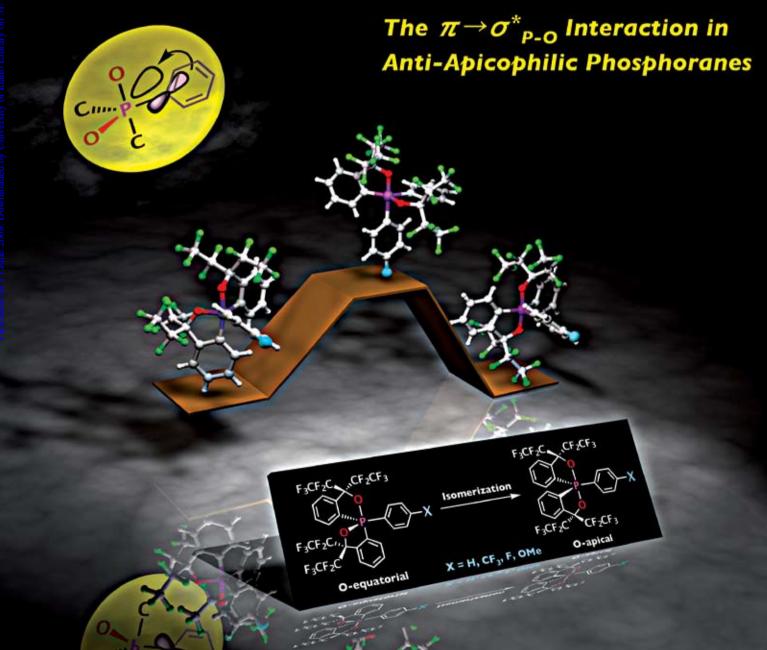
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Yamamoto *et al.* Synthesis, structure and isomerization of arylphosphoranes with anti-apicophilic bonding modes using a novel bidentate ligand with two C_2F_5 groups

PAPER

HOT PAPER

Linti and Seifert A decagallane(6) cluster Ga₁₀[Si(CMe₃)₃]₆ with an unprecedented structure

Synthesis, structure and isomerization of arylphosphoranes with anti-apicophilic bonding modes using a novel bidentate ligand with two C_2F_5 groups[†]

Xin-Dong Jiang, Shiro Matsukawa and Yohsuke Yamamoto*

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A series of anti-apicophilic pentacoordinate phosphoranes (with one chelating substituent in an O-equatorial, C-apical bonding mode at pentacoordinated phosphorus atom) bearing a *para*-substituted aryl group ($-C_6H_4(p-X)$; X = H, CF₃, F, OMe) or a mesityl (2,4,6-trimethylphenyl) group were isolated using a novel bulky bidentate ligand with two C_2F_5 groups. These phosphoranes were stable to isomerization at room temperature, and quantitatively converted into the corresponding more stable isomers (O-apical) at elevated temperatures in solution. On the basis of a kinetic study, the free energy of activation (ΔG^{\dagger}) of the stereomutation of the O-equatorial mesitylphosphorane to its O-apical isomer was higher than that of the CF₃ derivative by 2.6 kcal mol⁻¹, giving rise to a further example of the steric effect of the C₂F₅ group to freeze the isomerization of the O-equatorial *ortho*-unsubstituted derivatives ($-C_6H_4(p-X)$) to the corresponding O-apical isomers suggested that the O-equatorial isomers were stabilized by the $\pi \rightarrow \sigma^*_{P-O}$ interaction in the ground state.

Introduction

Hypervalent¹ 10-P-5² phosphorus compounds (phosphoranes) are regarded as intermediates (or transition states) in the formation or hydrolysis of biologically relevant phosphorus compounds³ such as DNA or RNA. Furthermore, they also play important roles as intermediates in the Wittig reaction.⁴ Phosphorane chemistry is crucially important to understand the mechanisms of the abovedescribed reactions, thus, a number of studies on phosphoranes have been carried out to establish many important properties.⁵ However, phosphorane chemistry is still complicated due to their kinetic and thermodynamic properties. Phosphoranes generally prefer a trigonal-bipyramid (TBP) structure which has two distinct bonds, *i.e.*, an apical bond and equatorial bond. The apical bond is described as a three-center-four-electron (hypervalent) bond whereas the equatorial bond is described as an sp² bond. The former is more polar than the latter, and the apical site is sterically more congested than the equatorial site,6 therefore, the electronegative and sterically less bulky groups prefer the apical site. The relative preference of substituents occupying the apical site is specifically known as apicophilicity.^{7,8} From a kinetic standpoint, on the other hand, a pentacoordinate molecule causes rapid stereomutations, giving rise to an exchange between the apical and equatorial ligand, therefore, such a molecule generally exists as an equilibrium mixture containing several (up to 20) stereoisomers in the solution state. This nondissociative intramolecular site exchange of substituents is usually advocated by the Berry pseudorotation (BPR).^{9,10} The barrier to the BPR is very low (calculated to be about 2-3 kcal mol⁻¹ for PH₅)¹¹ without any steric restrictions.

Using the rigid bidentate ligand called the Martin ligand (A),¹² our group succeeded in synthesizing the anti-apicophilic phosphoranes (with one chelating substituent in an O-equatorial, C-apical bonding mode at pentacoordinated phosphorus atom) having an alkyl monodentate ligand (O-equatorial 1) (Fig. 1).¹³ Unlike the case of other anti-apicophilic phosphoranes,¹⁴ these O-equatorial phosphoranes are kinetically stabilized products, and can still be converted into the corresponding more stable stereoisomers (O-apical 2). Although the Martin ligand is capable of stabilizing various types of hypervalent compounds both thermodynamically and kinetically, isolation of the O-equatorial phosphoranes bearing a small or electronegative monodentate ligand has still been troublesome because such compounds are isomerized too fast to be isolated. For example, the O-equatorial 1a (R = Me) could not be isolated in the pure form because the

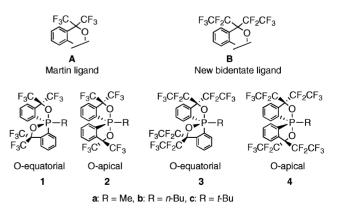


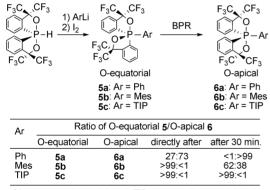
Fig. 1 Phosphoranes bearing bidentate ligands for freezing BPR.

Department of Chemistry, Graduate School of Science, Hiroshima University, 1-3-1 Kagamiyama, Higashi-Hiroshima, 739-8526, Japan. E-mail: yyama@sci.hiroshima-u.ac.jp; Fax: +81-82-424-0723

[†] CCDC reference numbers 672351 (12a), 672352 (12b), 672353 (12c), 672354 (12d), 672355 (12e), 672356 (13d) and 672357 (13e). For crystallographic data in CIF or other electronic format see DOI: 10.1039/b802947d

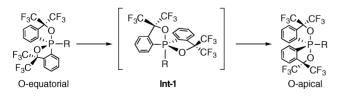
irreversible isomerization of **1a** to **2a** was relatively fast. Thus, we developed a bulky bidentate ligand with two C_2F_5 groups (**B**) in place of the CF₃ groups of the Martin ligand.¹⁵ This allowed us to isolate the O-equatorial methylphosphorane **3a** in the pure form, and the free energy of activation (ΔG^{\dagger}_{333}) of the isomerization of **3a** to **4a** was found to be greater than that of **1a** to **2a** by 3.6 kcal mol⁻¹. Furthermore, ligand **B** was also effective for freezing the isomerization of the "anti-apicophilic" 10-As-5 arsoranes (O-equatorial, C-apical) to the more stable stereoisomers (O-apical, C-equatorial).¹⁶

As a more electronegative monodentate ligand than the alkyl group, we chose any groups as the next targets. Although several attempts were made to isolate the O-equatorial arylphosphoranes 5 using the Martin ligand (A), it was found that these phosphoranes, except for the TIP (2,4,6-triisopropylphenyl) derivative 5c, rapidly isomerized to the O-apical counterparts (Scheme 1).¹⁷ For example, the isomerization of the phenyl derivative (5a to 6a) was completed within 30 min at r.t., which was in contrast to the O-equatorial alkyl derivatives (1). We assumed that the stereomutation of the O-equatorial to O-apical isomer passes through a high-energy intermediate (Int-1) which bears the monodentate ligand (R) at the apical site, therefore, the more apicophilic R could accelerate the isomerization (Scheme 2). Thus, the rapid isomerization was mainly due to the much higher apicophilicity of the aryl group than the alkyl group.^{7b,c,i,j} This required employing a very bulky TIP group to isolate the O-equatorial isomer.



Mes = 2,4,6-trimethylphenyl, TIP = 2,4,6-triisopropylphenyl.

Scheme 1 Synthesis of O-equatorial arylphosphoranes using the Martin ligand.



Scheme 2 High energy intermediate formed during the isomerization of the O-equatorial to O-apical isomer.

Theoretical studies have suggested that the π -donating substituents prefer to remain at the equatorial site of the TBP structure because such substituents are capable of interacting with the antibonding σ^* orbitals.⁸ Our group experimentally demonstrated that a dimethylamino group is less apicophilic than expected on the basis of its electronegativity, and this could be due to the existence of the $n_N \rightarrow \sigma^*_{P-C}$ interaction based upon the crystal structure of 7 (Fig. 2).^{7a} By use of a pair of O-equatorial and O-apical benzylphosphoranes, we subsequently demonstrated that the Oequatorial α -carbanion 8 was less basic than the corresponding O-apical isomer 9.¹⁸ This result was supported by the density functional calculations showing that the $n_C \rightarrow \sigma^*_{P-O}$ interaction of the O-equatorial α -anion 8 was much greater than the $n_C \rightarrow \sigma^*_{P-C}$ interaction of the corresponding O-apical isomer 9.¹⁸ Moreover, based on the kinetic study of the isomerization of the O-equatorial aminophosphorane 10 to the O-apical isomer 11, we succeeded in determining the energy of the $n_N \rightarrow \sigma^*_{P-O}$ interaction to be 4.4 kcal mol⁻¹, which was in good agreement with the calculated value (4.0 kcal mol⁻¹).¹⁹

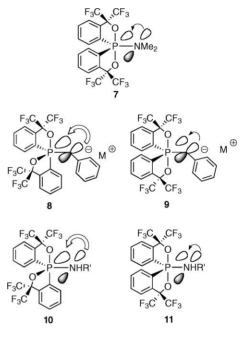
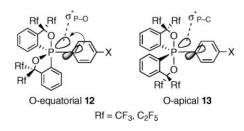
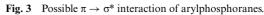


Fig. 2 The $n \to \sigma^*$ interaction of the $\alpha\text{-anions}$ and aminophosphoranes.

Consequently, the interaction between the π -orbital of the monodentate aromatic ligand and the σ^* orbital in the equatorial plane has been of significant interest. However, in the Martin ligand system (A), we isolated only the O-equatorial isomer with a TIP group (**5c**) as described above. Therefore, no insights into the $\pi \to \sigma^*$ interaction were established in the study. Thus, if the O-equatorial isomer (**12**) bearing an *ortho*-unsubstituted aryl monodentate ligand could be prepared (Fig. 3), and if the *para*-substituents (X) of the monodentate ligand affect the rate of the isomerization of the O-equatorial (**12**) to O-apical (**13**) isomer, the $\pi \to \sigma^*_{P-O}$ interaction in the O-equatorial isomer (**12**) could be estimated.



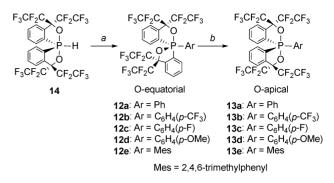


In this context, we determined to employ the bulky bidentate ligand B for stabilizing the anti-apicophilic O-equatorial arylphosphoranes (12). We now report the isolation of O-equatorial arylphosphoranes 12 using ligand B. These O-equatorial phosphoranes, even the ortho-unsubstituted derivatives (12a-d), were found to be stable at r.t., and could still be converted into the O-apical isomer 13 at elevated temperatures. The structures of the O-equatorial phosphoranes as well as the O-apical isomers were unambiguously determined by an X-ray crystallographic analysis. The kinetic study furnished the activation parameters for the stereomutation of the O-equatorial arylphosphoranes 12 to the O-apical isomers 13. Furthermore, the effect of the solvent and the para-substituent on the rate of the isomerization were also investigated in order to provide an insight into the $\pi \to \sigma^*_{P-0}$ interaction in the O-equatorial arylphosphoranes. Full details are given in the following sections.

Results and discussion

Synthesis and structure of the O-equatorial and O-apical arylphosphoranes

Using the new bidentate ligand **B**, the O-equatorial arylphosphoranes (12) were successfully synthesized by the reaction of the P–H spirophosphorane 14^{15a} with excess ArLi, followed by treatment with I₂ (Scheme 3).¹⁷ All of the O-equatorial 12 was quantitatively converted into the corresponding O-apical isomers 13 by heating in an organic solvent. Considering the fact that the O-equatorial arylphosphoranes 5 except for 5c (Ar = TIP) have not been isolated,¹⁷ it is noted that even the *ortho*-unsubstituted derivatives (12a–d) are isolable using ligand **B**, indicating that the



Scheme 3 Synthesis of the O-equatorial (12) and O-apical (13) arylphosphoranes. *Reagents and conditions*: (a) ArLi, Et₂O, r.t., 1 h, then I_2 , $-78 \degree C$ to 0 °C, 1 h, 12a: 85%, 12b: 58%, 12c: 76%, 12d: 48%, 12e: 83%; (b) C₆D₆, 80 °C, 8 h, 13a: 98%, 13b: 96%, 13c: 96%, 13d: 97%, 13e: 95%.

steric effect of the C_2F_5 groups for freezing the isomerization is also remarkable in the present case.

The structures of the O-equatorial (12a–e) and O-apical (13d and 13e) phosphoranes were confirmed by an X-ray crystallographic analysis (Fig. 4). All compounds have a distorted trigonalbipyramidal (TBP) structure, in which the aryl monodentate ligand occupies one of the equatorial sites. Two independent molecules were found in the unit cell for each of the 12a, 12c and 12e. The angles and distances involving the phosphorus atom of 12 and 13 (Table 1) were similar to those of the corresponding alkylphosphoranes 3 and 4, respectively. For the O-equatorial isomers, the equatorial O2–P–C1 angle of 12a–d (12a: 120.5°, 124.0°; 12b: 122.5°; 12c: 121.7°, 124.1°; 12d: 123.6°) is only slightly larger than those of the alkylphosphoranes (3a: 119.5°; 3b: 119.7°; 3c: 118.0°),^{15a} although those of the mesityl derivative 12e (112.8°,

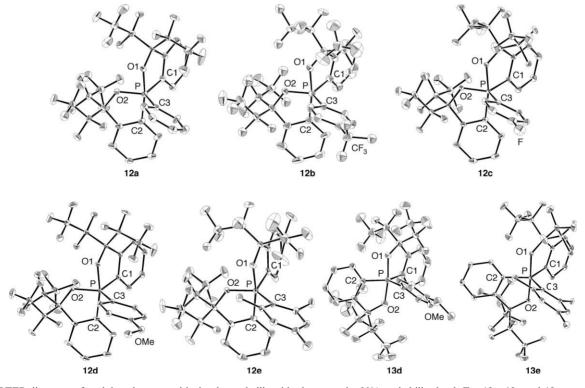


Fig. 4 ORTEP diagrams of arylphosphoranes with the thermal ellipsoids shown at the 30% probability level. For 12a, 12c and 12e, one of the two independent molecules is shown. All hydrogen atoms are omitted for clarity.

Table 1 Selected bond lengths (Å) and angles (°) for the O-equatorial (12) and O-apical (13) phosphoranes

Compound	12a ^{<i>a</i>}	12b	12c ^{<i>a</i>}	12d	12e ^{<i>a</i>}	13d	13e
P01	1.775(4), 1.807(4)	1.788(5)	1.7870(12), 1.7800(12)	1.7742(14)	1.7781(19), 1.7845(18)	1.7435(13)	1.7663(10)
PO2	1.666(5), 1.655(4)	1.643(5)	1.6501(12), 1.6564(12)	1.6654(13)	1.6636(18), 1.6722(18)	1.7526(13)	1.7663(10)
P-C1	1.824(7), 1.823(7)	1.813(7)	1.8215(17), 1.8229(17)	1.8264(17)	1.828(3), 1.817(3)	1.8335(18)	1.8298(15)
P-C2	1.889(6), 1.900(6)	1.878(7)	1.8805(17), 1.8763(18)	1.8810(19)	1.890(3), 1.887(3)	1.8294(17)	1.8282(14)
P-C3	1.825(6), 1.820(7)	1.841(7)	1.8245(17), 1.8191(17)	1.8125(18)	1.853(3), 1.850(2)	1.8045(18)	1.8422(15)
O1-P-O2	83.7(2), 81.32(19)	83.0(2)	82.26(6), 82.76(6)	82.34(6)	83.75(9), 82.60(9)	167.89(7)	174.02(5)
O1-P-C1	86.7(2), 86.6(2)	86.4(3)	87.06(7), 87.12(7)	86.95(7)	86.76(11), 87.12(10)	87.16(8)	86.85(6)
O1–P–C2	171.5(3), 168.6(2)	171.3(3)	170.13(7), 170.22(7)	169.59(8)	170.73(11), 169.61(10)	88.47(7)	90.68(6)
O1-P-C3	87.7(2), 89.5(2)	88.7(3)	89.17(7), 88.69(7)	89.20(7)	87.89(10), 88.06(10)	95.94(7)	93.52(6)
O2-P-C1	120.5(2), 124.0(2)	122.5(3)	121.70(7), 124.12(7)	123.63(7)	112.78(11), 115.18(10)	88.89(8)	90.28(6)
O2–P–C2	87.9(3), 87.3(2)	88.4(3)	87.97(7), 87.53(7)	87.25(8)	87.23(10), 87.14(10)	87.12(7)	86.72(6)
O2-P-C3	116.3(3), 116.9(3)	116.2(3)	118.47(7), 119.00(7)	117.75(7)	121.26(11), 125.08(11)	96.17(7)	92.46(6)
C1-P-C2	98.9(3), 99.4(3)	99.6(3)	99.32(8), 99.29(8)	99.12(8)	98.73(12), 98.81(11)	139.54(8)	125.52(7)
C1-P-C3	121.8(3), 117.4(3)	119.9(3)	118.48(8), 115.50(8)	117.24(8)	124.64(12), 118.23(11)	110.17(8)	118.85(6)
C2-P-C3	94.7(2), 96.2(3)	93.7(3)	94.32(7), 95.12(8)	95.43(8)	95.00(11), 96.52(11)	110.29(8)	115.63(7)

115.2°) are apparently smaller than those of **12a–d** and comparable to that of the O-equatorial TIP derivative **5c** (114.0°).¹⁷ Similarly, for the O-apical phosphoranes, the equatorial C1–P–C2 angle of **13e** (125.5°) is smaller than that of **13d** (139.5°). This is mainly due to the steric repulsions between the bulky mesityl group and the aromatic rings of the bidentate ligand.

The degree of the tilt of the monodentate aryl ring can be represented by the tilt angle θ , which is the dihedral angle between the root mean plane consisting of P and the equatorial atoms (C1, O2 and C3 for 12, C1, C2 and C3 for 13) and that consisting of the six atoms of the monodentate aromatic ring (Fig. 5 and Table 2). For the *ortho*-unsubtituted O-equatorial derivatives (12a, 12b, 12c and 12d), θ ranges from 14° to 33°, and the degree of the tilt is independent of the electronic property of the *para*-substituent. In addition, the two independent molecules for 12c have different tilt angles (27.7° and 14.1°). This implies that the $\pi \to \sigma^*$ interactions in these arylphosphoranes are not strong enough to dominate the tilt angle of the crystal structure. Comparing the

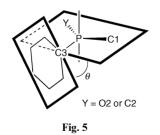


Table 2 Tilt angles θ of phosphoranes

Compound	Ar	Y	$\theta / ^{\circ}$
12a	Ph	O2	32.9, 29.8 ^a
12b	$C_6H_4(p-CF_3)$	O2	24.4
12c	$C_6H_4(p-F)$	O2	27.7, 14.1 ^a
12d	$C_6H_4(p-OMe)$	O2	18.8
12e	Mes	O2	$24.8, 20.4^{a}$
13d	$C_6H_4(p-OMe)$	C2	42.6
13e	Mes	C2	34.0

" Tilt angles for the two independent molecules are shown.

O-equatorial and O-apical isomers, the former has a smaller θ than the latter probably due to steric repulsions originating from the apical aromatic ring of the bidentate ligand. Overall, the crystal structures are mainly governed by steric reasons.

Kinetic study of the isomerization of O-equatorial 12 to O-apical 13

The rates of the isomerization of **12** to **13** were measured by monitoring the change in the integrals of the ¹⁹F NMR signals of the trifluoromethyl group, and the rates satisfied first-order kinetics. At first, in order to investigate the solvent effect for the isomerization, measurements of the stereomutation of **12a** to **13a** were carried out at 50 °C using DMSO- d_6 , CD₃CN, MeOH and C₆D₆. The rates and solvent parameters (ε_r , μ and E_T^N)²⁰ are summarized in Table 3. The rates were found to be slightly affected by the polarity of the solvents, in which the polar solvent decelerated the isomerization.

In order to investigate the *p*-substituent effect, the rates of the isomerization for **12a–d** were then measured in C₆D₆ at 50 °C (Table 4). Although the rates of **12a–d** ((2.77, 3.77, 2.57 and 2.02) × 10^{-4} s⁻¹ for **12a**, **12b**, **12c** and **12d**, respectively) were of the same order of magnitude, it was found that the stereomutation was decelerated by the electron-donating OMe group and accelerated by the electron-withdrawing CF₃ group. Table 4 shows the rate constants along with the substituent parameters (σ_p , σ_R and σ_1).²¹ Although the isomerization rates (*k*) of **12** to **13** do not nicely correlate with each substituent parameter, the resonance

Table 3The rates for the stereomutation from O-equatorial 12a to O-apical 13a in various solvents at 323 K

Solvent	$\mathcal{E}_{\mathrm{r}}^{a}$	$10^{30}\mu^a/\mathrm{C}\mathrm{m}$	$E_{\mathrm{T}}{}^{\mathrm{N}a}$	k^{b}/s^{-1}
$DMSO-d_6$ CD_3CN $MeOH$ C_6D_6	46.45 35.94 32.66 2.27	13.5 13.0 9.6 0.0	0.444 0.460 0.762 0.111	$\begin{array}{c} (0.84\pm 0.05)\times 10^{-4} \\ (1.51\pm 0.03)\times 10^{-4} \\ (1.90\pm 0.01)\times 10^{-4} \\ (2.77\pm 0.03)\times 10^{-4} \end{array}$

^a Values of undeuterated solvents are shown. ^b Error is given as the standard deviation.

Table 4 Effect of *p*-substituents on the isomerization rate in C_6D_6 at 323 K

Process	Х	σ_p	$\sigma_{ m R}$	σ_{I}	$10^4 k/s^{-1}$
$\begin{array}{c} 12a \rightarrow 13a \\ 12b \rightarrow 13b \\ 12c \rightarrow 13c \\ 12d \rightarrow 13d \end{array}$	H CF ₃ F OMe	0.00 0.51 0.06 -0.28	$0.00 \\ 0.11 \\ -0.48 \\ -0.58$	$0.00 \\ 0.40 \\ 0.54 \\ 0.30$	$\begin{array}{c} 2.77 \pm 0.03 \\ 3.77 \pm 0.03 \\ 2.57 \pm 0.03 \\ 2.02 \pm 0.02 \end{array}$

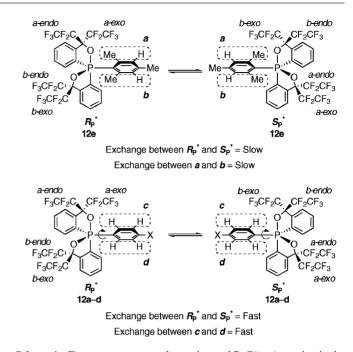
substituent parameter ($\sigma_{\rm R}$) seemed to be a greater contribution than the inductive constant ($\sigma_{\rm I}$). Thus, we carried out a multiregression analysis using $\sigma_{\rm R}$ and $\sigma_{\rm I}$ as variants, and obtained the following equation with a very good fit of $R^2 = 0.996$.

$$k = (2.31\sigma_{\rm R} + 1.72\sigma_{\rm I} + 2.80) \times 10^{-4}$$

This equation shows that the contribution of the resonance effect on the isomerization rate is 1.3 times greater than that of the inductive effect. Since the σ^*_{P-O} orbital of the O-equatorial phosphorane has found to be significantly lower in energy than the σ^*_{P-C} orbital of the O-apical isomer,¹⁸ the ground state (*i.e.*, TBP structure) of the O-equatorial *ortho*-unsubstituted derivatives (**12a–d**) should be somewhat stabilized by the $\pi \rightarrow \sigma^*_{P-O}$ interaction, whereas such an interaction can be negligible for the O-apical isomers (Fig. 3). In any case, this result agrees with the fact that the π -donating nature reduces the apparent apicophilicity of the substituent although the degree of the interaction is not significant.

The reason for the rather small interaction could be due to the small tilt angle described above (Fig. 5). Therefore, we investigated the structure in solution based on the ³¹P and ¹⁹F NMR. All the isolated phosphoranes showed singlets in ³¹P NMR (CDCl₃), and the chemical shifts for the O-equatorial phosphoranes ($\delta = -10.6$ (12a), -11.4 (12b), -11.1 (12c), -10.7 (12d) and -2.6 (12e) ppm) were shifted downfield compared to those of the O-apical isomers $(\delta = -26.9 \text{ (13a)}, -28.2 \text{ (13b)}, -27.7 \text{ (13c)} \text{ and } -27.1 \text{ (13d)} \text{ and}$ -23.6 (13e) ppm) similar to our previous results.^{13,15,17-19} The ¹⁹F NMR spectra of the O-apical phosphoranes (13) showed only two CF_3 signals due to their C_2 symmetrical structure. For the O-equatorial isomers, four CF₃ signals were observed for $12e (\delta =$ -78.1, -78.4, -79.1 and -79.2 ppm) at room temperature or even at 60 °C, whereas there were only two CF₃ signals for **12a–d**. As we have already reported, for the O-equatorial phosphoranes bearing a bulky monodentate ligand (5b and 5c), the four chemically nonequivalent CF3 groups have been observed distinctly in the 19F NMR. If a fast exchange between the $R_{\rm P}^*$ and $S_{\rm P}^*$ isomers takes place,²² the two endo-CF₃ groups (*a-endo*-CF₃ and *b-endo*-CF₃) coalesce into one, and the same occurs for the exo-CF₃ groups (aexo-CF₃ and *b*-exo-CF₃). Therefore, this observation implies that the one-step stereomutation between the $R_{\rm P}^*$ and $S_{\rm P}^*$ enantiomers of 12e is slow on the NMR timescale (Scheme 4).

In the ¹H NMR, two kinds of *ortho*-methyl protons ($\delta = 2.65$ (s, 3H) and 2.27 ppm (s, 3H)) as well as *meta*-protons ($\delta = 6.81$ (d, ⁴ $J_{\rm H,P} = 4.4$ Hz, 1H), 6.73 ppm (d, ⁴ $J_{\rm H,P} = 8.3$ Hz, 1H)) were clearly observed for **12e**, whereas only a set of *ortho*- and *meta*-aromatic protons was observed for **12a–d**. That is, sites *a* and *b* of **12e** are not equivalent, whereas sites *c* and *d* of **12a–d** are equivalent in the ¹H NMR (Scheme 4). If the conformation of the monodentate ligand is fixed, sites *a* (or *c*) and *b* (or *d*) are not exchanged with each other upon the one-step pseudorotation



Scheme 4 Frozen one-step pseudorotation and P–C(*ipso*) rotation in the O-equatorial mesitylphophorane (12e)

between R_P^* and S_P^* . These observations indicate that the P– C(*ipso*) rotation in the mesityl derivative (**12e**) is slow and that in the *ortho*-unsubstituted derivatives (**12a**–**d**) is fast on the NMR timescale. Therefore, the degree of freedom in the P–C(*ipso*) bond rotation is higher for **12a**–**d** than for **12e** in the solution state. Thus, the *ortho*-unsubstituted aryl group of **12a**–**d** should be capable of being nearly perpendicular to the equatorial plane, whereas the mesityl group of **12e** is most likely fixed as the crystal structure even in solution. Therefore, the $\pi \to \sigma^*_{P-O}$ interaction should work well for the O-equatorial *ortho*-unsubstituted derivative (**12a**–**d**), giving rise to stabilization of the ground states of **12a–d**.

In order to investigate the difference between the *ortho*unsubstituted aryl and mesityl groups, the rates of the isomerization of the phenyl (**12a** to **13a**) and the mesityl (**12e** to **13e**) derivatives were measured in the temperature range of 313–333 K (Table 5), and obtained the activation parameters from the Eyring plot (Fig. 6). The free energy of activation of the mesityl derivative

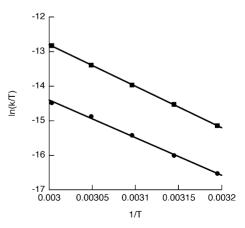


Fig. 6 Eyring plot for the isomerization in C_6D_6 . Squares: $12a \rightarrow 13a$; circles: $12e \rightarrow 13e$.

Process	T/K	k/s^{-1}	$\Delta H^{\ddagger}/\mathrm{kcal}\ \mathrm{mol}^{-1}$	ΔS^{\ddagger} /cal K ⁻¹ mol ⁻¹	$\Delta G^{\ddagger}_{_{298}}$ /kcal mol ⁻¹
$12a \rightarrow 13a (Ar = Ph)$	313	$(0.81 \pm 0.01) \times 10^{-4}$	23.9 ± 0.2	-0.6 ± 0.6	24.1
	318	$(1.56 \pm 0.01) \times 10^{-4}$			
	323	$(2.77 \pm 0.03) \times 10^{-4}$			
	328	$(5.00 \pm 0.04) \times 10^{-4}$			
	333	$(8.96 \pm 0.10) \times 10^{-4}$			
$12e \rightarrow 13e$ (Ar = Mes)	313	$(2.08 \pm 0.03) \times 10^{-5}$	21.6 ± 0.6	-10.8 ± 1.9	24.8
	318	$(3.56 \pm 0.02) \times 10^{-5}$			
	323	$(6.49 \pm 0.06) \times 10^{-5}$			
	328	$(11.3 \pm 0.03) \times 10^{-5}$			
	333	$(17.1 \pm 0.20) \times 10^{-5}$			

Table 5 Activation parameters of the stereomutation in $C_6 D_6^{a}$

 $(\Delta G^{\ddagger}_{298} = 24.8 \text{ kcal mol}^{-1})$ is slightly higher than that of the phenyl derivative (24.1 kcal mol}^{-1}). When compared to the CF₃ derivative, the free energy of activation for **12e** to **13e** ($\Delta G^{\ddagger}_{298} = 24.8 \text{ kcal mol}^{-1}$) was higher than that of **5b** to **6b** ($\Delta G^{\ddagger}_{298} = 22.2 \text{ kcal mol}^{-1}$) by 2.6 kcal mol}^{-1}, and was comparable to that of the TIP derivative **5c** to **6c** ($\Delta G^{\ddagger}_{298} = 24.1 \text{ kcal mol}^{-1}$).^{17b} This again proves the steric hindrance of the C₂F₅ group being more effective for freezing the isomerization than that of the CF₃ group.

As for the activation enthalpy (ΔH^{\ddagger}) of the phenyl derivative $(12a \rightarrow 13a)$ and of the mesityl derivative $(12e \rightarrow 13e)$, the former is greater than the latter by 2.3 kcal mol⁻¹ in clear contrast to the free energy of activation. When considering the high-energy intermediates in the three-step BPR from the O-equatorial 12 to the O-apical 13,¹³ the mesityl derivative (Int-2e) could be less stable than the phenyl derivative (Int-2a) because the former has a bulky mesityl group at the sterically congested apical site (Fig. 7). Therefore, the difference in enthalpy has to be explained by some factors in the ground state because the activation enthalpy is lower in the mesityl derivative $(12e \rightarrow 13e)$ than in the phenyl derivative $(12a \rightarrow 13a).$ The $\pi \rightarrow \sigma^*_{\text{P-O}}$ stabilizing interaction in 12a can be one of the reasons for the higher activation enthalpy for $12a \rightarrow$ 13a. The activation entropy (ΔS^{\ddagger}) for the mesityl derivative (12e \rightarrow 13e) is *lower* than that for the phenyl derivative $(12a \rightarrow 13a)$. The steric repulsion between the bulky mesityl group and the bidentate ligands even in Int-2e could be the reason.

Conclusions

The new bidentate ligand with two C_2F_5 groups was used to isolate the O-equatorial arylphosphoranes **12** (Ar = Ph, C₆H₄(*p*-CF₃), C₆H₄(*p*-F), C₆H₄(*p*-OMe) and Mes), which could be quantitatively converted into the corresponding O-apical isomers **13** by heating at elevated temperatures. The structures were confirmed by an X-ray analysis, showing that the tilt angle of the monodentate ligand was found to be independent of the *para*-substituents. However, the kinetic study showed a small *para*-substituent effect on the stereomutations. The multi-regression analysis for the *para*substituents revealed that the contribution of the resonance effect (σ_R) on the rate constant was superior to the inductive effect (σ_1). This indicates that the $\pi \rightarrow \sigma^*_{P-O}$ stabilizing interaction in the O-equatorial isomer plays some role in the isomerization. This result shows a general insight into the stereoelectronic effect of the pentacoordinated molecules.

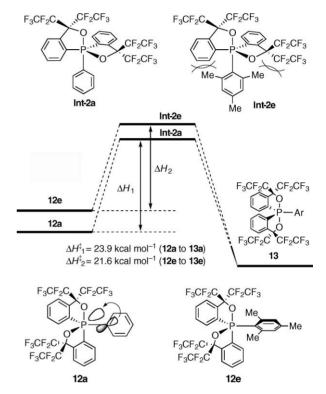


Fig. 7 Energy diagram for the isomerization of the O-equatorial phosphoranes to the O-apical isomers.

Experimental

General

Melting points were measured using a Yanaco micro melting point apparatus. The ¹H NMR (400 MHz), ¹⁹F NMR (376 MHz), ³¹P NMR (162 MHz) were recorded using JEOL EX-400 or JEOL AL-400 spectrometers. The ¹H NMR chemical shifts (δ) are given in ppm downfield from Me₄Si, determined by residual chloroform ($\delta = 7.26$ ppm). The ¹⁹F NMR chemical shifts are given in ppm downfield from the external CFCl₃. The ³¹P NMR chemical shifts are given in ppm downfield from the external 85% H₃PO₄. The elemental analyses were performed using a Perkin-Elmer 2400 CHN elemental analyzer. All reactions were carried out under N₂. Diethyl ether (Et₂O) was freshly distilled from Na/benzophenone, and the other solvents were distilled from CaH_2 . Merck silica gel $60GF_{254}$ was used for the preparative thin-layer chromatography.

General procedure for the synthesis of O-equatorial arylphosphoranes

Synthesis of 12a. PhLi (1.14 M cyclohexane-Et₂O solution, 0.19 mL, 0.216 mmol) was added to a solution of P-H phosphorane 14 (49.9 mg, 0.0696 mmol) in Et₂O (3.0 mL) at -78 °C. The mixture was then stirred for 1 h at room temperature. I_2 (52.0 mg, 0.204 mmol) was added to the mixture at -78 °C, and the mixture was stirred for 1 h at 0 °C. The reaction was then quenched with aqueous $Na_2S_2O_3$ (30 mL). The mixture was extracted with Et_2O (2 × 40 mL), and the organic layer was washed with brine $(2 \times 40 \text{ mL})$ and dried over anhydrous MgSO₄. After removing the solvents by evaporation, the resulting crude product was separated by preparative TLC (*n*-hexane– $CH_2Cl_2 = 7:1$) to afford 12a (47.0 mg, 0.0593 mmol, 85%) as a white solid. Colorless crystals of 12a suitable for the X-ray analysis were obtained by recrystallization from n-hexane-CH2Cl2. Mp 115.0-116.0 °C (decomp.). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.95-7.99$ (m, 2 H, Ar-H), 7.74 (br s, 2 H, Ar-H), 7.59-7.62 (m, 4 H, Ar-H), 7.49-7.55 (m, 2 H, Ar-H), 7.24-7.28 (m, 3 H, Ar-H) ppm. ¹⁹F NMR $(376 \text{ MHz}, \text{CDCl}_3)$: $\delta - 78.8$ (s, 6 F, CF₂CF₃), -79.4 (dd, ${}^{3}J_{\text{FF}} =$ 9.8 Hz, ${}^{5}J_{F,F} = 9.8$ Hz, 6 F, CF₂CF₃), -114.8 (d, ${}^{2}J_{F,F} = 295.6$ Hz, 2 F, CF_2CF_3), -116.1 (d, ${}^{2}J_{F,F}$ = 295.6 Hz, 4 F, CF_2CF_3), -118.4 $(dm, {}^{2}J_{F,F} = 295.6 \text{ Hz}, 2 \text{ F}, CF_{2}CF_{3}) \text{ ppm}. {}^{31}\text{P} \text{ NMR} (162 \text{ MHz},$ CDCl₃): δ –10.6 ppm. Anal. Found: C, 42.43; H, 1.26. Calc. for C₂₈H₁₃F₂₀O₂P: C, 42.44; H, 1.65%.

Synthesis of 12b. Following the above-described procedure using *p*-bromobenzotrifluoride (d = 1.607 g mL⁻¹, 0.040 mL, 0.29 mmol) and 14 (51.8 mg, 0.0723 mmol), 12b (36.0 mg, 0.0418 mmol, 58%) was isolated as a white solid. Colorless crystals of 12b suitable for the X-ray analysis were obtained by recrystallization from n-hexane-CH2Cl2. Mp 106.0-107.0 °C (decomp.). ¹H NMR (400 MHz, CDCl₃): δ 7.92–7.96 (m, 2 H, Ar-H), 7.76 (br s, 2 H, Ar-H), 7.62-7.66 (m, 4 H, Ar-H), 7.61 $(dd, {}^{3}J_{H,P} = 15.0 \text{ Hz}, {}^{3}J_{H,H} = 8.3 \text{ Hz}, 2 \text{ H}, \text{Ar-H}), 7.53 (d, {}^{4}J_{H,P} =$ 7.8 Hz, ${}^{3}J_{H,H} = 8.3$ Hz, 2 H, Ar–H) ppm. 19 F NMR (376 MHz, CDCl₃): δ -63.5 (s, 3 F, Ar-CF₃), -78.8 (s, 6 F, CF₂CF₃), -79.5 $(dd, {}^{3}J_{EF} = 9.8 \text{ Hz}, {}^{5}J_{EF} = 9.8 \text{ Hz}, 6 \text{ F}, \text{CF}_2\text{CF}_3), -114.8 (d, {}^{2}J_{EF} =$ 292.0 Hz, 2 F, CF_2CF_3), -116.2 (br dm, ${}^2J_{F,F} = 292.0$ Hz, 4 F, CF_2CF_3 , -118.4 (dm, ${}^{2}J_{E,F} = 292.0$ Hz, 2 F, CF_2CF_3) ppm. ${}^{31}P$ NMR (162 MHz, CDCl₃): δ –11.4 ppm. Anal. Found: C, 40.19; H, 1.01. Calc. for C₂₉H₁₂F₂₃O₂P: C, 40.49; H, 1.41%.

Synthesis of 12c. Following the above-described procedure using *p*-bromofluorobenzene (d = 1.593 g mL⁻¹, 0.040 mL, 0.36 mmol) and **14** (47.7 mg, 0.0665 mmol), **12c** (41.2 mg, 0.0508 mmol, 76%) was isolated as a white solid. Colorless crystals of **12c** suitable for the X-ray analysis were obtained by recrystallization from *n*-hexane–CH₂Cl₂. Mp 112.5–113.5 °C (decomp.). ¹H NMR (400 MHz, CDCl₃): δ 7.93–7.98 (m, 2 H, Ar–H), 7.76 (br d, ${}^{3}J_{\text{H,H}} = 8.0$ Hz, 2 H, Ar–H), 7.61–7.64 (m, 4 H, Ar–H), 7.54 (ddd, ${}^{3}J_{\text{H,F}} = 15.0$ Hz, ${}^{3}J_{\text{H,H}} = 9.0$ Hz, ${}^{4}J_{\text{H,F}} = 3.6$ Hz, 2 H, Ar–H) pm. ¹⁹F NMR (376 MHz, CDCl₃): δ -78.8 (s, 6 F, CF₂CF₃), -79.5 (dd, ${}^{3}J_{\text{H,F}} = 9.8$ Hz, ${}^{5}J_{\text{F,F}} = 9.8$ Hz, 6 F, CF₂CF₃), -111.0 (tt, ${}^{3}J_{\text{H,F}} = 9.0$ Hz, ${}^{4}J_{\text{H,F}} = 6.0$ Hz, 1 F,

Ar-F), -114.9 (d, ${}^{2}J_{F,F} = 292.0$ Hz, 2 F, $CF_{2}CF_{3}$), -116.1 (d, ${}^{2}J_{F,F} = 292.0$ Hz, 4 F, $CF_{2}CF_{3}$), -118.5 (dm, ${}^{2}J_{F,F} = 292.0$ Hz, 2 F, $CF_{2}CF_{3}$) ppm. 31 P NMR (162 MHz, CDCl₃): δ -11.1 ppm. Anal. Found: C, 41.48; H, 1.18. Calc. for $C_{28}H_{12}F_{21}O_{2}P$: C, 41.50; H, 1.49%.

Synthesis of 12d. Following the above-described procedure using *p*-bromoanisole (d = 1.494 g mL⁻¹, 0.060 mL, 0.48 mmol) and 14 (62.5 mg, 0.0872 mmol), 12d (34.7 mg, 0.0421 mmol, 48%) was isolated as a white solid. Colorless crystals of 12d suitable for the X-ray analysis were obtained by recrystallization from n-hexane-CH₂Cl₂. Mp 124.0-125.0 °C (decomp.). ¹H NMR (400 MHz, CDCl₃): δ 7.96-8.03 (m, 2 H, Ar-H), 7.76 (br s, 2 H, Ar–H), 7.58–7.61 (m, 4 H, Ar–H), 7.50 (dd, ${}^{3}J_{H,P} = 15.0$ Hz, ${}^{3}J_{H, H} = 8.8 \text{ Hz}, 2 \text{ H}, \text{Ar-H}), 6.78 (d, {}^{4}J_{H, P} = 7.8 \text{ Hz}, {}^{3}J_{H, H} = 8.8 \text{ Hz},$ 2 H, Ar-H), 3.74 (s, 3 H, Ar-OCH₃) ppm. ¹⁹F NMR (376 MHz, CDCl₃): δ -78.8 (s, 6 F, CF₂CF₃), -79.4 (dd, ${}^{3}J_{F,F} = 9.8$ Hz, ${}^{5}J_{\rm E,F} = 9.8$ Hz, 6 F, CF₂CF₃), -115.0 (d, ${}^{2}J_{\rm E,F} = 292.0$ Hz, 2 F, CF_2CF_3), -116.2 (br d, ${}^2J_{F,F}$ = 292.0 Hz, 4 F, CF_2CF_3), -118.5 $(dm, {}^{2}J_{E,F} = 292.0 \text{ Hz}, 2 \text{ F}, CF_{2}CF_{3}) \text{ ppm. }{}^{31}\text{P NMR} (162 \text{ MHz},$ $CDCl_3$): δ -10.7 ppm. Anal. Found: C, 42.05; H, 1.60. Calc. for C₂₉H₁₅F₂₀O₃P: C, 42.35; H, 1.84%.

Synthesis of 12e. Following the above-described procedure using 2-bromomesitylene ($d = 1.301 \text{ g mL}^{-1}$, 0.10 mL, 0.65 mmol) and 14 (71.3 mg, 0.0995 mmol), 12e (69.2 mg, 0.0829 mmol, 83%) was isolated as a white solid. Colorless crystals of 12e suitable for the X-ray analysis were obtained by recrystallization from CH₃CN. Mp 113.1–113.9 °C. ¹H NMR (400 MHz, CDCl₃): δ 7.95–7.98 (m, 1 H, Ar–H), 7.81 (br d, ${}^{3}J_{H, H} = 8.0$ Hz, 1 H, Ar–H), 7.69 (td, ${}^{3}J_{H,H} = 8.0$ Hz, ${}^{4}J_{H,H} = 1.2$ Hz, 1 H, Ar–H), 7.61 (br d, ${}^{3}J_{H,H} = 8.0$ Hz, 1 H, Ar–H), 7.44–7.53 (m, 4 H, Ar–H), 6.81 (d, ${}^{4}J_{H,P} = 4.4$ Hz, 1 H, Ar–H), 6.73 (d, ${}^{4}J_{H,P} = 8.3$ Hz, 1 H, Ar-H), 2.65 (s, 3 H, Ar-CH₃), 2.27 (s, 3 H, Ar-CH₃), 2.18 (s, 3 H, Ar-CH₃) ppm. ¹⁹F NMR (376 MHz, CDCl₃): δ -78.1 (s, 3 F, CF₂CF₃), -78.4 (s, 3 F, CF₂CF₃), -79.1 (s, 3 F, CF₂CF₃), -79.2 (s, 3 F, CF_2CF_3), -112.9 (d, ${}^2J_{F,F}$ = 292.0 Hz, 1 F, CF_2CF_3), -113.8 $(d, {}^{2}J_{F,F} = 292.0 \text{ Hz}, 1 \text{ F}, CF_{2}CF_{3}), -114.1 (d, {}^{2}J_{F,F} = 292.0 \text{ Hz},$ 2 F, CF₂CF₃), -114.9 (d, ${}^{2}J_{F,F} = 292.0$ Hz, 1 F, CF₂CF₃), -115.3 $(d, {}^{2}J_{E,F} = 292.0 \text{ Hz}, 1 \text{ F}, CF_{2}CF_{3}), -115.7 (d, {}^{2}J_{E,F} = 292.0 \text{ Hz}, 1$ F, CF₂CF₃), -117.8 (d, ${}^{2}J_{F,F} = 292.0$ Hz, 1 F, CF₂CF₃) ppm. ${}^{31}P$ NMR (162 MHz, CDCl₃): δ –2.7 ppm. Anal. Found: C, 44.54; H, 2.07. Calc. for C₃₁H₁₉F₂₀O₂P: C, 44.62; H, 2.30%.

Synthesis of 13a. A solution of **12a** (12.2 mg, 0.0153 mmol) in C₆D₆ (1.0 mL) was heated at 80 °C for 8 h. After concentration *in vacuo*, **13a** was obtained (12.0 mg, 0.0151 mmol, 98%) as a white solid. Mp 149.1–150.0 °C. ¹H NMR (400 MHz, CDCl₃): δ 8.70–8.78 (m, 2 H, Ar–H), 7.69–7.82 (m, 6 H, Ar–H), 7.41 (dd, ³*J*_{H,P} = 15.8 Hz, ³*J*_{H,H} = 8.0 Hz, 2 H, Ar–H), 7.18–7.27 (m, 3 H, Ar–H) ppm. ¹⁹F NMR (376 MHz, CDCl₃): δ –78.3 (s, 6 F, CF₂CF₃), –79.8 (dd, ³*J*_{E,F} = 19.7 Hz, ⁵*J*_{E,F} = 6.2 Hz, 6 F, CF₂CF₃), –116.7 (s, 2 F, CF₂CF₃), –116.8 (s, 2 F, CF₂CF₃), –116.8 (dq, ²*J*_{E,F} = 285.8 Hz, ³*J*_{E,F} = 19.7 Hz, 2 F, CF₂CF₃), –121.9 (dq, ²*J*_{E,F} = 285.8 Hz, ⁵*J*_{E,F} = 6.2 Hz, 2 F, CF₂CF₃) ppm. ³¹P NMR (162 MHz, CDCl₃): δ –26.9 ppm. Anal. Found: C, 42.30; H, 1.64. Calc. for C₂₈H₁₃F₂₀O₂P: C, 42.44; H, 1.65%.

Synthesis of 13b. A solution of 12b (14.0 mg, 0.0162 mmol) in C_6D_6 (1.0 mL) was heated at 80 °C for 8 h. After concentration *in vacuo*, 13b was obtained (13.5 mg, 0.0157 mmol, 96%) as a

white solid. Mp 115.0–116.0 °C. ¹H NMR (400 MHz, CDCl₃): δ 8.69-8.74 (m, 2 H, Ar-H), 7.71-7.84 (m, 6 H, Ar-H), 7.51 (dd, ${}^{3}J_{\text{H,P}} = 15.4 \text{ Hz}, {}^{3}J_{\text{H,H}} = 8.0 \text{ Hz}, 2 \text{ H}, \text{ Ar-H}), 7.45 \text{ (dd, } {}^{4}J_{\text{H,P}} =$ 7.6 Hz, ${}^{3}J_{H,H} = 8.0$ Hz, 2 H, Ar–H) ppm. 19 F NMR (376 MHz, CDCl₃): δ -63.5 (s, 3 F, Ar-CF₃), -78.2 (s, 6 F, CF₂CF₃), -79.9 $(dd, {}^{3}J_{EF} = 19.7 \text{ Hz}, {}^{5}J_{EF} = 6.2 \text{ Hz}, 6 \text{ F}, \text{ CF}_2\text{CF}_3), -116.7 \text{ (s,}$ 2 F, CF₂CF₃), -116.8 (s, 2 F, CF₂CF₃), -116.8 (dq, ${}^{2}J_{F,F}$ = 285.8 Hz, ${}^{3}J_{\rm EF} = 19.7$ Hz, 2 F, CF₂CF₃), -121.8 (dq, ${}^{2}J_{\rm EF} =$ 285.8 Hz, ${}^{5}J_{F,F} = 6.2$ Hz, 2 F, CF₂CF₃) ppm. 31 P NMR (162 MHz, CDCl₃): δ –28.2 ppm. Anal. Found: C, 40.13; H, 1.07. Calc. for C₂₉H₁₂F₂₃O₂P: C, 40.49; H, 1.41%.

Synthesis of 13c. A solution of 12c (11.8 mg, 0.0145 mmol) in C_6D_6 (1.0 mL) was heated at 80 °C for 8 h. After concentration in vacuo, 13c was obtained (11.4 mg, 0.0140 mmol, 96%) as a white solid. Mp 120.0-121.0 °C. ¹H NMR (400 MHz, CDCl₃): δ 8.67-8.72 (m, 2 H, Ar-H), 7.71-7.83 (m, 6 H, Ar-H), 7.37-7.45 (m, 2 H, Ar-H), 6.84-6.91 (m, 2 H, Ar-H) ppm. ¹⁹F NMR (376 MHz, CDCl₃): δ -78.3 (s, 6 F, CF₂CF₃), -79.8 (dd, ${}^{3}J_{\rm EF} = 19.7$ Hz, ${}^{5}J_{\text{F,F}} = 6.2 \text{ Hz}, 6 \text{ F}, \text{CF}_{2}\text{CF}_{3}), -110.2 \text{ to} -110.3 \text{ (m, 1 F, Ar-F)},$ -116.7 (s, 2 F, CF₂CF₃), -116.8 (s, 2 F, CF₂CF₃), -116.8 (dq, ${}^{2}J_{\rm EF} = 285.8$ Hz, ${}^{3}J_{\rm EF} = 19.7$ Hz, 2 F, CF₂CF₃), -121.8 (dq, ${}^{2}J_{\text{F,F}} = 285.8 \text{ Hz}, {}^{5}J_{\text{F,F}} = 6.2 \text{ Hz}, 2 \text{ F}, CF_2CF_3) \text{ ppm.} {}^{31}\text{P} \text{ NMR}$ (162 MHz, CDCl₃): δ –27.7 ppm. Anal. Found: C, 41.37; H, 1.33. Calc. for C₂₈H₁₂F₂₁O₂P: C, 41.50; H, 1.49%.

Synthesis of 13d. A solution of 12d (12.1 mg, 0.0147 mmol) in C_6D_6 (1.0 mL) was heated at 80 °C for 8 h. After concentration in vacuo, 13d was obtained (11.8 mg, 0.0143 mmol, 97%) as a white solid. Colorless crystals of 13d suitable for the X-ray analysis were obtained by recrystallization from *n*-hexane-CH₂Cl₂. Mp 117.0-117.8 °C. ¹H NMR (400 MHz, CDCl₃): δ 8.68-8.72 (m, 2 H, Ar-H), 7.67–7.78 (m, 6 H, Ar–H), 7.38 (dd, ${}^{3}J_{H,P} = 15.2$ Hz, ${}^{3}J_{H,H} =$ 8.8 Hz, 2 H, Ar–H), 6.69 (dd, ${}^{4}J_{H,P} = 3.4$ Hz, ${}^{3}J_{H,H} = 8.8$ Hz, 2 H, Ar-H), 3.73 (s, 3 H, Ar-OCH₃) ppm. ¹⁹F NMR (376 MHz, CDCl₃): δ -78.3 (s, 6 F, CF₂CF₃), -79.7 (dd, ³J_{F,F} = 19.7 Hz, ${}^{5}J_{F,F} = 6.2$ Hz, 6 F, CF_2CF_3), -116.7 (s, 2 F, CF_2CF_3), -116.8(s, 2 F, CF_2CF_3), -116.8 (dq, ${}^2J_{F,F} = 285.8$ Hz, ${}^3J_{F,F} = 19.7$ Hz, 2 F, CF₂CF₃), -121.8 (dq, ${}^{2}J_{EF} = 285.8$ Hz, ${}^{5}J_{EF} = 6.2$ Hz, 2 F, CF_2CF_3) ppm. ³¹P NMR (162 MHz, CDCl₃): δ –27.1 ppm. Anal. Found: C, 42.27; H, 1.46. Calc. for C₂₉H₁₅F₂₀O₃P: C, 42.35; H, 1.84%.

Synthesis of 13e. A solution of 12e (21.6 mg, 0.0258 mmol) in C₆D₆ (2.0 mL) was heated at 80 °C for 8 h. After concentration in vacuo, 13e was obtained (20.5 mg, 0.0245 mmol, 95%) as a white solid. Colorless crystals of 13e suitable for the X-ray analysis were obtained by recrystallization from n-hexane/CH₂Cl₂. 190.0-191.0 °C. ¹H NMR (400 MHz, CDCl₃): δ 8.56–8.61 (m, 2 H, Ar-H), 7.73 (br s, 2 H, Ar-H), 7.66-7.71 (m, 4 H, Ar-H), 6.58 (d, ${}^{4}J_{H,P} = 6.2$ Hz, 2 H, Ar–H), 2.15 (s, 3 H, Ar-CH₃), 2.00 (s, 6 H, Ar–CH₃) ppm. ¹⁹F NMR (376 MHz, CDCl₃): δ –77.0 (s, 6 F, CF₂CF₃), -79.7 (dd, ${}^{3}J_{F,F} = 16.0$ Hz, ${}^{5}J_{F,F} = 8.6$ Hz, 6 F, CF_2CF_3), -114.6 (dq, ${}^2J_{F,F} = 284.6$ Hz, ${}^3J_{F,F} = 16.0$ Hz, 2 F, CF_2CF_3), -115.9 (dd, ${}^2J_{F,F} = 284.6$ Hz, ${}^4J_{F,F} = 33.2$ Hz, 2 F, CF_2CF_3), -116.94 (d, ${}^2J_{F,F} = 284.6$ Hz, 1 F, CF_2CF_3), -116.97 $(d, {}^{2}J_{E,F} = 284.6 \text{ Hz}, 1 \text{ F}, CF_{2}CF_{3}), -118.1 (ddq, {}^{2}J_{E,F} = 284.6 \text{ Hz},$ ${}^{4}J_{F,F} = 33.2$ Hz, ${}^{5}J_{F,F} = 8.6$ Hz, 2 F, CF₂CF₃) ppm. 31 P NMR (162 MHz, CDCl₃): δ –23.6 ppm. Anal. Found: C, 44.47; H, 1.96. Calc. for C₃₁H₁₉F₂₀O₂P: C, 44.62; H, 2.30%.

Crystallographic data

Table 6

Plate 0.40 × 0.30 × 0.30 9.9960(2) 11.9660(3) 14.1910(3) 86.9670(10) 87.4260(10) 68.0970(10) 1572.10(6) Mo-Ka, 0.71073 173 +h, ±k, ±l 6952/0/490 0.0352 0.1256 1.126 C₃₁H₁₉F₂₀O₂P 834.43 Colorless Triclinic 1.763 Plate $0.60 \times 0.60 \times 0.40$ Mo-Ka, 0.71073 293 +h, +k, ±l 7443/0/479 0.0486 0.1689 1.153 $\begin{array}{c} C_{29}H_{15}F_{21}O_{3}P\\ 822.38\\ \end{array}$ $\begin{array}{c} 17.2000(3)\\ 18.8740(3)\\ 90\\ 101.8580(10)\\ 90\\ 3134.80(10)\\ 4\end{array}$ Monoclinic P2₁/c Colorless 9.8670(2) 1.742 0.240 1632 **3**d $\begin{array}{c} \mathbb{C}_{32}H_{20.5}F_{20}N_{0.5}O_2P\\ 854.96 \end{array}$ $0.60 \times 0.40 \times 0.40$ Mo-Ka, 0.71073 200 +h, ±k, ±l 13990/0/1007 0.0705 0.2089 1.036 2e-0.5CH3CN $\begin{array}{c} 11.11300(10)\\ 15.5850(3)\\ 19.1520(4)\\ 97.5780(10)\\ 97.1530(10)\\ 95.4110(10)\\ 3241.75(10)\\ 3241.75(10) \end{array}$ Colorless Plate **Friclinic** 1.752 0.234 1708 Colorless Plate $0.50 \times 0.10 \times 0.10$ 9.25000(10)Mo-Ka, 0.71073 173 +h, ±/ 6580/0/479 0.0421 0.1480 1.114 C₂₉H₁₅F₂₀O₃P 822.38 Triclinic 9.7100(2) 18.0170(5) 84.9520(10) 77.0590(10) 72.817(2) 1506.33(5) 1.813 0.250 816 **2**d $0.50\times0.40\times0.20$ Mo-Ka, 0.71073 173 +h, ±k, ±l 12473/0/937 0.0415 0.1196 1.076 C₂₈H₁₂F₂₁O₂P 810.35 16.9750(3) 19.6870(4) 88.1130(10) 82.1650(10) 81.3020(10) 8.94800(10) Colorless Plate 2928.09(9) **Friclinic** 1.838 0.258 1600 5 $\begin{array}{l} P2_1/c\\ Colorless\\ Plate\\ 0.50\times 0.10\times 0.10 \end{array}$ Mo-Ka, 0.71073 293 +h, +k, ±l 6824/0/497 0.1149 0.3878 1.109 C₂₉H₁₂F₂₃O₂P 860.36 $\begin{array}{c} 10.3050(3)\\ 18.2210(6)\\ 16.8820(4)\\ 90\\ 95.292(3)\\ 90\\ 3156.38(16)\\ 3156.38(16) \end{array}$ Monoclinic 0.254 $\mathbf{2b}$ $0.50\times0.06\times0.06$ Mo-Ka, 0.71073 173 $\begin{array}{c} C_{28}H_{13}F_{20}O_2P\\ 792.35\end{array}$ $\begin{array}{c} +h, +k, \pm l\\ 9091/0/919\\ 0.0839\\ 0.2914\\ 1.118\end{array}$ 37.4070(13) 16.1350(8) 90 96.6920(10) Monoclinic P2₁/n Colorless Plate 5776.9(4) 9.6370(2) 12a Data/restraints/parameters $R_1 (I > 2\sigma(I))$ vR_2 (all data) Crystal dimensions/mm **Trystal system** Adiation; λ/Å Data collected pace group Jolor Compound $D_{\rm c}/{\rm g~cm^-}$ formula

mm

Single-crystal X-ray analyses of 12a, 12b, 12c, 12d, 12e, 13d and 13e

Crystals suitable for the X-ray structural determination were mounted on a Mac Science DIP2030 imaging plate diffractometer and irradiated with graphite monochromated Mo-K α radiation ($\lambda = 0.71073$ Å) for the data collection. The unit cell parameters were determined by separately autoindexing several images in each data set using the DENZO program (MAC Science).²³ For each data set, the rotation images were collected in 3° increments with a total rotation of 180° about the ϕ axis. The data were processed using SCALEPACK. The structure was solved by a direct method using the SHELXS-97 program.²⁴ Refinement on F^2 was carried out using a full-matrix least-squares by the SHELXL-97 program.²⁴ All non-hydrogen atoms were refined using the anisotropic thermal parameters. The hydrogen atoms were included in the refinement along with isotropic thermal parameters. The crystallographic data are summarized in Table 6.

CCDC reference numbers 672351 (12a), 672352 (12b), 672353 (12c), 672354 (12d), 672355 (12e), 672356 (13d) and 672357 (13e).

For crystallographic data in CIF or other electronic format see DOI: 10.1039/b802947d

Kinetic measurements of the isomerization of 12 to 13

Samples (ca. 5 mg) in C_6D_6 (0.6 mL) were sealed in an NMR tube under N₂. Kinetic measurements of the pseudorotation process were carried out using a JEOL EX-400 spectrometer by monitoring the ¹⁹F NMR signals in a variable temperature mode, and the specified temperatures were maintained throughout each set of measurements (error within ± 1 °C). The observed temperatures were calibrated with the ¹⁹F NMR chemical shift difference in the signals of neat 1,3-propanediol (high temperature region) and MeOH (low temperature region). The data were analyzed on the basis of the first-order kinetics using the following equation: $\ln (C_0/C_{12}) = kT$, in which $C_0 = \text{ratio of } \mathbf{12}$ at t = 0, $C_{12} =$ ratio of 12 at arbitrary intervals. Here $C_0 = C_{12} + C_{13}$, $C_0/C_{12} =$ $(C_{12} + C_{13})/C_{12} = 1 + C_{13}/C_{12}$. The C_{13}/C_{12} ratio was monitored by the integration of the ¹⁹F NMR signals of the trifluoromethyl group at 40–60 °C. The rate constants and activation parameters for the stereomutation from 12 to 13 are shown in Tables 3-5.

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