitor required to reduce the concentration of the free radicals by half of their initial concentration measured at $\lambda_{max} = 517$ nm.

2.2 ABTS assay

To determine the antioxidant activities of the synthesized Schiff bases, we followed the ABTS method of Arnao $et\,al.$, ²² with a little amendment. The stock solutions were prepared as 7 mM ABTS solution and 2.4 mM potassium persulfate solution. Then the working solution for ABTS assay was prepared by mixing both the stock solutions of ABTS and potassium persulfate in equal quantities and allowing them to react for 14–16 h at room temperature in dark. The solution was then diluted by mixing 100 μ L ABTS solution with 6 mL methanol to obtain an absorbance around 0.706 \pm 0.01 units at 734 nm using a UV spectrophotometer. We prepared

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fresh ABTS solution for each assay. 100 μ L of the solutions (prepared in methanol) of each compound, as well as trolox (used as a standard) at four different concentrations (2, 4, 6, 8 10 mmol/L of methanol), were allowed to react with 2.5 mL of the ABTS solution. The absorbance was observed at 734 nm after 6 min for each sample for each concentration.

The percentage inhibitions of the synthesized products for both the methods were calculated using the following formula-

% Inhibition = $(A_B - A_S)/A_B \times 100$

Where, A_B is the absorbance of the ABTS/DPPH solution without sample and A_S is the absorbance of ABTS/ DPPH solution with the sample. The percentage inhibition was plotted against concentration and a straight line is obtained. From this graph we have calculated the IC_{50} values of the Schiff bases; that is, the amount of antioxidant Schiff bases necessary to decrease the 50% of the initial ABTS or DPPH radical concentration.

3. Results and Discussion

All the synthesized compounds were crystalline solids and found to be soluble in methanol and insoluble in dichloromethane. The reaction scheme, the time period of reactions, their percentage yields and the name of the synthesized products are mentioned in Table 1.

3.1 Characterization

The FT-IR Spectra of all the compounds gave characteristic bands for C=N stretching whereas the characteristic bands for carbonyl group of the aldehydes and the primary amino groups present in the starting materials did not occur in the IR spectra of each of the nine compounds, giving rise to the inference that the substrates were successfully converted into the desired di-Schiff bases. No characteristic peaks were observed for protons of the aldehyde or the amino group in the ¹H-NMR spectra, which affirms the formation of the desired dischiff base products and also by the ¹³C-NMR spectra of the compounds, the formation of imine bonds were confirmed. In the GCMS analysis of the compounds, the existence of molecular ion peak (M·⁺) with m/z value of 316.1 confirmed the formation of the desired products.

Schiff Base 1:

FT-IR (KBr, cm⁻¹): 3571.36 (Phenolic OH), 3108.42 (Aromatic C–H), 1612.56 (aromatic C=N), 1572.05–1465.96 (Aromatic C=C); 1 H NMR (DMSO-d₆; δ, ppm):8.55 (singlet, N=C–H), 5.21 (singlet, O–H), 7.30–6.56 (multiplet, Aromatic protons); 13 C NMR (DMSO-d₆; δ, ppm): 162.41 (Aromatic C=N), 161.01

(C–OH), 122.21 (Aromatic C–N); GCMS(m/z): $316.1(M\cdot^+)$

Schiff Base 2:

FT-IR (KBr, cm⁻¹): 3209.69 (Phenolic OH), 3039.94 (Aromatic C–H), 1607.74 (Aromatic C=N), 1514.19–1457.28 (Aromatic C=C); ¹H NMR DMSO-d₆; δ , ppm): 8.18 (singlet, N=C–H), 5.42 (singlet, O–H), 7.66–6.61 (multiplet, Aromatic protons); ¹³CNMR (DMSO-d₆; δ , ppm): 161.02 (C=N), 160.19 (C–OH), 153.87 (aromatic C–N); GCMS(m/z): 316.1(M·⁺)

Schiff Base 3:

FT-IR (KBr, cm $^{-1}$): 3229.04 (Phenolic OH), 3062.96 (Aromatic C–H), 1602.85 (Aromatic C=N), 1498.69–1458.18 (Aromatic C=C); 1 H NMR (DMSO-d₆; δ, ppm): 8.24 (singlet, N=C–H), 5.35(singlet, O–H), 7.76–6.92 (multiplet, Aromatic protons); 13 C NMR (DMSO-d₆; δ, ppm): 163.61 (C=N), 160.01 (C–OH), 142.59 (aromatic C–N); GCMS(m/z): 316.1(M· $^{+}$)

Schiff Base 4:

FT-IR (KBr, cm⁻¹): 3327.21 (Phenolic OH), 3024.38 (Aromatic C–H), 1603.05 (Aromatic C=N), 1504.48–1435.04 (Aromatic C=C); ¹HNMR (DMSO-d₆; δ , ppm): 8.42 (singlet, N=C–H), 5.44 (singlet, O–H), 7.68–6.33 (multiplet, Aromatic protons); ¹³CNMR (DMSO-d₆; δ , ppm): 161.80(C=N), 158.19 (C–OH), 122.95 (Aromatic C–N); GCMS(m/z): 316.1(M·⁺)

Schiff Base 5:

FT-IR (KBr, cm⁻¹): 3354.21 (Phenolic OH), 3076.48 (Aromatic C–H) 1600.92 (Aromatic C=N), 1500.62–1486.38 (Aromatic C=C); ¹HNMR (DMSO-d₆; δ , ppm): 8.36 (singlet, N=C–H), 5.21(singlet, O–H), 7.58–6.69 (multiplet, Aromatic protons); ¹³CNMR (DMSO-d₆; δ , ppm): 163.97 (C=N), 157.66 (C–OH), 153.97 (Aromatic C–N); GCMS(m/z): 316.1(M·⁺)

Schiff Base 6:

FT-IR (KBr, cm⁻¹): 3215.37 (Phenolic OH), 3018.04 (Aromatic C–H), 1660.00 (Aromatic C=N), 1593.20–1443.17 (Aromatic C=C); ¹HNMR (DMSO-d₆; δ , ppm): 8.51 (singlet, N=C–H), 5.20 (singlet, O–H), 7.79–6.88 (multiplet, Aromatic protons); ¹³CNMR (DMSO-d₆; δ , ppm): 163.15 (C=N), 158.88 (C–OH), 142.13 (aromatic C–N); GCMS(m/z): 316.1(M·⁺)

Schiff Base 7:

FT-IR (KBr, cm⁻¹): 3500.80 (Phenolic OH), 3058.57 (Aromatic C–H), 1610.56 (Aromatic C=N), 1496.78–1462.04 (Aromatic C=C); 1 H NMR (DMSO d₆; δ , ppm): 8.03 (singlet, N=C–H), 5.40 (singlet,O–H), 7.65–6.58 (multiplet. Aromatic protons); 13 C NMR (DMSO-d₆; δ , ppm): 163.07 (C=N), 158.98(C–OH), 120.96 (aromatic C–N); GCMS(m/z): 316.1(M·⁺)

Table 1. Schemes of synthesis of the desired Schiff bases catalysed by Stannous Chloride, their percentage yields and the IUPAC names of the products formed.

Compounds	Schemes of synthesis	Yield (%)	IUPAC Name of product
1	CHO OH 2 H 2-hydroxy benzeldehyde 1,2 diamino benzene NH2 OH HO (1)	77.5	2,2'-((1E,1'E)-(1,2- phenylenebis(azanylylide ne))bis(methanylylidene)) diphenol
2	CHO NH2 2-hydroxy 1,3 diamino benzaldehyde benzene OH HO (4)	86.9	2,2'-((1E,1'E)-(1,3- phenylenebis(azanylylide ne))bis(methanylylidene)) diphenol
3	CHO NH2 NH2 NH2 2-hydroxy benzeldehyde 1,4 diamino benzeldehyde 1,4 diamino benzele OH (7)	87.7	2,2'-((1E,1'E)-(1,4- phenylenebis(azanylylide ne))bis(methanylylidene)) diphenol
4	CHO NH2 2 NH2 3-hydroxy benzaldehyde 1,2-diamino benzene OH HO (2)	76.5	3,3'-((1E,1'E)-(1,2- phenylenebis(azanylylide ne))bis(methanylylidene)) diphenol
5	CHO NH2 2 + NH2 3-hydroxy 1,3-diamino benzene OH (5) OH	78.4	3,3'-((1E,1'E)-(1,3- phenylenebis(azanylylide ne))bis(methanylylidene)) diphenol
6	CHO NH2 2 H H H H H H H H H H H H H H H H H H	82.5	3,3'-((1E,1'E)-(1,4- phenylenebis(azanylylide ne))bis(methanylylidene)) diphenol
7	CHO NH ₂ 2 H NH ₂ OH 1,2-diamino benzene hO (3) HO (3)	84.3	4,4'-((1E,1'E)-(1,2- phenylenebis(azanylylide ne))bis(methanylylidene)) diphenol
8	2 HO NH2 2 H H J,3-diamino NH2 4-hydroxy benzene HO HO (6)	80.5	4,4'-((1E,1'E)-(1,3- phenylenebis(azanylylide ne))bis(methanylylidene)) diphenol
9	2 + Hodroxy 1,4-diamino benzaldehyde benzene HO	82.4	4,4'-((1E,1'E)-(1,4- phenylenebis(azanylylide ne))bis(methanylylidene)) diphenol

^{*}The molar ratio of aldehyde:amine is taken as 2:1 in all the reactions for the formation of the desired di-schiff bases and all the reactions are allowed to run for 6 h.

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Schiff Base 8:

FT-IR (KBr, cm⁻¹): 3512.37 (Phenolic OH), 3223.01 (Aromatic C–H), 1633.71 (Aromatic C=N), 1504.48–1483.76 (Aromatic C=C); 1 H NMR (DMSO-d₆; δ, ppm): 8.48 (singlet, N=C–H), 5.35 (singlet, O–H), 7.76–6.59 (multiplet, Aromatic protons); 13 C NMR (DMSO-d₆; δ, ppm): 163.62 (C=N), 160.01 (C–OH), 154.83 (aromatic C–N); GCMS(m/z): 316.1(M·⁺)

Schiff Base 9:

FT-IR (KBr, cm⁻¹): 3491.16 (Phenolic OH), 3290.56 (Aromatic C–H), 1649.14 (Aromatic C=N), 1536.84–1458.18 (Aromatic C=C); 1 H NMR (DMSO-d₆; δ , ppm) 8.52 (singlet, N=C–H), 5.33 (singlet, O–H), 7.80–6.88 (multiplet, Aromatic protons); 13 C NMR (DMSO-d₆; δ , ppm): 161.80 (C=N), 160.8 (C–OH), 148.19 (aromatic C–N); GCMS(m/z): 316.1(M·⁺)

3.2 Determination of antioxidant activity

The antioxidant activities of the hydroxyl di-Schiff bases and the parent aldehydes were evaluated by the ABTS and DPPH radical scavenging assay. Since the compounds are substituted with the phenolic OH groups, they show considerable radical scavenging activities. The working out of the radical scavenging capacity is based on the UV absorbance spectra of the compounds after reaction with the ABTS radical cation and DPPH radical. In our experiment, Trolox was taken as the standard for comparing the radical scavenging activities. Results are presented in Table 2.

3.2a *DPPH radical scavenging assay*: The observations found by using DPPH Radical Scavenging Assay are presented in Table 2 and Figures 1, 2 and 3.

It is observed that the IC_{50} values of the synthesized compounds decrease in the following order: 1 > 5 > 6 > 4 > 2 > 8 > 7 > 3 > 9. We know that the smaller the IC_{50} value, greater is the antioxidant activity of the compound. So, the order of Antioxidant activity is 9 > 3 > 7 > 8 > 2 > 4 > 6 > 5 > 1.

From the calculation of the IC₅₀, it is obvious that the IC₅₀ values of the parent aldehydes are greater than the synthesized products. Therefore, we can say that the synthesized compounds have better antioxidant activity than their corresponding parent aldehydes. However, there is an exception to the compound 1 showing higher IC₅₀ value and lower percentage inhibition than all the other synthesized compounds, and thus it is found to have the lowest antioxidant activity. This may be attributed to the possible intra-molecular H-bonding present in the compound. The Schiff bases formed using meta isomers of either the aldehyde or

the amine are found to have less antioxidant activity compared to the ones formed using ortho or para isomers of both the substrates (except compound 1). This may be due to the possibility of a lesser number of resonance structures in meta-isomer compared to para and ortho isomer. Thus, compound 5, the condensation product of meta-hydroxybenzaldehyde and 1,3-phenylenediamine is found to have the second highest IC_{50} value and thus the second lowest antioxidant activity.

Among the parent aldehydes, p-hydroxybenzaldehyde is found to have the highest antioxidant activity and the ortho isomer is found to have the lowest antioxidant activity. This also may be attributed to the intramolecular H-bonding present in the orthohydroxybenzaldehyde. Compound 9, 3 and 7 are found to have very low IC₅₀ values compared to the standard compound Trolox which is a very good antioxidant. So, we can say that these compounds can work as much better antioxidants than not only their parent aldehydes but also the standard Trolox. Compound 9, synthesized from, both the para isomers of aldehyde and amine is found to have the highest percentage inhibition and antioxidant activity. This may be due to the presence of a higher number of resonance structures compared to the compounds synthesized from meta-isomers, as well as due to the absence of intramolecular H-bonding and steric repulsion present in ortho-isomers.

3.2b *ABTS radical scavenging assay*: The observations of the experiments conducted to determine the antioxidant activities of the synthesized compounds by ABTS assay are presented in Table 2 and Figures 4, 5 and 6. It is observed that the IC_{50} values of the synthesized compounds decrease in the following order: 1 > 5 > 4 > 6 > 2 > 8 > 7 > 3 > 9. We know that the smaller the IC_{50} value, greater is the antioxidant activity of the compound. So, the order of Antioxidant activity is 9 > 3 > 7 > 8 > 2 > 6 > 4 > 5 > 1. From the calculation of IC_{50} values, we can conclude that synthesized compounds have better antioxidants activity than their corresponding parent aldehydes.

Similar to DPPH Assay, compound I is found to have the highest IC₅₀ value and thus the lowest antioxidant activity followed by compound 5. Compounds synthesized from the meta-isomers of the substrates are found to be weaker antioxidants than the products synthesized from the ortho and para isomer (except compound 1). On the other hand, compound 9 is found to have the highest percentage inhibition and thus the strongest antioxidant activity among all the compounds followed by compound 3. By following ABTS Assay calculations, we have seen that Compounds 9, 3, 7, 8, 2 and 6 are found

 $\textbf{Table 2.} \quad \text{Percentage inhibition and } IC_{50} \text{ values of synthesized Schiff bases by ABTS assay and DPPH assay.}$

Compound	Concentration (mM)	DPPH assay	ABTS assay		
		% inhibition	IC ₅₀ Value	% inhibition	IC ₅₀ Value
Compound 1	2	1.698	95.477	10.358	43.468
	4	3.043		12.632	
	6	4.015		14.886	
	8	4.948		16.364	
	10	5.897		17.987	
Compound 2	2	1.237	53.554	20.12	11.471
_	4	3.252		26.467	
	6	4.8		32.563	
	8	6.905		39.04	
	10	8.871		45.432	
Compound 3	2	19.937	28.466	30.829	4.390
1	4	22.523		44.444	
	6	24.925		62.404	
	8	26.922		82.914	
	10	29.026		102.799	
Compound 4	2	6.496	59.587	12.576	29.176
r	4	8.234		14.568	_,
	6	10.047		17.287	
	8	11.206		20.479	
	10	12.531		23.565	
Compound 5	2	1.197	80.682	8.262	38.152
Sompound 3	4	2.285	00.002	9.823	36.132
	6	4.001		13	
	8	4.936		14.993	
	10	6.076		17.29	
Compound 6	2	6.246	62.320	11.724	20.288
Compound o	4	7.985	02.320	17.67	20.200
		7.983 9.87			
	6			20.318	
	8	10.835		24.748	
G 1.7	10	12.031	20.010	28.736	5 551
Compound 7	2	2.435	29.910	24.5	5.551
	4	4.855		40.88	
	6	8.274		54.66	
	8	12.205		66.368	
	10	16.012		78.618	
Compound 8	2	7.113	52.667	27.7	7.287
	4	9.125		37.803	
	6	10.344		43.387	
	8	12.255		52.95	
	10	14		61.527	
Compound 9	2	12.527	17.594	30.184	3.552
	4	16.752		58.911	
	6	22.321		76.562	
	8	26.84		99.37	
	10	31.624		121	
2-hydroxybenzaldehyde (PA1)	2	5.954	134.276	2	59.823
-	4	6.553		4	
	6	7.385		5.6	
	8	7.95		7.14	
	10	8.588		8.7	
3-hydroxybenzaldehyde (PA2)	2	6.393	94.126	6	40.529
J = 1 J = 2	4	7.06		8	,
	6	8.03		11	
	8	9.09		12.85	
	10	10.128		15	
1-hydroxybenzaldehyde (PA3)	2	2.991	65.612	17.9	27.444
i ily dioxy ochizalachiyac (1713)	4	4.593	05.012	21.1	<i>△ /</i> . ⊤ ⊤ ⊤

Table 2. (contd.)

Compound	Concentration (mM)	DPPH assay		ABTS assay	
		% inhibition	IC ₅₀ Value	% inhibition	IC ₅₀ Value
	6	6.051		23.33	
	8	7.481		24.95	
	10	8.932		28.5	
Trolox	2	7.972	43.244	5.718	30.302
	4	10.01		9.823	
	6	12.048		13	
	8	14.086		15.565	
	10	16.124		18.29	

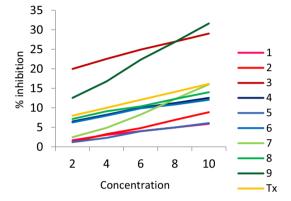


Figure 1. Percentage Inhibition Vs Concentration (mmol/L) of the Schiff Bases and Trolox by using DPPH assay.

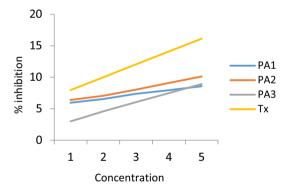


Figure 2. Percentage Inhibition Vs Concentration of the parent aldehydes and trolox (mmol/L) by using DPPH assay.

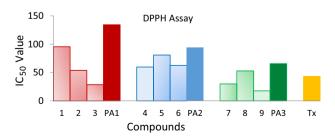


Figure 3. IC₅₀ Values of the Phenolic Schiff Bases in comparison to their parent aldehydes and Trolox by using DPPH Assay.

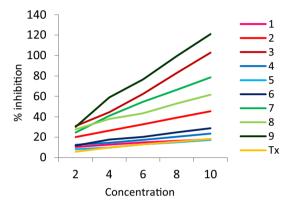


Figure 4. Percentage Inhibition Vs Concentration (mmol/L) of the Schiff Bases and Trolox by using ABTS assay.

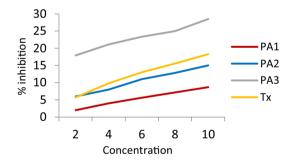


Figure 5. Percentage Inhibition Vs Concentration (mmol/L) of the parent aldehydes and Trolox by using ABTS assay.

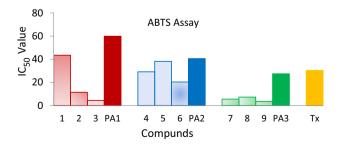


Figure 6. IC₅₀ Values of the Phenolic Schiff Bases in comparison to their parent aldehydes and Trolox by using ABTS Assay.

to have very less IC_{50} values and thus higher antioxidant activity compared to the standard compound Trolox.

4. Conclusions

In conclusion, we have synthesized a total of nine Schiff bases and studied their antioxidant activity by using DPPH and ABTS assay in order to compare the results obtained by both the two methods. The formation of the di-Schiff bases took considerable time because of the steric hindrance of the aromatic rings. All the synthesized products are found to have greater antioxidant activity than their corresponding parent aldehydes. This result may be attributed to the formation of the C=N bonds and increase in conjugation in the products compared to their parent aldehydes. Schiff bases formed from the ortho- or para-isomers of the substrates (except compound 1) are found to have better antioxidant activity compared to the ones formed from meta-isomers by using both the ABTS and DPPH Assay. Schiff base 9 is found to have the strongest antioxidant activity due to possibility of having higher number of resonance structures and lowest steric repulsion well as absence of intramolecular hydrogen bonding whereas Schiff base 1 is found to have the lowest antioxidant activity which may be due to the presence of steric repulsion or intramolecular H bonding leading to toughest removal of the phenolic H-atom.

Supplementary Information (SI)

General experimental procedure, IR, ¹HNMR and 13CNMR spectra for all compounds and graphs for calculation of IC50 values using DPPH and ABTS assay are available at www. ias.ac.in/chemsci.

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