

# Synthesis of 5-oxyquinoline derivatives for reversal of multidrug resistance

Torsten Dittrich<sup>1</sup>, Nils Hanekop<sup>2</sup>, Nacera Infed<sup>2</sup>, Lutz Schmitt<sup>2</sup> and Manfred Braun<sup>\*1</sup>

Full Research Paper	Open Access
Address:	Beilstein J. Org. Chem. 2012, 8, 1700–1704
<sup>1</sup> Institute of Organic and Macromolecular Chemistry, University of	doi:10.3762/bioc.8.103
	ddi. 10.37 02/bj00.8. 185
Dusseidorr, Universitätsstr. 1, D-40225 Dusseidorr, Germany and	
<sup>2</sup> Institute of Biochemistry, University of Düsseldorf, Universitätsstr. 1,	Received: 21 June 2012
D-40225 Düsseldorf, Germany	Accepted: 31 August 2012
	Published: 05 October 2012
Email <sup>.</sup>	
Manfred Dreum <sup>*</sup> has user @hbu de	
Manned Braun - braunn@nnu.de	Associate Eulior. S. Brase
* Corresponding author	© 2012 Dittrich et al; licensee Beilstein-Institut.
	License and terms: see end of document.
Keywords:	
acetals: enantionure compounds: beterocycles: inhibitor: multidrug	
resistance	

### Abstract

The inhibition of ABC (ATP binding cassette) transporters is considered a powerful tool to reverse multidrug resistance. Zosuquidar featuring a difluorocyclopropyl-annulated dibenzosuberyl moiety has been found to be an inhibitor of the P-glycoprotein, one of the best-studied multidrug efflux pumps. Twelve 5-oxyisoquinoline derivatives, which are analogues of zosuquidar wherein the dibenzosuberyl-piperazine moiety is replaced by either a diarylaminopiperidine or a piperidone-derived acetal or thioacetal group, have been synthesized as pure enantiomers. Their inhibitory power has been evaluated for the bacterial multidrug-resistance ABC transporter LmrCD and fungal Pdr5. Four of the newly synthesized compounds reduced the transport activity to a higher degree than zosuquidar, being up to fourfold more efficient than the lead compound in the case of LmrCD and about two times better for Pdr5.

### Introduction

The treatment of cancer is often severely hampered by efflux pumps, which are responsible for the extrusion of various chemotherapeutics from the tumor cell, an effect termed "multidrug resistance". Various transporters of the ATP-binding cassette family, so called ABC transporters, have been shown to be responsible for multidrug resistance [1]. Among these multidrug-resistance ABC transporters, the P-glycoprotein has been investigated most intensively [2]. As P-glycoprotein is considered to be a major player in multidrug resistance and has been found to be over expressed in tumor cells [3,4], considerable attempts have been made to develop inhibitors of P-glycoprotein in order to reverse multidrug resistance. These efforts were pursued for a longer period leading to a variety of smallmolecular compounds with different structures that act as P-glycoprotein inhibitors [5]. The drug zosuquidar (1a) emerged as the most promising among them [6-9]; however, so far it has not made its way into routine clinical application [10].

Apart from their role in multidrug resistance of tumor cells, ABC-type efflux pumps are also involved in multidrug resistance of pathogenic Gram-positive bacteria and fungi for which inhibitors have only rarely been studied. Therefore, we were interested in developing inhibitors of efflux pumps from bacterial and fungal organisms, namely LmrCD from *Lactococcus lactis* and Pdr5 from *Saccharomyces cerevisiae* [11,12]. Thus, we synthesized, guided by the lead structure zosuquidar (1a), a series of compounds that were evaluated as potential inhibitors of LmrCD and Pdr5. Whereas biochemical studies with the novel compounds will be disclosed separately [13], we present in this article the synthesis, in detail, of twelve new compounds that may provide reversal of multidrug-resistant activity.

### Results and Discussion Synthesis

As extended studies on modifications of zosuguidar (1a) had revealed both the 5-oxyquinoline and the hydroxypropane spacer as being crucial for inhibitory activity on P-glycoprotein [14-18], our strategy was based on modification or replacement of the dibenzosuberyl and the piperazine moieties, whereas the above-mentioned "southern part" of zosuquidar (1a) was planned to be maintained, including the absolute configuration of the stereogenic carbinol center. Thus, we replaced the piperazine motif in 1 by a piperidine ring carrying either nitrogen or oxygen as a hetero substituent(s) in the 4-position. Therefore, an initial series of 4-aminopiperidines 2a-4a and, in a second approach, new spirocyclic acetals 5a-12a as well as the thioacetal 13a were synthesized. All the new compounds, shown in Scheme 1, were obtained in enantiomerically pure form, with the stereogenic carbinol center adopting (R) configuration. The acetals 8a and 9a formed as mixtures of diastereomers, whereas





compound **10a** was enantiomerically and diastereomerically pure.

The commercially available chiral building block (*S*)-3-chloro-1,2-propanediol (14) was chosen as the starting material. It was converted to (*R*)-nosylate 15 in three steps by following a modified procedure [19], as shown in Scheme 2. The subsequent treatment with 5-hydroxyquinoline [20] led to the epoxide (*R*)-16, serving as the chiral building bloc in the final key step, which involves the nucleophilic attack of the secondary amino group of the precursors 2b-13b to the epoxide 16, to deliver the final products 2a-13a. The yields of the final coupling step are given in Scheme 2.

The precursor 2c was obtained by an N-arylation, as described in the literature.[21] A modified Buchwald–Hartwig amination reaction using tri-*tert*-butylphosphane as a ligand at palladium [22] was applied in order to couple 4-methylphenyl bromide and 4-fluorophenyl bromide with Boc-protected aminopiperidines 17 and 18, respectively. Thus, diarylaminopiperidine 3c and 4c became readily accessible, as outlined in Scheme 3. The cleavage of the protecting group occurred according to standard protocols by treatment with trifluoroacetic acid to liberate the amines 2b-4b, which were used without further purification in the coupling reaction with epoxide (R)-16. The spirocyclic compounds 6c-13c were obtained by standard acid-catalyzed reaction with the corresponding diols for the formation of acetals 6c-12c and propanedithiol for 13c, starting from *N*-acetyl-4-piperidone. Basic hydrolysis was found to be appropriate for the removal of the N-protecting group. Again, the free amines 6b-13b were used as crude products for the reaction with the oxirane (*R*)-16 to give the series of new compounds. Spiroacetal **5b** was commercially available.

#### **Biochemical studies**

The transport activity [13] of the compounds 2a-13a and the reference compound zosuguidar (1a) was determined for the ABC transporters LmrCD and Pdr5. The results are given in Table 1, which restricts itself to those compounds that displayed a significantly enhanced activity compared to that of 1a. The activity of the individual transporters in the absence of either zosuquidar (1a) or any of the new compounds 2a-13a is assigned as 100%, meaning that those substances that show a lower relative transport activity than zosuquidar (1a) are considered as potential improvements. Various new 5-oxyquinoline derivatives show activity that is comparable or inferior to that of 1a. For reasons of clarity, only those results are included in Table 1 that indicate a substantially higher reduction of transport activity than that for zosuquidar (1a). It turns out that, at the transporter LmrCD (first column), a derivative with a 4-aminopiperidine skeleton, compound **3a**, was distinctly more





Table 1: Relative transport activity of LmrCD and Pdr5 in the presence of zosuquidar (1a) and selected new 5-oxyquinoline derivatives (2a-13a). <sup>a</sup>		
Compound / Relative transport activity at LmrCD	Compound / Relative transport activity at Pdr5	
<b>1a</b> / 42.4%	<b>1a</b> / 23.7%	
<b>3a</b> / 19.8%	<b>2a</b> / 14.0%	
<b>10a</b> / 11.2%	<b>11a</b> / 12.5%	

<sup>a</sup>100% activity corresponds to the transport activity in the absence of either zosuquidar (**1a**) or any 5-oxyquinoline derivative. The reported values represent the average of three independent measurements.

active than zosuquidar (1a). Moreover, the acetal derivative 10a was found to be about four times more active than the lead compound 1. For the fungal transporter Pdr5 (second column), one compound of the 4-aminopiperidine series, 2a, is about twice as active as zosuquidar (1a). Similar activity is provided by catechol-derived acetal 13a, whereas, at this transporter, the acetals derived from 1,2-diphenylethanediol, are still active as transport inhibitors, but their activity is slightly lower than that of zosuquidar (31% relative transport activity). The full details of the results of transport inhibition for all the compounds 1a and 2a through 13a are given in a separate communication [13].

### Conclusion

The replacement of the benzosuberyl moiety of zosuquidar (1a) by diarylamino or acetal groups lead to a series of twelve new compounds, which have been tested as potential inhibitors of the multidrug-resistance ABC transporters LmrCD and Pdr5. As a result, four of the new compounds were identified as being superior to zosuquidar in terms of transport reduction. The compound 10a, which was found to be the most efficient of all, features a cyclic acetal moiety derived from (R,R)-1,2-diphenyl-1,2-ethanediol.

### Experimental

General: Melting points were determined with a Büchi 540 melting point apparatus and are not corrected. NMR spectra were recorded in CDCl<sub>3</sub> with a Bruker Avance DRX 200 and a Bruker Avance DRX 500 spectrometer. IR spectra were measured with a Bruker Vector 22 spectrometer. Mass spectra were recorded on a Thermo Finnigan Trace DSQ apparatus (GCMS), an ion-trap API mass spectrometer Finnigan LCQ Deca (ESI), a triple-quadrupole-mass spectrometer Finnigan TSQ 7000 (EI), and a sector field mass spectrometer Finnigan MAT 8200 (EI, 70 eV). High-resolution mass spectroscopy was carried out on Bruker FT-ICR APEX III (7.0 T) (MALDI) at the University of Bielefeld. Column chromatography was performed with Fluka silica gel 60 (230-400 mesh), and thinlayer chromatography was carried out by using Merck TLC silicagel 60 F<sub>254</sub> aluminium sheets. Toluene was freshly distilled from sodium under nitrogen.

## General procedure for the reaction of piperidines **2b–13b** with (*R*)-**16**

A solution of (*R*)-16 (0.101 g, 0.50 mmol) and a piperidine 2b-13b (0.50 mmol) in ethanol (10 mL) was heated under reflux in a 25 mL flask for 3 h. After cooling to room temperature, the solvent was removed in a rotary evaporator, and the residue was purified by column chromatography on silica gel (chloroform/methanol, 10:1). Thus, the products 2a-13a were obtained as white or yellowish solids or oils.

### General procedure for the preparation of acetals **6c–12c**

Under nitrogen, a mixture of *N*-acetyl-4-piperidinone (1.13 g, 8.0 mmol), the corresponding diol (or dithiol), and *p*-toluenesulfonic acid monohydrate (0.095 g, 0.5 mmol) in chloroform (60 mL) was heated under reflux in a 100 mL flask, equipped with a Soxhlet extraction apparatus, which was filled with molecular sieves (4 Å). After heating under reflux for 10 to 15 h, the mixture was cooled to room temperature, a saturated aqueous solution of sodium hydrogen carbonate (20 mL) was added, and the mixture was stirred for 10 min at room temperature. The aqueous layer was removed, and the organic layer was dried with sodium sulfate. After removal of the solvent in a rotary evaporator, the residue was purified by flash chromatography. For the preparation of catechol-derived acetals **11c** and **12c**, toluene was used instead of *p*-toluenesulfonic acid.

### General procedure for the N-arylation of *N*-Boc-4-arylaminopiperidines **17** and **18**

A 25 mL two-necked flask, equipped with a magnetic stirrer and a connection to a combined argon/vacuum line was charged with piperidine **19** or **20** (3.6 mmol), the corresponding aryl bromide (3.6 mmol), sodium *tert*-butoxide (0.43 g, 4.5 mmol) and palladium acetate (10.1 mg, 0.045 mmol). The flask was closed with a septum and the air in the flask was replaced by nitrogen. Dry toluene (5 mL) was added, and the mixture was stirred at room temperature for 30 min. Then, a solution of tri*tert*-butylphosphane (0.12 mL, 0.5 mmol) in toluene (1 mL) was added, and the mixture was refluxed for 16 h under argon. After cooling to room temperature, water (10 mL) was added, and the organic layer was separated, dried with sodium sulfate and evaporated. The crude product was purified by column chromatography (ethyl acetate/cyclohexane, 1:2) to give the yellowish solid products.

### Supporting Information

Supporting Information File 1

Procedures. Spectroscopic and analytical data. [http://www.beilstein-journals.org/bjoc/content/ supplementary/1860-5397-8-193-S1.pdf]

### Acknowledgements

We are grateful to the Volkswagenstiftung for generous support of the project "Conformational dynamics of the multidrug resistance ABC-transporter LmrA modulated by substrates and inhibitors".

### References

- Gottesman, M. M.; Fojo, T.; Bates, S. E. Nat. Rev. Cancer 2002, 2, 48–58.
  - See for a review.
- Ambudkar, S. V.; Dey, S.; Hrycyna, C. A.; Ramachandra, M.; Pastan, I.; Gottesman, M. M. Ann. Rev. Pharmacol. Toxicol. 1999, 39, 361–398. doi:10.1146/annurev.pharmtox.39.1.361 See for a review.
- Lage, H. Curr. Drug Targets 2006, 7, 813–821. doi:10.2174/138945006777709566
- Tan, B.; Piwnica-Worms, D.; Ratner, L. Curr. Opin. Oncol. 2000, 12, 450–458. doi:10.1097/00001622-200009000-00011
- Krishna, R.; Mayer, L. D. *Eur. J. Pharm. Sci.* 2000, *11*, 265–283. doi:10.1016/S0928-0987(00)00114-7
- Pfister, J. R.; Makra, F.; Muehldorf, A. V.; Wu, H.; Nelson, J. T.; Cheung, P.; Bruno, N. A.; Casey, S. M.; Zutshi, N.; Slate, D. L. *Bioorg. Med. Chem. Lett.* **1995**, *5*, 2473–2476. doi:10.1016/0960-894X(95)00426-T
- Starling, J. J.; Shepard, R. L.; Cao, J.; Law, K. L.; Norman, B. H.; Kroin, J. S.; Ehlhardt, W. J.; Baughman, T. M.; Winter, M. A.; Bell, M. G.; Shih, C.; Gruber, J.; Elmquist, W. F.; Dantzig, A. H. *Adv. Enzyme Regul.* **1997**, *37*, 335–347. doi:10.1016/S0065-2571(96)00021-0
- Dantzig, A. H.; Law, K. L.; Cao, J.; Starling, J. J. Curr. Med. Chem. 2001, 8, 39–50.
- Sorbera, L. A.; Castañer, J.; Silvestre, J. S.; Bayés, M. Drugs Future 2003, 28, 125–136. doi:10.1358/dof.2003.028.02.719469
- Ding, P.-R.; Tiwari, A. K.; Ohnuma, S.; Lee, J. W. K. K.; An, X.; Dai, C.-L.; Lu, Q.-S.; Singh, S.; Yang, D.-H.; Talele, T. T.; Ambudkar, S. V.; Chen, Z.-S. *PLoS One* **2011**, *6*, e19329. doi:10.1371/journal.pone.0019329
- Lubelski, J.; de Jong, A.; van Merkerk, R.; Agustiandari, H.; Kuipers, O. P.; Kok, J.; Driessen, A. J. M. *Mol. Microbiol.* 2006, *61*, 771–781. doi:10.1111/j.1365-2958.2006.05267.x
- Kolaczkowski, M.; van der Rest, M.; Cybularz-Kolaczkowska, A.;
  Soumillion, J.-P.; Konings, W. N.; Goffeau, A. J. Biol. Chem. 1996, 271, 31543–31548. doi:10.1074/jbc.271.49.31543

- Infed, N.; Smits, S. H. J.; Dittrich, T.; Braun, M.; Driessen, A. J. M.; Hanekop, N.; Schmitt, L. *Mol. Membrane Biol.* submitted.
- Pfister, J. R.; Slate, D. L. 10,11-Methanodibenzosuberane derivatives used as chemosensitizing agents. WO Patent 9424107, Oct 27, 1994.
- 15. Sircar, I.; Haleen, S. J.; Burke, S. E.; Barth, H. *J. Med. Chem.* **1992**, *35*, 4442–4449. doi:10.1021/jm00101a022
- Suzuki, T.; Fukazawa, N.; San-nohe, K.; Sato, W.; Yano, O.; Tsuruo, T. J. Med. Chem. 1997, 40, 2047–2052. doi:10.1021/jm960869l
- Fukazawa, N.; Odate, M.; Suzuki, T.; Otsuka, K.; Tsuruo, T.; Sato, W. Novel heterocyclic compounds and anticancer-drug reinforcing agents containing them as effective components. Eur. Pat. 0363212, April 11, 1990.
- Fukazawa, N.; Suzuki, T.; Otsuka, K.; Yano, O.; Sato, W.; Tsuruo, T. Novel quinoline derivative and carcinostatic effect-enhancing agent containing the same as active ingredient. Japanese Patent 04235983, Aug 25, 1992.
- Lee, H.-S.; Yun, J.-W.; Kim, S.-J.; Kim, H.-C. Y. Process for the preparation of gylycidyl derivatives. WO Patent 2006019202, Feb 23, 2006.
- Barnett, C. J.; Huff, B.; Kobierski, M. E.; Letourneau, M.; Wilson, T. M. J. Org. Chem. 2004, 69, 7653–7660. doi:10.1021/j0049051v
- Thoma, G.; Nuninger, F.; Schaefer, M.; Akyel, K. G.; Albert, R.; Beerli, C.; Bruns, C.; Francotte, E.; Luyten, M.; MacKenzie, D.; Oberer, L.; Streiff, M. B.; Wagner, T.; Walter, H.; Weckbecker, G.; Zerwes, H.-G. *J. Med. Chem.* **2004**, *47*, 1939–1955. doi:10.1021/jm031046g
- 22. Prashad, M.; Mak, X. Y.; Liu, Y.; Repič, O. J. Org. Chem. 2003, 68, 1163–1164. doi:10.1021/jo020609d

### License and Terms

This is an Open Access article under the terms of the Creative Commons Attribution License (<u>http://creativecommons.org/licenses/by/2.0</u>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The license is subject to the *Beilstein Journal of Organic Chemistry* terms and conditions: (http://www.beilstein-journals.org/bjoc)

The definitive version of this article is the electronic one which can be found at: doi:10.3762/bjoc.8.193