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Jaspine B (pachastrissamine) and 2-*epi*-jaspine B: synthesis and structural assignment

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Abstract—Jaspine B (pachastrissamine), which was isolated in 2002, is a naturally occurring anhydrophytosphingosine which displays potent biological activity, and as such has attracted much attention from synthetic chemists in recent years, with 14 syntheses reported to date. This review delineates the isolation of jaspine B, and laboratory total syntheses of jaspine B, 2-*epi*-jaspine B and analogues, which serves to confirm their structure and stereochemistry. © 2008 Elsevier Ltd. All rights reserved.

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1. Introduction

Phytosphingosines 1-4 are a sub-class of the sphingoid bases, and consist of a 1,3,4-trihydroxy-2-amino unit at the head of a long hydrocarbon chain. By far the most abundant phytosphingosine is D-ribo-phytosphingosine 1, with 18 carbon atoms in the hydrocarbon chain, although smaller amounts of other chain lengths have been detected. Sphingolipids are essential components of eukarvotic cells¹ and phytosphingolipids exhibit important physiological properties.^{2,3} As a result, there has been much interest in their synthesis, and this was the subject of a review in 2002.⁴ Jaspine B 5, also know as pachastrissamine, was the first naturally occurring anhydrophytosphingosine isolated,⁵ which also displays potent biological activity (vide infra). Since its isolation in 2002, there has been a great deal of interest from synthetic chemists concerning the total synthesis of jaspine B 5, its C(2)-epimer (2-epi-jaspine B 6) and their analogues (Fig. 1).



Figure 1. Structures of the C_{18} phytosphingosines 1–4 and the anhydrophytosphingosines jaspine B 5 and 2-*epi*-jaspine B 6.

2. Syntheses of anhydrophytosphingosines prior to the isolation of jaspine B

In 1959 the first report of an anhydrophytosphingosine 7 appeared, although a stereochemical assignment was not given.⁶ Subsequently, its structure and relative configuration were assigned by analogy with the 'truncated' analogue 16, bearing a $C_{12}H_{25}$ side chain. In this synthesis allylic alcohol 8, which was prepared from Grignard addition of dodecylmagnesium bromide to crotonaldehyde, was kinetically resolved under Sharpless asymmetric epoxidation conditions with (+)-diisopropyl tartrate [(+)-DIPT] to afford an enantioenriched sample of (R)-8 in 97% ee. which upon O-silvlation afforded (R)-9 in 41% yield over two steps. Ozonolysis of (R)-9 followed by Horner-Wadsworth–Emmons olefination of the resultant aldehvde 10 gave 11 in 60% overall yield over two steps. Reduction of the ester functionality within 11 was achieved with diisobutylaluminium hydride (DIBAL-H) to give allylic alcohol 12 in 90% yield, which was oxidised under Sharpless asymmetric epoxidation conditions with (-)-DIPT to give epoxide 13 in 97% yield and 96% de. Treatment of 13 with benzylisocyanate gave urethane 14 in 98% yield, and subsequent base-mediated epoxide opening gave tetrahydrofuran derivative 15 in 67% yield. The configuration of 15 was determined by conversion to the corresponding N,O-diacetyl derivative 17 by sequential desilylation, debenzylation and acetylation with Ac₂O to give 17 in 75% yield over two steps; subsequent ¹H NMR NOE studies on 17 supported a (2R,3S,4S)-configuration (Scheme 1).⁸

More recently (2001), the synthesis of an authentic sample of 2-*epi*-jaspine B **6** was reported by Kim et al., starting from D-*ribo*-phytosphingosine 1.⁹ Initial treatment of 1 with 3-nitrophthalic acid at reflux under Dean–Stark conditions is reported to give tetrahydrofuran derivative 18 in 67% isolated yield, with hydrazine-mediated N-deprotection giving **6** in 62% yield. In order to produce a crystalline derivative, tetrahydrofuran 18 was also treated with so-dium *p*-tolylthiolate via nucleophilic displacement of the aromatic nitro group to give 19,¹⁰ the structure of which was established by X-ray crystallography. Kim et al. also



Scheme 1. Reagents and conditions: (i) Ti($O^{i}Pr$)₄, (+)-DIPT, ^{*i*}BuO₂H, CH₂Cl₂, -20 °C, 15 h; (ii) TBDMSOTf, 2,6-lutidine, CH₂Cl₂, 0 °C, 12 h; (iii) O₃, CH₂Cl₂, -78 °C then Me₂S, -78 °C to rt; (iv) (EtO)₂P(O)CH₂CO₂Et, NaH, C₆H₆, 60 °C to rt, 15 min; (v) DIBAL-H, hexane, 0 °C to rt, 12 h; (vi) Ti($O^{i}Pr$)₄, (-)-DIPT, ^{*i*}BuO₂H, CH₂Cl₂, -20 °C, 21 h; (vii) BnNCO, Et₃N, CH₂Cl₂, rt, 12 h; (viii) NaH, THF, rt, 12 h; (ix) HF, H₂O, MeCN, 70 °C, 2 h then Pd/C, cyclohexene, HCl (1 M, aq), MeOH, reflux, 2 h; (x) Ac₂O, pyridine, 70 °C, 1.5 h.

report that treatment of *N*-trifluoroacetyl D-*ribo*-phytosphingosine **20** with *p*-toluenesulfonyl chloride (TsCl) in pyridine effected cyclisation to tetrahydrofuran derivative **21** in 75% yield, which upon deprotection also gave **6** in 67% yield, with identical spectroscopic data (Scheme 2).⁹



Scheme 2. Reagents and conditions: (i) 3-nitrophthalic acid, PhMe, reflux, 3 h; (ii) NH₂NH₂, H₂O, EtOH reflux, 2 h; (iii) NaH, HS(p-MeC₆H₄), DMF, 25 °C, 2 h; (iv) F₃CCO₂Et, EtOH, 25 °C, 16 h; (v) TsCl, pyridine, 25 °C, 16 h; (vi) K₂CO₃, MeOH, 25 °C, 16 h.

3. Isolation of jaspine B (pachastrissamine)

In 2002, studies on the marine sponge *Pachastrissa* sp. by Higa and co-workers⁵ led to the isolation of an anhydrophytosphingosine derivative which they named pachastrissamine **5** (Fig. 2). Shortly after, in an independent study,



Figure 2. Structures of jaspines A 22 and B (pachastrissamine) 5 and other synthetic analogues 6, 16 and 23–25.

Debitus and co-workers reported the isolation of two anhydrophytosphingosines from the marine sponge Jaspis sp.,¹¹ which they named jaspines A 22 and B 5; pachastrissamine and jaspine B being identical (Fig. 2). Both jaspines A 22 and B 5 display biological activity;⁵ jaspine B 5 in particular being the most potent compound isolated from the Jaspis genus to date against the A549 human lung carcinoma cell line. At the present time 11 enantiospecific syntheses of jaspine B 5 have been reported, utilising D-ribophytosphingosine,¹² L-serine,¹³ D-galactose,¹⁴ D-xylose,¹⁵ D-glucose,¹⁶ (R)-glycidol¹⁷ and D-tartaric acid,¹⁸ as the sources of chirality. Three asymmetric syntheses of jaspine B 5 have also been disclosed: in addition to the procedure reported by us (employing a diastereoselective lithium amide conjugate addition),¹⁹ two other strategies have appeared which both employ the Sharpless asymmetric dihydroxylation reaction.²⁰ Two syntheses of 'truncated' analogues 23 and 24 (bearing C_5H_{11} and C_8H_{17} side chains) have also emerged, based upon manipulation of L-xylose²¹ and Sharpless asymmetric epoxidation,²² respectively. Several syntheses of the C(2)-epimeric analogues, with C_8H_{17} , $C_{12}H_{25}$ and $C_{14}H_{29}$ side chains, 25,²² 16^{8a,b} and 6,^{9,12,19} are also known (Fig. 2).

3.1. Stereochemical assignment

In their original isolation paper, Higa et al. reported extensive NMR analyses suggesting a 2-alkyl-3-hydroxy-4-amino moiety. Importantly, in the key region of the ¹H and ¹³C NMR spectra corresponding to the tetrahydrofuran ring ($\delta_{\rm H}$ 3.3–4.2 ppm and $\delta_{\rm C}$ 51–86 ppm), the naturally occurring sample of jaspine B 5 did not match the data for the C(2)-epimer 6 (2-epi-jaspine B) or the $C_{12}H_{25}$ side-chain analogue 16 that had been reported previously,^{8a} and in fact showed marked differences. After the conversion of jaspine B 5 to the corresponding N-acetyl and N,O-diacetyl derivatives 26 and 27, Higa et al. determined the relative configuration of jaspine B 5 by ¹H NMR NOE analysis of the N.O-diacetvl derivative 27, which indicated the all-cis relationship of the substituents around the tetrahydrofuran ring. The (2S, 3S, 4S)-absolute configuration of the natural product was then established using the Mosher method, by conversion of N-acetyl jaspine B 26 to the corresponding (R)- and (S)-2-methoxy-2-trifluoromethylphenylacetyl (MTPA) derivatives.²³ In their independent



Figure 3. Structures of jaspine B **5**, 2-*epi*-jaspine B **6**, the 'truncated' $C_{12}H_{25}$ side-chain analogue **16** and the corresponding acetate derivatives **17** and **26–28**.

study, Debitus et al. also converted jaspine B 5 to the corresponding *N*-acetyl and *N*,*O*-diacetyl derivatives **26** and **27**, and compared the ¹H and ¹³C NMR data with that of the known $C_{12}H_{25}$ side-chain *N*,*O*-diacetyl derivative **17**,^{8a,b} which also indicated an all-*cis* relationship of the substituents around the tetrahydrofuran ring. The absolute configuration of the natural product was then determined, also by the Mosher method, as (2*S*,3*S*,4*S*) (Fig. 3).

4. Syntheses of jaspine B (pachastrissamine)

4.1. Synthesis of jaspine B via lithium amide conjugate addition

We have recently reported an asymmetric synthesis of jaspine B **5**, which employs the conjugate addition of a homochiral lithium amide as the key step.^{19b,c,24} Thus, α,β -unsaturated ester **29** was treated with lithium (*S*)-*N*-benzyl-*N*-(α -methylbenzyl)amide, with subsequent enolate oxidation with (+)-(camphorsulphonyl)-oxaziridine [(+)-CSO]²⁵ affording α -hydroxy- β -amino ester **30** in 75% yield and >98% de. Protecting group manipulation furnished oxazolidine ester **32** in 71% yield over two steps. Reduction of **32** with DIBAL-H, followed by re-oxidation with 2-iodoxybenzoic acid (IBX), gave aldehyde **34** which was immediately treated with tetradecylmagnesium bromide. This gave a chromatographically separable 90:10 mixture

of alcohols, with the major diastereoisomer (4S, 5S, 1'S)-36 isolated in 51% yield and >98% de.²⁶ The preferential addition of tetradecylmagnesium bromide to the Si face of aldehyde 34, giving alcohol (4S,5S,1'S)-36 as the major diastereoisomeric product, is consistent with the 1,2-addition reaction proceeding via a chelated Cram model 35.28 Desilylation of 36 with tetrabutylammonium fluoride (TBAF) gave diol 37 in 95% isolated yield. Optimum conditions for the selective formation of tetrahydrofuran 38 from diol 37 were heating at 80 °C with 3 equiv of tosyl chloride and 4-dimethylaminopyridine (DMAP) for 8 h, which gave quantitative conversion to an 82:18 mixture of diastereoisomeric tetrahydrofurans, with subsequent chromatographic separation giving the major diastereoisomer 38 in 52% vield and >98% de. Subsequent global deprotection of 38 and recrystallisation gave jaspine B 5 in 79% yield (10% overall yield in nine steps from α,β unsaturated ester 29) (Scheme 3). Our synthetic sample of jaspine B 5 showed good agreement with the isolation data, although there were significant discrepancies with some reported syntheses of jaspine B 5. To facilitate a full comparison with all the existing literature data (including 2-epi-jaspine B 6 and the N.O-diacetyl derivatives of both diastereoisomers 27 and 28), authentic samples of these compounds were synthesised. 2-epi-Jaspine B 6 was synthesised from intermediate alcohol 36 via initial mesylation with methanesulfonyl chloride (MsCl) to give 39, followed by desilylation with TBAF and in situ cyclisation of the



Scheme 3. Reagents and conditions: (i) lithium (*S*)-*N*-benzyl-*N*-(α -methylbenzyl)amide, THF, -78 °C, 2 h, then (+)-CSO, -78 °C to rt, 12 h; (ii) H₂ (5 atm), Pd(OH)₂/C, Boc₂O, EtOAc, rt, 12 h; (iii) 2,2-dimethoxypropane, BF₃·Et₂O, acetone, reflux, 12 h; (iv) DIBAL-H, CH₂Cl₂, 0 °C, 6 h; (v) IBX, DMSO, rt, 12 h; (vi) C₁₄H₂₉MgBr, THF, 0 °C to rt, 6 h; (vii) TBAF, THF, rt, 12 h; (viii) TsCl, DMAP, pyridine, reflux, 8 h; (ix) HCl (2 M, aq), MeOH, 50 °C, 6 h, then KOH (2 M, aq); (x) Ac₂O, DMAP, pyridine, rt, 12 h.

resultant primary oxyanion species to give tetrahydrofuran **40**. Tetrahydrofuran **40** proved inseparable by chromatography from the silicon-containing by-products formed in the desilylation reaction and therefore acidic hydrolysis was undertaken giving 2-*epi*-jaspine B **6**, after basification and recrystallisation, in 70% yield (14% overall yield in 10 steps from α,β -unsaturated ester **29**) and >98% de (Scheme 4). The corresponding *N*,*O*-diacetyl derivatives of both jaspine B **5** and 2-*epi*-jaspine B **6** were generated upon the treatment of the free bases with Ac₂O in pyridine



Scheme 4. Reagents and conditions: (i) MsCl, Et_3N , CH_2Cl_2 , rt, 12 h; (ii) TBAF, THF, rt, 12 h; (iii) HCl (2 M, aq), MeOH, 50 °C, 6 h, then KOH (2 M, aq); (iv) Ac₂O, DMAP, pyridine, rt, 12 h.

to give 27 and 28 in 95% and 81% yields, respectively (Schemes 3 and 4).

Our spectroscopic data for these compounds revealed that there are clear differences between the ¹H NMR spectra for jaspine B **5** and 2-*epi*-jaspine B **6**, and also between the ¹H NMR spectra for the corresponding *N*,*O*-diacetyl derivatives **27** and **28**. One characteristic difference between the spectra for jaspine B **5** and 2-*epi*-jaspine B **6** is that the natural product **5** has a peak at 3.91 ppm for one of the C(5) protons, whereas 2-*epi*-jaspine B **6** has a corresponding resonance at 4.11–4.15 ppm: there are no peaks in the ¹H NMR spectrum of jaspine B **5** above 4 ppm (Fig. 4). The most striking difference between the two diastereoisomeric *N*,*O*-diacetyl derivatives **27** and **28** concerns the peaks corresponding to C(3)*H*: for the *N*,*O*-diacetyl jaspine B **27** this appears at 5.38 ppm, whereas for the corresponding C(2)epimer **28** the peak is observed at 4.95–4.97 ppm (Fig. 5).

Within our laboratory, ¹H NMR NOE analyses of *N*,*O*diacetyl derivatives **27** and **28** were supportive of the assigned relative configurations of the tetrahydrofuran rings. Furthermore, single crystal X-ray analysis of **27** unambiguously confirmed the all-*cis* relationship of the substituents around the ring,^{19a} and determination of a Flack parameter²⁹ for the structure of -0.04(15), which satisfies the criterion for a reliable assignment of absolute configuration of a material known to be homochiral,²⁹ confirmed the reported absolute (2*S*,3*S*,4*S*)-configuration of the natural product (originally determined by Higa et al.⁵ using the Mosher method)²³ unambiguously (Fig. 6).



Figure 4. 400 MHz ¹H NMR spectra for (A) jaspine B 5 and (B) 2-epi-jaspine B 6 in CDCl₃.



Figure 5. 400 MHz ¹H NMR spectra for (C) *N*,*O*-diacetyl jaspine B 27 and (D) *N*,*O*-diacetyl-2-*epi*-jaspine B 28 in CDCl₃. [¹H NMR NOE studies have determined that for *N*,*O*-diacetyl jaspine B 27 C(5) H_A is *anti* to the *N*-acetylamino group and for *N*,*O*-diacetyl-2-*epi*-jaspine B 28 C(5) H_A is *syn* to the *N*-acetylamino group.]



Figure 6. Chem 3 D representation of the X-ray crystal structure of *N*,*O*-diacetyl jaspine B **27** (some H atoms omitted for clarity; CCDC deposition number 616170). A crystal structure of *N*,*O*-diacetyl jaspine B **27** has also been reported by Ramana et al. (CCDC deposition number 617243).¹⁶

4.2. Enantiospecific syntheses of jaspine B

In addition to our synthesis, 13 independent syntheses of jaspine B **5** have been reported. A number of enantiospecific syntheses were first to appear following the isolation of jaspine B **5**, with asymmetric approaches being disclosed later. Several different chiral pool starting materials have been used in the enantiospecific syntheses, including D-*ribo*-phytosphingosine,¹² L-serine,¹³ D-xylose,¹⁵ D-glucose,¹⁶ D-galactose,¹⁴ D-tartaric acid¹⁸ and (*R*)-glycidol.¹⁷ The strategies employed have used both starting materials that contain the desired stereochemistry at the N-bearing centre, such as D-*ribo*-phytosphingosine and L-serine, or installed this stereochemistry at a later stage in the synthesis, for example, by manipulation of a carbohydrate derivative.^{14–16} This is most frequently achieved by S_N2 displacement with a nitrogen nucleophile (e.g., azide),^{14–16,18,20b}

although directed C-H amination has also been employed.^{20a}

4.2.1. Manipulation of D-ribo-phytosphingosine: stereodivergent syntheses to both jaspine B and 2-epi-jaspine **B.** Overkleeft et al. have reported the synthesis of jaspine B 5 and 2-epi-jaspine B 6, and their derivatisation to the corresponding N, O-diacetyl derivatives 27 and 28, all based upon manipulation of D-ribo-phytosphingosine 1. Thus, treatment of 1 with triflic azide (TfN₃) resulted in efficient diazotransfer to produce azide 41 in 96% yield. Subsequent Lewis acid-promoted cyclisation upon treatment with BF₃ and trimethylorthoacetate (TMOA) gave tetrahydrofuran derivative 43 in 92% yield, presumably via oxonium ion 42. Removal of the acetate group was followed by Staudinger reduction of the azide group to give jaspine B 5 in 82% yield over the two steps (Scheme 5). The synthesis of 2-epi-jaspine B 6 from 1 was achieved via initial N-Boc protection, followed by selective tosylation of the primary C(1)-hydroxyl and concomitant cyclisation to give 45 in 76% yield over two steps, with N-Boc deprotection giving 2-epi-jaspine B 6 in 95% yield. Both jaspine B 5 and 2-epi-jaspine B 6 were converted into the corresponding N,O-diacetyl derivatives 27 and 28 by treatment with Ac₂O and pyridine (98% yield in both cases). Full characterisation data are reported for jaspine B 5, 2-epi-jaspine B 6 and their corresponding N,O-diacetyl derivatives 27 and 28: these data are in good agreement with both the data reported by Debitus et al.¹¹ for 5 and 27, and the data obtained in our laboratory (Scheme 5).^{12a}



Scheme 5. Reagents and conditions: (i) TfN₃, Na₂CO₃, CuSO₄, CH₂Cl₂, MeOH, H₂O, rt, 16 h; (ii) TMOA, BF₃·Et₂O, CH₂Cl₂, 0 °C to rt, 16 h; (iii) KO'Bu, MeOH; (iv) Me₃P, PhMe/H₂O (24:1), rt, 16 h; (v) Ac₂O, pyridine, rt, 16 h; (vi) (Boc)₂O, Et₃N, THF, rt, 30 min; (vii) TsCl, pyridine/CH₂Cl₂ (1:1), rt, 16 h; (viii) TFA, CH₂Cl₂, rt, 3 h.

Kim et al. also report a stereodivergent synthesis of jaspine B **5** and 2-*epi*-jaspine B **6** from D-*ribo*-phytosphingosine **1** via the key cyclic sulfate intermediate **47**.³⁰ Initial protection of the amino and primary hydroxyl groups of D-*ribo*-phytosphingosine **1** was achieved by sequential treatment with TfN₃ and CuSO₄, then *tert*-butyldiphenyl-silyl chloride (TBDPSCl) and DMAP to give azido *O*-TBDPS ether **46** in 78% yield. Subsequent treatment with SOCl₂ gave the corresponding cyclic sulfate **47** in 90% overall yield for two steps, from which both jaspine B **5**

and 2-*epi*-jaspine B **6** are derived. Desilylative cyclisation of **47** with TBAF resulted in the desired formation of the tetrahydrofuran core to give **48**, with subsequent reduction yielding jaspine B **5** in 56% overall yield from D-*ribo*-phytosphingosine **1**. In order to access 2-*epi*-jaspine B **6**, Kim et al. report the regioselective ring-opening of cyclic sulfate intermediate **47** with iodide, followed by acid-catalysed hydrolysis which is reported to give iodo alcohol **49** in 94% yield. Treatment of **49** with K₂CO₃ in MeOH followed by TBAF afforded epoxy alcohol **50** in 98% yield for two steps. Subsequent treatment of **50** with a catalytic amount



Scheme 6. Reagents and conditions: (i) TfN_3 , K_2CO_3 , $CuSO_4$, $MeOH/H_2O/CH_2Cl_2$, rt, 3 h; (ii) TBDPSCl, Et_3N , DMAP, CH_2Cl_2 , rt, 24 h; (iii) SOCl_2, Et_3N, CH_2Cl_2, 0 °C, 30 min; (iv) RuCl_3·3H_2O, $NaIO_4$, $CCl_4/MeCN/H_2O$, rt, 2 h; (v) TBAF, THF, rt, 1 h; (vi) H_2SO_4/H_2O, THF, rt, 1 h; (vii) H_2, Pd/C, MeOH, rt, 3 h; (viii) Bu_4NI, THF, 40 °C, 1 h; (ix) H_2SO_4, H_2O, THF; (x) K_2CO_3, MeOH, rt, 2 h; (xi) TBAF, THF, rt, 2 h; (xii) TsOH, PhMe, reflux, 12 h.

of *p*-toluenesulfonic acid (TsOH) at reflux promoted cyclisation to tetrahydrofuran derivative **52** in 79% yield, with hydrogenation giving 2-*epi*-jaspine **B 6** in 94% yield (48% yield in ten steps from *D*-*ribo*-phytosphingosine **1**). The mechanism of cyclisation to **52** is proposed by the authors to proceed via an unusual regioselective epoxide ring-opening³¹ with TsOH to give β -hydroxytosylate **51**, which cyclises in situ to give **52**. In support of this mechanism, the authors report the isolation of β -hydroxytosylate **51** upon treatment of **50** with stoichiometric TsOH at rt (Scheme 6).^{12b}

4.2.2. Manipulation of L-serine derivatives. The synthesis of jaspine B **5** reported by Rao and co-workers in 2005 used L-serine derived Garner's aldehyde **53** as the source of chirality.^{13a} Addition of vinylmagnesium bromide to aldehyde **53** gave a separable 86:14 mixture of diastereoisomeric alcohols in accordance with the well-known modified Felkin–Ahn selectivity for addition.³² The major diastereoisomer **54** was protected as the corresponding benzyl ether to give **55** in 92% isolated yield. Ozonolysis of **55** was followed by addition of tetradecylmagnesium bromide to give an inseparable 70:30 mixture of the diastereoisomeric alcohols **56** in 83% yield over two steps. Deprotection of the acetonide protecting group gave a 70:30 mixture of protected D-*ribo*- and D-*lyxo*-phytospingosine derivatives **57** in 91% yield, with subsequent selective silylation of the pri-

mary hydroxy with TBDMSCl giving a 70:30 mixture of diastereoisomeric *O*-silyl protected amino alcohols **58** in 86% yield. Mesylation followed by treatment of the corresponding mesylates with TBAF promoted desilylation and concomitant cyclisation to a separable 70:30 mixture of tetrahydrofurans, from which the all-*cis* diastereoisomer **59** was isolated in 28% yield and its C(2)-epimer **60** was isolated in 12% yield. Subsequent debenzylation and diacetylation of **59** and **60** gave *N*,*O*-diacetyl jaspine B **27** in 82% yield and *N*,*O*-diacetyl 2-*epi*-jaspine B **28** in 84% yield, respectively (Scheme 7).

Passiniemi and Koskinen report a synthesis of jaspine B hydrochloride **75** using L-serine-derived Garner's aldehyde **53** as the source of chirality, with Pd(0)-mediated cyclisation and Ru-mediated cross-metathesis reactions as the key steps in the synthesis. Iodide **66** was coupled with Garner's aldehyde **53** by treatment with BuLi and N,N'-dimethyl-N,N'-propylene urea (DMPU) to give a separable 92:8 mixture of diastereoisomers, with the major isomer **67** isolated in 63% yield. Subsequent benzylation, *O*-TBDMS deprotection and acetylation gave acetate **70** in 81% overall yield over three steps. Hydrolysis of the N,O-acetal followed by Pd(0)-mediated intramolecular 1,5-cyclisation³³ gave a separable 67:33 mixture of diastereoisomeric tetrahydrofuran derivatives in 95% combined yield. The all-*cis* isomer **72** was then coupled with 1-tetradecene (10 equiv) via a



Scheme 7. Reagents and conditions: (i) vinylmagnesium bromide, THF, 0 °C to rt, 12 h; (ii) BnBr, THF, NaH, 0 °C to rt, 12 h; (iii) O₃, CH₂Cl₂, -78 °C 1 h, Me₂S; (iv) C₁₄H₂₉MgBr, THF, 12 h, rt; (v) AcOH, 0 °C to rt, 12 h; (vi) TBDMSCl, imidazole, CH₂Cl₂, DMAP, 0 °C to rt, 12 h; (vii) MsCl, Et₃N, CH₂Cl₂, 0 °C to rt 12 h; (viii) TBAF, THF, rt, 2 h; (ix) Na, NH₃, THF, -78 °C, 30 min; (x) TFA:CH₂Cl₂ (1:1), rt, 6 h; (xi) Et₃N, Ac₂O, CH₂Cl₂, 0 °C to rt, 4 h.



Scheme 8. Reagents and conditions: (i) I₂, KOH, MeOH; (ii) KO₂CN=NCO₂K, AcOH, MeOH; (iii) TBDMSCl, imidazole, DMF; (iv) BuLi, DMPU, PhMe, -78 °C then 53, PhMe, -95 °C; (v) BnBr, TBAI, NaH, THF, 0 °C then Δ ; (vi) TBAF, CH₂Cl₂, rt; (vii) Ac₂O, DMAP, Et₃N, CH₂Cl₂, rt; (viii) FeCl₃–SiO₂, CHCl₃, rt; (ix) Pd(PPh₃)₄, PPh₃, THF, 55 °C; (x) Grubbs II 73, 1-tetradecene, CH₂Cl₂, 40 °C; (xi) H₂, Pd(OH)₂, MeOH/EtOAc (1:1), rt then HCl, MeOH/EtOAc (1:1).

cross-metathesis reaction with Grubbs II **73** to give **74** as the only diastereoisomer in 87% yield. Tandem hydrogenation/hydrogenolysis of **74** with Pd(OH)₂/C under H₂, followed by treatment with HCl gas was reported to afford jaspine B hydrochloride **75** in 76% yield (Scheme 8).^{13c}

4.2.3. Manipulation of carbohydrate derivatives. Several carbohydrate starting materials have also been employed for enantiospecific syntheses of jaspine B 5. Shaw et al. have reported a synthesis of jaspine B 5 using D-galactose as the source of chirality. Initial treatment of tri-Obenzyl-D-galactal 76 under Perlin hydrolysis conditions gave α,β -unsaturated aldehyde 77, which was immediately reduced under Luche conditions to afford allylic alcohol 78 in 65% yield over two steps. Allylic alcohol 78 was then treated under Sharpless asymmetric epoxidation conditions with (+)-diethyl tartrate [(+)-DET] to generate diol 79, via an intramolecular asymmetric ring-opening process,³⁴ with subsequent isopropylidene protection giving tetrahydrofuran derivative 80 in 48% yield over two steps.³⁵ Mesylation of 80 followed by S_N^2 displacement with NaN₃ gave azide 82 in 62% yield. Hydrolysis of the acetonide functionality within 82 and periodate cleavage of the resultant diol was followed by Wittig olefination to give 84 as a mixture of geometic isomers, in 82% yield. Global reduction was then achieved via transfer hydrogenation with HCO₂NH₄ in refluxing MeOH to yield jaspine B 5 in 72% yield (Scheme 9).¹⁴

Du and co-workers initially communicated their synthesis of jaspine B 5 in 2006 in which D-xylose 85 was protected as acetal 86, regioselectively tosylated on the primary

hydroxyl group and benzylated with benzyl trichloroacetimidate and trimethylsilyltriflate (TMSOTf) to give tosylate 88 in 57% overall yield for three steps. Subsequent treatment with HCl in EtOH gave 89, then mesylation and exposure to NaN₃ generated tetrahydrofuran derivative 91 in 63% yield. Hydrolysis of 91 with aqueous TFA afforded aldehyde 83, which was then subjected to Wittig olefination giving an inseparable mixture of (E)- and (Z)-isomers of the corresponding olefin 84 in 77% yield. Finally, 84 was fully reduced under hydrogenation conditions to reportedly give jaspine B 5 in 92% yield as the free base, although the data quoted actually correspond to those of jaspine B TFA salt 61 (Scheme 10).^{15a} Subsequently, an improved and scaleable synthesis was reported by Du et al., also starting from D-xylose 85.15b In this approach, protected D-xylose derivative 92 was treated with NaIO₄ to generate aldehyde 93, which was immediately subjected to a Wittig reaction to produce 94 in 82% yield over two steps. Iodine-promoted cyclofunctionalisation/debenzylation of 94 afforded iodide 95,³⁵ and subsequent oxidation of the iodomethyl group with DMSO followed by treatment with MsCl gave mesylate 96 in 74% yield over two steps from 95. Wittig olefination of 96 resulted in the generation of an inseparable mixture of (E)- and (Z)-isomers 97, which upon treatment with NaN₃ gave 84 in 72% yield over two steps (Scheme 11). Subsequent elaboration of 84 to jaspine B TFA salt 61 was reported to be directly analogous to the previous synthesis, affording jaspine B TFA salt 61 in 95% yield (Scheme 10). The ¹H and ¹³C NMR spectroscopic data reported in this article (for jaspine B TFA salt 61) are in excellent agreement with those reported by Higa et al. for the TFA salt of the natural product.⁵



Scheme 9. Reagents and conditions: (i) HgSO₄ (cat.), H₂SO₄ (0.01 M, aq), 1,4-dioxane, 0 °C to rt, 8 h; (ii) NaBH₄, (0.5 equiv), CeCl₃·7H₂O (1.0 equiv), EtOH, 0 °C to rt, 3 h; (iii) Ti(O'Pr)₄ (1.0 equiv), (+)-DET (1.2 equiv), 'BuO₂H (2.0 equiv), 4 Å MS, CH₂Cl₂, -25 °C to 0 °C, 2.5 h (iv) satd citric acid in acetone, 2 h; (v) MsCl, Et₃N, CH₂Cl₂, 0 °C to rt, 1.5 h; (vi) NaN₃, Bu₄NCl (cat.), DMF, 120 °C, 3 days; (vii) H₅IO₆ (1.3 equiv), EtOAc, rt, 3 h; (viii) [C₁₃H₂₇PPh₃]⁺[Br]⁻, BuLi, THF, -78 °C; (ix) HCO₂NH₄, MeOH, Pd/C, Δ , 18 h.



Scheme 10. Reagents and conditions: (i) acetone, H_2SO_4 (concd), then Na_2CO_3 ; (ii) tosylimidazolide, MeOTf, *N*-methylimidazole, THF; (iii) benzyl trichloroacetimidate, TMSOTf (1 mol %), CH₂Cl₂, -40 °C; (iv) HCl (5% in EtOH, v/v), Δ , 3 h; (v) MsCl, pyridine, rt, 4 h; (vi) NaN₃, NH₄Cl, DMF, 120 °C, 20 h; (vii) TFA (50% aq), CH₂Cl₂, rt, 30 min; (viii) [C₁₃H₂₇PPh₃]⁺[Br]⁻, BuLi, THF, -40 °C; (ix) H₂, Pd(OH)₂/C, MeOH/EtOAc, 5 h.

Ramana et al. have reported a stereodivergent synthesis of jaspine B 5 and *ent*-jaspine B *ent*-5 starting from D-glucose 98,¹⁶ in a directly analogous fashion to that reported by Du and co-workers.^{15a} Conversion of D-glucose 98 to aldehyde 99 (according to a literature procedure)³⁶ was followed by reduction with NaBH₄ and treatment with TsCl to give tos-

ylate **88** in 83% yield (over two steps from aldehyde **99**). Subsequent acid-mediated acetonide deprotection with concomitant cyclisation gave dimethylacetal **101** in 81% yield. Acid catalysed hydrolysis gave aldehyde **102**, which was then subjected to Ohira–Bestmann alkynylation to afford alkynol **103** in 54% yield. Conversion of **103** to the



Scheme 11. Reagents and conditions: (i) NaIO₄, MeOH, H₂O; (ii) $[CH_3PPh_3]^+[Br]^-$, BuLi, THF, -40°C to rt; (iii) I₂, NaHCO₃, MeCN; (iv) NaHCO₃, DMSO, 150°C, 6 min; (v) MsCl, pyridine, rt, 30 min; (iv) $[C_{13}H_{27}PPh_3]^+[Br]^-$, BuLi, THF, -40 °C to rt; (vii) NaN₃, NH₄Cl, DMF, 120 °C, 20 h; (viii) Pd/C, H₂, MeOH, TFA, 5 h.

corresponding azidoalkyne **104** was achieved by treatment with triflic anhydride (Tf₂O) in pyridine, followed by reaction with LiN₃ in DMF to give **104** in 81% yield over two steps. Alkylation of the terminal acetylene **104** with BuLi, hexamethylphosphoramide (HMPA) and $C_{12}H_{25}Br$ gave **105**, which was reduced under transfer hydrogenolysis conditions to give jaspine B 5 in 41% yield over two steps. Whilst no data are reported for the natural product, full characterisation data for the N,O-diacatate derivative 27, including X-ray crystal structure data, are given (Scheme 12). Aldehyde 99 was also converted into *ent*-jaspine B *ent*-5 by first subjecting it to Ohira–Bestmann alkynylation



Scheme 12. Reagents and conditions: (i) NaBH₄, MeOH, 0 °C, 2 h; (ii) TsCl, pyridine, DMAP, CH_2Cl_2 , 5 h; (iii) TsOH, MeOH, Δ , 6 h; (iv) H_2SO_4 (2.0 M, aq), AcOH (50% aq), 9 °C, 2 h; (v) (MeO)_2P(O)C(N_2)COCH_3, MeOH, K_2CO_3 , rt, 8 h; (vi) Tf_2O, pyridine, CH_2Cl_2 , 0 °C, 6 h; (vii) LiN₃, DMF, rt, 12 h; (viii) BuLi, THF/HMPA (7:1), $C_{12}H_{25}Br$, -78 °C to -40 °C, 1 h; (ix) Pd/C (10% w/w), HCO₂NH₄, MeOH, Δ , 10 h; (x) Ac₂O, Et₃N, DMAP, CH₂Cl₂, rt, 6 h.

to give **106** and then reductive deketalisation, using excess triethylsilane in the presence of $BF_3 \cdot Et_2O$, to give *ent*-**103** in 75% yield over two steps. The subsequent steps in the synthesis are directly analogous to that of the natural product with final diacetylation giving *ent*-*N*,*O*-diacetyl jaspine B *ent*-**27** in 31% overall yield in five steps from *ent*-**103** (Scheme 13).

Chandrasekhar et al. have published a synthesis of a 'truncated' version of jaspine B with a C_5H_{11} side chain 23,²¹ based upon manipulation of L-xylose-derived aldehyde ent-99.37 Aldehyde ent-99 was subjected to Wittig olefination with $[C_4H_9PPh_3]^+[Br]^-$, hydrolysis and in situ formation of the methyl glycoside to give 108 in 82% yield over two steps. Activation of the hydroxyl group as the corresponding imidazole sulfonate ester allowed S_N2-displacement with NaN₃ to give 109 in 58% yield. Subsequent reduction of 109 with Et₃SiH promoted by BF₃, followed by hydrogenation gave the 'truncated' version of the natural product 23 in 66% yield (Scheme 14). The truncated side chain has little effect on the $\delta_{\rm H}/\delta_{\rm C}$ signals in the ¹H and ¹³C NMR spectra for the resonances associated with the tetrahydrofuran core and, crucially, the highest field signal is at 3.85–3.95 ppm ($\delta_{\rm H}$ < 4.0 ppm for all ring protons in CDCl₃), which is in excellent agreement with an all-cis arrangement.

4.2.4. Syntheses based on manipulation of other chiral pool starting materials. In 2006, Marco et al. reported an enantiospecific synthesis of jaspine B 5 from (R)-glycidol. O-TBDPS protected (R)-glycidol 111 was initially treated with tridecylmagnesium bromide in the presence of CuI to afford the corresponding alcohol, which was then protected as its O-methoxymethyl (O-MOM) derivative 112. Sequential desilvlation, Swern oxidation and olefination then gave α . β -unsaturated ester (E)-114 in 81% yield over three steps. Ester reduction of 114 with DIBAL-H gave allylic alcohol 115, which was treated under Sharpless asymmetric epoxidation conditions, with (-)-DET, to provide epoxy alcohol 116 in 85% yield.³⁵ Epoxy alcohol 116 was then treated with trichloroacetonitrile in the presence of DBU to give imino ester derivative 117, which was then reacted with Et₂AlCl to generate oxazoline 118 in 72% yield. Subsequent hydrolysis and N-Boc protection gave diol 119. Removal of the O-MOM group was achieved with TMSBr to give triol 120, which was treated with TsCl, followed by K₂CO₃ in MeOH to induce cyclisation to give tetrahydrofuran derivative 122 in 50% yield over five steps. N-Boc Deprotection of 122 with TFA and subsequent basification with NaOH then gave jaspine B 5 in 75% yield. Full characterisation data for jaspine B 5 and the N,O-diacetyl derivative 27, which was prepared by



Scheme 13. Reagents and conditions: (i) $(MeO)_2P(O)C(N_2)COCH_3$, MeOH, K_2CO_3 , rt, 8 h; (ii) BF₃:Et₂O, Et₃SiH, CH₂Cl₂, -20 °C to rt, 6 h; (iii) Tf₂O, pyridine, CH₂Cl₂, 0 °C, 6 h; (iv) LiN₃, DMF, rt, 12 h; (v) BuLi, THF/HMPA (7:1), C₁₂H₂₅Br, -78 °C to -40 °C, 1 h; (vi) Pd/C (10% w/w), HCO₂NH₄, MeOH, Δ , 10 h; (vii) Ac₂O, Et₃N, DMAP, CH₂Cl₂, rt, 6 h.



Scheme 14. Reagents and conditions: (i) $[C_4H_9PPh_3^{++}[Br]^-$, BuLi, THF/HMPA, $-40 \degree C$, 4 h; (ii) AcCl, MeOH, $60 \degree C$, 8 h; (iii) SO₂Im₂, NaH, DMF, $-40 \degree C$, 3 h; (iv) NaN₃, Bu₄NCl, DMF, $110 \degree C$, 12 h; (v) BF₃·Et₂O, Et₃SiH, CH₂Cl₂, $0 \degree C$ to rt, 2 h; (vi) Pd/C, H₂, MeOH, rt, 6 h.



Scheme 15. Reagents and conditions: (i) $C_{13}H_{27}MgBr$, CuI, THF, -10 to 0 °C; (ii) MOMCl, EtN^{*i*}Pr₂, CH₂Cl₂, rt, 24 h; (iii) TBAF, THF, rt, 3 h; (iv) (COCl)₂, DMSO, Et₃N, CH₂Cl₂, -78 to -46 °C, 3 h; (v) (^{*i*}PrO)₂P(O)CH₂CO₂Et, LiBr, Et₃N, THF, rt; (vi) DIBAL-H, hexane, 0 °C, 2.5 h; (vii) 'BuO₂H, (-)-DET, Ti(O^{*i*}Pr)₄, CH₂Cl₂, -20 °C, 24 h; (viii) Cl₃CCN, DBU, CH₂Cl₂, 0 °C, 30 min; (ix) Et₂AlCl, CH₂Cl₂, 0 °C to rt, 5 h; (x) HCl (1.0 M, aq), THF, rt, 5 h; (xi) Boc₂O, NaHCO₃, THF, rt, 16 h; (xii) TMSBr, CH₂Cl₂, -78 °C, 30 min; (xii) TsCl, Et₃N, DMAP, CH₂Cl₂, rt, 20 min; (xiv) K₂CO₃, MeOH, rt, 16 h; (xv) TFA, CH₂Cl₂, 0 °C to rt, 45 min then NaOH, MeOH, 5 min; (xvi) Ac₂O, DMAP, Et₃N, CH₂Cl₂, rt, 16 h.

treatment of the free base with Ac_2O , Et_3N and DMAP, are reported (Scheme 15).¹⁷

Chandrakumar and Prasad have reported a synthesis of jaspine B 5 starting from D-tartaric acid-derived bisdimethylamide 123. Addition of tetradecylmagnesium bromide to bisdimethylamide 123 gave keto amide 124 in 86% yield, and stereoselective reduction of ketone 124 under Luche conditions furnished a mixture of diastereomeric alcohols (90% de) with 125 as the major diastereoisomer. Subsequent reaction of this mixture with excess 2,2-dimethoxypropane and TsOH gave a separable mixture of



Scheme 16. Reagents and conditions: (i) $C_{14}H_{29}MgBr$, THF, -15 °C, 30 min; (ii) NaBH₄, CeCl₃, MeOH, -15 °C, 2 h; (iii) 2,2-dimethoxypropane, TsOH, C_6H_6 , Δ , 12 h; (iv) NaBH₄, MeOH, -15 °C to rt, 3 h; (v) TBDMSCl, imidazole, DMF, rt, 12 h; (vi) TsCl, DMAP, CH₂Cl₂, rt, 14 h; (vii) NaN₃, DMF, 100 °C, 12 h; (viii) TBAF, THF, 0 °C to rt, 1 h; (ix) TsCl, Et₃N, CH₂Cl₂, 0 °C to rt, 5 h; (x) FeCl₃·6H₂O, CH₂Cl₂, rt, 8 h; (xi) K₂CO₃, MeOH, 0 °C to rt, 4 h; (xii) H₂, Pd/C, MeOH, CH₂Cl₂, rt, 3 h.

diastereoisomeric alcohols, from which the major diastereoisomer **126** was isolated in 86% yield. Alcohol **126** was reduced with NaBH₄ to afford diol **127**, which was first treated with TBDMSCl, then TsCl to give **128** in 85% yield (over three steps). Treatment of **128** with NaN₃ gave a mixture of the expected S_N2 displacement product and also resulted in partial desilylation, subsequent treatment with TBAF afforded azohydrin **129** in 92% yield from **128**. Tosylation of **129** followed by acid-promoted acetonide hydrolysis, then treatment with K₂CO₃ in MeOH gave the corresponding tetrahydrofuran derivative in 91% yield. Subsequent azide reduction with Pd/C gave jaspine B **5** in 94% yield (Scheme 16).¹⁸

4.3. Other asymmetric syntheses of jaspine B

In addition to the syntheses of jaspine B 5 highlighted above, there have also been two further asymmetric syntheses of the natural product, the first of these by Yakura et al. utilises the Sharpless asymmetric dihydroxylation of α , β unsaturated ester 131 to give diol 132 in 98% ee. Subsequent acetonide protection, reduction with DIBAL-H and Wittig olefination gave 135 as a single diastereoisomer in 85% yield. One-pot removal of the O-p-methoxybenzyl (O-PMB) group and olefin reduction was then achieved under hydrogenation conditions to give 136, which was treated with I₂ and PPh₃ to afford iodide 137 in 87% yield from olefin 135. Acid-catalysed deprotection of 137 followed by cyclisation gave tetrahydrofuran 138. Subsequent treatment with trichloroacetyl isocyanate and Rh-mediated C-H amination gave a mixture of products from which 139 was isolated in 19% yield from 138. Hydrolysis of 139 with KOH and acetylation with Ac_2O in pyridine gave N,O-diacetyl jaspine B 27 in 85% yield (Scheme 17).^{20a}

Venkatesan and Srinivasan have also reported an asymmetric synthesis of jaspine B 5 which utilises Sharpless asymmetric dihydroxylation and a chelation-controlled vinyl Grignard addition as the key steps in the synthetic strategy. 1-Pentadecanol 140 was subjected to oxidation with pyridinium chlorochromate (PCC), followed by Wittig olefination to give α , β -unsaturated ester 141 in 92% yield. α , β -Unsaturated ester 141 was then treated under Sharpless asymmetric dihydroxylation conditions to give diol 142 in 88% yield and 97% ee. Subsequent O-MOM protection, reduction and re-oxidation gave aldehyde 145 which was immediately treated with vinyl magnesium bromide, under chelation control, to give allylic alcohol 146 in >95% de and 89% yield from alcohol 144. O-Benzyl protection of 146, oxidative cleavage and reduction then gave alcohol 147 in 71% yield over four steps. Subsequent protecting group manipulation, tosylation and azide displacement then gave azide 149 in 75% yield over four steps. Acidic hydrolysis of 149 followed by treatment with TsCl gave tosylate 130 which was reacted according to the procedure outlined by Chandrakumar and Prasad¹⁸ (vide supra) to give jaspine B 5 (Scheme 18).^{20b}

The synthesis of another 'truncated' C_8H_{17} side chain analogue 24 is reported by Génisson et al., the synthesis of which is based on Sharpless asymmetric aminohydroxylation of allylic alcohol 150. Under these conditions, a 70:30 mixture of amino alcohols 151 and 152 is produced; subsequent treatment with methyl chloroformate generated a separable mixture of regioisomeric oxazolidinones 153 and 154, which were isolated in 60% and 23% yields, respectively; the *cis*-stereochemistry of the major product 153 was established by single crystal X-ray analysis. Oxidation of 153 with Dess-Martin periodinane (DMP) gave aldehyde 155 in 90% yield. The C_8H_{17} side chain was then



Scheme 17. Reagents and conditions: (i) AD-mix- α , BuOH–H₂O, 0 °C, 16 h; (ii) Me₂C(OMe)₂, CSA, rt, 12 h; (iii) DIBAL-H, CH₂Cl₂, -80 °C, 1 h; (iv) [Ph₃PC₁₃H₂₇]⁺[Br]⁻, BuLi, THF, -20 °C, 1 h; (v) H₂ (3 atm), Pd/C, EtOAc, rt, 3 h; (vi) I₂, PPh₃, imidazole, CH₂Cl₂, rt, 1 h; (vii) concd HCl, THF, rt, 2.5 h then K₂CO₃, MeOH, rt, 3 h; (viii) Cl₃CCON=C=O, CH₂Cl₂, rt, 1 h then neutral Al₂CO₃; (ix) Rh₂(OCOCPh₃)₄ (10 mol %), PhI(OAc)₂ (4.2 equiv), MgO (6.9 equiv), C₆H₆, reflux, 13 h; (x) KOH (aq), EtOH, reflux, 3 h; (xi) Ac₂O, pyridine, rt, 12 h.



Scheme 18. Reagents and conditions: (i) PCC, CH₂Cl₂, 0 °C, 3 h; (ii) Ph₃PCHCO₂Et, C₆H₆, Λ , 4 h; (iii) (DHQ)₂PHAL, K₂CO₃, K₃[Fe(CN)₆], MeSO₂NH₂, OsO₄ (0.1 M in PhMe), 'BuOH/H₂O (1:1), 0 °C, 24 h; (iv) MOMCl, CH₂Cl₂, EtNⁱPr₂, 0 °C to rt, 16 h; (v) DIBAL-H, CH₂Cl₂, -10 °C, 1 h; (vi) IBX, EtOAc, Λ , 3 h; (vii) vinylmagnesium bromide, MgBr₂·Et₂O, THF, -78 °C, 45 min; (viii) BnBr, NaH, THF, 0 °C, 4 h; (ix) OsO₄ (0.1 M in PhMe), 'BuOH/THF/H₂O (3:2:1), *N*-methylmorpholine-*N*-oxide (NMO), 16 h; (x) NaIO₄, NaHCO₃, H₂O, rt, 3 h; (xi) NaBH₄, MeOH, 0 °C to rt, 1 h; (xii) MOMCl, CH₂Cl₂, EtNⁱPr₂, 0 °C to rt, 6 h; (xiii) H₂, Pd/C, MeOH, rt, 16 h; (xiv) TsCl, CH₂Cl₂, Et₃N, DMAP, 0 °C to rt, 3 h; (xv) NaN₃, DMF, 95 °C, 14 h; (xvi) aq HCl, THF, 0 °C to rt, 5 h; (xvii) TsCl, CH₂Cl₂, DMAP, 0 °C to rt, 4 h; (xviii) K₂CO₃, MeOH, 0 °C to rt, 4 h; (xix) H₂, Pd/C, MeOH, rt, 3 h.

introduced via Grignard addition to aldehyde **155**; mesylation of the resultant alcohol **156** followed by hydrogenolytic O-debenzylation and in situ cyclisation gave tetrahydrofuran derivative **158** in 68% yield. N-Debenzylation of **158** was achieved via Birch reduction, and saponification of the resultant oxazolidinone **159** gave the 'truncated' version of the natural product **24** in 95% yield, with ¹H NMR data ($\delta_{\rm H} < 4.0$ ppm for all ring protons in CDCl₃) supporting an all-*cis* arrangement (Scheme 19).²²

4.4. Synthesis with inconsistent data

Datta et al. communicated their purported synthesis of jaspine B 5 in 2005;^{13b} in this synthesis L-serine 160



Scheme 19. Reagents and conditions: (i) $Ti(O'Pr)_4$, (-)-DET, 'BuO₂H, 4 Å MS, CH₂Cl₂, -23 °C, 3 days then Me₂S, rt, 3 h, then, $Ti(O'Pr)_4$, BnNH₂, rt, 1.5 days; (ii) K₂CO₃, MeOCOCl, THF, rt, 18 h; (iii) KOH, MeOH, rt, 5 h; (iv) DMP, CH₂Cl₂, rt, 3 h; (v) C₈H₁₇MgBr, CeCl₃, THF, -78 °C, 5 h then 0 °C, 1 h; (vi) MsCl, Et₃N, CH₂Cl₂, 0 °C to rt, 1.5 h; (vii) H₂ (10 bar), Pd(OH)₂, EtOH, 2 days; (viii) Na/NH₃, THF, -78 °C, 4 h; (ix) KOH, EtOH/H₂O, 85 °C, 7 h.



Scheme 20. Reagents and conditions: (i) HCO₂H, CH₂Cl₂, 0 °C then EtOAc, satd aq NaHCO₃; (ii) DIBAL-H, -78 °C; (iii) [Ph₃PC₁₂H₂₅]⁺[Br]⁻, BuLi, THF, -78 °C to rt; (iv) Pd/C, H₂, EtOAc, rt; (v) aq KOH, EtOH, Δ .

was converted into butenolide 161, the synthesis of which had previously been developed in the Datta laboratories.³⁸ Treatment of 161 with formic acid in dichloromethane enabled deprotection of the acetonide, with subsequent Michael addition of the free hydroxyl group giving cis-fused bicycle 162; the structure of which was supported by X-ray crystallographic analysis of related derivatives.^{13b,39} Subsequent elaboration of this compound was reported to furnish jaspine B 5 as the free base in 46% vield over four steps. Unfortunately upon inspection of the spectroscopic data reported by Datta et al., and comparison with the other natural and synthetic samples of jaspine B 5 discussed in this review, it is evident that the data are anomalous and in fact closely resemble those of 2-epi-jaspine B 6; a full comparison of the relevant spectroscopic data is given below. This discrepancy can be accounted for by a retro-Michael/Michael epimerisation pathway upon treatment of bicyclic lactone 162 with DIBAL-H and excess Wittig reagent. Enolate 166, which is generated under these conditions, may undergo a retro-Michael reaction to generate α,β -unsaturated aldehyde 167. Recyclisation of the resultant oxyanion species would generate the thermodynamically favoured 2,3-anti-aldehyde 168, which is trapped out as the corresponding olefin 170. Subsequent hydrogenation of 170 and cleavage of the resultant oxazolidinone 171 with ethanolic KOH afforded 2-epi-jaspine B 6 in 77% yield (Scheme 20).

5. Comparison of spectroscopic data

5.1. Specific rotation

A summary of the reported specific rotation data for jaspine B 5, 2-epi-jaspine B 6 (both free bases and salts) and the corresponding \hat{N} , O-diacetyl derivatives 27 and 28, excluding the data reported in the anomalous syntheses highlighted above, ^{13b,15a} is given below (Table 1). It is known that hydrogen-bonding systems do not always give consistent measurements for specific rotation,⁴⁰ and as such judgements based upon specific rotation alone should be regarded with caution. In this system, specific rotation is not diagnostic of relative stereochemistry due to the reported similarities between the specific rotations for jaspine B 5, 2-epi-jaspine B 6 and their derivatives. There is seemingly only one major inconsistency within this set of data $\{[\alpha]_D^{24} = +76.0 \ (c \ 0.2, CHCl_3)$ reported by Kim et al.^{12b} for the free base of jaspine B 5} otherwise the values for jaspine B 5, 2-epi-jaspine B 6 and the TFA salts of both diastereoisomers are typically in the range +10 to +20, and upon conversion to the corresponding N,O-diacetyl derivatives, lie approximately within the range -15 to -30 (Table 1).

5.2. ¹H and ¹³C NMR spectroscopic data

A thorough inspection of the ¹H and ¹³C NMR data for all reported synthetic and the naturally occurring samples of

 Table 1. Specific rotation data for jaspine B 5, 2-epi-jaspine B 6, and the corresponding N,O-diacetyl derivatives 27 and 28, TFA salts 61 and 62, and HCl salts 75 and 172

Compound	Source	Specific rotation
H ₂ N, OH $O^{(1)}$, $C_{14}H_{29}$ jaspine B 5	Higa (Ref. 5) Shaw (Ref. 14) Srinivasan (Ref. 20b) Prasad (Ref. 18) Davies (Refs. 19a,19c) Overkleeft (Ref. 12a) Shaw (Ref. 14) Kim (Ref. 12b) Davies ^d Marco (Ref. 17) Debitus (Ref. 11) Kim (Ref. 12b) ^a	$\begin{bmatrix} \alpha \end{bmatrix}_{D} = +18.0 \ (c \ 0.1, \ EtOH) \\ [\alpha]_{D}^{28} = +13.3 \ (c \ 0.03, \ EtOH) \\ [\alpha]_{D}^{25} = +17.7 \ (c \ 0.4, \ EtOH) \\ [\alpha]_{D}^{2} = +17.5 \ (c \ 0.4, \ EtOH) \\ [\alpha]_{D}^{2} = +17.5 \ (c \ 0.4, \ EtOH) \\ [\alpha]_{D}^{22} = +4.8 \ (c \ 1.0, \ MeOH) \\ [\alpha]_{D}^{23} = +8.5 \ (c \ 0.047, \ MeOH) \\ [\alpha]_{D}^{23} = +19.7 \ (c \ 0.5, \ MeOH) \\ [\alpha]_{D}^{25} = +9.4 \ (c \ 0.3, \ MeOH) \\ [\alpha]_{D}^{25} = +9.0 \ (c \ 1.5, \ CHCl_3) \\ [\alpha]_{D}^{20} = +7.0 \ (c \ 0.1, \ CHCl_3) \\ [\alpha]_{D}^{24} = +76.0 \ (c \ 0.2, \ CHCl_3) \\ \hline \begin{bmatrix} \alpha \end{bmatrix}_{D}^{24} = +6.5 \ (c \ 0.4, \ CHCl_3) \\ \hline \begin{bmatrix} \alpha \end{bmatrix}_{D}^{24} = +6.5 \ (c \ 0.4, \ CHCl_3) \\ \hline \begin{bmatrix} \alpha \end{bmatrix}_{D}^{24} = +6.5 \ (c \ 0.4, \ CHCl_3) \\ \hline \begin{bmatrix} \alpha \end{bmatrix}_{D}^{24} = +6.5 \ (c \ 0.4, \ CHCl_3) \\ \hline \begin{bmatrix} \alpha \end{bmatrix}_{D}^{24} = +6.5 \ (c \ 0.4, \ CHCl_3) \\ \hline \end{bmatrix} $
H_2N_{II} OH $C_{14}H_{29}$ 2-epi-jaspine B 6	Davies ^d Kim (Ref. 9) Davies ^d Kim (Ref. 12b) Overkleeft (Ref. 12a) Davies (Ref. 19c) Datta (Ref. 13b) ^c Davies ^d	$[\alpha]_{D}^{10} = +6.5 (c \ 0.47, \text{CHCl}_3)$ $[\alpha]_{D}^{23} = +14.8 (c \ 0.002, \text{EtOH})$ $[\alpha]_{D}^{18} = +12.7 (c \ 0.65, \text{EtOH})$ $[\alpha]_{D}^{23} = +23.2 (c \ 1.0, \text{MeOH})$ $[\alpha]_{D}^{22} = +15.0 (c \ 1.0, \text{MeOH})$ $[\alpha]_{D}^{21} = +16.4 (c \ 0.85, \text{MeOH})$ $[\alpha]_{D}^{25} = +9.6 (c \ 0.11, \text{CHCl}_3)$ $[\alpha]_{D}^{18} = +11.8 (c \ 0.65, \text{CHCl}_3)$
AcHN, OAc 0, ''''C ₁₄ H ₂₉ 27	Kim (Ref. 12b) Ramana (Ref. 16) Marco (Ref. 17) Overkleeft (Ref. 12a) Davies (Ref. 19c) Ramana (<i>ent.</i> , Ref. 16) Rao (Ref. 13a) ^b	$\begin{split} & [\mathbf{x}]_{D}^{24} = -18.5 \ (c \ 0.7, \ CHCl_3) \\ & [\mathbf{x}]_{D}^{25} = -34.6 \ (c \ 1.0, \ CHCl_3) \\ & [\mathbf{x}]_{D}^{25} = -28.4 \ (c \ 1.0, \ CHCl_3) \\ & [\mathbf{x}]_{D}^{22} = -22.6 \ (c \ 1.0, \ CHCl_3) \\ & [\mathbf{x}]_{D}^{23} = -26.4 \ (c \ 0.5, \ CHCl_3) \\ & [\mathbf{x}]_{D}^{25} = +29.0 \ (c \ 1.0, \ CHCl_3) \\ & [\mathbf{x}]_{D}^{25} = -28.8 \ (c \ 1.1, \ CHCl_3) \end{split}$
AcHN, OAc	Overkleeft (Ref. 12a) Davies (Ref. 19c)	$[\alpha]_{\rm D}^{22} = -15.4 \ (c \ 1.0, \ {\rm CHCl}_3)$ $[\alpha]_{\rm D}^{22} = -14.6 \ (c \ 0.5, \ {\rm CHCl}_3)$
$H_3N^{\textcircled{O}}_{14}H_{29}$	Rao (Ref. 13a) Du (Ref. 15b)	$[\alpha]_{D}^{25} = +17.1 \ (c \ 0.4, \text{ EtOH})$ $[\alpha]_{D}^{25} = +11.0 \ (c \ 1.0, \text{ CHCl}_3)$
$H_{3}N_{4}^{\oplus}$ $TFA_{0}^{\oplus}C_{14}H_{29}$ 62	Rao (Ref. 13a)	$[\alpha]_{\rm D}^{25} = +14.5 \ (c \ 1.5, \ {\rm EtOH})$
H ₃ N [⊕] OH Cl [⊖] /′′′′C ₁₄ H ₂₉ 75	Davies (Ref. 19c)	$[\alpha]_{\rm D}^{23} = +2.6 \ (c \ 0.4, \ {\rm MeOH})$
$H_{3}N_{1}^{\oplus}OH$ $CI^{\Theta}C_{14}H_{29}$ 172	Davies (Ref. 19c)	$[\alpha]_{\rm D}^{21} = +15.5 \ (c \ 0.9, \ {\rm MeOH})$

^a A personal communication from the corresponding author has confirmed the value for this specific rotation.

^b Data kindly supplied in a personal communication from the corresponding author.

^c Stereochemistry re-assigned in this review (see Section 4.4).

^d Value obtained for this review.

Table 2. Comparison of reported ¹H NMR data for jaspine B 5, 2-epi-jaspine B 6, and the corresponding N,O-diacetyl derivatives 27 and 28, TFA salts 61 and 62, and HCl salts 75 and 172 ____

Compound	Entry	Source	Solvent	C(2) <i>H</i>	C(3) <i>H</i>	C(4)H	$C(5)H_A$	$C(5)H_B$
	1	Higa (Ref. 5)	MeOD	3.81 dt	3.86 dd	3.51 dt	3.43 dd	3.90 dd
	2	Shaw (Ref. 14) ^a	MeOD	3.73–3.78 m	3.92 qt	3.49–3.69 m	3.49–3.69 m	4.01–4.03 m
	3	Srinivasan (Ref. 20b) ^a	MeOD	3.69–3.75 m	3.84–3.88 m	3.63–3.67 m	3.49–3.53 m	3.89–3.94 m
H ₂ N OH	4	Debitus (Ref. 11)	CDCl ₃	3.75 ddd	3.88 dd	3.68 dt	3.54 dd	3.95 dd
4 3	5	Kim (Ref. 12b) ^a	CDCl ₃	3.73 dt	3.86 dd	3.64 dt	3.50 dd	3.92 dd
⁵ ² '''''C ₁₄ H ₂₉	6	Marco (Ref. 17) ^a	CDCl ₃	3.73 ddd	3.86 dd	3.65 m	3.51 dd	3.92 dd
i aspine B 5	7	Overkleeft (Ref. 12a)	CDCl ₃	3.75 ddd	3.87 dd	3.59 br m	3.53 dd	3.92 dd
Juspine D 0	8	Prasad (Ref. 18) ^{a,b}	CDCl ₃	3.71–3.76 m	3.86–3.88 m	6.62–3.70 m	3.50–3.53 m	3.91–3.95 m
	9	Davies (Refs. 19a, 19c)	CDCl ₃	3.74 td	3.87 t	3.61–3.69 m	3.50 dd	3.91 t
	10	Koskinen (Ref. 13c)	CDCl ₃	3.73 ddd	3.87 dd	3.64 m	3.51 dd	3.92 dd
	11	Datta (Ref. 13b) ^{a,d}	CDCl ₃	Not reported	3.63 m	3.40–3.48 m	3.40–3.48 m	4.14 dd
	12	Kim (Ref. 12b) ^a	CDCl ₃	3.60 dd	3.62 m	3.45 m	3.39 dd	4.11 dd
$\begin{pmatrix} 4 & 3 \\ 5 & 2 \end{pmatrix}$	13	Kim (Ref. 9) ^a	CDCl ₃	3.58–3.62 m	3.58–3.62 m	3.45 q	3.36 ABq	4.20 ABq
C ₁₄ H ₂₉	14	Davies (Refs. 19a, 19c)	CDCl ₃	3.60–3.64 m	3.60–3.64 m	3.46–3.49 m	3.37–3.47 m	4.11–4.15 m
2-epi-jaspine B 6	15	Overkleeft (Ref. 12a)	CDCl ₃ /MeOD	3.74 m	4.04 t	3.67 quin	3.75 dd	4.13 dd
	16	Higa (Ref. 5)	CDCl ₃	3.90 ddd	5.38 dd	4.81 dq	3.60 dd	4.15 dd
	17	Debitus (Ref. 11)	CDCl ₃	3.93 ddd	5.42 dd	4.85 dq	3.62 dd	4.11 dd
AcHN OAc	18	Kim (Ref. 12b) ^a	CDCl ₃	3.90 m	5.38 dd	4.81 dddd	3.59 t	4.07 t
4 3	19	Ramana (Ref. 16) ^a	CDCl ₃	3.85–3.93 m	5.37 dd	4.80 dq	3.58 dd	4.06 dd
5 ² ""C+Han	20	Marco (Ref. 17) ^a	CDCl ₃	3.90 ddd	5.40 dd	4.83 qd	3.60 t	4.09 t
0 ⁰ 14 ¹ 29	21	Overkleeft (Ref. 12a) Valuera (\mathbf{R} of $(20a)^a$	CDCl ₃	3.90 m	5.38 dd	4.81 ddd	3.59 dd	4.07 dd
21	22	Davies (Refs. $10a 10c$)	CDCl ₃	3.00–.393 III 3.01 ddd	5.38 dd	4.82 qu 4.82 ddd	3.00 t	4.08 t 4.10 t
	23 24	B_{ao} (Ref. 13a) ^{a,e}	CDCl ₃	3.80_3.90 m	5.30_5.40 m	4.82 ddd 4 70_4 80 m	3.50-3.60 m	4.10 t 4.00_4.10 m
	24	Ruo (Rui: 154)	ebelg	5.00 5.90 m	5.50 5.40 m	4.70 4.00 m	5.50° 5.00° II	4.00 4.10 m
AcHN OAc								
4 3	25	Overkleeft (Ref. 12a)	CDCl ₃	3.86 m	4.91 dd	4.65 m	3.52 dd	4.17 dd
5 2	26	Davies (Refs. 19a, 19c)	CDCl ₃	3.90–3.93 m	4.95–4.97 m	4.67–4.73 m	3.55–3.58 m	4.21–4.24 m
O C ₁₄ H ₂₉								
20								
_								
H₃N, OH								
	27	Higa (Ref. 5)	MeOD	3.79 dd	4.25 dd	3.91 dd	3.71 dt	3.88 m
5 ² ''''Cutheo	28	Du (Ref. 15b) ^a	MeOD	3.79 dd	4.23 dd	3.82–3.93 m	3.70 dt	3.82-3.93 m
0 ⁻¹⁴⁻¹²⁹		Rao (Ref. 13a) ^{a,e}	MeOD	3.75–3.80 m	4.20–4.25 m	3.80–3.90 m	3.65–3.75 m	3.80-3.90 m
01								
H₃N, OH								
$ \Theta \left[4 - 3 \right]$								
	29	Rao (Ref. 13a) ^a	MeOD	3.64–3.75 m	4.10–4.17 m	3.64–3.75 m	3.64–3.75 m	3.98–4.04 m
O ^{14H29}								
62								
u N⊕ ou								
$CI = \begin{pmatrix} 4 & 3 \\ 5 & 2 \end{pmatrix}$	30	Davies (Ref. 19c)	MeOD	3.73 ddd	4.28 dd	3.88–3.96 m	3.83 q	3.88-3.96 m
O ^{-'''''} C ₁₄ H ₂₉								
75								
.								
H₃N ⊂ OH								
$Cl^{\Theta^{4}}$	21	Davias (Pef 10a)	MaOD	270 277	1 01 1 06	2 70 2 77	2 70 2 77	1 16 1 10 m
	51	Davies (Rel. 190)	MEOD	3.70-3.77 m	4.04–4.00 m	5.70-5.77 m	5.70-5.77 m	4.10–4.19 M
0 ⁰ 14 ¹¹ 29								
112								

^a Protons assigned in this review.

^bResonance at 3.62–3.70 ppm confirmed in a personal communication from the corresponding author. ^c Data kindly supplied in a personal communication from the corresponding author.

^d Stereochemistry re-assigned in this review (see Section 4.4).

^eSpectra kindly supplied in a personal communication from the corresponding author (peaks quoted \pm 0.05 ppm).

jaspine B **5** and comparison with that of 2-*epi*-jaspine B **6** reveals that Datta's proposed data for the natural product (Tables 2 and 3, entry 11) in fact bears close resemblance to that of the 2-*epi*-jaspine B **6** (Tables 2 and 3, entries 12-

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14) and displays a characteristic peak at 4.14 ppm in the ${}^{1}\text{H}$ NMR spectrum corresponding to one of the C(5) protons; the all-*cis* stereochemistry of 2-alkyl-3-hydroxy-4-amino-tetrahydrofuran derivatives is characterised by the absence

Table 3. Comparison of reported ¹³C NMR data for jaspine B 5, 2-*epi*-jaspine B 6, and the corresponding *N*, *O*-diacetyl derivatives 27 and 28, TFA salts 61 and 62, and HCl salts 75 and 172

Compound	Entry	Source	Solvent	C(2)	C(3)	C(4)	C(5)
	1	Higa (Ref. 5)	MeOD	84.3	73.3	56.0	72.1
	2	Shaw (Ref. 14) ^a	MeOD	83.5	71.3	54.4	70.5
	3	Srinivasan (Ref. 20b) ^a	MeOD	83.2	72.3	54.4	71.8
H ₂ N OH	4	Debitus (Ref. 11)	CDCl ₃	83.1	72.2	54.2	71.6
	5	Kim (Ref. 12b) ^a	CDCl ₃	83.2	72.3	54.3	71.8
$\begin{pmatrix} 5 & 2 \end{pmatrix}$	6	Marco (Ref. 17) ^a	CDCl ₃	83.3	72.4	54.4	71.8
O ''C ₁₄ H ₂₉	7	Overkleeft (Ref. 12a)	CDCl ₃	82.9	71.8	54.3	71.7
jaspine B 5	8	Prasad (Ref. 18) ^a	CDCl ₃	84.3	70.9	54.4	68.9
	9	Davies (Refs. 19a, 19c)	CDCl ₃	83.1	72.4	54.4	71.8
	10	Koskinen (Ref. 13c) ^{a,b}	CDCl ₃	83.2	72.3	54.3	71.7
H ₂ N, QH	11	Datta (Ref. 13b) ^{a,c}	CDCl ₃	85.1	74.6	52.5	73.1
	12	Kim (Ref. 12b) ^a	CDCl ₃	85.2	74.8	52.7	73.1
5 2	13	Kim (Ref. 9) ^a	CDCl ₃	85.2	74.7	52.6	73.1
C ₁₄ H ₂₉	14	Davies (Refs. 19a, 19c)	CDCl ₃	85.3	74.8	52.6	73.2
2- <i>epi</i> -jaspine B 6	15	Overkleeft (Ref. 12a)	CDCl ₃ /MeOD	84.4	73.1	52.0	68.0
	16	Kim (Ref. 12b) ^a	CDCl ₃	81.2	73.5	51.3	69.9
	17	Ramana (Ref. 16) ^a	CDCl ₃	81.2	73.5	51.3	69.9
ACHIN OAC	18	Marco (Ref. 17) ^a	CDCl ₃	81.2	73.6	51.4	70.0
	19	Overkleeft (Ref. 12a) ^a	CDCl ₃	81.1	71.4	51.2	69.7
⁵ ² ''''C ₁₄ H ₂₉	20	Yakura (Ref. 20a) ^a	CDCl ₃	81.2	73.6	51.3	70.0
27	21	Davies (Refs. 19a, 19c)	CDCl ₃	81.2	73.5	51.3	69.8
21	22	Rao (Ref. 13a) ^{a,d}	CDCl ₃	81.0	73.5	51.5	70.0
	23	Overkleeft (Ref. 12a) ^a	CDC1.	84.0	76.6	49.8	69.7
$\begin{pmatrix} 4 & 3 \\ 5 & 2 \end{pmatrix}$	23	Davies (Refs. $10a 10c$)	CDCl ₃	84.2	70.0	49.0	60.0
28 C ₁₄ H ₂₉	24	Davids (Rels. 194,196)	CDCI3	04.2	//.1	49.9	09.9
H ₃ N OH	25	Higa (Ref. 5) ^a	MeOD	84.3	70.9	54.3	68.9
$TFA^{\ominus/4}$	26	Du (Ref. 15b) ^a	MeOD	84.4	70.9	54.3	68.9
0 ⁵ / ¹ /'''C ₁₄ H ₂₉		Rao (Ref. 13a) ^{a,d}	MeOD	85.0	71.5	55.0	69.5
61							
$TFA^{\ominus}/\overset{4}{\overset{3}{\overset{1}{\overset{1}{\overset{1}{\overset{1}{\overset{1}{\overset{1}{1$	27	Rao (Ref. 13a) ^a	MeOD	85.2	74.3	53.7	69.4
C ₁₄ H ₂₉							
62							
H₃N⊕_OH							
	•				(0.0	52.4	60.0
CT (5 2) (1) (C ₁₄ H ₂₉	28	Davies (Ref. 19c)	MeOD	83.4	69.9	53.4	68.0
75							
æ							
H ₃ N ^Y OH							
$CI \xrightarrow{(-)} \sqrt{4 - 3}_{5 - 2}$	29	Davies (Ref. 19c)	MeOD	83.8	73.0	52.3	68.0
O C ₁₄ H ₂₉							
172							

^a Carbons assigned in this review.

^b Data kindly supplied in a personal communication from the corresponding author.

^c Stereochemistry re-assigned in this review (see Section 4.4).

^d Spectra kindly supplied in a personal communication from the corresponding author (peaks quoted \pm 0.5 ppm).

of peaks above 4 ppm in the ¹H NMR spectra in CDCl₃, as found for jaspine B 5 (Table 2, entries 4-10).

6. Conclusion

Since the isolation of jaspine B (pachastrissamine) was first communicated (2002), there have been 13 independent syntheses of this natural product. This review has highlighted some inconsistencies between the data reported for some synthetic samples of jaspine B and that of the naturally occurring compound. However, the majority of syntheses (including those of 2-*epi*-jaspine B) corroborate the original structural assignment. The wealth of data summarised in this review serves to confirm the structure and (2S,3S,4S)-absolute configuration of this natural product, which was correctly assigned by Higa et al. in the first instance and then independently by Debitus and co-workers.

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- 26. The configurations of the newly formed C(1')-stereocentres within alcohols (4*S*,5*S*,1'*S*)-**36** and (4*S*,5*S*,1'*R*)-**173** could not be assigned a priori; they were established via conversion to the corresponding *N*,*O*,*O*,*O*-tetra-acetyl phytosphingosines **174** and **175**. Global hydrolysis of the protecting groups within the major diastereoisomeric alcohol (4*S*,5*S*,1'*S*)-**36** was achieved upon treatment with HCl in MeOH/H₂O, with subsequent acetylation giving *N*,*O*,*O*,*O*-tetra-acetyl D-*lyxo*phytosphingosine **174** with spectroscopic properties in excellent agreement with those previously reported {[α]_D²¹ = -3.1 (*c* 0.7 in CHCl₃); lit.²⁷ [α]_D²² = -3.1 (*c* 1.1 in CHCl₃)}. Analogous treatment of the minor diastereoisomeric alcohol (4*S*,5*S*,1'*R*)-**173** gave *N*,*O*,*O*,*O*-tetra-acetyl D-*ribo*-hytosphingosine **175** {[α]_D²² = +18.2 (*c* 1.0 in CHCl₃); lit.²⁷ [α]_D²² = +21.9 (*c* 1.1 in CHCl₃)} (Scheme 21).



Scheme 21. Reagents and conditions: (i) HCl (3 M, aq), MeOH, 50 $^{\circ}$ C, 3 h, then Ac₂O, DMAP, pyridine, rt, 12 h.

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