



Short communication

Azide-enolate 1,3-dipolar cycloaddition in the synthesis of novel triazole-based miconazole analogues as promising antifungal agents



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ABSTRACT

Seven miconazole analogs involving 1,4,5-tri and 1,5-disubstituted triazole moieties were synthesized by azide-enolate 1,3-dipolar cycloaddition. The antifungal activity of these compounds was evaluated *in vitro* against four filamentous fungi, including *Aspergillus fumigatus*, *Trichosporon cutaneum*, *Rhizopus oryzae*, and *Mucor hiemalis* as well as three species of *Candida* spp. as yeast specimens. These pre-clinical studies suggest that compounds **4b**, **4d** and **7b** can be considered as drug candidates for future complementary biological studies due to their good/excellent antifungal activities.

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1. Introduction

Miconazole is an azole-type drug with a broad spectrum of antifungal activity [1]. Due to the development of fungal resistance to this drug [2], the medicinal chemistry of anti-fungal agents has become an important field of study in organic synthesis [3]. To design new agents free of antibiotic resistance, the modification of functional groups in lead molecules has been an efficient strategy [4].

The triazole ring system, is a very well-recognized pharmacophore [5], this nitrogen heterocycle is prominent among U.S. FDA approved pharmaceuticals [6]. In particular, the 1,2,3-triazole core has been an increasingly important heterocycle with successful application in medicinal chemistry [7]. There are reports in literature about the biological activity of 1,2,3-triazole derivatives against cancer [8], malaria [9], tuberculosis [10], trypanosomiasis

[11], leishmaniasis [12], HIV [13], influenza [14], dengue [15], pain (analgesic) [16], epilepsy [17], obesity [18], inflammation [19] and bacterial infection [20]. On the other hand, the study of 1,2,3-triazole scaffolds for the synthesis of antifungals [21], and particularly for miconazole analogs [22] has represented an ongoing and promising field of research in the last few years.

Cu-catalyzed azide-alkyne cycloaddition (CuAAC) represents the conventional method for obtaining 1,2,3-triazole moieties [23]. In recent years, azide-enolate 1,3-dipolar cycloaddition has emerged as a novel and potent tool for the synthetic approach to these valuable heterocycles [24]. Its application in medicinal chemistry has already been demonstrated [25].

We previously reported the antifungal activity of benzyloxy derivatives of miconazole [26]. As part of our ongoing research, we herein describe the synthesis/evaluation of triazolic analogs (1,4,5- and 1,5-substituted derivatives) that maintain the 1-(2-phenylethyl)imidazole core responsible for the biological activity of this compound [27] (Fig. 1).

2. Chemistry

From 2,4-dichlorobenzaldehyde **1** [Eq. (1)], the 1-(2,4-

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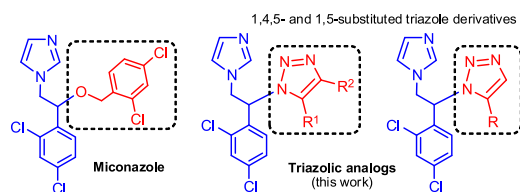
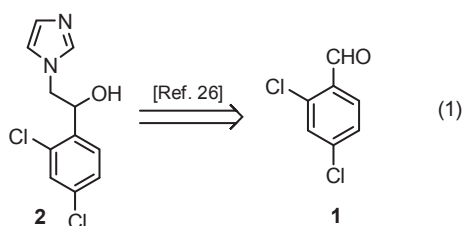
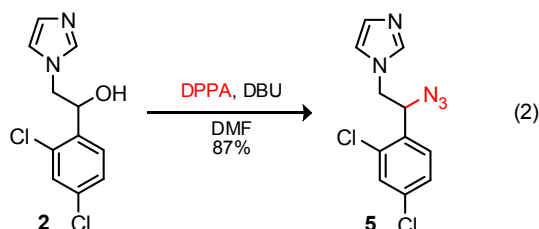


Fig. 1. Proposed triazolic analogs of miconazole.

dichlorophenyl)-2-(1H-imidazol-1-yl)ethanol **2** (key precursor) was obtained in two steps according to our previous report [26]. Previously we published a novel method for preparing 1,4,5-trisubstituted 1,2,3-triazoles from benzylic alcohols *via* an azide-enolate 1,3-dipolar cycloaddition [28]. For this purpose we used diphenylphosphoryl azide (DPPA) as an azidating agent, followed by an efficient cycloaddition in the presence of active ketones. Miconazole analogs **4a–d** (1,4,5-trisubstituted derivatives) were synthesized in good yields (Table 1) by coupling acetylacetone **3a**, 2-benzoylacetophenone **3b**, benzoylacetone nitrile **3c** and 1-(phenylsulfonyl)heptan-2-one **3d** under the aforementioned protocol.



Recently [29] we reported a novel method for obtaining 1,5-disubstituted triazoles from azides by coupling them with β -keto-phosphonates. Therefore, we decided to begin with the synthesis of benzyl azide **5** [Eq. (2)] as a precursor. The azidation of benzyl alcohol **2** was achieved using DPPA and DBU in dry DMF with good yields [30]. The synthesis of alkyl (**7a** and **7b**) and aryl (**7c**) 1,5-disubstituted triazole derivatives was carried out *via* an azide-enolate 1,3-dipolar cycloaddition (Table 2).



An outstanding aspects for compounds **7a–c** is a singlet signal in the range δ 7.6–7.4 ppm (^1H NMR spectra) attributable to the triazolic hydrogen [Eq. (3)]. Likewise, for all final compounds, the hydrogens H^a , H^b and H^c on the imidazole moiety can be observed in the ranges δ 7.7–7.4, 7.0–6.9 and 6.9–6.6 ppm respectively.

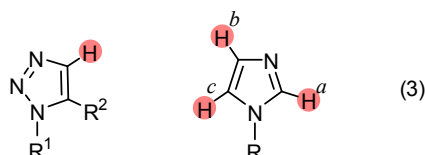
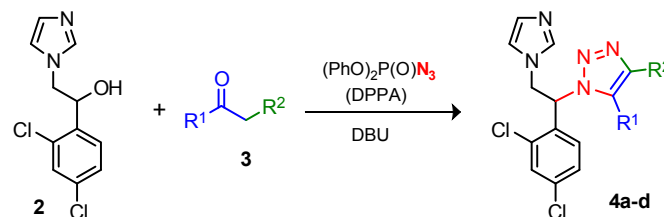


Table 1

Synthesis of 1,4,5-trisubstituted 1,2,3-triazole **4a–d** (miconazole analogs) from alcohol **2** by coupling with active ketones **3**.



Entry ^a	Ketone	Triazole ^b (Yield%) ^c
1	3a : $\text{R}^1 = \text{CH}_3$, $\text{R}^2 = \text{COCH}_3$	4a (75%)
2	3b : $\text{R}^1 = \text{Ph}$, $\text{R}^2 = \text{COPh}$	4b (67%)
3	3c : $\text{R}^1 = \text{Ph}$, $\text{R}^2 = \text{CN}$	4c (63%)
4	3d : $\text{R}^1 = \text{CH}_3(\text{CH}_2)_4-$, $\text{R}^2 = \text{SO}_2\text{Ph}$	4d (78%)

^a Reaction conditions: A mixture of compound **2** (1.0 eq), DPPA (1.1 eq), and DBU (2.0 eq) in DMF was stirred at r.t. for 3 h. Then **3** (1.0 eq) was added and the reaction continued at 60 °C for 3 h.

^b Confirmed by ^1H NMR, ^{13}C NMR, and MS.

^c Yields refer to chromatographically pure isolated compounds.

3. Microbiology

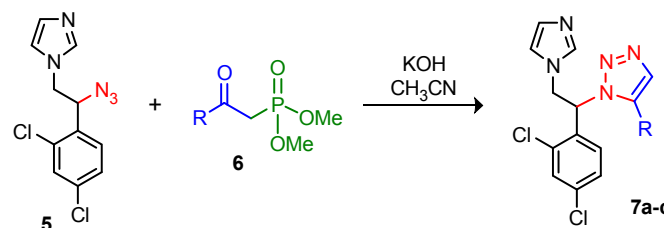
Compounds **4a–d** and **7a–c** were evaluated for their *in vitro* antifungal activity against four filamentous fungi (*Aspergillus fumigatus* ATCC-16907, *Trichosporon cutaneum* ATCC-28592, *Rhizopus oryzae* ATCC-10329 and *Mucor hiemalis* ATCC-8690) as well as three yeast specimens (*Candida utilis* ATCC-9226, *Candida albicans* ATCC-10231 and *Candida tropicalis* ATCC-13803).

CLSI standardized methods were adopted to carry out the microbiological tests. The M38-A microdilution method [31] was used to determine the sensitivity of filamentous fungi, and the M27-A3 method [32] for *Candida* yeasts.

The antifungal activity of compounds **4a–d** and **7a–c** was compared with itraconazole, a standard antifungal drug. The minimum inhibitory concentration (MIC) values of the compounds and standard drugs, expressed in micrograms per millilitre, were determined in 96-well plates by using RPMI 1640 medium buffered with MOPS (3-[N-morpholino]propane sulfonic acid; Sigma-Aldrich).

Table 2

Synthesis of 1,5-disubstituted 1,2,3-triazole **7a–c** (miconazole analogs) from azide **5** by coupling with β -ketophosphonates **6**.



Entry ^a	Ketone	Triazole ^b (Yield%) ^c
1	6a : $\text{R} = \text{cyclohexyl}$	7a (64%)
2	6b : $\text{R} = \text{CH}_3(\text{CH}_2)_3\text{C}(\text{CH}_3)_2-$	7b (70%)
3	6c : $\text{R} = p\text{-(CH}_3\text{S)phenyl}$	7c (72%)

^a Reaction conditions: A mixture of compound **5** (1.0 eq), **6** (1.0 eq), and KOH (3.0 eq), in CH_3CN was stirred at 60 °C for 5 h.

^b Confirmed by ^1H NMR, ^{13}C NMR, and MS.

^c Yields refer to chromatographically pure isolated compounds.

Table 3
In vitro antifungal activities of synthesized compounds (MIC, µg/mL).

Compound	Yeast fungi			Filamentous fungi			
	<i>C. uti.</i>	<i>C. alb.</i>	<i>C. trop.</i>	<i>A. fum.</i>	<i>T. cut.</i>	<i>R. ory.</i>	<i>M. hie</i>
4a	16	16	16	16	16	8	16
4b	16	0.03	0.06	16	16	8	16
4c	16	16	16	16	16	16	16
4d	0.25	0.06	0.06	2	0.12	0.5	0.25
7a	16	16	8	16	16	2	0.12
7b	2	0.03	0.03	0.5	8	8	16
7c	16	4	1	16	16	16	16
Standard ^a	0.06	0.03	0.03	0.25	1	0.5	1

Abbreviations: *C. uti.*, *Candida utilis*; *C. alb.*, *Candida albicans*; *C. trop.*, *Candida tropicalis*; *A. fum.*, *Aspergillus fumigatus*; *T. cut.*, *Trichosporon cutaneum*; *R. ory.*, *Rhizopus oryzae*; *M. hie.*, *Mucor hiemalis*.

^a Itraconazole.

4. Results and discussion

The antifungal activity of the evaluated compounds is summarized in Table 3. Compounds **4b**, **4d** and **7b** showed good activity against *C. albicans* and *C. tropicalis* (MIC 0.03–0.06 µg/mL) as compared to itraconazole (MIC 0.03 µg/mL). Such compounds proved to be ‘sensitive’¹ according to the sensitivity parameters of document M27-A3 (Table 4). On the other hand, the antifungal screening of compound **4d** showed that it was either better than or comparable to itraconazole against filamentous fungi *T. cutaneum*, *R. oryzae*, and *M. hiemalis* (MIC 0.12 versus 1.0, 0.5 versus 0.5, and 0.25 versus 1.0 µg/mL respectively). Compound **7b** demonstrated moderate growth inhibition of *A. fumigatus* (MIC 0.5 µg/mL) compared with the standard drug (MIC 0.25 µg/mL). In contrast with the results observed for an aryl substituent, these outcomes clearly indicate that an alkyl group in the 5-substituted triazole promotes the biological activity of this type of compound. Additionally, substituent probably allows for a better interaction with the 14- α -demethylase (P450_{14DM}, CYP51) enzyme [33], leading to its selective inhibition and therefore the growth inhibition of the fungal cell.

5. Conclusion

In summary, azide-enolate 1,3-dipolar cycloaddition allowed for the synthesis of seven miconazole analogs with 1,4,5-tri and 1,5-disubstituted triazole moieties. The pre-clinical studies showed that compounds **4b**, **4d** and **7b** have a good scope against *C. albicans* and *C. tropicalis*. A broader spectrum of **4d** against filamentous fungi (*T. cutaneum*, *R. oryzae*, and *M. hiemalis*) has been demonstrated. Due to their good/excellent activity, these miconazole analogs can be considered as drug candidates for future complementary biological studies.

6. Experimental section

6.1. General

Flash column chromatography: SiO₂ 60 (230–400 mesh). TLC: Silica-gel plates (SiO₂; 0.20-mm thickness); visualization with UV

Table 4
Determination of the sensitivity of yeast (according to document M27-A3): Susceptible (S), dose-dependent sensitive (SDD) and resistant (R).

Compound	<i>C. uti.</i>	<i>C. alb.</i>	<i>C. trop.</i>
4a	R	R	R
4b	R	S	S
4c	R	R	R
4d	SDD	S	S
7a	R	R	R
7b	R	S	S
7c	R	R	SDD
Standard ^a	S	S	S

^a Itraconazole. Interpretive criteria: Breakpoints (MIC, µg/mL) = 0.12 [S], 0.25–0.5 [SDD], 1 [R].

light at 254 nm *m.p.*: Fischer-Johns Scientific melting point apparatus; uncorrected. ¹H and ¹³C-NMR spectra: Bruker Avance 300 MHz and Varian 500 MHz; δ in ppm rel. to Me₄Si as internal standard, *J* in Hz. MS: Shimadzu GCMS-QP2010 Plus; in *m/z* (rel. %).

6.2. Experimental procedures

6.2.1. 1-(1-(1-(2,4-Dichlorophenyl)-2-(1H-imidazol-1-yl)ethyl)-5-methyl-1H-1,2,3-triazol-4-yl)ethanone **4a**

To a cold solution (0 °C) of benzylic alcohol **2** (0.35 g, 1.36 mmol) and diphenylphosphoryl azide (0.32 mL, 1.5 mmol) in anhydrous DMF (3.5 mL) was added DBU (0.4 mL, 2.72 mmol). The solution was stirred for 15 min at 0 °C under nitrogen atmosphere, and then brought to room temperature with continuous stirring for 3 h. At this time, TLC indicated the disappearance of the starting material. Acetylacetone **3a**, (0.14 mL, 1.36 mmol) was then added to the reaction mixture, which was stirred for 3 h at 60–70 °C. Brine (~40 mL) was added and then the reaction mixture was washed with EtOAc (3 × 10 mL). The organic layer was dried (Na₂SO₄) and the solvent evaporated under reduced pressure. The crude extract was purified by flash column chromatography, eluting with DCM/MeOH 95/5 to afford the thick yellow oil **4a** (0.37 g, 75%). *R*_f: 0.3 (DCM/MeOH 95/5). ¹H NMR: (500 MHz, CDCl₃) δ = 7.51 (d, *J* = 1.7 Hz, 1Hm-H), 7.33–7.26 (m, 3Ar-H), 7.01 (s, 1Hm-H), 6.86 (s, 1Hm-H), 6.03 (dd, *J* = 9.9, 4.0 Hz, 1H), 5.25 (dd, *J* = 14.7, 10.0 Hz, 1H), 4.62 (dd, *J* = 14.7, 3.9 Hz, 1H), 2.69 (s, 3H), 2.31 (s, 3H) ppm. ¹³C NMR: (125 MHz, CDCl₃) δ = 193.91 (C=O), 143.86 (C), 137.94 (C–Cl), 136.42 (C), 133.09 (C), 131.03 (CH), 130.91 (C–Cl), 130.20 (CH), 129.94 (CH), 129.18 (CH), 128.78 (CH), 128.72 (CH), 59.52 (CH), 49.25 (CH₂), 27.77 (CH₃), 8.56 (Ar–CH₃) ppm. MS-EI⁺ *m/z* (%): 364 [M⁺+1], 212 (100), 203 (45), 149 (63), 81 (20), 57 (19), 43 (83).

6.2.2. 1-(1-(2,4-Dichlorophenyl)-2-(1H-imidazol-1-yl)ethyl)-5-phenyl-1H-1,2,3-triazol-4-yl(phenyl)methanone **4b**

Following the synthetic procedure for **4a**, compound **2** (0.35 g, 1.36 mmol) and **3b** (0.305 g, 1.36 mmol) were coupled in the presence of diphenylphosphoryl azide (0.32 mL, 1.5 mmol) and DBU (0.4 mL, 2.72 mmol). The crude extract was purified by flash column chromatography, eluting with DCM/MeOH 95/5 to afford the thick yellow oil **4b** (0.445 g, 67%). *R*_f: 0.35 (DCM/MeOH 95/5). ¹H NMR: (500 MHz, CDCl₃) δ = 8.28–8.20 (m, 2Ar-H), 7.73 (d, *J* = 8.5 Hz, 1Hm-H), 7.64–7.56 (m, 1Ar-H), 7.53–7.32 (m, 8Ar-H), 7.01 (s, 1Hm-H), 6.87–6.80 (m, 2Ar-H), 6.71 (s, 1Hm-H), 5.98 (dd, *J* = 10.6, 3.7 Hz, 1H), 5.18 (dd, *J* = 14.6, 10.6 Hz, 1H), 4.48 (dd, *J* = 14.6, 3.8 Hz, 1H) ppm. ¹³C NMR: (125 MHz, CDCl₃) δ = 185.81 (C=O), 143.73 (C), 143.37 (C–Cl), 137.31 (C), 136.77 (C), 136.23 (C), 133.23 (C), 132.93 (C–Cl), 131.34 (CH), 130.57 (2CH), 130.40 (CH), 130.13 (C), 129.89 (CH), 129.59 (CH), 129.57 (CH), 129.39 (CH), 129.35 (2CH), 128.92 (2CH), 128.58 (CH), 128.28 (2CH), 60.02 (CH), 49.84 (CH₂) ppm. MS-EI⁺ *m/z* (%): 487 [M⁺], 105 [C₇H₅O⁺] (100), 84 (25), 77 [C₆H₅⁺] (61), 43 (47).

¹ ‘S’, ‘SDD’ and ‘R’ are represented by standardized values (breakpoints) used to appreciate the clinical value of the *in vitro* antifungal testing result and predicting the response of patients infected. Sensitivity is dependent on achieving the maximum dosages in plasma (breakpoints) to obtain optimal response. For itraconazole, an MIC within the susceptible-dose dependent (SDD) range indicates the need for plasma concentrations 0.25–0.5 µg/mL for an optimal response. Actual breakpoints are described in Table 4 (See Ref. [32b]).

6.2.3. 1-(1-(2,4-Dichlorophenyl)-2-(1H-imidazol-1-yl)ethyl)-5-phenyl-1H-1,2,3-triazole-4-carbonitrile **4c**

Following the synthetic procedure for **4a**, compound **2** (0.35 g, 1.36 mmol) and **3c** (0.197 g, 1.36 mmol) were coupled in the presence of diphenylphosphoryl azide (0.32 mL, 1.5 mmol) and DBU (0.4 mL, 2.72 mmol). The crude extract was purified by flash column chromatography, eluting with DCM/MeOH 95/5 to afford the thick yellow oil **4c** (0.35 g, 63%). *R*_f: 0.3 (DCM/MeOH 95/5). ¹H NMR: (500 MHz, CDCl₃) δ = 7.68 (d, *J* = 8.5 Hz, 1Ar-H), 7.59–7.55 (m, 1Ar-H), 7.52–7.47 (m, 3Ar-H), 7.39 (dd, *J* = 8.5, 2.1 Hz, 1Ar-H), 7.34 (s, 1Ar-H), 7.01 (s, 1Ar-H), 6.95–6.90 (m, 2Ar-H), 6.64 (s, 1Ar-H), 6.07 (dd, *J* = 10.5, 3.7 Hz, 1H), 5.13 (dd, *J* = 14.7, 10.5 Hz, 1H), 4.52 (dd, *J* = 14.7, 3.8 Hz, 1H) ppm. ¹³C NMR: (125 MHz, CDCl₃) δ = 145.35 (C), 136.76 (C–Cl), 132.92 (C), 131.73 (CH), 130.57 (C), 130.23 (C–Cl), 130.15 (CH), 129.79 (2CH), 129.59 (CH), 129.43 (CH), 128.83 (CH), 128.75 (2CH), 124.73 (CH), 121.97 (CH), 120.89 (C), 111.21 (C≡N), 60.97 (CH), 49.99 (CH₂) ppm. MS-EI⁺ *m/z* (%): 409 [M⁺], 373 (20), 299 (30), 229 (31), 203 (100), 149 (36), 81 (85).

6.2.4. 1-(1-(2,4-Dichlorophenyl)-2-(1H-imidazol-1-yl)ethyl)-5-pentyl-4-(phenylsulfonyl)-1H-1,2,3-triazole **4d**

Following the synthetic procedure for **4a**, compound **2** (0.35 g, 1.36 mmol) and **3d** (0.346 g, 1.36 mmol) were coupled in the presence of diphenylphosphoryl azide (0.32 mL, 1.5 mmol) and DBU (0.4 mL, 2.72 mmol). The crude extract was purified by flash column chromatography, eluting with DCM/MeOH 95/5 to afford the thick yellow oil **4d** (0.54 g, 78%). *R*_f: 0.4 (DCM/MeOH 9/1). ¹H NMR: (500 MHz, CDCl₃) δ = 8.07–8.01 (m, 2Ar-H), 7.68–7.63 (m, 1Im-H), 7.60–7.54 (m, 2Ar-H), 7.51 (dd, *J* = 1.6, 0.8 Hz, 1Ar-H), 7.39 (s, 1H), 7.31–7.29 (m, 2Ar-H), 6.92 (s, 1Im-H), 6.70 (s, 1Im-H), 6.03 (dd, *J* = 9.7, 4.2 Hz, 1H), 5.15 (dd, *J* = 14.7, 9.7 Hz, 1H), 4.56 (dd, *J* = 14.9, 4.3 Hz, 1H), 2.88–2.68 (m, 2H), 1.23–1.09 (m, 6H), 0.78 (t, *J* = 6.9 Hz, 3H) ppm. ¹³C NMR: (125 MHz, CDCl₃) δ = 145.12 (C–Cl), 141.44 (C), 140.43 (C), 137.25 (C), 136.62 (C), 134.00 (C), 132.82 (CH), 130.90 (C–Cl), 129.91 (CH), 129.35 (2CH), 129.21 (CH), 128.91 (CH), 128.79 (CH), 127.90 (2CH), 118.76 (CH), 60.31 (CH), 49.52 (CH₂), 31.29 (CH₂), 28.49 (CH₂), 22.50 (CH₂), 22.02 (CH₂), 13.70 (CH₃) ppm. MS-EI⁺ *m/z* (%): 518 [M⁺], 376 (13), 280 (19), 203 (64), 172 (36), 159 (37), 149 (49), 125 (58), 77 (100), 41 (49).

6.2.5. 1-(2-Azido-2-(2,4-dichlorophenyl)ethyl)-1H-imidazole **5**

To a cold solution (0 °C) of benzylic alcohol **2** (1.5 g, 5.83 mmol) and diphenylphosphoryl azide (1.25 mL, 5.83 mmol) in anhydrous DMF (13.0 mL), was added DBU (0.872 mL, 5.83 mmol). The solution was stirred for 15 min at 0 °C under nitrogen atmosphere, and then brought to room temperature with continuous stirring for 3 h. Brine (~100 mL) was added to the reaction mixture and washed with EtOAc (3 × 30 mL). The organic layer was dried (Na₂SO₄) and the solvent evaporated under reduced pressure. The crude extract was purified by flash column chromatography, eluting with DCM/MeOH 95/5 to afford the thick yellow oil **5** (1.43 g, 87%). *R*_f: 0.4 (DCM/MeOH 95/5). ¹H NMR: (300 MHz, CDCl₃) δ = 7.46 (d, *J* = 2.0 Hz, 1Im-H), 7.38–7.20 (m, 3Ar-H), 7.05 (s, 1Im-H), 6.91 (s, 1Im-H), 5.26 (dd, *J* = 7.6, 3.5 Hz, 1H), 4.22 (dd, *J* = 14.4, 3.5 Hz, 1H), 4.01 (dd, *J* = 14.4, 7.6 Hz, 1H) ppm. ¹³C NMR: (75 MHz, CDCl₃) δ = 137.62 (C–Cl), 135.46 (C), 133.03 (CH), 132.32 (C–Cl), 129.81 (CH), 129.68 (CH), 128.81 (CH), 128.14 (CH), 119.45 (CH), 62.65 (CH–N₃), 50.51 (CH₂) ppm.

6.2.6. 5-Cyclohexyl-1-(1-(2,4-dichlorophenyl)-2-(1H-imidazol-1-yl)ethyl)-1H-1,2,3-triazole **7a**

To a solution of benzyl azide **5** (0.35 g, 1.24 mmol) and β-keto-phosphonate **6a** (0.29 g, 1.24 mmol) in acetonitrile grade reagent (3.5 mL) was added potassium hydroxide (0.2 g, 3.73 mmol). The solution was stirred for 5 h at 60 °C. Brine (~40 mL) was added to

the reaction mixture and washed with EtOAc (3 × 30 mL). The organic layer was dried (Na₂SO₄) and the solvent evaporated under reduced pressure. The crude extract was purified by flash column chromatography, eluting with DCM/MeOH 95/5 to afford the thick yellow oil **7a** (0.31 g, 64%). *R*_f: 0.3 (DCM/MeOH 95/5). ¹H NMR: (300 MHz, CDCl₃) δ = 7.50 (d, *J* = 2.0 Hz, 1Im-H), 7.45 (s, 1Ar-H), 7.42 (s, 1Ar-H), 7.31–7.29 (m, 2Ar-H), 6.96 (s, 1Im-H), 6.76 (s, 1Im-H), 6.03 (dd, *J* = 9.9, 3.9 Hz, 1H), 5.22 (dd, *J* = 14.5, 9.9 Hz, 1H), 4.55 (dd, *J* = 14.5, 3.9 Hz, 1H), 2.64–2.50 (m, 1H), 1.90–1.47 (m, 5H), 1.43–1.01 (m, 5H) ppm. ¹³C NMR: (75 MHz, CDCl₃) δ = 143.80 (C–Cl), 135.94 (C), 132.75 (C), 132.19 (CH), 131.23 (C–Cl), 129.80 (CH), 129.67 (CH), 129.58 (CH), 128.64 (CH), 122.56 (CH), 118.99 (CH), 59.41 (CH), 51.56 (CH₂), 33.22 (CH), 32.37 (2CH₂), 25.71 (CH), 25.44 (2CH₂) ppm.

6.2.7. 1-(1-(2,4-Dichlorophenyl)-2-(1H-imidazol-1-yl)ethyl)-5-(2-methylhexan-2-yl)-1H-1,2,3-triazole **7b**

Following the synthetic procedure for **7a**, compound **5** (0.35 g, 1.24 mmol) and **6b** (0.311 g, 1.24 mmol) were coupled in the presence of KOH (0.2 g, 3.73 mmol). The crude extract was purified by flash column chromatography, eluting with DCM/MeOH 95/5 to afford the thick yellow oil **7b** (0.35 g, 70%). *R*_f: 0.3 (DCM/MeOH 9/1). ¹H NMR: (500 MHz, CDCl₃) δ = 7.54–7.53 (m, 1Im-H), 7.44 (s, 1Ar-H), 7.28–7.27 (m, 1Ar-H), 7.27–7.25 (m, 2Ar-H), 6.96 (s, 1Im-H), 6.78 (s, 1Im-H), 6.23 (dd, *J* = 10.6, 3.0 Hz, 1H), 5.29 (dd, *J* = 14.6, 10.6 Hz, 1H), 4.50 (dd, *J* = 14.6, 3.0 Hz, 1H), 1.37–1.21 (m, 2H), 1.10 (s, 3H), 1.03 (s, 3H), 0.97–0.82 (m, 2H), 0.75–0.64 (m, 1H), 0.61–0.54 (m, 3H), 0.43–0.31 (m, 1H) ppm. ¹³C NMR: (75 MHz, CDCl₃) δ = 145.93 (C–Cl), 137.33 (C), 135.81 (CH), 133.29 (CH), 132.53 (C–Cl), 132.26 (CH), 130.90 (C), 129.73 (CH), 129.50 (CH), 128.45 (CH), 118.89 (CH), 62.20 (CH), 49.80 (CH₂), 41.07 (CH₂), 33.47 (C), 27.77 (CH₃), 27.67 (CH₃), 26.73 (CH₂), 22.76 (CH₂), 13.65 (CH₃) ppm. MS-EI⁺ *m/z* (%): 406 [M⁺], 296 (11), 240 (28), 203 (53), 172 (25), 159 (33), 124 (34), 81 (45), 57 (100), 41 (71).

6.2.8. 1-(1-(2,4-Dichlorophenyl)-2-(1H-imidazol-1-yl)ethyl)-5-(4-(methylthio)phenyl)-1H-1,2,3-triazole **7c**

Following the synthetic procedure for **7a**, compound **5** (0.35 g, 1.24 mmol) and **6c** (0.341 g, 1.24 mmol) were coupled in the presence of KOH (0.2 g, 3.73 mmol). The crude extract was purified by flash column chromatography, eluting with DCM/MeOH 95/5 to afford the thick yellow oil **7c** (0.385 g, 72%). *R*_f: 0.3 (DCM/MeOH 9/1). ¹H NMR: (300 MHz, CDCl₃) δ = 7.69 (m, 1Im-H), 7.68 (s, 1Ar-H), 7.47 (d, *J* = 2.1 Hz, 2Ar-H), 7.36–7.19 (m, 4Ar-H), 6.96 (s, 1Im-H), 6.75 (d, *J* = 8.4 Hz, 2Ar-H), 6.64 (s, 1Im-H), 6.01 (dd, *J* = 10.5, 3.5 Hz, 1H), 5.15 (dd, *J* = 14.5, 10.5 Hz, 1H), 4.48 (dd, *J* = 14.5, 3.6 Hz, 1H), 2.49 (s, 3H) ppm. ¹³C NMR: (75 MHz, CDCl₃) δ = 141.81 (C), 139.43 (C–Cl), 136.05 (C–S), 133.21 (C), 132.72 (CH), 131.83 (CH), 130.91 (C–Cl), 129.83 (CH), 129.77 (C), 129.40 (CH), 128.94 (2CH), 128.79 (CH), 128.55 (CH), 126.14 (2CH), 118.83 (CH), 59.98 (CH), 50.20 (CH₂), 15.05 (CH₃) ppm.

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Appendix A. Supplementary data

Supplementary data related to this article can be found at <http://dx.doi.org/10.1016/j.ejmech.2016.02.013>

References

- [1] M. Grudzień, A. Król, G. Paterek, K. Stępień, F. Pluciński, A.P. Mazurek, The structure–bioavailability approach in antifungal agents, *Eur. J. M. Chem.* 44 (2009) 1978–1981.
- [2] (a) Z.A. Kanafani, J.R. Perfect, Resistance to antifungal agents: mechanisms and clinical impact, *Clin. Infect. Dis.* 46 (2008) 120–128; (b) Michael A. Pfaffler, Antifungal drug resistance: mechanisms, epidemiology, and consequences for treatment, *Am. J. Med.* 125 (2012) S3–S13; (c) Giulia Morace, Federica Perdoni, Elisa Borghi, Antifungal drug resistance in *Candida* species, *Am. J. Med.* 2 (2014) 254–259; (d) G. Morace, F. Perdoni, E. Borghi, Antifungal drug resistance in *Candida* species, *J. Glob. Antim. Res.* 2 (2014) 254–259; (e) J.B. Anderson, Evolution of antifungal-drug resistance: mechanisms and pathogen fitness, *Nat. Rev. Microbiol.* 3 (2005) 547–556.
- [3] For complete Reviews about this issue see: (a) M.K. Kathiravan, A.B. Salake, A.S. Chothe, P.B. Dudhe, R.P. Watode, M.S. Mukta, S. Gadhwe, The biology and chemistry of antifungal agents: a review, *Bioorg. Med. Chem.* 20 (2012) 5678–5698; (b) A.B. Petersen, M.H. Rønne, T.O. Larsen, M.H. Clausen, The chemistry of griseofulvin, *Chem. Rev.* 114 (2014) 12088–12107; (c) J. Heeres, L. Meerpoel, P. Lewi, Conazoles, *Molecules* 15 (2010) 4129–4188.
- [4] W.R.J.D. Galloway, A. Bender, M. Welch, D.R. Spring, The discovery of antibacterial agents using diversity-oriented synthesis, *Chem. Commun.* (2009) 2446–2462.
- [5] (a) A. Massarotti, S. Aprile, V. Mercalli, E. Del Grosso, G. Grosa, G. Sorba, G.C. Tron, Are 1,4- and 1,5-disubstituted 1,2,3-triazoles good pharmacophoric groups? *ChemMedChem* 9 (2014) 2497–2508; (b) S.G. Agalave, S.R. Maujan, V.S. Pore, Click Chemistry: 1,2,3-Triazoles as Pharmacophores, 6, 2011, pp. 2696–2718; (c) C.H. Zhou, Y. Wang, Recent researches in triazole compounds as medicinal drugs, *Curr. Med. Chem.* 19 (2012) 239–280.
- [6] (a) E. Vitaku, D.T. Smith, J.T. Njardarson, Analysis of the structural diversity, substitution patterns, and frequency of nitrogen heterocycles among U.S. FDA approved pharmaceuticals, *J. Med. Chem.* 57 (2014) 10257–10274; (b) H.X. Ding, C.A. Leverett, R.E. Kyne, K.K.C. Liu, S.J. Fink, A.C. Flick, C.J. O'Donnell, Synthetic approaches to the 2013 new drugs, *Bioorg. Med. Chem.* 23 (2015) 1895–1922.
- [7] For concise Reviews about this issue see: (a) F. de Carvalho da Silva, M.F. do Carmo Cardoso, P. Garcia-Ferreira, V.F. Ferreira, Biological properties of 1H-1,2,3- and 2H-1,2,3-triazoles, in: W. Dehaen, V.A. Bakulev (Eds.), *Chemistry of 1,2,3-triazoles*, Springer International Publishing, Switzerland, 2015, pp. 117–165, http://dx.doi.org/10.1007/7081_2014_124; (b) A. Lauria, R. Delisi, F. Mingoa, A. Terenzi, A. Martorana, G. Barone, A.M. Almerico, 1,2,3-Triazole in heterocyclic compounds, endowed with biological activity, through 1,3-dipolar cycloadditions, *Eur. J. Org. Chem.* (2014) 3289–3306; (c) P. Thirumurugan, D. Matosiuk, K. Jozwiak, Click chemistry for drug development and diverse chemical–biology applications, *Chem. Rev.* 113 (2013) 4905–4979; (d) J.F. Lutz, Z. Zarafshani, Efficient construction of therapeutics, bioconjugates, biomaterials and bioactive surfaces using azide–alkyne “click” chemistry, *Adv. Drug Deliv. Rev.* 60 (2008) 958–970; (e) F. Amblard, J.H. Cho, R.F. Schinazi, Cu(I)-Catalyzed Huisgen azide–alkyne 1,3-dipolar cycloaddition reaction in nucleoside, nucleotide, and oligonucleotide Chemistry, *Chem. Rev.* 109 (2009) 4207–4220.
- [8] (a) L.Y. Ma, L.P. Pang, B. Wang, M. Zhang, B. Hu, D.Q. Xue, K.P. Shao, B.L. Zhang, Y. Liu, E. Zhang, H.M. Liu, Design and synthesis of novel 1,2,3-triazole-pyrimidine hybrids as potential anticancer agents, *Eur. J. Med. Chem.* 86 (2014) 368–380; (b) L.Y. Ma, B. Wang, L.P. Pang, M. Zhang, S.Q. Wang, L.C. Zheng, K.P. Shao, D.Q. Xue, H.M. Liu, Design and synthesis of novel 1,2,3-triazole–pyrimidine–urea hybrids as potential anticancer agents, *Bioorg. Med. Chem. Lett.* 25 (2015) 1124–1128; (c) B.E. Gryder, M.J. Akbashev, M.K. Rood, E.D. Raftery, W.M. Meyers, P. Dillard, S. Khan, A.K. Oyeler, Selectively targeting prostate cancer with antiandrogen equipped histone deacetylase inhibitors, *ACS Chem. Biol.* 8 (2013) 2550–2560.
- [9] (a) K. Kumar, B. Pradines, M. Madamet, R. Amalvict, N. Benoit, V. Kumar, 1H-1,2,3-triazole tethered isatin-ferrocene conjugates: synthesis and in vitro antimalarial evaluation, *Eur. J. Med. Chem.* 87 (2014) 801–804; (b) R. Raj, J. Gut, P.J. Rosenthal, V. Kumar, 1H-1,2,3-Triazole-tethered isatin-7-chloroquinoline and 3-hydroxy-indole-7-chloroquinoline conjugates: Synthesis and antimalarial evaluation, *Bioorg. Med. Chem. Lett.* 24 (2014) 756–759.
- [10] (a) T. Yempala, J.P. Sridevi, P. Yogeeswari, D. Sriram, S. Kantevari, Rational design and synthesis of novel dibenzol[b,d]furan-1,2,3-triazole conjugates as potent inhibitors of *Mycobacterium tuberculosis*, *Eur. J. Med. Chem.* 71 (2014) 160–167; (b) M.H. Shaikh, D.D. Subhedar, L. Nawale, D. Sarkar, F.A. Kalam Khan, J.N. Sangshetti, B.B. Shingate, 1,2,3-Triazole derivatives as antitubercular agents: synthesis, biological evaluation and molecular docking study, *Med. Chem. Commun.* 6 (2015) 1104–1116.
- [11] P. de Andrade, O.A. Galo, M.R. Carvalho, C.D. Lopes, Z.A. Carneiro, R. Sesti-Costa, E.B. de Melo, J.S. Silva, I. Carvalho, 1,2,3-Triazole-based analogue of benzimidazole displays remarkable activity against *Trypanosoma cruzi*, *Bioorg. Med. Chem.* 23 (2015) 6815–6826.
- [12] T.T. Guimarães, M.C.F.R. Pinto, J.S. Lanza, M.N. Melo, R.L. do Monte-Neto, I.M.M. de Melo, E.B.T. Diogo, V.F. Ferreira, C.A. Camara, W.O. Valença, R.N. de Oliveira, F. Frézard, E.N. da Silva Jr., Potent naphthoquinones against antimony-sensitive and -resistant *Leishmania* parasites: Synthesis of novel α - and α -nor- α -lapachone-based 1,2,3-triazoles by copper-catalyzed azide–alkyne cycloaddition, *Eur. J. Med. Chem.* 63 (2013) 523–530.
- [13] (a) S.K.V. Vernekar, L. Qiu, J. Zacharias, R.J. Geraghty, Z. Wang, Synthesis and antiviral evaluation of 4'-(1,2,3-triazol-1-yl)thymidines, *Med. Chem. Commun.* 5 (2014) 603–608; (b) R. Aneja, A.A. Rashad, H. Li, R.V.K. Sundaram, C. Duffy, L.D. Bailey, I. Chaiken, Peptide triazole inactivators of HIV-1 utilize a conserved two-cavity binding site at the junction of the inner and outer domains of Env gp120, *J. Med. Chem.* 58 (2015) 3843–3858.
- [14] B.H. Fraser, S. Hamilton, A.M. Krause-Heuer, P.J. Wright, I. Greguric, S.P. Tucker, A.G. Draffan, V.V. Fokin, K.B. Sharpless, Synthesis of 1,4-triazole linked zanamivir dimers as highly potent inhibitors of influenza A and B, *Med. Chem. Commun.* 4 (2013) 383–386.
- [15] S.K.V. Vernekar, L. Qiu, J. Zhang, J. Kankanala, H. Li, R.J. Geraghty, Z. Wang, 5'-Silylated 3'-1,2,3-triazolyl thymidine analogues as inhibitors of West Nile virus and dengue virus, *J. Med. Chem.* 58 (2015) 4016–4028.
- [16] J.L. Diaz, U. Christmann, A. Fernández, A. Torrens, A. Port, R. Pascual, I. Álvarez, J. Burguño, X. Monroy, A. Montero, A. Balada, J.M. Vela, C. Almansa, Synthesis and structure–activity relationship study of a new series of selective σ 1 receptor ligands for the treatment of pain: 4-aminotriazoles, *J. Med. Chem.* 58 (2015) 2441–2451.
- [17] (a) S.K. Kessler, A. McCarthy, A. Cnaan, D.J. Dlugos, Retention rates of rufinamide in pediatric epilepsy patients with and without Lennox–Gastaut Syndrome, *Epilepsy Res.* 112 (2015) 18–26; (b) J. Gilchrist, S. Dutton, M. Diaz-Bustamante, A. McPherson, N. Olivares, J. Kalia, A. Escayg, F. Bosmans, Nav1.1 Modulation by a novel triazole compound attenuates Epileptic seizures in rodents, *ACS Chem. Biol.* 9 (2014) 1204–1212.
- [18] H.H. Kinfe, Y.H. Belay, J.S. Joseph, E. Mukwehvo, Evaluation of the influence of thiosemicarbazone–triazole hybrids on genes implicated in lipid oxidation and accumulation as potential anti-obesity agents, *Bioorg. Med. Chem. Lett.* 23 (2013) 5275–5278.
- [19] S. Shafi, M.M. Alam, N. Mulakayala, C. Mulakayala, G. Vanaja, A.M. Kalle, R. Pallu, M.S. Alam, Synthesis of novel 2-mercapto benzothiazole and 1,2,3-triazole based bis-heterocycles: their anti-inflammatory and antinociceptive activities, *Eur. J. Med. Chem.* 49 (2012) 324–333.
- [20] (a) M.R. El Sayed Aly, H.A. Saad, M.A.M. Mohamed, Click reaction based synthesis, antimicrobial, and cytotoxic activities of new 1,2,3-triazoles, *Bioorg. Med. Chem. Lett.* 25 (2015) 2824–2830; (b) H. Kuhn, D. Gutelius, E. Black, C. Nadolny, A. Basu, C. Reid, Anti-bacterial glycosyl triazoles – identification of an N-acetylglucosamine derivative with bacteriostatic activity against *Bacillus*, *Med. Chem. Commun.* 5 (2014) 1213–1217.
- [21] (a) S.Z. Ferreira, H.C. Carneiro, H.A. Lara, R.B. Alves, J.M. Resende, H.M. Oliveira, L.M. Silva, D.A. Santos, R.P. Freitas, Synthesis of a new peptide–coumarin conjugate: A potential agent against cryptococcosis, *ACS Med. Chem. Lett.* 6 (2015) 271–275; (b) M. Irfan, B. Aneja, U. Yadava, S.I. Khan, N. Manzoor, C.G. Daniluc, M. Abid, Synthesis, QSAR and anticandidal evaluation of 1,2,3-triazoles derived from naturally bioactive scaffolds, *Eur. J. Med. Chem.* 93 (2015) 246–254; (c) Z. Jiang, J. Gu, C. Wang, S. Wang, N. Liu, Y. Jiang, G. Dong, Y. Wang, Y. Liu, J. Yao, Z. Miao, W. Zhang, C. Sheng, Design, synthesis and antifungal activity of novel triazole derivatives containing substituted 1,2,3-triazole-piperidine side chains, *Eur. J. Med. Chem.* 82 (2014) 490–497.
- [22] (a) K. Pericherla, P. Khedar, B. Khungar, A. Kumar, Click chemistry inspired structural modification of azole antifungal agents to synthesize novel ‘drug like’ molecules, *Tetrahedron Lett.* 53 (2012) 6761–6764; (b) S. Kim, S.N. Cho, T. Oh, P. Kim, Design and synthesis of 1H-1,2,3-triazoles derived from econazole as antitubercular agents, *Bioorg. Med. Chem. Lett.* 22 (2012) 6844–6847.
- [23] For complete Reviews about this issue see: (a) S. Hassan, T.J.J. Müller, Multi-component syntheses based upon copper-catalyzed alkyne–azide cycloaddition, *Adv. Synth. Catal.* 357 (2015) 617–666; (b) J. Totobenazara, A.J. Burke, New click-chemistry methods for 1,2,3-triazoles synthesis: recent advances and applications, *Tetrahedron Lett.* 56 (2015) 2853–2859; (c) B. Dervaux, F.E. Du Prez, Heterogeneous azide–alkyne click chemistry: towards metal-free end products, *Chem. Sci.* 3 (2012) 959–966; (d) M. Meldal, C.W. Tornøe, Cu-catalyzed azide–alkyne cycloaddition, *Chem. Rev.* 108 (2008) 2952–3015; (e) V.D. Bock, H. Hiemstra, J.H. van Maarseveen, Cu^I-catalyzed alkyne–azide “Click” cycloadditions from a mechanistic and synthetic perspective, *Eur. J. Org. Chem.* (2006) 51–68.

- [24] For complete Reviews about this issue see: (a) C.G.S. Lima, A. Ali, S.S. van Berkel, B. Westermann, M.W. Paixão, Emerging approaches for the synthesis of triazoles: beyond metal-catalyzed and strain-promoted azide–alkyne cycloaddition, *Chem. Commun.* 51 (2015) 10784–10796; (b) J. John, J. Thomas, W. Dehaen, Organocatalytic routes toward substituted 1,2,3-triazoles, *Chem. Commun.* 51 (2015) 10797–10806; (c) S.S.V. Ramasastry, Enamine/Enolate-mediated organocatalytic azide–carbonyl [3+2] cycloaddition reactions for the synthesis of densely functionalized 1,2,3-triazoles, *Angew. Chem. Int. Ed.* 53 (2014) 14310–14312.
- [25] (a) D.V. Demchuk, A.V. Samet, N.B. Chernysheva, V.I. Ushkarov, G.A. Stashina, L.D. Konyushkin, M.M. Raihstat, S.I. Firgang, A.A. Philchenkov, M.P. Zavelevich, L.M. Kuiava, V.F. Chekhun, D.Y. Blokhin, A.S. Kiselyov, M.N. Semenova, V.V. Semenov, Synthesis and antiproliferative activity of conformationally restricted 1,2,3-triazole analogues of combretastatins in the sea urchin embryo model and against human cancer cell lines, *Bioorg. Med. Chem.* 22 (2014) 738–755; (b) J.B. He, L.L. Feng, J. Li, R.J. Tao, Y.L. Ren, J. Wan, H.W. He, Design, synthesis and molecular modeling of novel N-acylhydrazone derivatives as pyruvate dehydrogenase complex E1 inhibitors, *Bioorg. Med. Chem.* 22 (2014) 89–94.
- [26] A. Ramírez-Villalva, D. González-Calderón, C. González-Romero, M. Morales-Rodríguez, B. Jauregui-Rodríguez, E. Cuevas-Yañez, A. Fuentes-Benites, A facile synthesis of novel miconazole analogues and the evaluation of their antifungal activity, *Eur. J. Med. Chem.* 97 (2015) 275–279.
- [27] (a) G. La Regina, F.D. D'Auria, A. Tafi, F. Piscitelli, S. Olla, F. Caporuscio, L. Nencioni, R. Cirilli, F. La Torre, N. Rodrigues De Melo, S.L. Kelly, D.C. Lamb, M. Artico, M. Botta, A.T. Palamara, R. Silvestri, 1-[(3-Aryloxy-3-aryl)propyl]-1H-imidazoles, new imidazoles with potent activity against *Candida albicans* and dermatophytes. Synthesis, structure–activity relationship, and molecular modeling studies, *J. Med. Chem.* 51 (2008) 3841–3855; (b) A. Davood, M. Iman, Molecular docking and QSAR study on imidazole derivatives as 14 α -demethylase inhibitors, *Turk. J. Chem.* 37 (2013) 119–133.
- [28] D. González-Calderón, I. Santillán-Iniesta, C.A. González-González, A. Fuentes-Benites, C. González-Romero, A novel and facile synthesis of 1,4,5-trisubstituted 1,2,3-triazoles from benzylic alcohols through a one-pot, three-component system, *Tetrahedron Lett.* 56 (2015) 514–516.
- [29] D. González-Calderón, A. Fuentes-Benites, E. Díaz-Torres, C.A. González-González, C. González-Romero, An azide–enolate 1,3-dipolar cycloaddition as an efficient approach for the synthesis of 1,5-disubstituted 1,2,3-triazoles from alkyl/aryl azides and β -ketophosphonates, *Eur. J. Org. Chem.* (2016) 668–672.
- [30] A.S. Thompson, G.R. Humphrey, A.M. DeMarco, D.J. Mathre, E.J.J. Grabowski, Direct conversion of activated alcohols to azides using diphenyl phosphorazidate. A practical alternative to Mitsunobu conditions, *J. Org. Chem.* 58 (1993) 5886–5888.
- [31] (a) National Committee for Clinical Laboratory Standards Institute (CLSI), Document M38–A2: Reference Method for Broth Dilution Antifungal Susceptibility Testing of Filamentous Fungi, Approved Standard–Second Edition, Clinical and Laboratory Standards Institute, Wayne, PA, 2002; (b) A.L. Barry, An overview of the Clinical and Laboratory Standards Institute (CLSI) and its impact on antimicrobial susceptibility tests, in: R. Schwalbe, L. Steele-Moore, A.C. Goodwin (Eds.), *Antimicrobial Susceptibility Testing Protocols*, CRC Press Taylor & Francis Group, Florida, 2007, pp. 1–6. ISBN: 9780824741006; (c) A. Espinel-Ingroff, E. Cantón, Antifungal susceptibility testing of filamentous fungi, in: R. Schwalbe, L. Steele-Moore, A.C. Goodwin (Eds.), *Antimicrobial Susceptibility Testing Protocols*, CRC Press Taylor & Francis Group, Florida, 2007, pp. 209–241. ISBN: 9780824741006.
- [32] (a) National Committee for Clinical and Laboratory Standards Institute (CLSI), M27–A3: Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts, Approved Standard–Third Edition, Clinical and Laboratory Standards Institute, Wayne, PA, 2008. ISBN: 1-56238-666-2; (b) A. Espinel-Ingroff, E. Cantón, Antifungal susceptibility testing of yeasts, in: R. Schwalbe, L. Steele-Moore, A.C. Goodwin (Eds.), *Antimicrobial Susceptibility Testing Protocols*, CRC Press Taylor & Francis Group, Florida, 2007, pp. 173–208. ISBN: 9780824741006; (c) A.W. Fothergill, Antifungal Susceptibility Testing: Clinical Laboratory and Standards Institute (CLSI) methods, in: G.S. Hall (Ed.), *Interactions of Yeasts, Moulds, and Antifungal Agents. How to Detect Resistance*, Springer Science-Business Media, 2012, pp. 65–74, http://dx.doi.org/10.1007/978-1-59745-134-5_2.
- [33] (a) C. Sheng, W. Zhang, H. Ji, M. Zhang, Y. Song, H. Xu, J. Zhu, Z. Miao, Q. Jiang, J. Yao, Y. Zhou, J. Zhu, J. Lü, Structure-based optimization of azole antifungal agents by CoMFA, CoMSIA, and molecular docking, *J. Med. Chem.* 49 (2006) 2512–2525; (b) H. Ji, W. Zhang, Y. Zhou, M. Zhang, J. Zhu, Y. Song, J. Lü, J. Zhu, A three-dimensional model of lanosterol 14 α -demethylase of *Candida albicans* and its interaction with azole antifungals, *J. Med. Chem.* 43 (2000) 2493–2505.