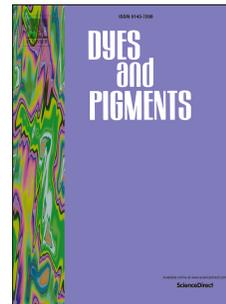


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Unique fluorescence of boronic acid derived salicylidenehydrazone complexes with two perpendicular ICT: Solvent effect on PET process

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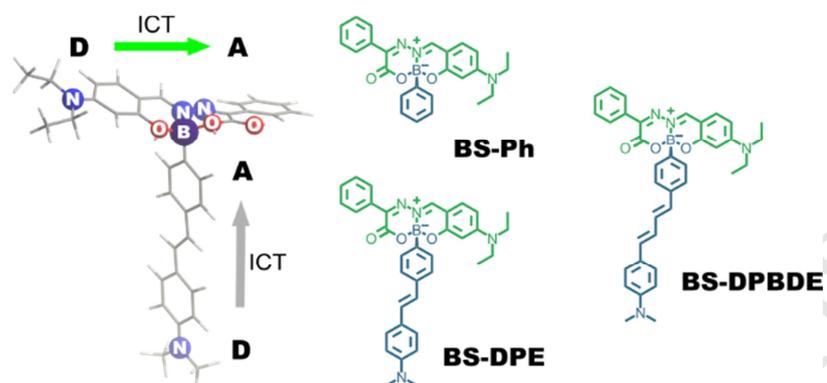
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Graphical Abstracts

1 **Unique fluorescence of boronic acid derived salicylidenehydrazone**
2 **complexes with two perpendicular ICT: solvent effect on PET**
3 **process**

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7 **Abstract:** Solvent effects on the absorption and emission spectra of organic
8 compounds have been widely observed. It is usually found that the positions,
9 intensities and shapes of its bands are usually modified by these solvents. The solvent
10 effects on intramolecular charge transfer (ICT) process have been extensively
11 reported, whereas the solvent effect on photoinduced electron transfer (PET) process
12 is scarce. In this contribution, it was disclosed that boronic acid derived
13 salicylidenehydrazone complexes (BSs), BS-DPE and BS-DPBDE, with two
14 perpendicular ICT states developed a non-fluorescent PET compounds through
15 separating its two ICT states by boron node. Relative strong acidic proton in non-
16 hydrogen-bond accepting solvent were proved to actuate the vanishment of PET and
17 reinstatement of its fluorescence.

18 **Keywords:** Solvent effects; Boronic acid; Salicylidenehydrazone; Photoinduced
19 electron transfer; T-shaped molecular geometry

20

21 1. Introduction

22 Solvent effects[1, 2] have been widely observed in different areas of chemistry such as
23 catalysis,[3] biochemistry[4] and photochemistry.[5] Of these observation, most concern its effects
24 on the position of chemical equilibria and/or the rates of chemical reactions. Furthermore, from a
25 photophysical point of view, the ground and excited state of solvated chromophore-containing
26 molecules in different solvents might undergo a physical perturbation as compared with its
27 isolated ones (gas state)[1]. Vibrational relaxation of electronic state with surrounding solvent are
28 coincided with energy loss both from Franck-Condon excited (ground) state to equilibrium excited
29 (ground) state, leading to Stokes shift.[6] Many types of positive, negative and inverted
30 fluorosolvatochromistic dyes have been synthesized and widely applied in bioimaging and
31 biosensing,[7] such as studying lipid domains,[8] apoptosis and endocytosis.[9] The theory of
32 solvent effects on the fluorescence assumes that the fluorophore is a point dipole residing in the
33 centre of a spherical cavity in a homogeneous and isotropic dielectric, which is so-called Lippert-
34 Mataga theory[10] but deviated from authentic condition. Therefore, it is of paramount
35 importance to come up with and fully comprehend specific fluorophores solute/solvent interaction,
36 especially involving hydrogen bonding, electron-pair donor/electron-pair acceptor interactions or
37 others.[1, 11, 12]

38 Over the last few years were recognized a special solvent effect on ground-state reversible
39 isomerization of silica-rhodamines (SiR),[13] in which SiR-carboxyl exists predominantly
40 in its spiro lactone form in dioxane-water mixtures of dielectric constant less than 30. This
41 seminal fluorogenic behaviour enabled multicolour imaging of target proteins in wash-free
42 conditions.[14] Also, based on a hypothesis which proposed that the break of fluorophore-
43 solvent hydrogen bond in the excited state account for fluorescence quenching,[15, 16]
44 flavones-based fluorophores with carbonyl as hydrogen bond acceptors were demonstrated
45 to be ideal wash-free bioprobe for mitochondria[17] and endoplasmic reticulum
46 imaging.[18] Our group also recently found that boronic acid derived
47 salicylidenehydrazone complexes (BSs) were ideal candidates for wash-free fluorescence
48 imaging of cellular organelles.[19] A remarkably hypsochromic shift of intramolecular

49 charge-transfer (ICT) absorption band in water was observed than in apolar solvent, which
50 were attributed to lower the n-state energy of oxygen lone pairs of the carbonyl group
51 through hydrogen bonding with water.

52 In this contribution, we report that a peculiar fluorescence turn-on property of two BSs
53 involved donor-acceptor diphenylpolyenes[20-22] as auxochrome. The versatile
54 fluorescent dye platform BSs were first reported last year, which were constructed from
55 modular assembly of boronic acids with schiff base ligand salicylidenehydrazone by
56 groups of Goris and Pischel.[23] Subsequently, they thoroughgoingly explored the
57 photophysical scope of the dyes through the variation of both the push-pull ICT character
58 at the ligand backbone and the electronic nature of the boronic acid derived moiety, the
59 latter of which have been demonstrated to be no influence on the Uv-vis absorption and
60 fluorescence properties.[24] The T-shaped molecular geometry of BSs sparked our
61 imagination on what photophysical properties will come into view if two perpendicular
62 ICT states were located in BSs (Scheme 1). To the extent of our knowledge, there has been
63 little research on fluorophore with this type molecular architecture.[25]

64 2. Experimental

65 2.1 General Information

66 All reagents were purchased from Aldrich or ShenZhen Dieckmann Technology Development co.,
67 LTD and were used without further purification, unless otherwise stated. HPLC grade solvent
68 were used in Uv-vis and fluorescence studies. Uv-vis spectra were recorded on a SHIMADZU Uv-
69 1800 spectrophotometer, with a quartz cuvette (path length 1 cm). The fluorescence spectra were
70 recorded with a HORIBA Fluorolog[®]-3 spectrofluorimeter. ¹H and ¹³C NMR spectra were
71 recorded on a BRUKER instrument (400 MHz and 100 MHz, respectively) and internally
72 referenced to tetramethylsilane signal or residual protio solvent signals. Data for ¹H NMR are
73 recorded as follows: chemical shift (δ , ppm), multiplicity (s, singlet; d, doublet; t, triplet; q, quartet;
74 m, multiplet), intergration, coupling constant (Hz). Data for ¹³C NMR are reported in terms of
75 chemical shift (δ , ppm). High resolution mass spectra for all the new compounds were done by an

76 a Xevo G2-XS QToF spectrometer (Waters, USA). The fluorescence quantum yields (QYs) were
77 quantified using Fluorescein as the standard ($\Phi_f = 0.89$, in 0.1 M NaOH). The QYs can be
78 calculated through adopting the following equation:

$$79 \Phi_s = \Phi_f(A_r n_s^2 F_s)/(A_s n_r^2 F_r)$$

80 where the subscripts s and r denote the sample and the standard (Fluorescein), respectively, Φ is
81 the quantum yield, F is the integrated emission intensity, A is the absorbance, and n is the
82 refractive index.

83 Fluorescence lifetimes were detected by a DeltaTime™ TCSPC on Fluorolog®. The monitored
84 wavelength was 545 nm. Fluorescence decay histograms were recorded using the time-correlated
85 single photon counting technique in 4095 channels through a HORIBA Fluorolog®-3
86 spectrofluorimeter equipped with a HORIBA NanoLED source (N-455 nm). Histograms of the
87 instrument response functions and sample decays were obtained until it typically reached 1.0×10^4
88 counts. The fitting parameters (decay times and pre-exponential factors) were decided by
89 minimizing the reduced chisquare χ^2 .

90 For a two-exponential decay the average lift time is given by following equation:

$$\bar{\tau} = \frac{\alpha_1 \tau_1^2 + \alpha_2 \tau_2^2}{\alpha_1 \tau_1 + \alpha_2 \tau_2}$$

91 The time-dependent density functional theory (TD-DFT/B3LYP) calculations of BS-DPE, BS-
92 DPBDE were performed. All calculations were performed with the G03 software. The TD-DFT
93 calculation of the lowest 25 singlet-singlet excitation energies was calculated with a basis set
94 composed of 6-31G (d, p) for C, N, H, O, B atoms. The lowest 25-spin allowed singlet-singlet
95 transitions, up to the energy of about 5 eV, were taken into account for the calculation of the
96 absorption spectra.

97 2.2 Synthesis

98 **(Z)-2-(((E)-4-(diethylamino)-2-hydroxybenzylidene)hydrazono)-2-phenylacetic acid (Ligand)**
99 **(Scheme 2-1)**. Adopting a literature procedure with some reasonable modification,[23]

100 hydrazonoe (2.5 mmol, 490 mg) was completely dissolved in 5 mL water firstly and 20 mL
101 methanol was added. After shaking moderately, this solution was eluted through a columned
102 packed with Amberlyst[®] 15 (5 g wet resin exchanged with 10 mL methanol for three times) and
103 dropped into a methanolic solution of SA (2.5 mmol, 483 mg in 5 mL methanol). The ultimate
104 solution was stirred for 2 h at room temperature and then incubation at 4 °C in refrigerator
105 overnight. The slurry was filtered and washed with cold methanol (5 mL) three times, and
106 afterwards collected as orange powder: yield 65%; ¹H NMR (400 MHz, CDCl₃) δ 8.16 (s, 1H),
107 8.09 – 8.01 (m, 2H), 7.71 (d, *J* = 2.3 Hz, 1H), 7.43 (td, *J* = 7.0, 5.8, 3.0 Hz, 3H), 7.19 (d, *J* = 9.2
108 Hz, 1H), 6.36 (dd, *J* = 9.2, 2.3 Hz, 1H), 3.59 (q, *J* = 7.1 Hz, 4H), 1.31 (t, *J* = 7.1 Hz, 6H). ¹³C
109 NMR (101 MHz, DMSO-d₆) δ 167.3, 164.9, 161.9, 159.5, 152.4, 134.7, 131.9, 131.7, 129.6,
110 127.3, 106.5, 104.9, 97.3, 44.5, 13.0. HRMS-ESI calcd for C₁₉H₂₂N₃O₃ ([M+H]⁺), 340.1661;
111 found, 340.1703.

112 **(4-boronobenzyl)triphenylphosphonium bromide.** A solution containing (4-
113 (bromomethyl)phenyl)boronic acid (5.2 mmol, 1.12 g) and triphenylphosphane (5.72 mmol, 1.5 g)
114 in 160 mL MeCN was refluxed for 12 h. After the completion of reaction, the solvent was
115 removed in vacuo and the residue was triturated with diethyl ether (3 × 10 mL) and collected as
116 white precipitate: yield 95%; ¹H NMR (400 MHz, DMSO-d₆) δ 7.88 – 7.78 (m, 3H), 7.71 – 7.51
117 (m, 17H), 6.90 (dd, *J* = 8.1, 2.5 Hz, 2H), 4.98 (d, *J* = 15.4 Hz, 2H). ¹³C NMR (101 MHz, DMSO-
118 d₆) δ 135.6, 135.6, 134.9, 134.8, 134.6, 134.5, 130.6, 130.5, 130.4, 130.4, 118.7, 117.9, 29.0.
119 HRMS-ESI calcd for C₂₅H₂₃BO₂P ([M-Br]⁺), 397.1523; found, 397.1613.

120 **8-(diethylamino)-2,5-diphenyl-3H,5H-514,1214-benzo[5,6][1,3,2]oxazaborinino[2,3-**
121 **b][1,3,4,2]oxadiazaborinin-3-one (BS-Ph) (Scheme 2-4).** The derivate of phenylboronic acid
122 (0.3 mmol) was dissolved in CH₃CN (3 mL), toward which the ligand (0.33 mmol, 112 mg) was
123 added. The resulting solution was refluxed until the termination of reaction. And then, the solution
124 was concentrated and the residue was triturated with cold methanol for washing away minor
125 impurities. [19] Orange solid; Yield 95%; ¹H NMR (400 MHz, CDCl₃) δ 8.21 (s, 1H), 7.96 – 7.88
126 (m, 2H), 7.41 – 7.27 (m, 5H), 7.18 – 7.08 (m, 4H), 6.29 (dd, *J* = 9.1, 2.4 Hz, 1H), 6.13 (d, *J* = 2.3

127 Hz, 1H), 3.37 (ddt, $J = 16.6, 14.8, 7.4$ Hz, 4H), 1.16 (t, $J = 7.2$ Hz, 6H). ^{13}C NMR (101 MHz,
128 CDCl_3) δ 161.6, 157.3, 155.8, 154.0, 153.6, 134.6, 132.7, 130.8, 130.8, 129.5, 128.2, 127.8,
129 127.6, 107.7, 106.4, 98.7, 45.4, 12.8. HRMS-ESI calcd for $\text{C}_{25}\text{H}_{25}\text{BN}_3\text{O}_3$ ($[\text{M}+\text{H}]^+$), 426.1989;
130 found, 426.1873.

131 **(E)-8-(diethylamino)-5-(4-(4-(dimethylamino)styryl)phenyl)-2-phenyl-3H,5H-5I4,12I4-**
132 **benzo[5,6][1,3,2]oxazaborinino[2,3-b][1,3,4,2]oxadiazaborinin-3-one (BS-DPE) (Scheme 2-**
133 **5).** (E)-(4-(4-(dimethylamino)styryl)phenyl)boronic acid in Scheme 2-2 was synthesized on the
134 previous reports (Spectroscopic sugar sensing by a stilbene derivative with push (Me_2N)-pull
135 $(\text{HO})_2\text{B}$ -) type substituents). The derivate of phenylboronic acid (0.3 mmol) was dissolved in
136 CH_3CN (3 mL), toward which the ligand (0.33 mmol, 112 mg) was added. The resulting solution
137 was refluxed until the termination of reaction. And then, the solution was concentrated and the
138 residue was dissolved in tiny amount toluene and then purified by column chromatography
139 (Hexane : EtOAc=10:1). Pale orange solid: yield 65%. ^1H NMR (400 MHz, Chloroform-*d*) δ 8.31
140 (s, 1H), 8.07 – 7.99 (m, 2H), 7.51 – 7.30 (m, 9H), 7.24 (d, $J = 9.1$ Hz, 1H), 6.98 (d, $J = 16.3$ Hz,
141 1H), 6.85 (d, $J = 16.2$ Hz, 1H), 6.78 – 6.66 (m, 2H), 6.39 (dd, $J = 9.1, 2.4$ Hz, 1H), 6.24 (d, $J = 2.3$
142 Hz, 1H), 3.46 (ddt, $J = 23.8, 14.5, 7.0$ Hz, 4H), 2.98 (s, 6H), 1.26 (t, $J = 7.2$ Hz, 6H). ^{13}C NMR
143 (101 MHz, CDCl_3) δ 161.6, 157.3, 155.8, 154.1, 153.5, 150.0, 137.6, 134.5, 132.7, 131.1, 130.8,
144 129.5, 128.2, 128.1, 127.4, 126.1, 125.3, 124.8, 112.5, 107.7, 106.4, 98.7, 45.4, 40.5, 12.7.
145 HRMS-ESI calcd for $\text{C}_{35}\text{H}_{36}\text{BN}_4\text{O}_3$ ($[\text{M}+\text{H}]^+$), 571.2880; found, 571.2463.

146 **8-(diethylamino)-5-(4-((1E,3E)-4-(4-(dimethylamino)phenyl)buta-1,3-dien-1-yl)phenyl)-2-**
147 **phenyl-3H,5H-5I4,12I4-benzo[5,6][1,3,2]oxazaborinino[2,3-b][1,3,4,2]oxadiazaborinin-3-one**
148 **(BS-DPBDE) (Scheme 2-6).** The derivate of phenylboronic acid (0.3 mmol) was dissolved in
149 CH_3CN (3 mL), toward which the ligand (0.33 mmol, 112 mg) was added. The resulting solution
150 was refluxed until the termination of reaction. And then, the solution was concentrated and the
151 residue was dissolved in tiny amount toluene and then purified by column chromatography
152 (Hexane : EtOAc=10:1). Pale orange solid: yield 54%. ^1H NMR (400 MHz, Chloroform-*d*) δ 8.31
153 (s, 1H), 8.06 – 7.98 (m, 2H), 7.51 – 7.20 (m, 10H), 6.88 (dd, $J = 15.3, 10.5$ Hz, 1H), 6.81 – 6.66
154 (m, 3H), 6.54 (t, $J = 16.1$ Hz, 2H), 6.39 (dd, $J = 9.2, 2.4$ Hz, 1H), 6.24 (d, $J = 2.4$ Hz, 1H), 3.47

155 (qt, $J = 14.4, 7.3$ Hz, 4H), 2.98 (s, 6H), 1.27 (t, $J = 7.0$ Hz, 6H). ^{13}C NMR (101 MHz, CDCl_3) δ
156 161.6, 157.3, 155.8, 154.1, 153.5, 137.3, 134.5, 132.7, 132.6, 131.1, 130.8, 129.5, 129.4, 128.2,
157 127.4, 125.6, 125.4, 112.5, 107.6, 106.4, 98.7, 45.4, 40.5, 29.7, 12.7. HRMS-ESI calcd for
158 $\text{C}_{37}\text{H}_{38}\text{BN}_4\text{O}_3$ ($[\text{M}+\text{H}]^+$), 597.3037; found, 597.2697.

159 3. Results and discussion

160 3.1 Synthesis

161 Two derivatives of diphenylethene (DPE) or diphenylbutadiene (DPBDE), possessing a
162 dimethylamino and a boronic acid group as electron-donor and electron-withdrawing
163 groups, were settled on assembly with Schiff base ligand to create targeted molecules, BS-
164 DPE and BS-DPBDE. The B-N dative bond in BSs could furnish an extra molecular
165 stability,[26] which might also modify the electronic properties of auxiliary ICT state.
166 DPE-BA (boronic acid) and DPBDE-BA were synthesized by the Wittig reaction (Scheme
167 2) between the corresponding benzaldehydes and the para-boronic acid derivative of the
168 benzyltriphenylphosphonium (TPP) bromide,[27] which were then converted into BS-
169 DPE and BS-DPBDE by simple condensation with salicylidenehydrazone ligand.[23, 24]

170 3.2 Distinctive optical properties

171 Unexpectedly, light tangelo BS-DPE and BS-DPBDE are essentially nonfluorescent in the
172 solid state (Fig. 1b) and show very weak fluorescence in chloroform (Fig. 1c), which is in
173 marked contrast to BS-Ph with high fluorescence quantum yield (QY ca. 0.5-0.7) and
174 brightness in chloroform as previous reports.[19, 23] Serendipitously, it was found BS-
175 DPE and BS-DPBDE are highly luminescent in deuterated chloroform (CDCl_3) (Fig. 1d)
176 during NMR test, in striking contrast to in chloroform. It spurred us into a detailed test on
177 its absorption and emission spectra in different solvent. Fig. 1e shows that BS-DPE has a
178 dual absorption band trait in nine selected solvents, where the low-energy band from 400
179 nm to 525 nm were attributed to ICT absorption of the ligand backbone, and the high-
180 energy band within the range of 300 nm – 400 nm originated from ICT state of DPE.[20]

181 The spectra in CDCl_3 and HEPES buffer were unpredicted, the former of which shows a
182 large hypsochromic shift of high-energy band from 357 nm to 320 nm in comparison with
183 that in CHCl_3 , the latter exhibits that both of bands broadened and stretched out to 410 nm
184 and 550nm, indicating water molecules have obvious influence on both of ICT ground
185 states. Our earlier report revealed that BS-Ph had just one local chromophore absorption
186 band located at 400 nm, which is an outcome of interaction of backbone ICT state with
187 water.[19] Based on the above facts in HEPES buffer, it could be concluded that the
188 horizontal and vertical ICT states have independent features, but also interact with each
189 other and offer an ensemble solvatochromism. Fluorescence spectra follow the same trend
190 as the absorption spectra (Fig. 1f). High QY up to 0.43 was attained in CDCl_3 , whereas no
191 fluorescent signal was detected in HEPES. A red shift was observed in high polar solvent
192 in comparison with apolar solvents (average 520 nm), up to 15 nm, 32 nm and 17 nm for
193 PEG400, DMSO and CH_3OH , respectively. For BS-DPBDE, the spectral features are
194 comparable to that of BS-DPE, but with three distinctions: (1) High energy-band is located
195 at 344 nm for CDCl_3 and near 385 nm for others; (2) The intensities of high energy-band
196 are higher than that of low energy-band in all solvents except CDCl_3 ; (3) The QY of BS-
197 DPBD (0.52) is higher than BS-DPE in CDCl_3 (Table 1). Due to much higher brightness (ϵ
198 $\times \Phi_s$) of BS-DPBDE, it was selected for further discussion.

199 3.3 Turn-on phenomena and characterization.

200 The extraordinary brightness in CDCl_3 made us think about the concealed factor in
201 fluorescence turn-on. Hydrochloric acid as the main decomposed product of CHCl_3 was
202 tested first.[28] As shown in Fig. 2a-b and Fig. S1-S2, the absorption and emission spectra
203 of BS-DPBDE in different solvents were compared with that of corresponding solvent
204 added HCl up to 15 μM . Distinctly, the spectral feature of BS-DPBDE in CHCl_3 with HCl
205 was identical with that in CDCl_3 , which indicates that it is easy for CDCl_3 to release DCl as
206 acid for actuating fluorescence of BS-DPBDE. For other solvents with adding HCl,
207 unexpectedly, there are no significant enhancement of fluorescence intensities even in
208 apolar solvent toluene, and also no occurrence of high-energy absorption band blue-shift.

209 In toluene and THF, addition of HCl even caused the decrease in fluorescence a little bit.
210 This fact points out that the fluorescent turn-on is not only an acid-actuation process, but
211 also dependence on solvent. Next, the chloride solvents $C_2H_4Cl_2$ and CH_2Cl_2 were used to
212 further investigate this solvent effect on fluorescence turn-on. From **Fig. 2c**, it can be seen
213 that the low-energy band were heightened, and also that the low energy band lost its
214 vibrational fine-structure when HCl was added into $CHCl_3$ and $C_2H_4Cl_2$, which are
215 consonant with intensified fluorescence in $CHCl_3$ and $C_2H_4Cl_2$ with HCl (**Fig. 2d**).
216 Afterwards, UV-vis and emission titration experiments of BS-DPBDE (10 μ M) in $CHCl_3$
217 were recorded with increasing concentrations of HCl (**Fig. 2e-f**). The continuous addition of
218 HCl from 1 μ M to 20 μ M induced a dramatic change in the spectra that accompanied two
219 isosbestic points at 350 and 445 nm. The presence of an isosbestic point signified the
220 presence of an equilibrium process between turn-on and turn-off states. No further spectral
221 change was registered when HCl was added beyond the mole ratio of 1.25 equiv. The
222 fluorescence intensity increase caused by addition of HCl to BS-DPBDE is linearly
223 proportional to HCl concentration in the 1-7.5 μ M range (**Fig. 2f**). Ensuingly, acetic acid,
224 Trifluoroacetate (TFA) and sulfuric acid were selected for testing whether or not all acids
225 could actuate the fluorescence of BS-DPBDE. **Fig. 2g** presents the fluorescence intensity
226 change (I/I_0) upon adding acid into $CHCl_3$ solution, which manifest that acids with pKa
227 lower than 3.0 enable the $CHCl_3$ solution of BS-DPBDE to be emitted. The spectral
228 properties of BS-DPE were comparable with BS-DPBDE and the data were shown in Fig.
229 S5.

230 Fluorescence decay parameters of the BS-DPBDE in different solvents without or with
231 acids are listed in **Table 2**. Fluorescence decay profiles were satisfactorily fitted with a
232 two-exponential model and lifetimes show apparent effect of solvent, much shorter
233 lifetimes for polar solvent THF (1.23 ns) and DMSO (0.61 ns).^[29] The faster component
234 of the lifetime can be attributed to the ICT species, and the slower component is due to the
235 presence of the LE molecules^[30]. Remarkably, roughly twice prolongation of τ_1 were
236 registered in $CHCl_3$ upon adding HCl and H_2SO_4 over others, especially in contrast to that

237 of TFA and acetic acid (**Fig. 3a**). It indicates that there might have some fresh species with
238 longer lifetime, associated with high fluorescence. Schiff-base backbone and two
239 dialkylamino-group confused us as to whether BS-DPBDE was degraded and which
240 dialkylamino base group was vulnerable to attack by acid. Further, proton NMR titrations
241 were employed to disclose the underlying mechanism. DMSO-*d*₆ with 1.2 M acids was
242 added to CDCl₃ solution of BS-DPBDE (10 mM), ultimately with a volume ratio of 1: 100,
243 circumventing the solvent effects on chemical shifts. The signals of proton 7, 8 and 9 are of
244 non-movement under all tests, regarded as reference peak together with residual signal of
245 CDCl₃ (**Fig. 3c**), signifying that Schiff-base backbone is stable in CDCl₃ with a small
246 amount of acid although it might be unstable in acidic aqueous solution (Fig. S8). With
247 gradual addition of HCl, the larger downfield shift of the dimethyl group proton 1 and
248 aromatic protons 4 were observed, whereas the smaller downfield shift of that as replacing
249 HCl with TFA. It is well-known that chemical shift is a measure of electron density near
250 the proton being measured. And thus, it can be concluded that stronger acidic proton as
251 electron acceptor makes protons 1 and 4 in BS-DPBDE deshielded heavily.[31] In contrast,
252 proton 2 and 3 of the diethyl-amino group are less affected even though its high pK_a value
253 (N-diethylaniline, 6.56 vs N-dimethylaniline, 5.06[32]). The peaks appeared under
254 addition of HCl to 0.8 equiv. (asterisk peaks) were contributed to slow-proton-exchange of
255 protonated diethyl-amino group on the NMR time scale.[33]

256 **3.4 Molecular mechanism and calculations.**

257 To understand the fluorosolvatochromic behaviour, the fluorescence intensity of BS-
258 DPBDE in different solvent with 1.5 equiv. HCl were associated with three solvatochromic
259 parameters: polarity parameter ($E_T(30)$), hydrogen-bond-donating parameter (α), and
260 hydrogen-bond-donating parameter (β) (Fig. 3b). It is apparent that no correlation is
261 observed for $E_T(30)$ and α , whereas indicates that the fluorescence of BS-DPBDE is
262 lighted up only in strong acid containing non-hydrogen-bond accepting solvents,
263 chloroform, whose β value approximate zero. Density functional theory (DFT) calculations
264 revealed that the HOMOs, for both of BS-DPE and BS-DPBDE, are located on the donor-

265 acceptor diphenylpolyenes, whereas the LUMOs on the Schiff-base backbone (**Fig. 4** and
266 **Table 3**). It is in sharp contrast with HOMO and LUMO of BS-Ph, both of which located
267 on the Schiff-base backbone.[23] The non-superimposed HOMO and LUMO distribution
268 support the photoinduced electron transfer (PET) process in these two molecules, which
269 weakened and even quenched the fluorescence of the Schiff base backbone fluorophore.
270 All of clues demonstrate that hydrogen-bond accepting solvents might enfeeble the
271 electron withdrawing ability of acidic proton, and hence impeded the vanishment of PET
272 process. We believe that this unique fluorescence phenomena might broaden our notion of
273 fluorosolvatochromism and be applied in diverse fields.[34]

274 **4. Conclusions**

275 In closing, BSs derivatives, BS-DPE and BS-DPBDE, with two perpendicular ICT states
276 have been synthesized and its spectroscopic and photophysical properties are described.
277 We disclosed that boron in BSs as a node separated its two ICT states (Schiff base
278 backbone ICT and diphenylpolyenes ICT states) and developed non-fluorescent PET
279 compounds. Peculiarly, our work provides indisputable prerequisite for its fluorescent turn-
280 on, that is, stronger acidic proton in non-hydrogen-bond accepting solvents. We believe
281 that the unique fluorescence of BS-DPE and BS-DPBDE would not only spur new
282 application in diverse field, but also provide the foundation for modification of
283 conventional theories.

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288 **Appendix A. Supplementary data**

289 Supplementary data related to this article can be found at

290 **References**

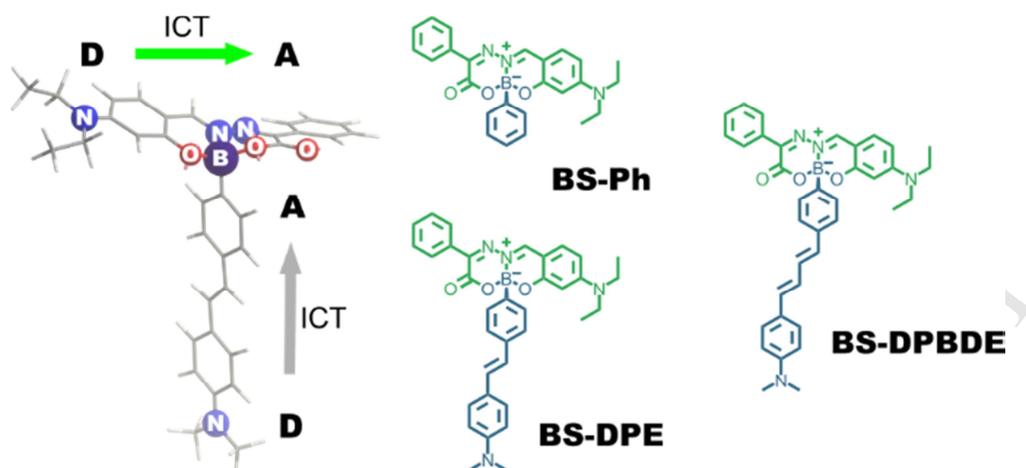
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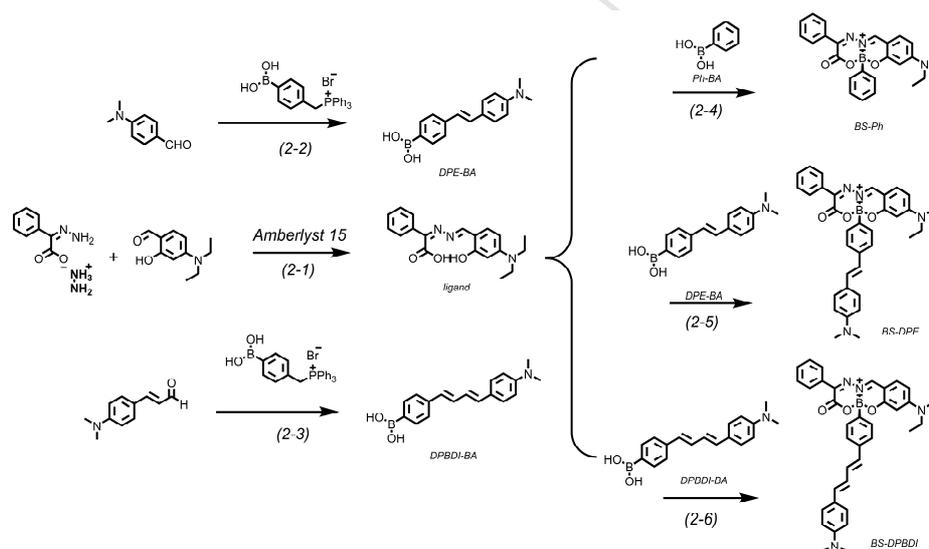
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Scheme 1 T-shape molecular geometry of BSs with two perpendicular ICT states (The DFT optimized structure) and the structures of BS-Ph, BS-DMA, BS-DPE and BS-DPBDE

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Scheme 2 Synthetic routine of BS-Ph, BS-DPE and BS-DPBDE

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371 **Table 1** Spectral Properties and Fluorescence Quantum Yield (Φ_s) of the BS-DPE and BS-
 372 DPBDE in different solvents.

Compd	solvent	λ_{abs} (nm)	ϵ ($\text{M}^{-1}\text{cm}^{-1}$)	$\lambda_{\text{em}}^{\text{b}}$ (nm)	Δ^{c} (nm)	Φ_s	
BS-DPE	CDCl_3	321	478	53000	518	40	0.43
	CHCl_3	357	478	52000	517	39	<0.01
	CH_2Cl_2	359	477	52000	522	45	<0.01
	Toluene	355	472	54000	508	36	<0.01
	THF	355	471	55000	517	46	<0.01
	PEG400	361	478	43000	535	57	<0.01
	DMSO	363	478	42000	552	74	<0.01
	CH_3OH	351	473	43000	537	64	<0.01
	20 mM HEPES	356	471	38000	— ^a		
BS-DPBDE	CDCl_3	344	469	58000	518	49	0.52
	CHCl_3	387	478	56000	517	39	0.02
	CH_2Cl_2	390	477	55000	522	45	0.02
	Toluene	387	472	59000	509	37	0.02
	THF	384	472	55000	520	48	<0.01
	PEG400	387	477	48000	533	56	<0.01
	DMSO	386	479	47000	544	65	<0.01
	CH_3OH	379	473	46000	553	80	<0.01
	20 mM HEPES	389	465	40000	— ^a		

373 ^a Not determined due to the weak fluorescence. ^b λ_{ex} = at 470 nm. ^c Stokes shift.

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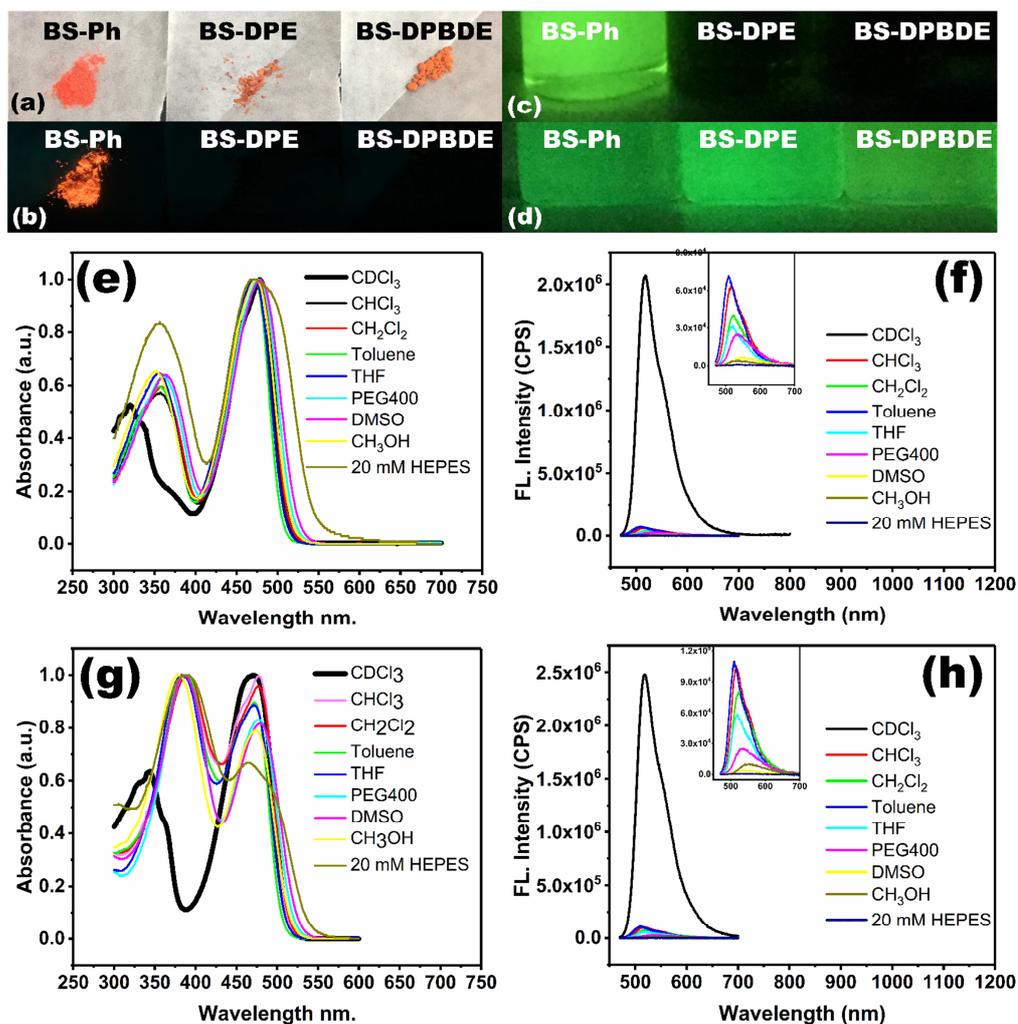


Fig. 1 Image of compounds BS-Ph, BS-DPE and BS-DPBDE (a) under vis light; (b) in solid states, (c) dissolved in CHCl_3 or (d) in CDCl_3 under 365 nm UV light. Absorption (e, g) and emission spectra (f, h) of BS-DPE (e, f) and BS-DPBDE (10 μM) (g, h) in CDCl_3 , CHCl_3 , CH_2Cl_2 , toluene, THF, PEG400, DMSO, CH_3OH and HEPES buffer (20 mM). (excitation at 450 nm, slit: 1 nm/1 nm)

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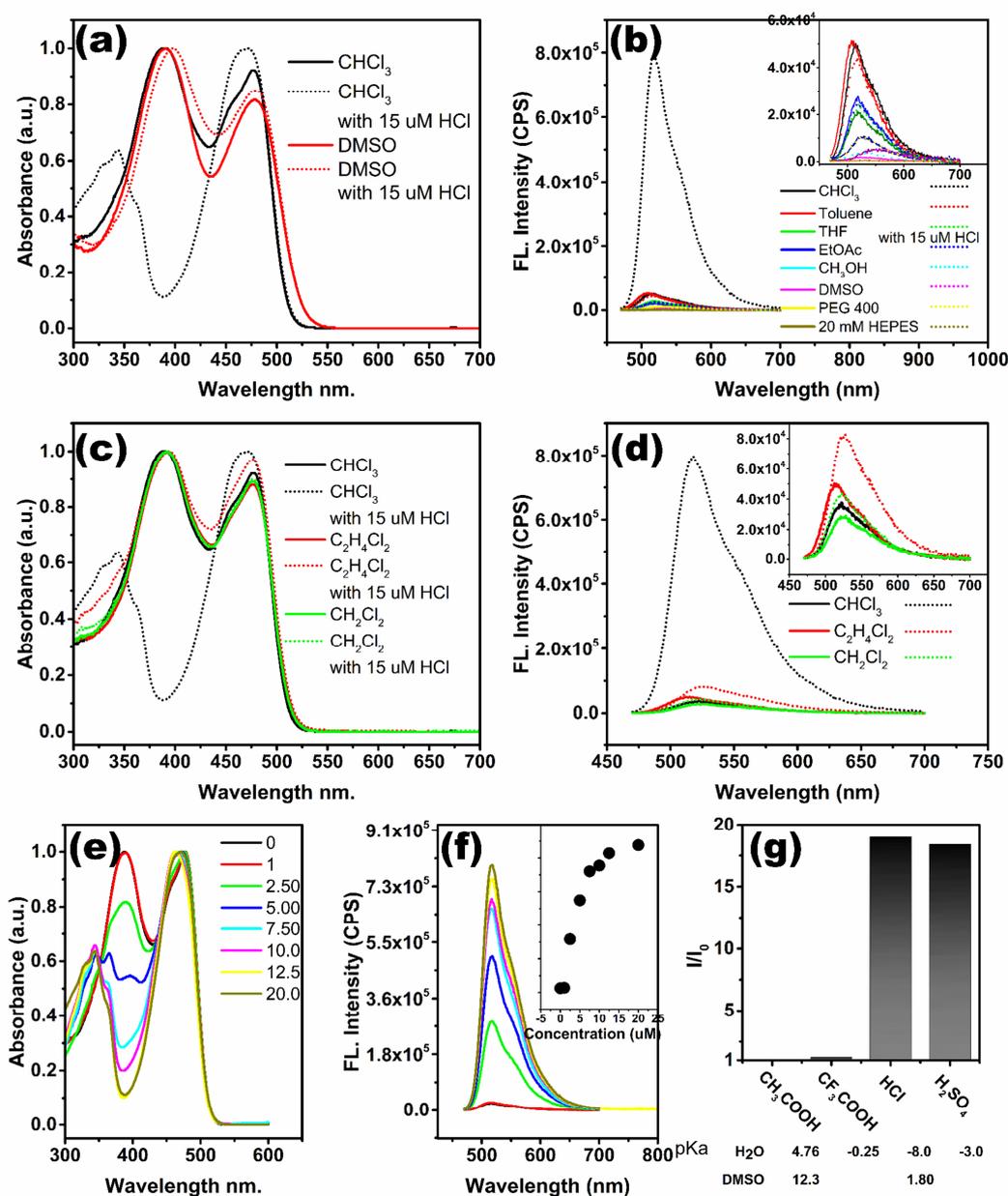


Fig. 2 Absorption (a, c) and emission (b, d) spectra of 10 μM BS-DPBDE in different solvents with or without 15 μM HCl. Notes: for (b), CHCl_3 (black), toluene (red), THF (green), EtOAc (blue), CH_3OH (cyan), DMSO (magenta), PEG400 (yellow) and 20 mM HEPES buffer (dark yellow); short dot line, with 15 μM HCl. Absorption (e) and emission (f) titration spectra in the presence of varying concentrations of HCl in CHCl_3 . (g) the emission ratio (I/I_0) upon adding acid into CHCl_3 solution. (excitation at 450 nm, slit: 1 nm/1 nm)

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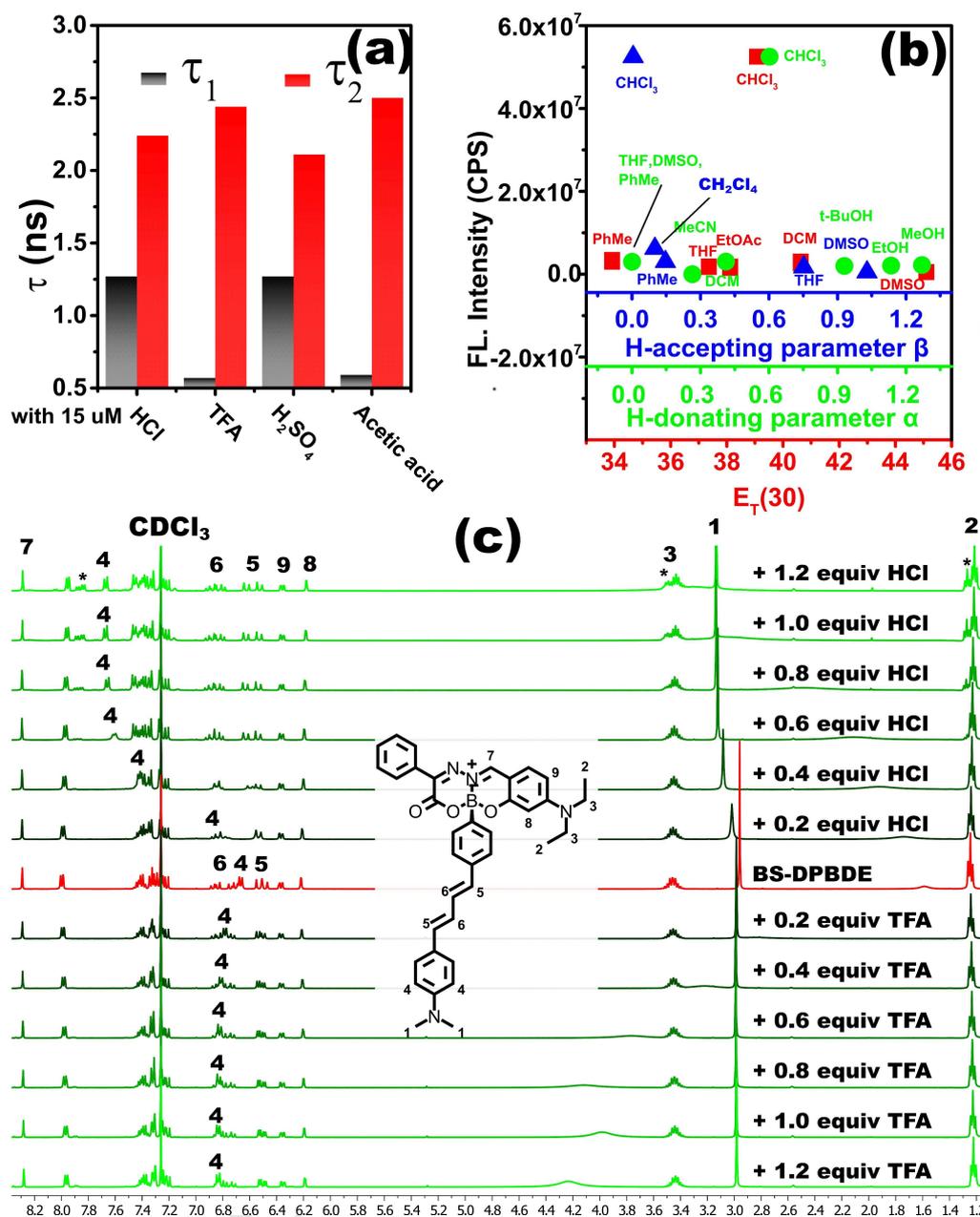


Fig. 3 (a) The distribution of fluorescence decay times of BS-DPBDE in CHCl_3 with 15 μM acids (BS-DPBDE, 10 μM), data from **Table 2** and Fig S3-4 (b) Relationship between the fluorescent intensity of BS-DPBDE in 15 μM HCl solution and the solvatochromic parameters $E_T(30)$, hydrogen-bond-donating parameter (α), and hydrogen-bond-accepting parameter (β) of solvents. Partial Data are repeated from Fig.2; (c) ^1H NMR titration of BS-DPBDE with HCl and TFA in CDCl_3 (BS-DPBDE, 10 mM).

380 **Table 2** Fluorescence Decay Parameters of the BS-DPBDE investigated in different solvent with
 381 or without acid

		τ_1 (ns)	τ_2 (ns)	α_1	α_2	$\bar{\tau}$ (ns)	χ_R^2
1	CHCl ₃	0.60	2.45	0.18	0.82	2.36	1.7
2	CHCl ₃ with 15 uM HCl	1.27	2.24	0.18	0.82	2.13	1.9
3	CH ₂ Cl ₂	0.74	2.41	0.32	0.68	2.20	1.9
4	CH ₂ Cl ₂ with 15 uM HCl	0.64	2.22	0.38	0.62	1.98	1.7
5	Toluene	0.90	2.23	0.20	0.80	2.11	1.6
6	Toluene with 15 uM HCl	0.87	2.10	0.39	0.61	1.84	1.6
7	THF	0.05	1.25	0.26	0.74	1.23	3.8
8	THF with 15 uM HCl	0.09	1.06	0.22	0.78	1.04	6.0
9	DMSO	0.34	0.80	0.62	0.38	0.61	2.0
10	DMSO with 15 uM HCl	0.51	2.24	0.97	0.03	0.72	2.6
11	CHCl ₃ with 15 uM TFA	0.57	2.44	0.26	0.74	2.30	1.5
12	CHCl ₃ +15 uM H ₂ SO ₄	1.27	2.11	0.42	0.58	1.86	1.6
13	CHCl ₃ +15 uM Acetic acid	0.59	2.50	0.17	0.83	2.41	1.8

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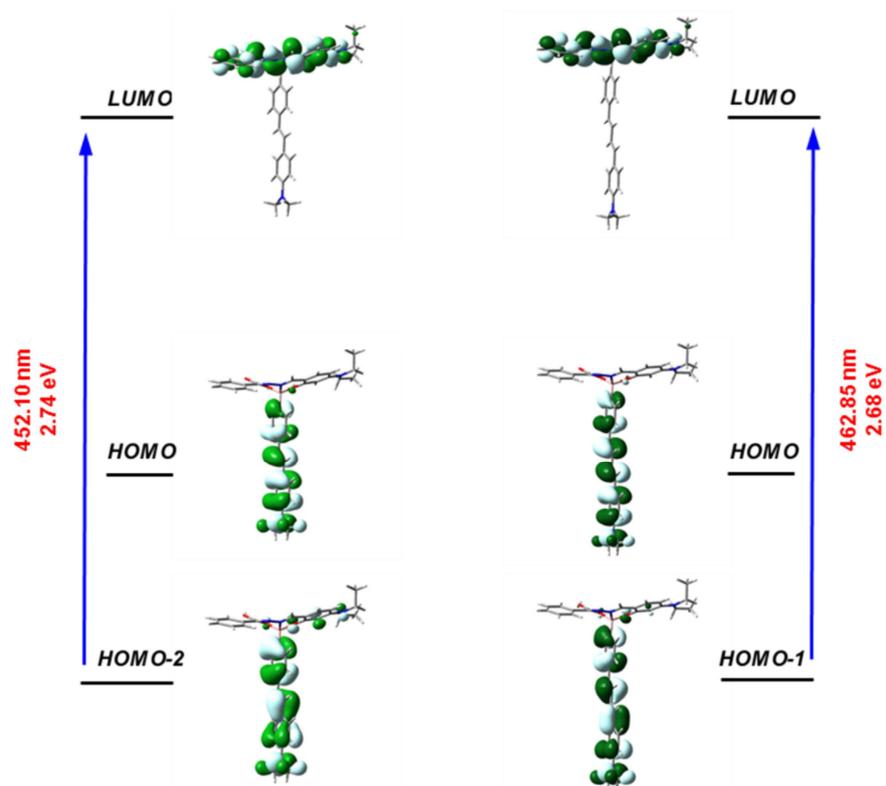


Fig. 4 Molecular orbitals (LUMO and HOMO) and HOMO/LUMO energy gaps of BS-DPE and BS-DPBDE.

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387 **Table 3** Calculated single-photon related spectral properties of BS-DPE, BS-DPBDE in the gas
 388 phase.

Comp.	$\lambda_{\max}(\text{nm})^a$	$\Delta E_1(\text{eV})^b$	f	OI ^c
	452.10	2.74	0.1196	149 → 152 (H-2 → L)
2	462.85	2.68	0.0296	157 → 159 (H-1 → L)

389 *a.* Peak position of the longest absorption band; *b.* The energy gap of the one-photon
 390 absorption band; *c.* TD-DFT method with the orbitals Involved (OI).

Highlights

- Two boronic acid derived salicylidenehydrazone complexes (BSs) with two perpendicular ICT states, BS-DPE and BS-DPBDE, were synthesized and characterized.
- The non-fluorescent PET characteristic of BS-DPE and BS-DPBDE were disclosed.
- Relative strong acidic proton in non-hydrogen-bond accepting solvent were proved to actuate the vanishment of PET and reinstatement of its fluorescence.